

A supplementary document of

**Estimation of the Mechanism of Adrenal Action of
Endocrine-disrupting Compounds Using a Computational Model of
Adrenal Steroidogenesis in NCI-H295R Cells.**

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1. Abbreviations

Steroid hormones

PREG: pregnenolone

HPREG: 17 α -hydroxypregnenolone

DHEA: dehydroepiandrosterone

PROG: progesterone

HPROG: 17 α -hydroxyprogesterone

DIONE: androstenedione

DCORTICO: 11-deoxycorticosterone

DCORT: 11-deoxycortisol

CORTICO: corticosterone

CORT: cortisol

ALDO: aldosterone

TESTO: testosterone

E1: estrone

E2: 17 β -estradiol

Proteins of adrenal steroidogenic pathway

CYP11A1: cholesterol side chain cleavage enzyme

CYP17H: 17 α -hydroxylase of CYP17

CYP17L: C₁₇₋₂₀ lyase of CYP17

HSD3B2: 3 β -hydroxysteroid dehydrogenase

CYP21A2: 21-hydroxylase

CYP11B1: 11 β -hydroxylase

CYP11B2: 18-hydroxylase

HSD17B3: 17 β -hydroxysteroid dehydrogenase.

CYP19A1: aromatase

2. A computational model of adrenal steroidogenesis in NCI-H295R cells

Cell proliferation

Equations to estimate the proliferation of NCI-H295R cells have been proposed by previous studies [1, 2]. In this model, the dynamics of the total number of cells (N_{cell}) and total cell volume (V_{cell}) were implemented by the following equations:

$$N_{\text{cell}}(t) = N_{\text{cell}}(-72) \cdot \exp[k_p \cdot (t + t_{\text{incubation}})] \quad \dots \text{(Eq. 1)}$$

$$V_{\text{cell}}(t) = V_{\text{indiv}} \cdot N_{\text{cell}} \quad \dots \text{(Eq. 2)}$$

where, $N_{\text{cell}}(-72)$ is the initial number of NCI-H295R cells per well before incubation (6×10^5 cells), k_p is the cell proliferation rate of NCI-H295R cells (0.00878 1/h), $t_{\text{incubation}}$ is the incubation time until stimulation (72 h), t is the culture time after stimulation, and V_{indiv} is the mean cell volume of individual NCI-H295R cells ($1,499 \mu\text{m}^3$).

Cholesterol transport and intracellular distribution

Cholesterol transport and the intracellular localization pathway were modified as a part of the ACTH-stimulated cortisol secretion model described by Dempsher and colleagues [3]. Intracellular cholesterol was separated into five states based on localization in the adrenal cells. There are stored cholesterol esters in the lipid droplets (CHOS), free cytosolic cholesterol (CHOC), free mitochondrial cholesterol (CHOM), free mitochondrial cholesterol close to CYP11A1 enzymes (CHON), and free mitochondrial cholesterol remote from CYP11A1 enzymes (CHOR). Cholesterols (CHOL) almost always exist as a cholesterol ester (CE) in extracellular culture medium. Therefore, imported CHOL from the culture medium is first stored in lipid droplets (transition to CHOS from medium CHOL). CHOS is transformed to free CHOL by cholesterol ester hydrolase (CEH) and distributed to the cytosolic space (transition from CHOS to CHOC). CHOC is transported into mitochondria from the cytosol by StAR protein (transition from CHOC to CHOM). CHOM is continuously translocated in the vicinity of CYP11A1 enzymes by StAR (transition from CHOM to CHON), so that CHON is available for the adrenal steroidogenesis pathway. On the other hand, CHOM also passively recedes from CYP11A1 enzymes (transition from CHOM to CHOR). Moreover, the oxysterol biosynthesis pathway was incorporated as a bypass of the steroidogenesis pathway, which was proposed by Breen et al [2]. In fact, the total mass balance of cholesterol and all steroids is not preserved in

this *in vitro* system. However, the oxysterol biosynthesis pathway and/or bypass pathway are expected to exist in NCI-H295R cells. In this model, the oxysterol biosynthesis pathway was defined to branch from CHOC to oxysterol (OXY). These variable parameters belonging to cholesterol transport and the intracellular transport system were described by the following ordinary differential equations (ODEs):

$$\frac{d[\text{CHOL}]_{\text{med}}}{dt} = -\frac{V_{\text{cell}}}{V_{\text{med}}} \cdot v_{\text{CholesterolTransport}} \quad \dots \text{ (ODE 1)}$$

$$\frac{d\text{CHOS}}{dt} = V_{\text{cell}} \cdot (v_{\text{CholesterolTransport}} - v_{\text{CEH}}) \quad \dots \text{ (ODE 2)}$$

$$\frac{d\text{CHOC}}{dt} = V_{\text{cell}} \cdot (v_{\text{CEH}} - v_{\text{MTR}} - v_{\text{OxysterolSynthesis}}) \quad \dots \text{ (ODE 3)}$$

$$\frac{d\text{CHOM}}{dt} = V_{\text{cell}} \cdot (v_{\text{MTR}} - v_{\text{acc}} - v_{\text{loc}}) \quad \dots \text{ (ODE 4)}$$

$$\frac{d\text{CHON}}{dt} = V_{\text{cell}} \cdot (v_{\text{loc}} - v_{\text{CYP11A1}}) \quad \dots \text{ (ODE 5)}$$

$$\frac{d\text{CHOR}}{dt} = V_{\text{cell}} \cdot v_{\text{acc}} \quad \dots \text{ (ODE 6)}$$

$$\frac{d\text{OXY}}{dt} = V_{\text{cell}} \cdot v_{\text{OxysterolSynthesis}} \quad \dots \text{ (ODE 7)}$$

where, V_{med} is the volume of culture medium (2 mL), $v_{\text{CholesterolTransport}}$ is the cholesterol import rate into the cytosolic space from the extracellular culture medium, v_{CEH} is the enzymatic reaction rate of CEH, v_{MTR} is the mitochondrial transport rate by the StAR protein, $v_{\text{OxysterolSynthesis}}$ is the reaction rate of oxysterol biosynthesis, v_{acc} is the passive diffusion rate from CYP11A1 enzymes, v_{loc} is the translocation rate close to CYP11A1 enzymes by StAR, and v_{CYP11A1} is the enzymatic reaction rate of the first adrenal steroidogenic enzyme, CYP11A1.

Net cholesterol concentration in intracellular space was calculated by a following equation:

$$[\text{CHOL}]_{\text{cell}} = \frac{(\text{CHOS} + \text{CHOC} + \text{CHOM} + \text{CHON} + \text{CHOR})}{V_{\text{cell}}} \quad \dots \text{ (Eq. 3)}$$

C₂₁-steroid hormone biosynthesis pathway

The C₂₁-steroid hormone biosynthesis pathway includes 14 steroid hormones, PREG, HPREG, DHEA, PROG, HPROG, DIONE, TESTO, DCORTICO, DCORT, CORTICO, CORT, ALDO, E1 and E2, and 17 enzymatic reactions catalyzed by nine steroidogenic enzymes, CYP11A1, CYP17H, CYP17L, HSD3B2, CYP21A2, CYP11B1, CYP11B2, HSD17B3, and CYP19A1.

These variable parameters belonging to the adrenal steroidogenesis system were described by the following ODEs:

$$V_{\text{cell}} \cdot \frac{d[\text{PREG}]_{\text{cell}}}{dt} = \frac{V_{\text{cell}}^2}{V_{\text{cell}} + V_{\text{med}} \cdot q_{\text{PREG}}} \cdot (v_{\text{CYP11A1}} - v_{\text{A,CYP17H}} - v_{\text{A,HSD3B2}}) \quad \dots \text{ (ODE 8)}$$

$$V_{\text{cell}} \cdot \frac{d[\text{HPREG}]_{\text{cell}}}{dt} = \frac{V_{\text{cell}}^2}{V_{\text{cell}} + V_{\text{med}} \cdot q_{\text{HPREG}}} \cdot (v_{\text{A,CYP17H}} - v_{\text{A,CYP17L}} - v_{\text{B,HSD3B2}}) \quad \dots \text{ (ODE 9)}$$

$$V_{\text{cell}} \cdot \frac{d[\text{DHEA}]_{\text{cell}}}{dt} = \frac{V_{\text{cell}}^2}{V_{\text{cell}} + V_{\text{med}} \cdot q_{\text{DHEA}}} \cdot (v_{\text{A,CYP17L}} - v_{\text{C,HSD3B2}}) \quad \dots \text{ (ODE 10)}$$

$$V_{\text{cell}} \cdot \frac{d[\text{PROG}]_{\text{cell}}}{dt} = \frac{V_{\text{cell}}^2}{V_{\text{cell}} + V_{\text{med}} \cdot q_{\text{PROG}}} \cdot (v_{\text{A,HSD3B2}} - v_{\text{B,CYP17H}} - v_{\text{A,CYP21A2}}) \quad \dots \text{ (ODE 11)}$$

$$V_{\text{cell}} \cdot \frac{d[\text{HPROG}]_{\text{cell}}}{dt} = \frac{V_{\text{cell}}^2}{V_{\text{cell}} + V_{\text{med}} \cdot q_{\text{HPROG}}} \cdot (v_{\text{B,CYP17H}} + v_{\text{B,HSD3B2}} - v_{\text{B,CYP17L}} - v_{\text{B,CYP21A2}}) \quad \dots \text{ (ODE 12)}$$

$$V_{\text{cell}} \cdot \frac{d[\text{DIONE}]_{\text{cell}}}{dt} = \frac{V_{\text{cell}}^2}{V_{\text{cell}} + V_{\text{med}} \cdot q_{\text{DIONE}}} \cdot (v_{\text{B,CYP17L}} + v_{\text{C,HSD3B2}} - v_{\text{A,CYP19A1}} - v_{\text{A,HSD17B3}}) \quad \dots \text{ (ODE 13)}$$

$$V_{\text{cell}} \cdot \frac{d[\text{TESTO}]_{\text{cell}}}{dt} = \frac{V_{\text{cell}}^2}{V_{\text{cell}} + V_{\text{med}} \cdot q_{\text{TESTO}}} \cdot (v_{\text{A,HSD17B3}} - v_{\text{B,CYP19A1}}) \quad \dots \text{ (ODE 14)}$$

$$V_{\text{cell}} \cdot \frac{d[\text{DCORTICO}]_{\text{cell}}}{dt} = \frac{V_{\text{cell}}^2}{V_{\text{cell}} + V_{\text{med}} \cdot q_{\text{DCORTICO}}} \cdot (v_{\text{A,CYP21A2}} - v_{\text{A,CYP11B1}}) \quad \dots \text{ (ODE 15)}$$

$$V_{\text{cell}} \cdot \frac{d[\text{DCORT}]_{\text{cell}}}{dt} = \frac{V_{\text{cell}}^2}{V_{\text{cell}} + V_{\text{med}} \cdot q_{\text{DCORT}}} \cdot (v_{\text{B,CYP21A2}} - v_{\text{B,CYP11B1}}) \quad \dots \text{ (ODE 16)}$$

$$V_{\text{cell}} \cdot \frac{d[\text{CORTICO}]_{\text{cell}}}{dt} = \frac{V_{\text{cell}}^2}{V_{\text{cell}} + V_{\text{med}} \cdot q_{\text{CORTICO}}} \cdot (v_{\text{A,CYP11B1}} - v_{\text{CYP11B2}}) \quad \dots \text{ (ODE 17)}$$

$$V_{\text{cell}} \cdot \frac{d[\text{CORT}]_{\text{cell}}}{dt} = \frac{V_{\text{cell}}^2}{V_{\text{cell}} + V_{\text{med}} \cdot q_{\text{CORT}}} \cdot v_{\text{B,CYP11B1}} \quad \dots \text{ (ODE 18)}$$

$$V_{\text{cell}} \cdot \frac{d[\text{ALDO}]_{\text{cell}}}{dt} = \frac{V_{\text{cell}}^2}{V_{\text{cell}} + V_{\text{med}} \cdot q_{\text{ALDO}}} \cdot v_{\text{CYP11B2}} \quad \dots \text{ (ODE 19)}$$

$$V_{\text{cell}} \cdot \frac{d[\text{E1}]_{\text{cell}}}{dt} = \frac{V_{\text{cell}}^2}{V_{\text{cell}} + V_{\text{med}} \cdot q_{\text{E1}}} \cdot (v_{\text{A,CYP19A1}} - v_{\text{B,HSD17B3}}) \quad \dots \text{ (ODE 20)}$$

$$V_{\text{cell}} \cdot \frac{d[\text{E2}]_{\text{cell}}}{dt} = \frac{V_{\text{cell}}^2}{V_{\text{cell}} + V_{\text{med}} \cdot q_{\text{E2}}} \cdot (v_{\text{B,HSD17B3}} + v_{\text{B,CYP19A1}}) \quad \dots \text{ (ODE 21)}$$

where, q_x (dimensionless) is the equilibrium constant of steroid hormone x between the cytosol and extracellular culture medium and v_x ($\mu\text{M/h}$) is the enzymatic reaction rate of

steroidogenic enzyme X.

Diffusional transport of steroid hormones

Descriptions of all passive transport reactions of steroid hormones were based on the quasi-equilibrium reaction. Therefore, steroid concentrations in the culture medium were calculated from each cytosolic concentration and equilibrium constant.

$$[\text{PREG}]_{\text{med}} = q_{\text{PREG}} \cdot [\text{PREG}]_{\text{cell}} \quad \dots \text{ (Eq. 4)}$$

$$[\text{HPREG}]_{\text{med}} = q_{\text{HPREG}} \cdot [\text{HPREG}]_{\text{cell}} \quad \dots \text{ (Eq. 5)}$$

$$[\text{DHEA}]_{\text{med}} = q_{\text{DHEA}} \cdot [\text{DHEA}]_{\text{cell}} \quad \dots \text{ (Eq. 6)}$$

$$[\text{PROG}]_{\text{med}} = q_{\text{PROG}} \cdot [\text{PROG}]_{\text{cell}} \quad \dots \text{ (Eq. 7)}$$

$$[\text{HPROG}]_{\text{med}} = q_{\text{HPROG}} \cdot [\text{HPROG}]_{\text{cell}} \quad \dots \text{ (Eq. 8)}$$

$$[\text{DIONE}]_{\text{med}} = q_{\text{DIONE}} \cdot [\text{DIONE}]_{\text{cell}} \quad \dots \text{ (Eq. 9)}$$

$$[\text{TESTO}]_{\text{med}} = q_{\text{TESTO}} \cdot [\text{TESTO}]_{\text{cell}} \quad \dots \text{ (Eq. 10)}$$

$$[\text{DCORTICO}]_{\text{med}} = q_{\text{DCORTICO}} \cdot [\text{DCORTICO}]_{\text{cell}} \quad \dots \text{ (Eq. 11)}$$

$$[\text{DCORT}]_{\text{med}} = q_{\text{DCORT}} \cdot [\text{DCORT}]_{\text{cell}} \quad \dots \text{ (Eq. 12)}$$

$$[\text{CORTICO}]_{\text{med}} = q_{\text{CORTICO}} \cdot [\text{CORTICO}]_{\text{cell}} \quad \dots \text{ (Eq. 13)}$$

$$[\text{CORT}]_{\text{med}} = q_{\text{CORT}} \cdot [\text{CORT}]_{\text{cell}} \quad \dots \text{ (Eq. 14)}$$

$$[\text{ALDO}]_{\text{med}} = q_{\text{ALDO}} \cdot [\text{ALDO}]_{\text{cell}} \quad \dots \text{ (Eq. 15)}$$

$$[\text{E1}]_{\text{med}} = q_{\text{E1}} \cdot [\text{E1}]_{\text{cell}} \quad \dots \text{ (Eq. 16)}$$

$$[\text{E2}]_{\text{med}} = q_{\text{E2}} \cdot [\text{E2}]_{\text{cell}} \quad \dots \text{ (Eq. 17)}$$

Kinetic equations

In this mathematical model of adrenal steroidogenesis in NCI-H295R cells, the flux velocities of molecular transportation and enzymatic reaction rates of steroidogenic enzymes were defined based on the first-order reaction and rapid-equilibrium enzyme kinetics, respectively, by the following equations:

$$v_{CholesterolTransport} = k^{CholesterolTransport} \cdot [CHOL]_{med} \cdot \frac{V_{med}}{V_{cell}} \quad \dots \text{(Eq. 18)}$$

$$v_{CEH} = k^{CEH} \cdot [CHOS]_{cell} \quad \dots \text{(Eq. 19)}$$

$$v_{OxysterolSynthesis} = k^{OxysterolSynthesis} \cdot [CHOC]_{cell} \quad \dots \text{(Eq. 20)}$$

$$v_{MTR} = k_f^{MTR} \cdot [CHOC]_{cell} - k_b^{MTR} \cdot [CHOM]_{cell} \quad \dots \text{(Eq. 21)}$$

$$v_{acc} = k_f^{acc} \cdot [CHOM]_{cell} - k_b^{acc} \cdot [CHOR]_{cell} \quad \dots \text{(Eq. 22)}$$

$$v_{loc} = k_f^{loc} \cdot [CHOM]_{cell} - k_b^{loc} \cdot [CHON]_{cell} \quad \dots \text{(Eq. 23)}$$

$$v_{CYP11A1} = \frac{V_{max}^{CYP11A1} \cdot [CHON]_{cell}}{[CHON]_{cell} + K_m^{CYP11A1}} \quad \dots \text{(Eq. 24)}$$

$$v_{A,CYP17H} = \frac{V_{maxA}^{CYP17H} \cdot \frac{[PREG]_{cell}}{K_{mA}^{CYP17H}}}{1 + \frac{[PREG]_{cell}}{K_{mA}^{CYP17H}} + \frac{[PROG]_{cell}}{K_{mB}^{CYP17H}}} \quad \dots \text{(Eq. 25)}$$

$$v_{B,CYP17H} = \frac{V_{maxB}^{CYP17H} \cdot \frac{[PROG]_{cell}}{K_{mB}^{CYP17H}}}{1 + \frac{[PREG]_{cell}}{K_{mA}^{CYP17H}} + \frac{[PROG]_{cell}}{K_{mB}^{CYP17H}}} \quad \dots \text{(Eq. 26)}$$

$$v_{A,CYP17L} = \frac{V_{maxA}^{CYP17L} \cdot \frac{[HPREG]_{cell}}{K_{mA}^{CYP17L}}}{1 + \frac{[HPREG]_{cell}}{K_{mA}^{CYP17L}} + \frac{[HPROG]_{cell}}{K_{mB}^{CYP17L}}} \quad \dots \text{(Eq. 27)}$$

$$v_{B,CYP17L} = \frac{V_{maxB}^{CYP17L} \cdot \frac{[HPROG]_{cell}}{K_{mB}^{CYP17L}}}{1 + \frac{[HPREG]_{cell}}{K_{mA}^{CYP17L}} + \frac{[HPROG]_{cell}}{K_{mB}^{CYP17L}}} \quad \dots \text{(Eq. 28)}$$

$$v_{A,HSD3B2} = \frac{V_{maxA}^{HSD3B2} \cdot \frac{[PREG]_{cell}}{K_{mA}^{HSD3B2}}}{1 + \frac{[PREG]_{cell}}{K_{mA}^{HSD3B2}} + \frac{[HPREG]_{cell}}{K_{mB}^{HSD3B2}} + \frac{[DHEA]_{cell}}{K_{mC}^{HSD3B2}}} \quad \dots \text{(Eq. 29)}$$

$$v_{B,HSD3B2} = \frac{V_{\max B}^{HSD3B2} \frac{[HPREG]_{\text{cell}}}{K_{mB}^{HSD3B2}}}{1 + \frac{[PREG]_{\text{cell}}}{K_{mA}^{HSD3B2}} + \frac{[HPREG]_{\text{cell}}}{K_{mB}^{HSD3B2}} + \frac{[DHEA]_{\text{cell}}}{K_{mC}^{HSD3B2}}} \quad \dots \text{ (Eq. 30)}$$

$$v_{C,HSD3B2} = \frac{V_{\max C}^{HSD3B2} \frac{[DHEA]_{\text{cell}}}{K_{mC}^{HSD3B2}}}{1 + \frac{[PREG]_{\text{cell}}}{K_{mA}^{HSD3B2}} + \frac{[HPREG]_{\text{cell}}}{K_{mB}^{HSD3B2}} + \frac{[DHEA]_{\text{cell}}}{K_{mC}^{HSD3B2}}} \quad \dots \text{ (Eq. 31)}$$

$$v_{A,CYP21A1} = \frac{V_{\max A}^{CYP21A1} \frac{[PROG]_{\text{cell}}}{K_{mA}^{CYP21A1}}}{1 + \frac{[PROG]_{\text{cell}}}{K_{mA}^{CYP21A1}} + \frac{[HPROG]_{\text{cell}}}{K_{mB}^{CYP21A1}}} \quad \dots \text{ (Eq. 32)}$$

$$v_{B,CYP21A1} = \frac{V_{\max B}^{CYP21A1} \frac{[HPROG]_{\text{cell}}}{K_{mB}^{CYP21A1}}}{1 + \frac{[PROG]_{\text{cell}}}{K_{mA}^{CYP21A1}} + \frac{[HPROG]_{\text{cell}}}{K_{mB}^{CYP21A1}}} \quad \dots \text{ (Eq. 33)}$$

$$v_{A,CYP11B1} = \frac{V_{\max A}^{CYP11B1} \frac{[DCORTICO]_{\text{cell}}}{K_{mA}^{CYP11B1}}}{1 + \frac{[DCORTICO]_{\text{cell}}}{K_{mA}^{CYP11B1}} + \frac{[DCORT]_{\text{cell}}}{K_{mB}^{CYP11B1}}} \quad \dots \text{ (Eq. 34)}$$

$$v_{B,CYP11B1} = \frac{V_{\max B}^{CYP11B1} \frac{[DCORT]_{\text{cell}}}{K_{mB}^{CYP11B1}}}{1 + \frac{[DCORTICO]_{\text{cell}}}{K_{mA}^{CYP11B1}} + \frac{[DCORT]_{\text{cell}}}{K_{mB}^{CYP11B1}}} \quad \dots \text{ (Eq. 35)}$$

$$v_{CYP11B2} = k^{CYP11B2} \cdot [CORTICO]_{\text{cell}} \quad \dots \text{ (Eq. 36)}$$

$$v_{A,HSD17B3} = \frac{V_{\max A}^{HSD17B3} \frac{[DIONE]_{\text{cell}}}{K_{mA}^{HSD17B3}}}{1 + \frac{[DIONE]_{\text{cell}}}{K_{mA}^{HSD17B3}} + \frac{[E1]_{\text{cell}}}{K_{mB}^{HSD17B3}}} \quad \dots \text{ (Eq. 37)}$$

$$v_{B,HSD17B3} = \frac{V_{\max B}^{HSD17B3} \frac{[E1]_{\text{cell}}}{K_{mB}^{HSD17B3}}}{1 + \frac{[DIONE]_{\text{cell}}}{K_{mA}^{HSD17B3}} + \frac{[E1]_{\text{cell}}}{K_{mB}^{HSD17B3}}} \quad \dots \text{ (Eq. 38)}$$

$$v_{A,CYP19A1} = \frac{V_{\max A}^{CYP19A1} \frac{[DIONE]_{\text{cell}}}{K_{mA}^{CYP19A1}}}{1 + \frac{[DIONE]_{\text{cell}}}{K_{mA}^{CYP19A1}} + \frac{[TESTO]_{\text{cell}}}{K_{mB}^{CYP19A1}}} \quad \dots \text{ (Eq. 39)}$$

$$v_{B,CYP19A1} = \frac{V_{\max B}^{CYP19A1} \frac{[TESTO]_{\text{cell}}}{K_{mB}^{CYP19A1}}}{1 + \frac{[DIONE]_{\text{cell}}}{K_{mA}^{CYP19A1}} + \frac{[TESTO]_{\text{cell}}}{K_{mB}^{CYP19A1}}} \quad \dots \text{ (Eq. 40)}$$

where k^X (1/h) is the rate constant of the first order reaction for metabolic flux X, $V_{\max A}^X$ ($\mu\text{M}/\text{h}$) is the maximum velocity of enzyme X for substrate A, and K_{mA}^X is the equilibrium dissociation constant of enzyme X for substrate A.

3. Model parameters

All static parameters in this model are described in Supplementary Table 1. Rate constants and the maximum velocity were estimated by fitting to experimental time-course data of the concentrations of cholesterol and all steroids. Equilibrium dissociation constants were extracted from previous biochemical studies. Equilibrium constants of each steroid transport were estimated in previous studies by Breen and colleagues [1, 2]. All initial values of variable parameters in this model are described in Supplementary Table 2. Initial values of cholesterol and the 14 steroid concentrations were used in each experimentally measured value, and every steroid concentration was assumed to rapidly reach the equilibrium state between the culture medium and intracellular space.

Table S1. Fixed parameters in the adrenal steroidogenesis model of NCI-H295R cells

Parameter number	Model parameter	Optimized value	Units	Sensitivity for the fitting objective function	Reference
1	$k^{CholesterolTransport}$	0.0197596	1/h	4.6234	optimized
2	k^{CEH}	0.302850	1/h	0.0331	optimized
3	k_f^{MTR}	11.7203	1/h	1.4953	optimized
4	k_b^{MTR}	202.531	1/h	1.7737	optimized
5	k_f^{acc}	5.17768	1/h	3.1793	optimized
6	k_b^{acc}	0.0404461	1/h	1.8855	optimized
7	k_f^{loc}	9.97234	1/h	8.0399	optimized
8	k_b^{loc}	2339.88	1/h	5.8004	optimized
9	$k^{OxysterolSynthesis}$	0.0654339	1/h	1.3421	optimized
10	$K_m^{CYP11A1}$	0.5	μM	5.8000	(4)
11	$V_{max}^{CYP11A1}$	675.701	$\mu\text{M}/\text{h}$	22.4750	optimized
12	K_{mA}^{CYP17H}	0.25	μM	0.2342	(5)
13	K_{mB}^{CYP17H}	0.45	μM	0.2570	(5)
14	V_{maxA}^{CYP17H}	108.063	$\mu\text{M}/\text{h}$	3.8388	optimized
15	V_{maxB}^{CYP17H}	1050.21	$\mu\text{M}/\text{h}$	0.2985	optimized
16	K_{mA}^{CYP17L}	0.270	μM	0.1274	(6)
17	K_{mB}^{CYP17L}	0.525	μM	0.1597	(7)

18	$V_{\max A}^{\text{CYP17L}}$	10.8710	$\mu\text{M/h}$	0.2107	optimized
19	$V_{\max B}^{\text{CYP17L}}$	69.7546	$\mu\text{M/h}$	0.3717	optimized
20	K_{mA}^{HSD3B2}	2.8	μM	0.3626	(8)
21	K_{mB}^{HSD3B2}	3.5	μM	0.4001	(8)
22	K_{mC}^{HSD3B2}	3.7	μM	0.0929	(8)
23	$V_{\max A}^{\text{HSD3B2}}$	235.152	$\mu\text{M/h}$	30.9490	optimized
24	$V_{\max B}^{\text{HSD3B2}}$	1901.65	$\mu\text{M/h}$	0.6361	optimized
25	$V_{\max C}^{\text{HSD3B2}}$	525.917	$\mu\text{M/h}$	0.1046	optimized
26	K_{mA}^{CYP21A2}	1.5	μM	0.1652	(9)
27	K_{mB}^{CYP21A2}	1.6	μM	0.3853	(9)
28	$V_{\max A}^{\text{CYP21A2}}$	213.911	$\mu\text{M/h}$	7.0023	optimized
29	$V_{\max B}^{\text{CYP21A2}}$	544.496	$\mu\text{M/h}$	1.9225	optimized
30	K_{mA}^{CYP11B1}	2.50	μM	0.1095	(10)
31	K_{mB}^{CYP11B1}	0.882	μM	0.1042	(11)
32	$V_{\max A}^{\text{CYP11B1}}$	214.848	$\mu\text{M/h}$	0.2427	optimized
33	$V_{\max B}^{\text{CYP11B1}}$	36.9996	$\mu\text{M/h}$	0.1781	optimized
34	k^{CYP11B2}	0.0698923	1/h	0.1634	optimized
35	K_{mA}^{HSD17B3}	0.7	μM	0.1022	(12)
36	K_{mB}^{HSD17B3}	3.3	μM	0.1111	(12)
37	$V_{\max A}^{\text{HSD17B3}}$	0.81059	$\mu\text{M/h}$	0.1666	optimized
38	$V_{\max B}^{\text{HSD17B3}}$	6.15932	$\mu\text{M/h}$	0.1769	optimized
39	K_{mA}^{CYP19A1}	0.215	μM	0.0035	(13)
40	K_{mB}^{CYP19A1}	0.370	μM	0.0003	(13)
41	$V_{\max A}^{\text{CYP19A1}}$	10.0091	$\mu\text{M/h}$	0.7052	optimized
42	$V_{\max B}^{\text{CYP19A1}}$	1.0e-6	$\mu\text{M/h}$	0.0000	optimized
43	q_{PREG}	0.0129	dimensionless	---	(2)
44	q_{HPREG}	0.0605	dimensionless	---	(2)
45	q_{DHEA}	0.0377	dimensionless	---	(2)
46	q_{PROG}	0.0052	dimensionless	---	(2)
47	q_{HPROG}	0.0212	dimensionless	---	(2)
48	q_{DIONE}	0.0400	dimensionless	---	(2)
49	q_{DCORTICO}	0.0412	dimensionless	---	(2)
50	q_{DCORT}	0.0422	dimensionless	---	(2)
51	q_{CORTICO}	0.0558	dimensionless	---	(2)

52	q_{CORT}	0.0676	dimensionless	---	(2)
53	q_{ALDO}	0.0911	dimensionless	---	(2)
54	q_{TESTO}	0.0443	dimensionless	---	(2)
55	q_{E1}	0.0267	dimensionless	---	(2)
56	q_{E2}	0.0351	dimensionless	---	(2)

Table S2. Initial values of variable parameters in the adrenal steroidogenesis model of NCI-H295R cells.

Compartment	Molecular parameter name	Initial value	Unit
Culture medium	CHOL	81.1	μM
Culture medium	PREG	0.00085	μM
Culture medium	HPREG	0.06945	μM
Culture medium	DHEA	0.0	μM
Culture medium	PROG	0.00003	μM
Culture medium	HPROG	0.0	μM
Culture medium	DIONE	0.00080	μM
Culture medium	DCORTICO	0.0	μM
Culture medium	DCORT	0.0	μM
Culture medium	CORTICO	0.00011	μM
Culture medium	CORT	0.00003	μM
Culture medium	ALDO	0.00091	μM
Culture medium	TESTO	0.00080	μM
Culture medium	E1	0.00011	μM
Culture medium	E2	0.00121	μM
Intracellular space	OXY	0.0	nmol
Intracellular space	CHOS	16.08	nmol
Intracellular space	CHOC	0.3503	nmol
Intracellular space	CHOM	0.02943	nmol
Intracellular space	CHON	0.0006431	nmol
Intracellular space	CHOR	0.6350	nmol
Intracellular space	PREG	0.008580	nmol
Intracellular space	HPREG	0.0	nmol
Intracellular space	DHEA	0.003165	nmol

Intracellular space	PROG	0.00002892	nmol
Intracellular space	HPROG	0.00009361	nmol
Intracellular space	DIONE	0.002115	nmol
Intracellular space	DCORTICO	0.0007598	nmol
Intracellular space	DCORT	0.06871	nmol
Intracellular space	CORTICO	0.002065	nmol
Intracellular space	CORT	0.003114	nmol
Intracellular space	ALDO	0.0	nmol
Intracellular space	TESTO	0.0	nmol
Intracellular space	E1	0.001895	nmol
Intracellular space	E2	0.0003870	nmol

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