

mean \pm sd derived from 3 replicate samples. Panels b and d are images of agarose gel electrophoresis showing single band of the PCR products. Ctrl, control siRNA; IP, immunoprecipitation; F/F, *Frs2 α ^{fllox}*; CN, *Frs2 α ^{CN}*; *, $P < 0.05$.

Online Fig. 1. Depletion of FRS2 α compromises the tumorigenicity of MDA PCa 118b cells. **A.** Real-time RT-PCR analyses of MDA PCa 118b cells infected with shFRS2 α (n=6) control (n=6) adenoviruses. **B&C.** X-ray analyses of mouse femurs 7 weeks after injection of 1.5×10^6 of the indicated cells. Investigators were blinded when assessing the bone lesions and blood vessels. The tumor incidence rates were calculated and expressed as mean \pm sd (B). Representative images are shown in (C). **D.** Immunostaining of tumor sections with anti-CD31 antibodies. Panel c is the average percent of CD31 stained areas in randomly viewed areas. Ctrl, control virus infected PC3 cells; *, $P < 0.05$.

Online Fig. 2. Validation of antibodies used for immunostaining. **A&B.** Immunostaining of HIF1 α , FRS2 α , phosphorylated FRS2 α , and cJun. Expression of HIF1 α in PC3 cells was induced by treating the cells with 150 μ M CoCl₂ in DEMEM containing 2% serum for 24 hours. Untreated cells were used as a negative control for anti-HIF1 α antibody. For anti-FRS2 α staining anti-phosphorylated FRS2 α , and anti-cJUN, the siRNA depleted cells were used as negative controls. **C.** The human PCa tissue section was co-stained with anti-CD31 antibody and isolectin. The images were captured with a confocal microscope. The data showed that both anti-CD31 and

isolectin stained the same areas of the section, indicating that anti-CD31 antibody specifically recognized endothelial cells in the tissue. To-Pro3 (TP3) was used for nuclear counterstaining. Ctrl, control.



