

Supplementary Methods

Constructs for CRISPR-Cas9 and transformation:

The vector *pPcas9-rU3-gRNA* from the ViewSolid Biotech was used, and two 20 bp fragments from the second exon of *NF-YB1* (sequence as showed in Figure 1C) was introduced into the vector, and then these plasmids were transformed into rice (Zhonghua 11) as described previously [1]. Principles for designing the target was described previously [2].

Constructs for RNAi and transformation:

RNA interference (RNAi) construct with a 350-bp fragment (nucleotides from 168 to 517 counted from ATG of coding sequence of *NF-YB1*) amplified from cDNA of caryopses of Zhonghua 11 with sequences carrying restriction enzyme sites *KpnI-SpeI* (forward primer), *BamHI-SacI* (reverse primer) and cloned into the vector pEASY-Blunt Simple vector. The plasmid was then digested by *KpnI-BamHI* to obtain the sense fragment and by *SpeI-SacI* to obtain the antisense fragment. These sense and antisense fragments were introduced into the target vector *pTCK303* together to generate the *pUbi::NF-YB1RNAi* construct. The protocols and principles for construct the RNAi vector as described previously [3].

Yeast one-hybrid experiments:

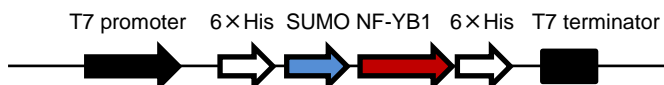
The cDNA sequence of the *NF-YB1* was amplified from cDNA isolated from caryopses of Zhonghua 11, and cloned into the vector *pB42AD* to generate *GAD-NF-YB1*. The 5' upstream regulatory sequences of *SUT1* (3 177 bp), *SUT3* (2 821 bp), *SUT4* (1 026 bp), *MST4* (3 074 bp) and *SUT2* (1 055 bp) were introduced into *pLacZ2u* to generate the LacZ reporter plasmids. These five constructs were transformed individually into yeast strain EGY48, together either with the *pB42AD* empty vector or *GAD-NF-YB1*. Yeast cells were grown on a synthetic dropout medium without tryptophan and uracil. 5-bromo-4-chloro-3-indolyl-b-D-galactopyranoside was used for color assay.

Luciferase (LUC) activity assay in rice protoplasts:

For the transient expression assay, the cDNA sequence of *NF-YB1* was amplified from cDNA of caryopses of Zhonghua 11, and cloned into the vector *pCambia1301-35S::tNOS* to get the effector construct; the 5' regulatory sequences of *SUT1*, *SUT3*, *SUT4*, *MST4* and *SUT2* (sequences are same as used in the yeast one-hybrid experiments described above) were introduced into the vector *pUC-35sLUC* to generate the reporter plasmids. *p35S::GUS* was used as an internal control. The effector (*p35S::NF-YB1::tNOS*) or the empty vector were co-transfected with one of the reporter constructs (*pSUT1::LUC*, *pSUT3::LUC*, *pSUT4::LUC*, *pMST4::LUC* or *pSUT2::LUC*) and *p35S::GUS*. Empty vector co-transfected with the reporter construct was normalized to 1, and luciferase activities in protoplasts co-transfected with the effector and these reporter constructs were relative to this value. The protoplasts were incubated in the dark for 12 hours before harvesting, and the LUC and GUS activities were determined with a luminometer / fluorometer (Promega). The protocols for isolating protoplasts and transfection were used as described previously [4].

Purification of His-NF-YB1 recombinant protein and EMSA:

The cDNA sequence of the *NF-YB1* was amplified from cDNA of Zhonghua 11, and cloned into the expression vector *pET28-SUMO-XF1024* to generate the *pHis-NF-YB1* expression vector, and the His-NF-YB1 protein was produced in the *Escherichia coli* BL21 (Transetta) strain and the recombinant protein were then purified by Ni-charged resin (Bio-Rad). The 59 bp fragments containing the CCAAT box (sequences showed below) were identified from the 1 kb 5' upstream region of *SUT1*, *SUT3* or *SUT4*, labeled with biotin. The unlabeled fragments were used as the competitive probes. *SUT4Δ* was made with a mutated CCAAT box (CCAAT changed to TGATA), which was used as a negative control. Then the recombinant protein with the labeled probes, or labeled probes with 200-folds, 500-folds unlabeled competitive probes were incubated on the ice for 20 minutes, the binding buffer used was Tris-HCl (pH 8.5). The electrophoresis was performed on 6% native polyacrylamide gel for 70 minutes, and then transferred to membrane, cross-linked, detected as described in the instruction book of the Light Shift Chemiluminescent EMSA Kit (Thermo Scientific). The rough map of the *pHis-NF-YB1* is illustrated below.



Supplementary information

Putative CCAAT elements identified in 5' upstream sequences of *SUT1*, *SUT3* and *SUT4*, and sequences used for EMSA: putative CCAAT elements are underlined, and sequences used for EMSA are colored in red.

The 5' upstream sequence of *SUT1*:

GGAGGAGTTAACAGCAAAGACAAGAAAGGATAATCGAGATCCTCTACAATACTATATGTGCAGTACTACTTACTGACATG
TGAATCAACTGTCAGTGAGTCGATATGTCGTGAACAGTACTGCACATGTATGGCACTACAAGAGTACTTGCATCGAAGGA
AGAAGCGTCAACCCCCCCCCCCCCCGGGGCGGGGCGAGCACACGCGTAGTCGCGGTAGCGGCCCTGCGATTCTTC
CTTATCCATTCATGCATGCATGGCAGACCATTCCCTCCATTCCATTTGATCGATTTTAACTCCTCGGTATTGCAAACCCG
ATAGCCTCGGTATGCCAAATCTTGGTGTTACGGATGGATGCATGGATGATGTGGTTGACCAACCCGTCGTCAGTTGTTCA
TTGATCATAACGCATCGGAGAAAGAAAGAGACTTTCCGTCTTTCTTTACCTTCCAATCCCCGTTGACTTGGATCATGTCAT
GCAGCTATCTGAGTGAATTGTTAATTAAGCAAGAGACATTCCTCGCTTGCAGTGCACCCGGCCGGGCACAGAACAAATGT
CACTGTCGCGGTACTGAATTATTCAGACAATCTGGCTGCATCTCCTCATTTTACAGCATCAGTTAGACAAGCCACCTTTT
TTCCAGTAGTATGGGTTATGTGTAGGTTTGCTGCATACTCCTATATAGGAGTATCGCTTAAAGGAATCAATGTGCAGGCT
CCCATAGAGTTCAGCTGAATGCTTCAGAAGGAATTGAATGTCCATACTGCAGATTTGTAGATGTAGATATCGAGAAGCA
TAGCCTTATATCTGCATGCATGGACTTGGATATTTCTCTTATCTGATAGCCCATGTATAGCTCTATATCTGCACAACA
CTTTACCATGCACATATATTTGCATCTAGCTACAACCTCCGTGAGACATCTAGCTAGCTAGGAGGTGTGTGCAACAAGAAC
AATATCATATCTCACACCTTTTCCGAAAACCTCAATGTCAAACCCGGCATCATATCTCCATTGACGTGGTTTTCTCTT
TTTTCCAGAGAAAAAAAACATGCATGTTGAGTAGTACGAATTATAGACTCTACTGCAAATGCCAAGAAATATTAATTGGG
ACAAAAACGACGAAATGAACACTGTATTAACCCGATGCAAACCTGCTAAAAGTAGCGTTAATTAAGCTGACCATTGCGA
CCGACCGACCGAATTGAATCGAGTCTCCAAAAGAAAAAGAGAAAAATGCCCTTCTCTGCATGCATGCAGTCTTGGTGC
AGCAGTGACCTCACTGAATTTGTTTGTCTTGGCTAGGGACTGCTTGAATTGGATTTTAGTCTAGCAAAATTTGACTC
CAACATGGGAGCATGTGAGATACATGTCATTGATCACATCACTATACTGCTGTGTATCTTTTCATACCAACTACCATA
ATATGGTAGCCCAAGTAGTGGAAATTAAGAAAGCACAACCTCCATATCTCGATCCATCTATCCCTTACTATATATAT
CTATATCATTATCCCGTAGCTATTTACAATATCACAATATATAAAAAGATTTAGTTTTTTTTAAGGAGTGAATATGTTT
GTTTCAGTTTGACTATAATTTCTGAACTTTTTTCAGTGACCGACCAGGCGACCACCCGTTACTAAGATCAGGGTAAAAT
GTGAGGCCGACATGTGTATTACTGGTCAATGGCAGAACTGAAATTCGAAAATGTATTGATGTATTATTCCTCTTTTTTTT
TTGTTCAAATTAAGATGACACGTAGGCTCCAGCAACCATCCACGTGGCGGAGGTTGTGCACGTCCTCCATCTCCAACA
CATCAGCATTTTTAGCAAAGAAAGAAAAAACTGTAGTGGACTTTGCAGAAAACGCTGACGACGAGTACGTACGAATTAAT
GCCTTAATCATATGATATCTCCAAAGATTAATTAATAAGTGGCTTGCTTAATTTTGTACTAATCGCTATCTCGACGTAG
CCAGAGAGCCATGCATGCATCACAGCGAGCCCACGGCCGACGAGCACTCGCCAACGAATTCGCTAGGCGTACACCGTGA
ATGATTTGATGCGTTGATTACGGGTATTCATATTCCTTTATGAAAGGTTATTGTCAGACTTTTTTTTATCCACAAGATCG
ATCATACTACAAAGTTATTCTACAATAGTTTAGAACACTTATCCAGTTGTGTTAGAATATAATAATGATGGATGGATATG
TATGCCATATTAACAATCTAAATTTCCCAAAAACATATAAAAAGAACTATAATAAACTATGGTTTATCCAACATGGA
CATATATTTAAATGAAGTGCATCTCCGGTGCTCTTTACTGGTAGGATGAATGATGATAGAGATAAAAGCGTTTAAACAAA
TATGGCCTCAAGCGAAATTCGTTATATTAATTAATCAATGAAAACATTTACTGGATTAATAAAAACCTCATGCTACTCCA
TTATAAATGAACGCACACCTATATATAGCAAATTCCTATTTGCCAGTAGGTCCAATACTTCGGATCTGTTTTTTTTCT
TTTAAATATCCAAATTTGATTTGGATACTACTCGACGTACAACGAATTAACCAGCTATTACAACGTCGAGTGGAT
TTAAAACACTCCTCTATTAATTCACCTACAGAAAGTCGTTCCCGTGAAATAATCGCACCGTCTAGAAGCTCGGCAAGC
GTGTCGCTAATCCGATACTAACTCCATTAATTCATTTCAATAATTGTTGAAGTTATTACTGCACTGGAATAA
TAAAGGCAGGGGGTGTAACTGGGTGTGTACAAAGTGTGGTGAGCATAGCAGTTGGCAGGTGCACCACCTTTATTATA

The 5' upstream sequence of *SUT4*:

TGGCAGACTACAGAGGAATGAATGGTCCATTGTTGGGGACATAGGGTGTGGCGTTTGATTTAGTTTGTGTTTGTGGTGTAT
TGCTATCTGCTAATGTTAGTAGCTTAATTATGTACACTGTTGAGCTAAGTTGTCATTACTATAGAGCTCTCTCCTATACA
TGAGGGGAGGTGACATGATTTTGGTGTGCCTTTTTCTGTGCGGTGGTGGTCTGCTGGGTTGTGGTCATACAATCCAGGTG
CAGACATGATGTATTGTTTGCGAATTTATTATATTTTTTTCAGTCTAACCCCTGAGGAAAAGGAATCTTTGGTGGTGTGGAA
TAGGCAAATACGTTGGGTGGCATAATGTTAGATGAAGTGATTGATACACAGGTGGTTGTAGAGGTAACGTTGAGTGATC
AAGTGCAGGCTTGCTGGAGCAAAAAGAAGATGATGCCTCACTCAGTCGTTGACGGCACAAATCTAGCTAACCTGCTGCCAT
TGTGGTGATTCATGCCTCCTCTCCGAAGACTCTTTGACTTGACTAATGGAGTGGACTGCTACCAAACACGCATGAATAG
ACGACCAGATCATCATACAATTCATCAAAATTAGCACTCCTAGTTTCAAACACTTAAACAAAGGAAATTC AACAAACTTC
ATTTAAATAGTGTAGTGTATTTTTTTCTTTAAAAAAAATTGCCAACGCCAATCGCGGAGTTTACATGGAGGAGGGAAG
AATGGAATGAGCGCCAGGCGTCGCGTCGGCGCGCGTGAGTGCCTGATTTGTGTTGGTTGTCTTCCTCTTGGCTTCGCT
TCGCTGCTTTGCCGCGCTTCGCCGAGGGAGGCCACCAACGCATCAATCAAACACACAAGCACACCACGCGGACGCAGC
AGCAGGGGAGGAGACAATTTCTATTCTTCCTCGCCCCGCGTCGCCTCGCCTGAGTCTGACTCTCAAACGCCGACCAGT
GACGCCGCGAGCCTTGCCCCTTGCCC GCGCAGATCTACCAAACCCTACCAGATCTGCGCCCCGCC

Supplementary information

References:

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