

1 **Relative abundance of *Nitrotoga* spp. in a biofilter of a cold-freshwater aquaculture**
2 **plant appears to be stimulated by slightly acidic pH**

3
4 Jennifer Hüpeden^a, Simone Wegen^a, Sandra Off^{a*}, Sebastian Lücker^{b,c}, Yvonne Bedarf^a,
5 Holger Daims^c, Carsten Kühn^d and Eva Spieck^{a#}

6 ^aBiocenter Klein Flottbek, Microbiology & Biotechnology, University of Hamburg, Hamburg,
7 Germany

8 ^bDepartment of Microbiology, Faculty of Science, Radboud University, Nijmegen, the
9 Netherlands

10 ^cDepartment of Microbiology and Ecosystem Science, Division of Microbial Ecology,
11 University of Vienna, Vienna, Austria

12 ^dState Research Centre of Agriculture and Fisheries Mecklenburg-Vorpommern, Institute of
13 Fisheries, Rostock, Germany

14
15 #Address correspondence to Eva Spieck, eva.spieck@uni-hamburg.de

16 *Present address: Hamburg University of Applied Sciences, Department of Life Sciences,
17 Hamburg, Germany

18
19
20 **Running title:** Abundance of *Nitrotoga* in aquaculture biofilters

23 **Supplemental Material**

24

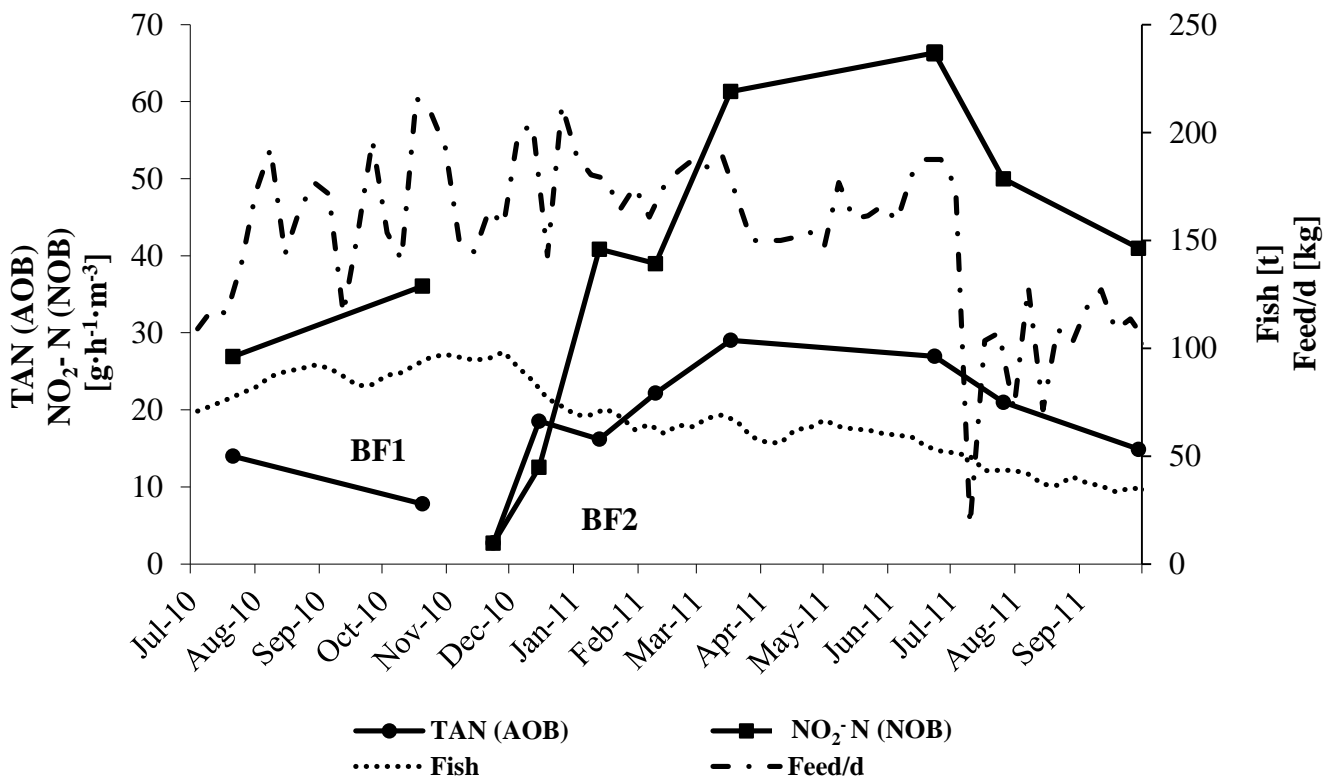


25

26

27 Fig. S1: Well covered carrier element type BCN009 of BF2 sampled in March 2011 after an
28 operating time of four months, illustrating the dense microbial biofilm on the inside.

29



30

31 Fig. S2: Temporal development of potential nitrification rates in BF1 (left) and BF2 (right).

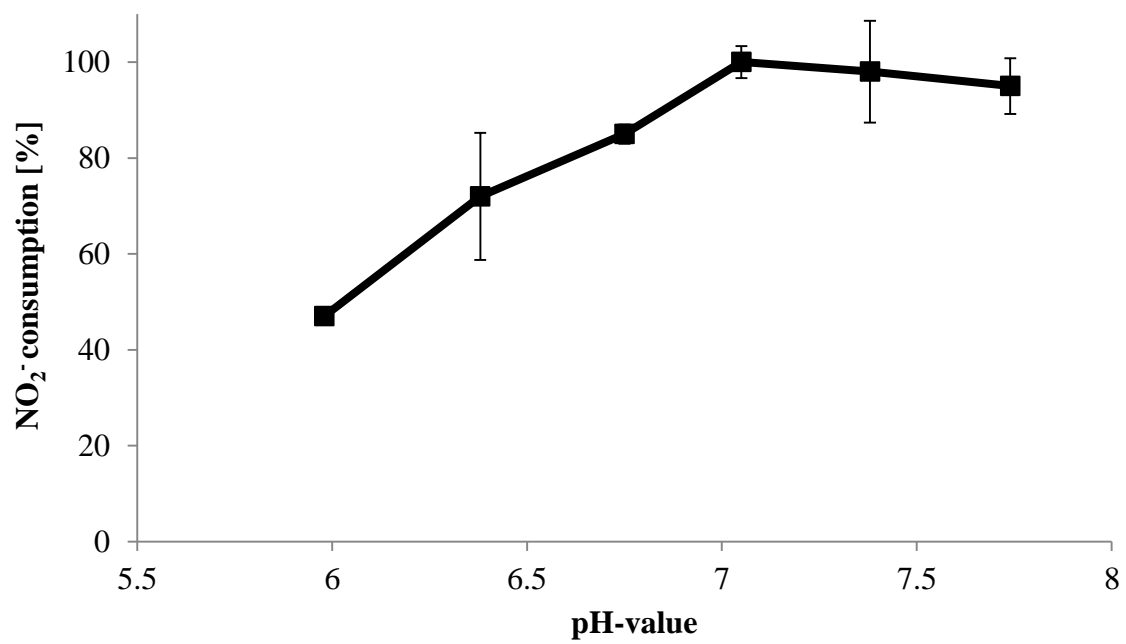
32 Turnover rates refer to the conversion of ammonia via AOB (TAN) and nitrite via NOB

33 (NO₂-N) within 8 hours. The amount of fish and the daily addition of feed inside the

34 respective aquaculture ponds of BF1 and BF2 were evaluated as weekly average stocking and

35 feeding rates.

36



37

38 Fig. S3: Short-term nitrite consumption of biofilm inhabiting NOB of BF2 in dependence on

39 the pH, incubated at 17°C.

40