

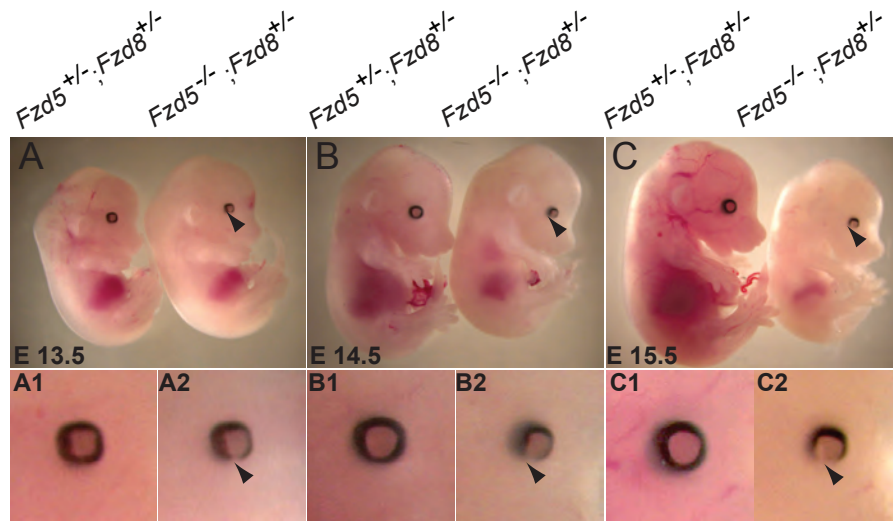
Supplementary table 1: Rare *FZD5* variant identified in coloboma patients

Patient id	cDNA change	Protein change	dbSNP id	1000G/EVS MAF	Polyphen prediction	SIFT prediction
49	c.A290T	*D97V	NA	NA	Possibly damaging	Damaging

* D97V variant has not been detected in control genomes, SNP database and 1000 Genome Project. Primary tests show that this variant increases *FZD5*-mediated canonical Wnt activity.

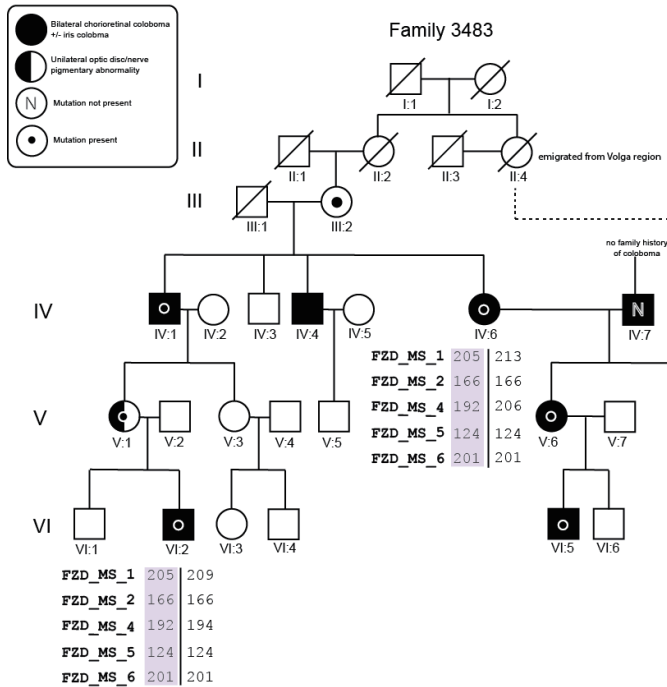
Supplementary table 2: *FZD5* gene amplification primers (5'>3')

<i>FZD5</i> Primer1F	tgccaggcgcgctcgcctcc
<i>FZD5</i> Primer2F	taaccgtctctcccagccctatc
<i>FZD5</i> Primer3F	cgcgacgccgaggctcctctgcatg
<i>FZD5</i> Primer4F	gcagtactccacctggctgcgt
<i>FZD5</i> Primer5R	caccactacctctcaggcac
<i>FZD5</i> Primer6R	agaaacgcaaaatagaatacac
<i>FZD5</i> Primer7R	cgtcttggtgccgcctgcttg
<i>FZD5</i> Primer8R	ggtagcagggtaccgcgcag
<i>FZD5</i> Primer9R	gatgggtcaggccgagccatc
<i>FZD5_ex2_1F</i>	gtagcgcgacggccagtgagattggagacagctcgc
<i>FZD5_ex2_2F</i>	gtagcgcgacggccagtctggaggtgcaccagtct
<i>FZD5_ex2_3F</i>	gtagcgcgacggccagtgccttcagtgccgacgag
<i>FZD5_ex2_4F</i>	gtagcgcgacggccagtgaaccagaacctgaactcg
<i>FZD5_ex2_1R</i>	cagggcgcagcgatgacgcagacagatgggcgtgta
<i>FZD5_ex2_2R</i>	cagggcgcagcgatgactccatgctgatgaggaaggt
<i>FZD5_ex2_3R</i>	cagggcgcagcgatgacgaagagcgacacgaagcc
<i>FZD5_ex2_4R</i>	cagggcgcagcgatgactaaacggaagtaccttggc

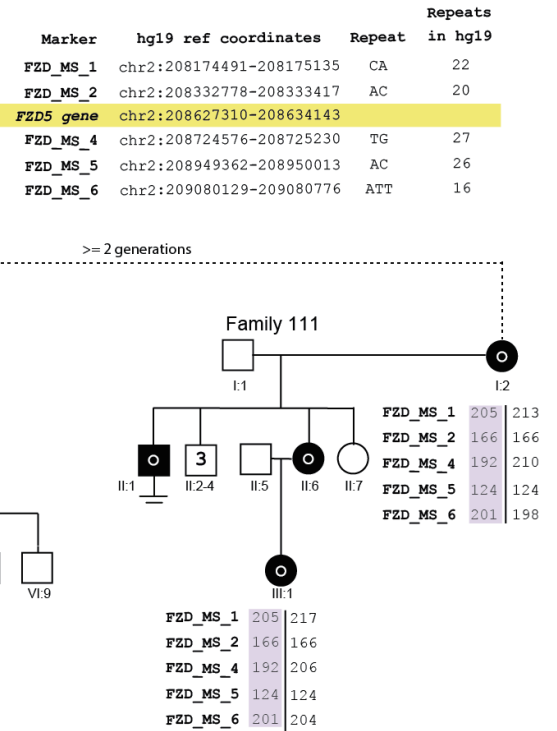


Supplementary Figure 1

A



B



Supplementary Figure 2

Frizzled 5 protein N-terminal alignment across species

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Protein Domains [-----signal peptide(aa1-26)-----]
Human WT      MARPD----PSAPPS-----LLLLLLAQLVGRAAAASKAPVCEQITVPMCRGIGYNL 48
cow           MARPD----PSAPPS-----LLLLLLAQLAGRAAAAASKAPACQEQITVPMCRGIGYNL 48
pig           MARPD----PSAPPS-----LLLLLLAQLAGRATAASKAPVCEQITVPMCRGIGYNL 48
dog           MARPD----PCAPPS-----LLLLLLAQLLGRAAAASKAPVCEQITVPMCRGIGYNL 48
elephant     MARLD----PSAPPP-----LLLLLLAQLVGRAAAASKAPVCEQITVPMCRGIGYNL 48
gorilla      MARPD----PSAPPS-----LLLLLLAQLVGRAAAASKAPVCEQITVPMCRGIGYNL 48
chimp        MARPD----PSAPPS-----LLLLLLAQLVGRAAAASKAPVCEQITVPMCRGIGYNL 48
mouse        MARPD----PSAPPS-----LLLLLLAQLVGRAAAASKAPVCEQITVPMCRGIGYNL 48
zebrafish    MRKPADEHHFTMETSGMHLVGFVLLHVLVLLFQLSGLGDSASKDIVCEPITVPMCKGIGYNH 60
* :          . .          * :*** ** * . :*** .*: *****:*****.

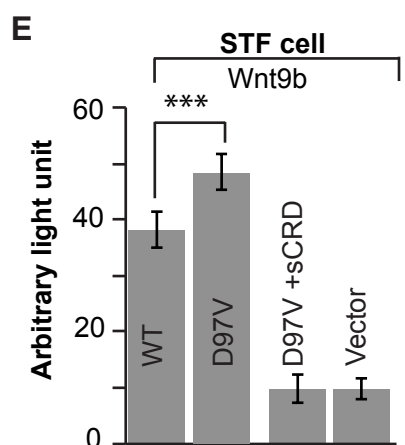
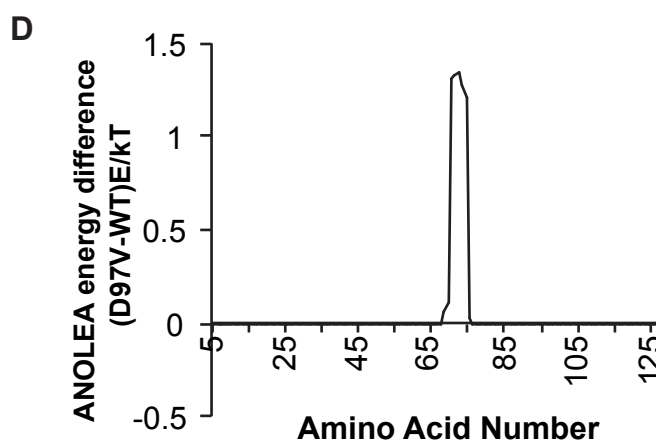
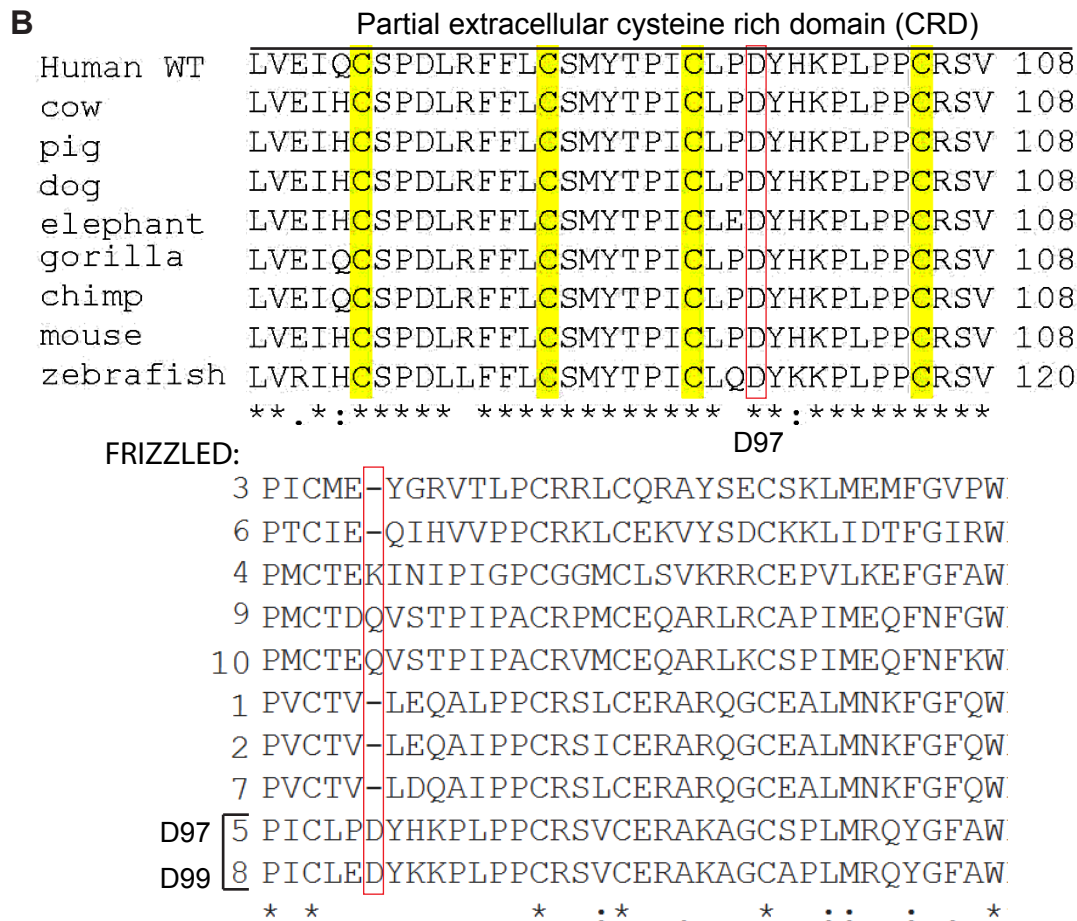
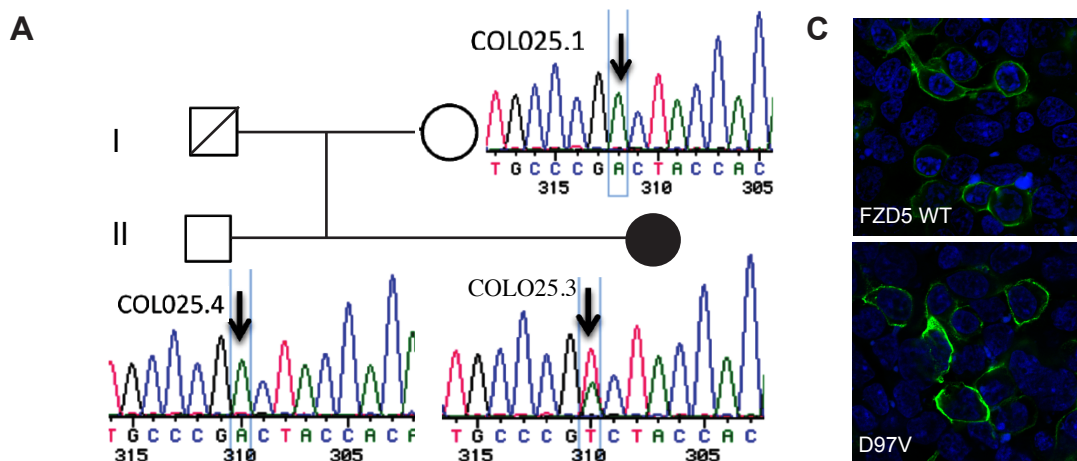
-----frizzled domain(~aa30-152) (aka cysteine rich domain)---
Human WT      THMPNQFNHDTQDEAGLEVHQFWPLVEIQCSPLDRFFFLCSMYTPICLPDYHKPLPPCRSV 108
cow           THMPNQFNHDTQDEAGLEVHQFWPLVEIHCSPDLRFFFLCSMYTPICLPDYHKPLPPCRSV 108
pig           THMPNQFNHDTQDEAGLEVHQFWPLVEIHCSPDLRFFFLCSMYTPICLPDYHKPLPPCRSV 108
dog           THMPNQFNHDTQDEAGLEVHQFWPLVEIHCSPDLRFFFLCSMYTPICLPDYHKPLPPCRSV 108
elephant     THMPNQFNHDTQDEAGLEVHQFWPLVEIHCSPDLRFFFLCSMYTPICLEDYHKPLPPCRSV 108
gorilla      THMPNQFNHDTQDEAGLEVHQFWPLVEIQCSPLDRFFFLCSMYTPICLPDYHKPLPPCRSV 108
chimp        THMPNQFNHDTQDEAGLEVHQFWPLVEIQCSPLDRFFFLCSMYTPICLPDYHKPLPPCRSV 108
mouse        THMPNQFNHDTQDEAGLEVHQFWPLVEIHCSPDLRFFFLCSMYTPICLPDYHKPLPPCRSV 108
zebrafish    TYMPNQFNHDTQDEAGLEVHQFWPLVRIHCSPDLLFFFLCSMYTPICLQDYKKPLPPCRSV 120
*:*****.***.*****.*:***** ***** **:******

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Human WT      CERAKAGCSPLMRQYGFAPPERMSCDRLPVLGRDAEVL CMDYNRSEATTAPPRPFPKPT 168
cow           CERAKAGCSPLMRQYGFAPPERMSCDRLPVLGRDAEVL CMDYNRSEATTAPPRPFPKPT 168
pig           CERAKAGCSPLMRQYGFAPPERMSCDRLPVLGRDAEVL CMDYNRSEATTAPPRPFPKPT 168
dog           CERAKAGCSPLMRQYGFAPPERMSCDRLPVLGRDAEVL CMDYNRSEATTAPPRPFPKPT 168
elephant     CERAKAGCSPLMRQYGFAPPERMSCDRLPVLGRDAEVL CMDYNRSEATTAPPRPFTAKPT 168
gorilla      CERAKAGCSPLMRQYGFAPPERMSCDRLPVLGRDAEVL CMDYNRSEATTAPPRPFPKPT 168
chimp        CERAKAGCSPLMRQYGFAPPERMSCDRLPVLGRDAEVL CMDYNRSEATTAPPRPFPKPT 168
mouse        CERAKAGCSPLMRQYGFAPPERMSCDRLPVLGGDAEVL CMDYNRSEATTASPKSFPKPT 168
zebrafish    CERAKRGCSPMLIQYGFAPPERMSCDRLPMLG-DTDRLCMDRNSSETTTLSP-PFP-KPT 177
***** ***** **** *****:***:*** *: : **** * **:*** .* *. ***

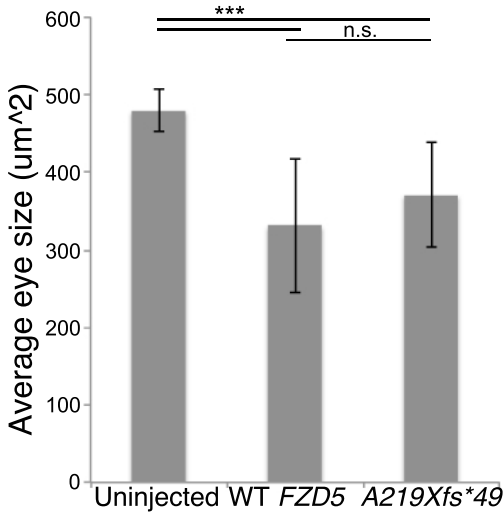
First altered aa (A219X) in frameshift
Human WT      LPGPPGAPASGGGECFAG-GPFVCKREPFVPIPKESHPLYNKVVRTGQVPNCVPCYQP-- 227
cow           LSLPLGSPASGNDCAAG-GPSVCKREPFVPIPKESHPLYNKVVRTGQVPNCVPCYQP-- 227
pig           LSGPPGAPASGSDCVAG-GPSVCKREPFVPIPKESHPLYNKVVRTGQVPNCVPCYQP-- 227
dog           LPGPAGGAPASGAECAAG-APSVCKREPFVPIPKESHPLYNKVVRTGQVPNCVPCYQP-- 227
elephant     HPGLPGAPASGVECAAG-GPSVCKREPFVPIPKESHPLYNKVVRTGQVPNCVPCYQP-- 227
gorilla      LPGPPGAPASGGGECFAG-GPFVCKREPFVPIPKESHPLYNKVVRTGQVPNCVPCYQP-- 227
chimp        LPGPPGAPASGGGECFAG-GPFVCKREPFVPIPKESHPLYNKVVRTGQVPNCVPCYQP-- 227
mouse        LPGPPGAPASSGGGECPSG-GPSVCTCREPFVPIPKESHPLYNKVVRTGQVPNCVPCYQP-- 227
zebrafish    PKGTPRHRATAKSAPPQKCDRECHCRGPLVPIKKEAHPLHNRVNTGSLPNCALPCHQP-- 237
* .      :.. . .      * ** :*** **:***:***.***:***:*** ** *

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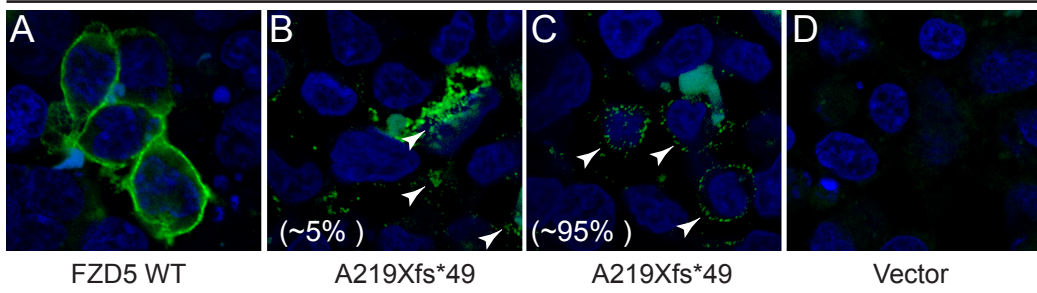
Supplementary Figure 3



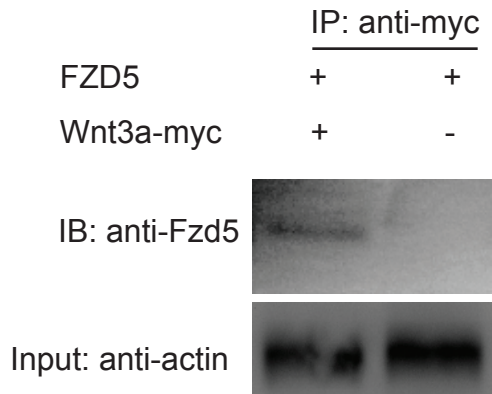
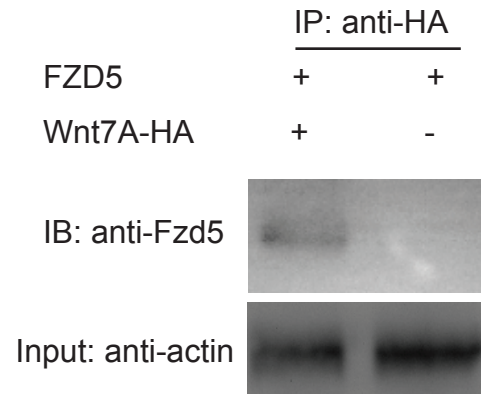
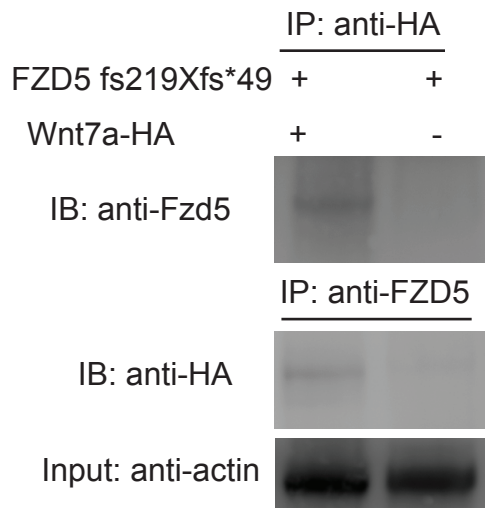
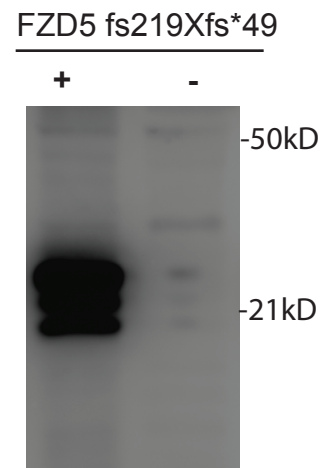
Supplementary Figure 4

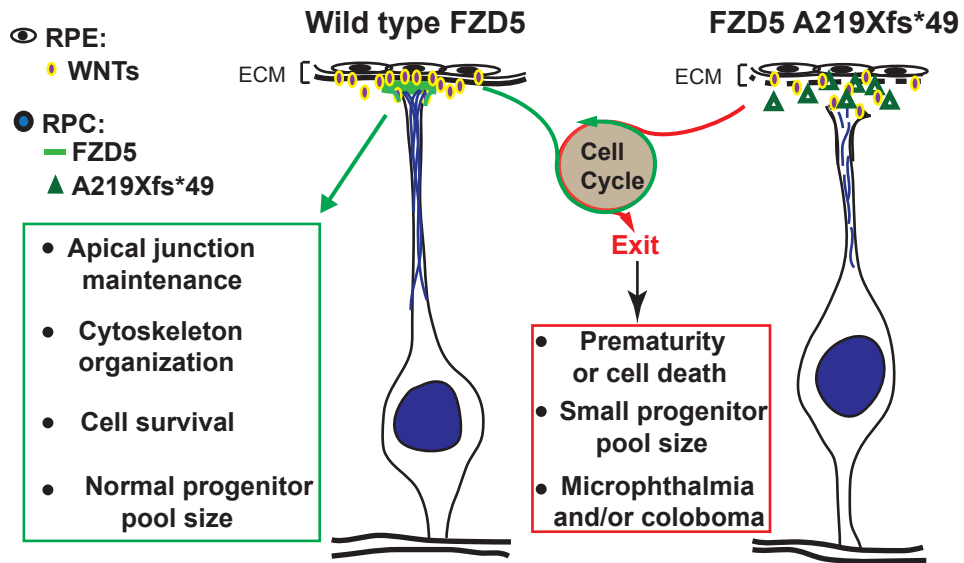


Anti-Fzd5 antibody live cell surface staining



Supplementary Figure 6

A**B****C****D****Supplementary Figure 7**



Supplementary Figure 8

Supplementary Figure Legends

Supplementary Figure 1. Microphthalmia and retinal coloboma manifested in the compound mutants of homozygous *Fzd5* conditional knockout (*Fzd5*^{-/-}) with one allele of *Fzd8* mutation (*Fzd8*^{+/-}). Lower panels are larger views of developing eyes corresponding to upper panels. Arrowheads indicate open optic fissure/coloboma. Transgenic *Sox2-Cre* was used to excise loxP sites. A, A1, A2, E13.5; B, B1, B2, E14.5; C, C1, C2, E15.5.

Supplementary Figure 2. Microsatellite markers and Haplotype analysis of Families 3484 and 111. **A.** Shows the same pedigree as Figure 1. **B.** Gives the absolute genomic coordinates of the five microsatellite markers used in this analysis and shows their position relative to the *FZD5* gene (yellow highlight). Under four affected individuals (Family 3483 IV:6 & VI:2 and Family 111 1:2 & III:1) are shown the most plausible locus haplotypes for each individual. The purple highlight indicates the identical haplotype shared by each of the affected individual suggesting recent common ancestry. It should be noted that markers FZD_MS_2 and FZD_MS_5 are not informative for haplotype construction.

Supplementary Figure 3. Conservation of FZD5 Wnt-binding cysteine rich domain (CRD) across species. Yellow bars highlighted 10 conserved cysteines in FZD5 CRD. The purple bar indicates mutant protein insertion/deletion point (p219A).

Supplementary Figure 4. Identification of a novel missense mutation -FZD5

D97V. A. Sequences of a two-generation family with the daughter OC patient carrying D97V mutation. **B.** Upper panel: Alignment of Frizzled 5 CRD region from multiple species showing the conservation of D97. Lower panel: Alignment of ten FZD CRDs shows that D97 is variable except for FZD8 and FZD5. **C.** FZD5 D97V protein is correctly localized in transfected cells. **D.** Atomic non-local environment assessment (ANOLEA) predicted that the D97V variant perturbs local interactions. **E.** Slightly increase in Wnt9b induced canonical Wnt activity by D97V can be abolished by FZD5 sCRD. Student t-test was used for statistical analysis. ***, $P < 0.0001$.

Supplementary Figure 5. Eye size analysis of zebrafish embryos injected

with FZD5 mRNA. Zebrafish embryos were injected at the 1-cell stage with 200 pg of either WT *FZD5* or *A219Xfs*49 FZD5* mRNA and imaged at 3 dpf. Ocular area measurements were taken 3 times with ImageJ and averaged for each eye. Measurements for eyes in each injection group were then averaged (Un-injected, N=12 embryos; WT *FZD5*, N=20 embryos; *A219Xfs*49 FZD5*, N=30 embryos). ***, $p < 0.0003$; n.s.=not significant; t-test with Bonferroni correction for multiple comparisons.

Supplementary Figure 6. Cellular localization of wild type and mutant FZD5

protein. Immunofluorescence detection of FZD5 proteins (green) on cell surface.

Images were merged with DAPI indicating the nucleus. **A**, Wild type FZD5 was localized on the cell membrane. **B-C**, A majority of A219Xfs*49 mutant protein was present in extracellular space, presumptively, ECM (arrowheads). The distribution of the mutant proteins appeared to be uneven with about 5% cells showing locally heavy and/or dispersed deposition (**B**, arrowhead), whilst the rest (~95% **C**, arrowhead) showing local (near-membrane) FZD5 distribution. **D**, Negative control with vector transfection was shown in right panel. To avoid the cytoplasm staining, live cells were first incubated with anti-FZD5 antibody in cultured medium, washed with PBS, and then post-fixed with PFA for further immunohistochemistry.

Supplementary Figure 7. FZD5 A219Xfs*49 binds to Wnt. **A**, FZD5 binds to myc-tagged Wnt3a. HEK293T cells co-transfected with *Wnt3a-myc* and *FZD5*. Cell extracts were immunoprecipitated (IP) with anti-myc antibody, and immunoblotted (IB) with anti-FZD5 antibody. **B**, FZD5 binds to HA-tagged WNT7A. HEK293T cells co-transfected with *WNT7A-HA* and *FZD5*. Cell extracts were immunoprecipitated with anti-HA antibody, and the immunoblot was probed with anti-FZD5 antibody. **C**, FZD5219Xfs*49 protein binds to HA-tagged WNT7A. HEK293 cells were co-transfected with *WNT7A-HA* and *FZD5219Xfs*49* constructs. Cell extracts were immunoprecipitated with anti-HA antibody, and the protein blot was probed with anti-FZD5 antibody. Reverse IP was conducted with anti-FZD5 antibody and the blot was probed with anti-HA antibody. **D**, Multiple bands of FZD5219Xfs*49 were detected under reducing conditions using anti-

FZD5 antibody.

Supplementary Figure 8. A model of coloboma disease mechanism caused by *FZD5 A219Xfs*49*. During development, WNT signaling is crucial for maintenance of neuroblast apical junction, cell polarity, cell survival and proliferation. By competing for Wnt ligands, A219Xfs*49 mutant protein (dark green triangles) may intercept WNTs (yellow circles), which are secreted from RPE at the apical extracellular matrices during development (brackets), preventing FZD5 (from RPC apical membrane, bright green lines) - evoked WNT signaling in the neuroblasts (blue nuclear cells). Consequently, insufficient WNT-FZD5 signaling leads to early cell cycle exit, prematurity and cell death, and reduced progenitor pool size, resulting in microphthalmia and/or coloboma. The model is modified from Liu et al., 2012 (32).