# Web appendix: Supplementary material

# Performance of alternative strategies for primary cervical cancer screening in sub-Saharan Africa: systematic review and meta-analysis of diagnostic test accuracy studies

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1. PICOS components

1.1. Research Question

Among the most studied alternative methods to cytology for cervical cancer screening (Visual inspection with

Acetic Acid, Visual inspection with Lugol's Iodine and Human Papillomavirus testing), which tool performs

better in the context of primary screening in sub-Saharan Africa?

1.2. PICOS components

**Population**: women apparently healthy, previously unscreened, participating in a primary screening program in

sub-Saharan Africa

**Intervention** (screening with index tests): each screening tool considered independently

I1: Visual inspection with Acetic Acid

I2: Visual inspection with Lugol's Iodine

I3: Human Papillomavirus testing

Comparison (reference standard): either random biopsy without colposcopy, or colposcopy and colposcopy-

directed biopsies, loop excision or endocervical curettage performed in all women of the study population ('Gold

standard all' group, GSA) or in a proportion of women including all screen-positive women to the considered

index text ('Gold standard partial' group, GSP)

**Outcomes:** 

Outcome 1: absolute sensitivity and specificity in detecting CIN2+ for VIA, VILI and HPV testing

Outcome 2: relative sensitivity and specificity for CIN2+ detection of

VIA versus VILI

VIA versus HPV testing

VILI versus HPV testing

Outcome 3: prevalence of CIN2+, positivity rate of VIA, VILI and HPV testing

Studies:

Diagnostic test accuracy studies conducted in sub-Saharan Africa, where one of the index tests was

performed using a cross-sectional design

Diagnostic test accuracy studies conducted in sub-Saharan Africa, where two index tests were

performed independently in women, using a cross-sectional design

1

- Randomized trial studies with either of the index texts in one arm, when both screening and reference tests were performed at enrollment

#### 2. Literature review strings

#### 2.1. In Medline (PubMed)

#1: Africa OR "Africa South of the Sahara" OR "sub-Saharan-Africa" OR "Low resource setting"

#2: Angola OR Benin OR Botswana OR "Burkina Faso" OR "Upper Volta" OR Burundi OR Urundi OR Cameroon OR Cameroons OR "Cape Verde" OR "Central African Republic" OR Chad OR Comoros OR "Comoro Islands" OR Comores OR Mayotte OR Congo OR "Cote d'Ivoire" OR "Ivory Coast" OR "Democratic Republic of the Congo" OR Zaire OR Djibouti OR "French Somaliland" OR Eritrea OR Ethiopia OR Gabon OR "Gabonese Republic" OR Gambia OR Ghana OR "Gold Coast" OR Guinea OR Kenya OR Lesotho OR Basutoland OR Liberia OR Madagascar OR "Malagasy Republic" OR Malawi OR Nyasaland OR Mali OR Mauritania OR Mauritius OR Mozambique OR Namibia OR Niger OR Nigeria OR Rwanda OR "Sao Tome" OR Seychelles OR Senegal OR "Sierra Leone" OR Somalia OR "South Africa" OR Sudan OR "South Sudan" OR Swaziland OR Tanzania OR Togo OR "Togolese Republic" OR Uganda OR Zambia OR Zimbabwe OR Rhodesia

#3: Cervical cancer OR cervical neoplasia OR cervical neoplasm OR cancer of the uterine cervix

#4: screening OR early detection

#5: (#1 OR #2) AND #3 AND #4

#### With the following Filters:

- Publication dates : from 01/01/1994 to 30/06/2014

- Species: Humans

#### 2.2. In Embase

#1: low+resource+setting OR 'africa'/exp OR africa OR 'sub+saharan+africa'/exp OR sub+saharan+Africa

#2: angola OR benin OR botswana OR 'burkina faso' OR 'upper volta' OR burundi OR urundi OR cameroon OR cameroons OR 'cape verde' OR 'central african republic' OR chad OR comoros OR 'comoro islands' OR comores OR mayotte OR congo OR 'cote ivoire' OR 'ivory coast' OR 'democratic republic of the congo' OR zaire OR djibouti OR 'french somaliland' OR eritrea OR ethiopia OR gabon OR 'gabonese republic' OR gambia OR ghana OR 'gold coast' OR guinea OR kenya OR lesotho OR basutoland OR liberia OR madagascar OR 'malagasy republic' OR malawi OR nyasaland OR mali OR mauritania OR mauritius OR mozambique OR namibia OR niger OR nigeria OR rwanda OR 'sao tome' OR seychelles OR senegal OR 'sierra leone' OR somalia OR 'south africa' OR sudan OR 'south sudan' OR swaziland OR tanzania OR togo OR 'togolese republic' OR uganda OR zambia OR zimbabwe OR Rhodesia

#3: 'cervical+cancer'/exp OR cervical+cancer

#4: 'screening'/exp OR screening OR 'early detection of cancer'/exp OR 'early detection of cancer'

#### #5: (#1 OR #2) AND #2 AND #3

#### With the following Filters:

- Publication dates: from 01/01/1994 to 30/06/2014
- Map to preferred terminology
- Also search as free text
- Include sub-terms/derivatives

#### 2.3. In Cochrane library

#1: 'cervical cancer' or 'cervical neoplasia'

#2: Visual inspection with acetic acid

#3: Visual inspection with Lugol's Iodine

#4: 'Human papillomavirus' or 'Human papillomavirus testing'

#4: 'Low resource setting' or 'Africa' or 'sub-saharan Africa'

#### With the following Filters:

- Cochrane reviews
- Other reviews

#### 3. Excluded studies

## List of studies excluded after reading of abstracts or methods of full texts

First Author, Year	Country		Screening Test (s)	Code exclusion
Untiet, 2014 <sup>1</sup>	Cameroon		HPV	1
Dartell, 2014 <sup>2</sup>	Tanzania		HPV, VIA	1
Ajenifuja, 2013 <sup>3</sup>	Nigeria		VIA	1
Ugwu, 2013 <sup>4</sup>	Nigeria		NS	2
Ogilvie, 2013 <sup>5</sup>	Uganda		HPV	3
Firnhaber, 2013 <sup>6</sup>	South Africa		HPV, VIA	4
Mwanahamuntu, 2013 <sup>7</sup>	Zambia		VIA	4
De Vuyst, 2013	Kenya		HPV	4
Chigbu, 2013 <sup>9</sup>	Nigeria		VIA	2
Busingye, 2012 <sup>10</sup>	Uganda		VIA, VILI	2
Denny, 2012 11	South Africa	3	HPV, VIA, VILI	2
Moon, 2012 <sup>12</sup>	Mozambique	_	NS	4
Horo, 2012 <sup>13</sup>	Cote d'Ivoire		VIA	4
Mingo, 2012 <sup>14</sup>	Botswana		NS	2
Audet, 2012 <sup>15</sup>	Mozambique		VIA	2
Jemal, 2012 <sup>16</sup>	NS		NS	2
Gage, 2012 <sup>17</sup>	Nigeria		CareHPV	5
Ramogola-Masire, 2012 <sup>18</sup>	Bostwana		VIA	4
Kahesa, 2012 <sup>19</sup>	Tanzania		NS	2
Teguete, 2012 <sup>20</sup>	Mali		VIA, VILI	6
Cronje, 2011 <sup>21</sup>	NA		VIA, HPV	2
Awodele, 2011 <sup>22</sup>	Nigeria		NS	2
Balandya, 2011 <sup>23</sup>	Tanzania		VIA	4
Lewis, 2011 <sup>24</sup>	Kenya		VILI	5
Saleh, 2011 <sup>25</sup>	Tanzania		NS	2
Kuhn, 2010 <sup>26</sup>	South Africa		HPV	4
Hovland, 2010 <sup>27</sup>	DRC		HPV	7
Were, 2010 <sup>28</sup>	Kenya		VIA, VILI	2
Batra, 2010 <sup>29</sup>	South Africa		Cytology	2
Peters, 2010 <sup>30</sup>	Tanzania		NS	2
Denny, 2010 <sup>31</sup>	South Africa		HPV, VIA	3
Koffi, 2010 <sup>32</sup>	Central african		cytology	2
	Republic			
Qureshi, 2010 <sup>33</sup>	India		VIA, VILI	8
Hassan, 2009 <sup>34</sup>	Sudan		Cytology	2
Hoque, 2009 <sup>35</sup>	Botswana		NS	2
Akinwuntan, 2008 <sup>36</sup>	Nigeria		VIA	4
Anorlu, 2008 <sup>37</sup>	NA		NS	2
Kawonga, 2008 <sup>38</sup>	South Africa		Cytology	2
Hoque, 2008 <sup>39</sup>	South Africa		Cytology	2
Arbyn, 2008 <sup>40</sup>	5 countries		VIA, VILI	2
Kamaté, 2008 <sup>41</sup>	Mali		VIA, VILI	2

Roblyer, 2007 <sup>42</sup>	Nigeria	NA	2
Akinola, 2007 <sup>43</sup>	Nigeria	VIA	1
Anorlu, 2007 <sup>44</sup>	Nigeria	NS	2
Kamal, 2007 <sup>45</sup>	India	VIA, cytology	8
Cronje, 2007 <sup>46</sup>	South Africa	Cytology	2
Sodhani, 2006 <sup>47</sup>	India	VIA	8
Cronje, 2005 <sup>48</sup>	NA	NS	2
Denny, 2005 <sup>49</sup>	NA	NS	2
Denny, 2005 <sup>50</sup>	NA	HPV	2
Doh, 2005 <sup>51</sup>	Cameroon	VIA	1
Millogo, 2004 <sup>52</sup>	Burkina Faso	VIA, VILI	9
El Shalakany, 2004 <sup>53</sup>	Egypt	VIA	8
Bhatla, 2004 <sup>54</sup>	India	VIA, VILI	8
Okewole, 2003 <sup>55</sup>	Nigeria	NS	2
Petry, 2003 <sup>56</sup>	Tanzania	HPV	2
Hawes, 2003 <sup>57</sup>	Senegal	HPV	2
Claeys, 2003 <sup>58</sup>	Nicaragua	VIA	8
Cronjé, 2003 <sup>59</sup>	South Africa	VIA	6
Adanu, 2002 <sup>60</sup>	Ghana	NS	2
Blumenthal, 2001 <sup>61</sup>	Zimbabwe	VIA, HPV	10
Cronjé, 2001 <sup>62</sup>	South Africa	VIA	6
Singh, 2001 <sup>63</sup>	India	VIA	8
Cronje, 2000 <sup>64</sup>	South Africa	VIA	1
Denny, 2000 <sup>65</sup>	South Africa	VIA, HPV	10
Womack, 2000 <sup>66</sup>	Zimbabwe	HPV	10
Chirenje, 1999 <sup>67</sup>	Zimbabwe	VIA	1
Thistle, 1997 <sup>68</sup>	Zimbabwe	cytology	2
Sitas, 1997 <sup>69</sup>	South Africa	cytology	2
Megevand, 1996 <sup>70</sup>	South Africa	Cytology	10
Nolting, 1995 <sup>71</sup>	South Africa	VIA	2

HPV: human papillomavirus testing. NA: not applicable. NS: not specified. VIA: visual inspection with acetic acid. VILI: visual inspection with Lugol's Iodine.

#### Reasons for exclusion of studies

Code	Reason for exclusion	Number
1	Inappropriate gold standard or disease threshold	7
2	No accuracy study or no primary data (review, comment, letter)	35
3	Inappropriate design (including self-sampling for HPV testing)	2
4	HIV positive women	9
5	no primary screening (test used for triage)	2
6	Many screening tests performed not independently of each other	3
7	Symptomatic patients	1
8	Studies not conducted in sub-Saharan Africa	7
9	Screening performed by physicians	1
10	Double reporting	4

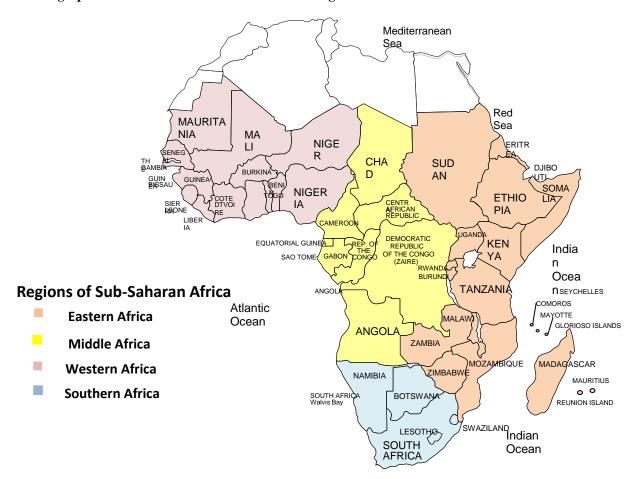
4. Items included in the systematic review of accuracy of alternative methods for cervical cancer screening in SSA according to the PRISMA guidelines  $^{72}$ 

## Supplementary Table 1: Checklist of items included in the systematic review

n/topic	Checklist item	
	Identifying the general as a systematic goviery, moto analysis, on both	
CITE	Identifying the report as a systematic review, meta-analysis, or both.	
CT		
summary	Providing a structured summary including, as applicable: background; objectives; data sources; study eligibility criteria, participants, and interventions; study appraisal and synthesis methods; results; limitations; conclusions and implications of key findings.	
UCTION		
	Describing the rationale for the review in the context of what is already known.	
	Providing an explicit statement of questions being addressed with reference to participants, interventions, comparisons, outcomes, and study design (PICOS).	
OS		
criteria	Specifying study characteristics (e.g., PICOS, length of follow-up) and report characteristics (e.g., years considered, language, publication status) used as criteria for eligibility, giving rationale.	
n sources	Describing all information sources (e.g., databases with dates of coverage, contact with study authors to identify additional studies) in the search and date last searched.	
	Presenting full electronic search strategy for at least one database, including any limits used, such that it could be repeated.	
ction	Stating the process for selecting studies (i.e., screening, eligibility, included in systematic review, and, if applicable, included in the meta-analysis).	
ction	Describing method of data extraction from reports (e.g., piloted forms, independently, in duplicate) and any processes for obtaining and confirming data from investigators.	
	Listing and defining all variables for which data were sought (e.g., PICOS, funding sources) and any assumptions and simplifications made.	
ıs in studies	Describing methods used for assessing risk of bias of individual studies (including specification of whether this was done at the study or outcome level).	
measures	Stating the principal summary measures (e.g., risk ratio, difference in means).	
of results	Describing the methods of handling data and combining results of studies, if done, including measures of consistency (e.g., I <sup>2</sup> ) for each meta-analysis.	
is across	Specifying any assessment of risk of bias that may affect the cumulative evidence (e.g., publication bias, selective reporting within studies).	
analyses	Describing methods of additional analyses (e.g., sensitivity or subgroup analyses, meta- regression), if done, indicating which were pre-specified.	

n/topic	Checklist item
S	
ction	Giving numbers of studies screened, assessed for eligibility, and included in the review, with reasons for exclusions at each stage, ideally with a flow diagram.
racteristics	For each study, presenting characteristics for which data were extracted (e.g., study size, PICOS, follow-up period) and providing the citations.
as within	Presenting data on risk of bias of each study and, if available, any outcome-level assessment.
of results	Presenting results of each meta-analysis done, including confidence intervals and measures of consistency.
as across	Presenting results of any assessment of risk of bias across studies.
analysis	Give results of additional analyses, if done (e.g., sensitivity or subgroup analyses, meta-regression).
ION	
of evidence	Summarize the main findings including the strength of evidence for each main outcome; consider their relevance to key groups (e.g., health care providers, users, and policy makers).
S	Discuss limitations at study and outcome level (e.g., risk of bias), and at review level (e.g., incomplete retrieval of identified research, reporting bias).
ns	Provide a general interpretation of the results in the context of other evidence, and implications for future research.
<b>5</b>	
	Describe sources of funding for the systematic review and other support (e.g., supply of data); role of funders for the systematic review.

#### 5. Geographical areas in sub-Saharan Africa according to United Nations 73



Supplementary figure 1: Countries in sub-Saharan Africa, by geographic region

Sub-Saharan Africa includes countries of Eastern Africa (Burundi, Comoros, Djibouti, Eritrea, Ethiopia, Kenya, Madagascar, Malawi, Mauritius, Mayotte, Mozambique, Uganda, Tanzania, Reunion, Rwanda, Seychelles, Somalia, South Sudan, Zambia, and Zimbabwe); Central or Middle Africa (Angola, Cameroon, Congo, Gabon, Equatorial Guinea, Central African Republic, Democratic Republic of Congo, Sao Tome and Principe, and Chad); Southern Africa (South Africa, Botswana, Lesotho, Namibia, and Swaziland) and Western Africa (Benin, Burkina Faso, Cape Verde, Ivory Coast, Gambia, Ghana, Guinea, Guinea-Bissau, Liberia, Mali, Mauritania, Niger, Nigeria, Saint Helena, Senegal, Sierra Leone, and Togo). Sudan which belongs to Northern Africa is also part of sub-Saharan Africa. As part of this work, Sudan was linked to South Sudan (Eastern Africa), as the only study included in these two countries was conducted when they were still a single nation.

Supplementary Table 2: Assessment of methodological validity of selected studies

G. P.	Screening		Risk of bias									Applicability concerns					
Studies	test(s)	Patie Selec		Inde	ex test	Re	ferenc	e test			Flow and	timin	g		Patient Selection	Index test	Reference test
		P 1	P2	T1	T2	R1	R2	R3	F1	F2	F3	F4	F5	F6			
Megevand, 1996 <sup>74</sup>	VIA	Y	U	Y	Y	N	U	N	Y	N	N	N	N	N	L	Н	L
Univ. Zimbabwe, 1999 <sup>75</sup>	VIA	U	U	Y	Y	N	Y	N	Y	Y	N	Y	y	У	L	L	L
Womack, 2000 <sup>76</sup>	HPV test	U	U	Y	Y	N	Y	Y	Y	Y	Y	Y	Ň	Ý	L	L	L
Wright, 2000 <sup>77</sup>	HPV test	U	U	Y	Y	N	Y	Y	N	N	Y	N	N	N	L	L	L
Kuhn, $2000^{78}$	HPV test	U	U	Y	U	N	U	Y	U	N	Y	N	N	N	L	L	L
Denny, 2000 <sup>79</sup>	VIA	U	U	Y	Y	N	U	N	Y	N	N	Y	N	N	L	Н	L
Denny, 2002 <sup>80</sup>	VIA	U	U	Y	Y	N	U	N	N	N	N	Y	N	Y	L	H	L
Sankaranarayan, 2004 <sup>81</sup>	VIA/VILI	Y	U	Y	Y	N	Y	N	Y	Y	N	Y	N	Y	L	L	L
De Vuyst, 2005 <sup>82</sup>	VIA/HPV test	Y	U	Y	Y	N	Y	N/Y*	Y	Y	N/Y*	N	N	N	Н	L	Н
Sangwa-Lugoma, 2006 <sup>83</sup>	VIA/VILI	Y	U	Y	Y	N	Y	N	Y	Y	N	N	N	Y	L	L	L
Muwonge, 2010 <sup>84</sup>	VIA/VILI	U	U	Y	Y	N	Y	N	Y	Y	N	Y	N	Y	L	L	L
Ngoma, 2010 <sup>85</sup>	VIA/VILI	Y	U	Y	Y	N	Y	N	Y	Y	N	Y	N	Y	L	U	L
Ibrahim, 2012 <sup>86</sup>	VIA	U	U	Y	Y	N	U	N	Y	N	N	U	N	N	L	H	L
Mahmud, 2012 <sup>87</sup>	HPV test	Y	U	Y	Y	N	Y	Y	Y	N	Y	Y	Y	Y	L	L	L
Jeronimo, 2014 <sup>88</sup>	VIA/HPV test	U	U	Y	Y	N	U	N/Y*	Y	N	N/Y*	Y	N	N	L	U	L

QUADAS items<sup>89</sup>: P1=acceptable enrolment method, P2=inappropriate exclusions avoided, T1=pre-specified test cut-off or criteria for a positive result, T2=results of the index test are masked towards the reference test, R1=imperfect gold standard bias avoided, R2=results of the reference test are masked towards the index test, R3=incorporation bias avoided, F1=acceptable delay between index test and reference test, F2=partial verification avoided, F3=differential verification avoided, F4=withdrawals explained, F5=uninterpretable results reported for reference test. Each quality item is judged with the following: Y=fulfilled, U=unclear and N=not fulfilled for risk of bias and L=low risk, U=unclear risk and H=high risk for concerns of applicability. \* N for VIA and Y for HPV test. HPV test: Human Papillomavirus testing, VIA: visual inspection with acetic acid, VILI: visual inspection with Lugol's Iodine.

#### 6. Forest plot of the prevalence of CIN2+ and positivity rate of screening tests

#### 6.1. In the GSA Group

#### Supplementary figure 2a: Prevalence of disease (CIN2+) in the GSA group, by screening test

GSA: gold standard (colposcopy followed by colposcopy directed biopsies) performed in all women of the study population. CIN2+: cervical intraepithelial neoplasia grade 2 or worse. N: number of patients. VIA: visual inspection with acetic acid. VILI: visual inspection with Lugol's iodine. HPV: Human papillomavirus.

Study groups	Cases	N				Prevalence	95% CI
1 - VIA studies							
Univ. Zimbabwe 1999 (Zimbabwe)	206	2130				0.097	[0.084; 0.110]
Sankaranarayan 2004 (Burkina)	50	2051	-			0.024	[0.018, 0.032]
Sankaranarayan 2004 (Congo)	389	6935		-		0.056	[0.051; 0.062]
Sankaranarayan 2004 (Guinea)	168	8627				0.019	[0.017; 0.023]
Sankaranarayan 2004 (Mali)	164	5552	+			0.030	[0.025; 0.034]
Sankaranarayan 2004 (Niger)	20		+			0.008	[0.005; 0.012]
De Vuyst 2005 (Kenya)	60					0.092	[0.071; 0.117]
Sangwa-Lugoma 2006 (DRC)	29	1528	-			0.019	[0.013; 0.027]
Muwonge 2010 (Angola)	132	8849				0.015	[0.012, 0.018]
Ngoma 2010 (Tanzania)	233	10374				0.022	[0.020; 0.025]
Random effects model		49233	0	-		0.033	[0.021; 0.047]
Heterogeneity: I-squared-98.4%, tau-squared-0.0033, Q=559.9, df-9, p<0.0001							
2 - VILI studies							
Sankaranarayan 2004 (Burkina)	50	2051	-			0.024	[0.018; 0.032]
Sankaranarayan 2004 (Congo)	388	6934		+		0.056	[0.051; 0.062]
Sankaranarayan 2004 (Guinea)	168	8627	+			0.019	[0.017; 0.023]
Sankaranarayan 2004 (Mali)	164	5552	+			0.030	[0.025; 0.034]
Sankaranarayan 2004 (Niger)	20	2534	+			0.008	[0.005, 0.012]
Sangwa-Lugoma 2006 (DRC)	29	1528				0.019	[0.013; 0.027]
Muwonge 2010 (Angola)	132	8842				0.015	[0.013; 0.018]
Ngoma 2010 (Tanzania)	233	10367				0.022	[0.020; 0.026]
Random effects model		46435	0			0.023	[0.015; 0.033]
Heterogeneity: I-squared=97.6%, tau-squared=0.0018, Q=293.7, df=7, p<0.0001							
3 - HPV studies							
Womack 2000 (Zimbabwe)	207	2145				0.097	[0.084; 0.110]
De Vuyst 2005 (Kenya)	53	453				0.117	[0.089; 0.150]
Mahmud 2012 (DRC)	24	1352	-			0.018	[0.011; 0.026]
Random effects model		3950	_	_	_	0.069	[0.017; 0.152]
Heterogeneity: I-squared=98.4%, tau-squared=0.0138, Q=127.4, df=2, p<0.0001							
				T.	10.50		
		1	0 (	0.05	0.1 0.	15	

#### Supplementary figure 2b: Prevalence of disease (CIN2+) in the GSA group, by geographic region

GSA: gold standard (colposcopy followed by colposcopy directed biopsies) performed in all women of the study population. CIN2+: cervical intraepithelial neoplasia grade 2 or worse. N: number of patients. VIA: visual inspection with acetic acid. VILI: visual inspection with Lugol's iodine. HPV: Human papillomavirus.

Study groups	Cases	N				Prevalence	95% CI
Eastern Africa							
Univ. Zimbabwe 1999 (Zimbabwe)	206	2130			-	0.097	[0.084; 0.110]
Womack 2000 (Zimbabwe)	207	2145				0.097	
De Vuyst 2005 (Kenya)	60	653				0.092	
Ngoma 2010 (Tanzania)	233	10374	*			0.022	
Random effects model		15302				- 0.072	
Heterogeneity: I-squared=99.2%, tau-squared=0.0117, Q=362, df=3, p<0.000							
Middle Africa							
Sankaranarayan 2004 (Congo)	389	6935		-		0.056	[0.051; 0.062]
Sangwa-Lugoma 2006 (DRC)	29	1528	-#-			0.019	
Muwonge 2010 (Angola)	132	8849	*			0.015	
Mahmud 2012 (DRC)	24	1352	-			0.018	
Random effects model	1.70	18664	-	-		0.025	
Heterogeneity: I-squared=98.7%, tau-squared=0.0048, Q=225.6, df=3, p<0.00	01					,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	
Western Africa							
Sankaranarayan 2004 (Burkina)	50	2051	-4-			0.024	[0.018; 0.032]
Sankaranarayan 2004 (Guinée)	168	8627				0.019	
Sankaranarayan 2004 (Mali)	164	5552	-			0.030	
Sankaranarayan 2004 (Niger)	20	2534	*			0.008	
Random effects model		18764	0			0.019	
Heterogeneity: I-squared=94.1%, tau-squared=0.001, Q=50.9, df=3, p<0.0001							
			1	1	-1	1	
			0	0.05	0.1	0.15	

#### Supplementary figure 3: Positivity rate of VIA, VILI and HPV testing in the GSA group

GSA: gold standard (colposcopy followed by colposcopy directed biopsies) performed in all women of the study population. CIN2+: cervical intraepithelial neoplasia grade 2 or worse. N: overall number of patients. VIA: visual inspection with acetic acid. VILI: visual inspection with Lugol's iodine. HPV: Human papillomavirus.

Study groups	Positive	N		Postivity rate	95% CI
1 - VIA studies					
Univ. Zimbabwe 1999 (Zimbabwe)	849	2130	+	0.399	[0.378; 0.420]
Sankaranarayan 2004 (Burkina)	561	2051	+	0.274	[0.254; 0.293]
Sankaranarayan 2004 (Congo)	1845	6935		0.266	[0.256; 0.277]
Sankaranarayan 2004 (Guinea)	680	8627	Til.	0.079	[0.073; 0.085]
Sankaranarayan 2004 (Mali)	624	5552		0.112	[0.104; 0.121]
Sankaranarayan 2004 (Niger)	152	2534	*	0.060	[0.051; 0.070]
De Vuyst 2005 (Kenya)	177	653		0.271	[0.237; 0.307]
Sangwa-Lugoma 2006 (DRC)	555	1528	-	0.363	[0.339, 0.388]
Muwonge 2010 (Angola)	581	8849		0.066	[0.061; 0.071]
Ngoma 2010 (Tanzania)	399	10374	4	0.038	[0.035; 0.042]
Random effects model		49233		0.174	[0.104; 0.256]
Heterogeneity: I-squared-99.8%, tau-squared-0.0264, Q-4448.6, df-9, p<0.00	001				. MORE TRANSPORT
2 - VILI studies					
Sankaranarayan 2004 (Burkina)	588	2051	+	0.287	[0.267; 0.307]
Sankaranarayan 2004 (Congo)	1094	6934		0.158	[0.149; 0.167]
Sankaranarayan 2004 (Guinea)	974	8627		0.113	[0.106; 0.120]
Sankaranarayan 2004 (Mali)	727	5552		0.131	[0.122; 0.140]
Sankaranarayan 2004 (Niger)	228	2534	•	0.090	[0.079; 0.102]
Sangwa-Lugoma 2006 (DRC)	401	1528	-	0.262	[0.241; 0.285]
Muwonge 2010 (Angola)	2832	8842		0.320	[0.311; 0.330]
Ngoma 2010 (Tanzania)	500	10367	8	0.048	[0.044; 0.053]
Random effects model		46435		0.165	[0.098; 0.247]
Heterogeneity: I-squared=99.8%, tau-squared=0.0213, Q=3315, df=7, p<0.000	1				3 / O
3 - HPV studies					
Womack 2000 (Zimbabwe)	917	2145	-	0.428	[0.406; 0.449]
De Vuyst 2005 (Kenya)	193	453		0.426	[0.380; 0.473]
Mahmud 2012 (DRC)	169	1352	+	0.125	[0.108; 0.144]
Random effects model		3950			[0.115; 0.558]
Heterogeneity: I-squared=99.5%, tau-squared=0.048, Q=438.3, df=2, p<0.0001	(	159900		1507.100	
		1		7	
			0 0.1 0.2 0.3 0.4 0.5	0.6	

#### 6.2. In the GSP group

#### Supplementary figure 4a: Prevalence of disease (CIN2+) in the GSP group, by screening test

GSP: gold standard (colposcopy followed by directed biopsies) performed in screen positive women and only a portion of screen negative women. CIN2+: cervical intraepithelial neoplasia grade 2 or worse. N: overall number of patients. VIA: visual inspection with acetic acid. HPV: Human papillomavirus.

Study groups	Cases	N			Prevalence	95% CI
1 - VIA studies						
Megevand 1996 (South Africa)	31	2426	+		0.01	3 [0.009; 0.018]
Denny 2000 (South Africa)	86	2944	+		0.02	9 [0.023; 0.036]
Denny 2002 (South Africa)	117	2698	+		0.04	3 [0.036; 0.052]
Ibrahim 2012 (Sudan)	76	934			0.08	1 [0.065; 0.101]
Jeronimo 2014 (Uganda)	87	3146	*		0.02	8 [0.022; 0.034]
Random effects model		12148	0		0.03	5 [0.021; 0.054]
Heterogeneity: I-squared=96%, tau-squared=0.0025, Q=99.6, df=4, p<0.0001						
3 - HPV studies						
Wright 2000 (South Africa)	56	1365			0.04	1 [0.031; 0.053]
Kuhn 2000 (South Africa)	94	2861	+		0.03	3 [0.027; 0.040]
Jeromino 2014 (Uganda)	87	3146	-		0.02	8 [0.022; 0.034]
Random effects model		7372			0.03	3 [0.026; 0.040]
Heterogeneity: I-squared=62.2%, tau-squared=0.0002, Q=5.3, df=2, p=0.0708						
			-	- 1	-	
		- 8	0.0	5 0.1	0.15	

#### Supplementary figure 4b: Prevalence of disease (CIN2+) in the GSP group, by geographic region

GSP: gold standard (colposcopy followed by directed biopsies) performed in screen positive women and only a portion of screen negative women. CIN2+: cervical intraepithelial neoplasia grade 2 or worse. N: overall number of patients. VIA: visual inspection with acetic acid. HPV: Human papillomavirus.

Study groups	Cases	N			Prevalen	e	95% CI
Eastern Africa							
Jeronimo 2014 (Uganda)	87	3146	-		0.0	28 [0.022	: 0.034]
Random effects model		3146	-		0.0	28 [0.022	0.034]
Heterogeneity: I-squared=NaN%, tau-squared=0, Q=0, df=0, p=1						•	
Southern Africa							
Megevand 1996 (South Africa)	31	2426	4		0.0	13 10.009	0.018]
Denny 2000 (South Africa)	86	2944			0.0		0.036
Kuhn 2000 (South Africa)	94	2861	-4-		0.0	33 [0.027	: 0.040]
Wright 2000 (South Africa)	56	1365			0.0	The second second	: 0.053]
Denny 2002 (South Africa)	117	2698			0.0		0.0521
Ibrahim 2012 (Sudan)	76	934			0.0		0.101
Random effects model		13228			0.0	And the second second	0.053]
Heterogeneity: I-squared=95%, tau-squared=0.0022, Q=101, df=5, p<0.0001		-500000					
		i	-	- 1			
			0.0	5 0.1	0.15		

#### Supplementary figure 5: Positivity rate of VIA, VILI and HPV testing in the GSP group

GSP: gold standard (colposcopy followed by colposcopy directed biopsies) performed in screen positive women and only a portion of screen negative women. CIN2+: cervical intraepithelial neoplasia grade 2 or worse. N: overall number of patients. VIA: visual inspection with acetic acid. HPV: Human papillomavirus.

Study groups	Positive	N		Postivity rate	95% CI
1 - VIA studies					
Megevand 1996 (South Africa)	76	2426	E	0.031	[0.025; 0.039]
Denny 2000 (South Africa)	534	2944	-		[0.168; 0.196]
Denny 2002 (South Africa)	657	2698	+	0.244	[0.227; 0.260]
Ibrahim 2012 (Sudan)	71	934	*		[0.060; 0.095]
Jeronimo 2014 (Uganda)	1085	3146			[0.328; 0.362]
Random effects model		12148		0.158	[0.060; 0.290]
Heterogeneity: I-squared=99.7%, tau-squared=0.0334, Q=1268.6, df=4, µ	><0.0001				
3 - HPV studies					
Wright 2000 (South Africa)	275	1365	*	0.201	[0.180; 0.224]
Kuhn 2000 (South Africa)	634	2861	+		[0.206; 0.237]
Jeromino 2014 (Uganda)	619	3146	+		[0.183; 0.211]
Random effects model		7372			[0.191; 0.224]
Heterogeneity: I-squared=66.3%, tau-squared=0.0002, Q=5.9, df=2, p=0.	0516				•
		Γ			
		0	0.1 0.2 0.	3 0.4 0.5 0.6	

#### 7. Forest plots of absolute sensitivity and specificity for CIN2+ detection of VIA and HPV testing in the GSP group

#### Supplementary figure 6: Absolute sensitivity and specificity to detect CIN2+ in the GSP group, by screening test

Heterogeneity analysis across studies provided the following Cochran's Q p-values: in VIA studies, p=0.62 and p<0.0001 for sensitivity and specificity, respectively; and in HPV studies, p=0.13 and p=0.08 for sensitivity and specificity, respectively. GSP: gold standard (colposcopy followed by directed biopsies) performed in screen positive women and only a portion of screen negative women. CIN2+: cervical intraepithelial neoplasia grade 2 or worse. N: overall number of patients. VIA: visual inspection with acetic acid. HPV: Human papillomavirus.

Study groups	N	Sensitivity [95%CI]	N	N Specificity [95%CI]		
1 - VIA Studies						
Megevand 1996 (South Africa) — Denny 2000 (South Africa) Denny 2002 (South Africa) Ibrahim 2012 (Sudan)	31 86 117 76	0.645 [0.454;0.808] 0.674 [0.565;0.772] 0.726 [0.636;0.805] 0.645 [0.527;0.751]	2395 2858 2581 858	0.977 [0.970;0.982] 0.833 [0.819;0.847] 0.778 [0.762;0.794] 0.974 [0.961;0.984]		
Jeronimo 2014 (Uganda) Pooled estimates	87	0.736 [0.630;0.824] 0.686 [0.624;0.742], 12=0.0%	3059	0.666 [0.649;0.683] 0.898 [0.720;0.968], I2=99.4%		
3 - HPV Studies  Wright 2000 (South Africa) Kuhn 2000 (South Africa) Jeromino 2014 (Uganda) Pooled estimates	56 94 87	0.839 [0.717;0.924] 0.883 [0.800;0.940] 0.770 [0.668;0.854] 0.830 [0.742;0.893], 12=50.3%	1309 2767 3059	0.826 [0.804;0.846] 0.801 [0.785;0.816] 0.820 [0.805;0.833] 0.814 [0.799;0.828], 12=59.6%		
0.40	0.60 0.80 1.00 Sensitivity	0.40 0.60 0.80 specificity	1.00			

# 8. Pooled positive predictive value (PPV) and negative predictive value (NPV) of VIA, VILI and HPV testing in sub-Saharan Africa, by geographic region Supplementary table 3: Estimated predictive values of screening tests to detect CIN2+ in sub-Saharan Africa, by geographic region

		Positive predictive Value (95% CI)			Negative predictive Value (95% CI)			
Geographical Area	Pooled prevalence	VIA	VILI	HPV testing	VIA	VILI	HPV testing	
Western Africa	2.0%	11.7% (6.5 to 20.8)	13.1% (7.8 to 21.7)	6.4% (3.4 to 13.6)	99.6% (99.4 to 99.7)	99.9% (99.7 to 99.9)	99.7% (99.2 to 99.9)	
Middle Africa	2.5%	14.3% (8.0 to 24.8)	15.9% (9.6 to 25.8)	8.0% (4.2 to 16.5)	99.5% (99.2 to 99.6)	99.9% (99.7 to 99.9)	99.6% (99.0 to 99.8)	
Southern Africa	3.7%	20.0% (11.5 to 33.1)	22.1% (13.8 to 34.3)	11.5% (6.2 to 22.9)	99.2% (98.9 to 99.5)	99.8% (99.5 to 99.9)	99.4% (98.5 to 99.8)	
Eastern Africa	6.2%	30.1% (18.3 to 46.0)	32.8% (21.6 to 47.3)	18.2% (10.2 to 33.8)	98.7% (98.1 to 99.1)	99.6% (99.2 to 99.8)	99.0% (97.5 to 99.6)	

We included all selected studies in this analysis, as the prevalence of disease was not different between the GSA and GSP groups, by screening test. VIA: visual inspection with acetic acid. VILI: visual inspection with Lugol's iodine. HPV: Human papillomavirus. CIN2+: cervical intraepithelial neoplasia grade 2 or worse. CI: confidence interval.

#### References

- 1. Untiet S, Vassilakos P, McCarey C, et al. HPV self-sampling as primary screening test in sub-Saharan Africa: Implication for a triaging strategy. *Int J Cancer* 2014; **135**(8): 1911–7.
- 2. Dartell M, Rasch V, Iftner T, et al. Performance of visual inspection with acetic acid and human papillomavirus testing for detection of high-grade cervical lesions in HIV positive and HIV negative Tanzanian women. *Int J Cancer* 2014; **135**(4): 896–904.
- 3. Ajenifuja K, Gage J, Adepiti A, et al. A population-based study of visual inspection with acetic acid (VIA) for cervical screening in rural Nigeria. *Int J Gynecol Cancer* 2013; **23**(3): 507–12.
- 4. Ugwu E, Obi S, Ezechukwu P, Okafor II, Ugwu AO. Acceptability of human papilloma virus vaccine and cervical cancer screening among female health-care workers in Enugu, Southeast Nigeria. *Niger J Clin Pract* 2013; **16**(2): 249–52.
- 5. Ogilvie G, Mitchell S, Sekikubo M, et al. Results of a community-based cervical cancer screening pilot project using human papillomavirus self-sampling in Kampala, Uganda. *Int J Gynaecol Obstet* 2013; **122**(2): 118–23.
- 6. Firnhaber C, Mayisela N, Mao L, et al. Validation of cervical cancer screening methods in HIV positive women from Johannesburg South Africa. *PLoS One* 2013; **8**(1): e53494.
- 7. Mwanahamuntu M, Sahasrabuddhe V, Blevins M, et al. Utilization of cervical cancer screening services and trends in screening positivity rates in a 'screen-and-treat' program integrated with HIV/AIDS care in Zambia. *PLoS One* 2013; **8**(9): e74607.
- 8. De Vuyst H, Chung M, Baussano I, et al. Comparison of HPV DNA testing in cervical exfoliated cells and tissue biopsies among HIV-positive women in Kenya. *Int J Cancer* 2013; **133**(6): 1441–6.
- 9. Chigbu CO, Onyebuchi A, Ajah L, Onwudiwe E. Motivations and preferences of rural Nigerian women undergoing cervical cancer screening via visual inspection with acetic acid. *Int J Gynaecol Obstet* 2013; **120**(3): 262–5.
- 10. Busingye P, Nakimuli A, Nabunya E, Mutyaba T. Acceptability of cervical cancer screening via visual inspection with acetic acid or Lugol's iodine at Mulago Hospital, Uganda. *Int J Gynaecol Obstet* 2012; **119**(3): 262–5.
- 11. Denny L. Cervical cancer prevention: new opportunities for primary and secondary prevention in the 21st century. *Int J Gynaecol Obstet* 2012; **119 Suppl 1**: S80–4.
- 12. Moon T, Silva-Matos C, Cordoso A, Baptista A, Sidat M, Vermund S. Implementation of cervical cancer screening using visual inspection with acetic acid in rural Mozambique: successes and challenges using HIV care and treatment programme investments in Zambezia Province. *J Int AIDS Soc* 2012; **15**(2): 17406.
- 13. Horo A, Jaquet A, Ekouevi D, et al. Cervical cancer screening by visual inspection in Cote d'Ivoire, operational and clinical aspects according to HIV status. *BMC Public Health* 2012; **12**: 237.
- 14. Mingo A, Panozzo C, DiAngi Y, et al. Cervical cancer awareness and screening in Botswana. *Int J Gynecol Cancer* 2012; **22**(4): 638–44.
- 15. Audet C, Silva-Matos C, Blevins M, Cardoso A, Moon T, Sidat M. Acceptability of cervical cancer screening in rural Mozambique. *Health Educ Res* 2012; **27**(3): 544–51.
- 16. Jemal A, Bray F, Forman D, et al. Cancer burden in Africa and opportunities for prevention. *Cancer* 2012; **118**(18): 4372–84.
- 17. Gage J, Ajenifuja K, Wentzensen N, et al. Effectiveness of a simple rapid human papillomavirus DNA test in rural Nigeria. *Int J Cancer* 2012; **131**(12): 2903–9.
- 18. Ramogola-Masire D, de Klerk R, Monare B, Ratshaa B, Friedman H, Zetola. Cervical cancer prevention in HIV-infected women using the "see and treat" approach in Botswana. *J Acquir Immune Defic Syndr* 2012; **59**(3): 308–13.
- 19. Kahesa C, Kjaer S, Mwaiselage J, et al. Determinants of acceptance of cervical cancer screening in Dar es Salaam, Tanzania. *BMC Public Health* 2012; **12**: 1093.

- 20. Teguete I, Muwonge R, Traore C, Dolo A, Bayo S, Sankaranarayanan R. Can visual cervical screening be sustained in routine health services? Experience from Mali, Africa. *BJOG* 2012; **119**(2): 220–6.
- 21. Cronje H. Cervical screening strategies in resourced and resource-constrained countries. *Best Pract Res Clin Obstet Gynaecol* 2011; **25**(5): 575–84.
- 22. Awodele O, Adeyomoye A, Awodele D, Kwashi V, Awodele I, Dolapo D. A study on cervical cancer screening amongst nurses in Lagos University Teaching Hospital, Lagos, Nigeria. *J Cancer Educ* 2011; **26**(3): 497–504.
- 23. Balandya B, Pembe A, Mwakyoma H. Cervical pre-malignant lesions in HIV infected women attending Care and Treatment Centre in a tertiary hospital, Dar es Salaam, Tanzania. *East Afr J Public Health* 2011; **8**(3): 185–9.
- 24. Lewis K, Tsu V, Dawa A, Kidula N, Chami I, Sellors J. A comparison of triage methods for Kenyan women who screen positive for cervical intraepithelial neoplasia by visual inspection of the cervix with acetic acid. *Afr Health Sci* 2011; **11**(3): 362–9.
- 25. Saleh F. Cervical cancer: a missed health priority in Tanzania. *East Afr J Public Health* 2011; **8**(4): 247–9.
- 26. Kuhn L, Wang C, Tsai W, Wright T, Denny L. Efficacy of human papillomavirus-based screenand-treat for cervical cancer prevention among HIV-infected women. *AIDS* 2010; **24**(16): 2553–61.
- 27. Hovland S, Arbyn M, Lie A, et al. A comprehensive evaluation of the accuracy of cervical precancer detection methods in a high-risk area in East Congo. *Br J Cancer* 2010; **102**(6): 957–65.
- 28. Were E, Nyaberi Z, Buziba N. Integrating cervical cancer and genital tract infection screening into mother, child health and family planning clinics in Eldoret, Kenya. *Afr Health Sci* 2010; **10**(1): 58–65.
- 29. Batra P, Kuhn L, Denny L. Utilisation and outcomes of cervical cancer prevention services among HIV-infected women in Cape Town. *S Afr Med J* 2010; **100**(1): 39–44.
- 30. Peters L, Soliman A, Bukori P, Mkuchu J, Ngoma T. Evidence for the need of educational programs for cervical screening in rural Tanzania. *J Cancer Educ* 2010; **25**(2): 153–9.
- 31. Denny L, Kuhn L, Hu C, Tsai W, Wright T. Human papillomavirus-based cervical cancer prevention: long-term results of a randomized screening trial. *J Natl Cancer Inst* 2010; **102**(20): 1557–67.
- 32. Koffi B, Serdouma E, Mbolissa-Nguerekoudou W, et al. Cervical dysplasia in women with human immunodeficiency virus (HIV) in areas of high prevalence of infection. *Gynecol Obstet Fertil* 2010; **38**(10): 576–80.
- 33. Qureshi S, Das V, Zahra F. Evaluation of visual inspection with acetic acid and Lugol's iodine as cervical cancer screening tools in a low-resource setting. *Trop Doct* 2010; **40**(1): 9–12.
- 34. Hassan F, Khirelseed M. Cervical cancer screening among Sudanese women. *Gulf J Oncolog* 2009; (6): 28–34.
- 35. Hoque M, Ibekwe C, Ntuli-Ngcobo B. Screening and perceived severity of cervical cancer among women attending Mahalapye District Hospital, Botswana. *Asian Pac J Cancer Prev* 2009; **10**(6): 1095–100.
- 36. Akinwuntan A, Adesina O, Okolo C, et al. Correlation of cervical cytology and visual inspection with acetic acid in HIV-positive women. *J Obstet Gynaecol* 2008; **28**(6): 638–41.
- 37. Anorlu R. Cervical cancer: the sub-Saharan African perspective. *Reprod Health Matters* 2008; **16**(32): 41–9.
- 38. Kawonga M, Fonn S. Achieving effective cervical screening coverage in South Africa through human resources and health systems development. *Reprod Health Matters* 2008; **16**(32): 32–40
- 39. Hoque M, Hoque E, Kader S. Evaluation of cervical cancer screening program at a rural community of South Africa. *East Afr J Public Health* 2008; **5**(2): 111–6.

- 40. Arbyn M, Sankaranarayanan R, Muwonge R, et al. Pooled analysis of the accuracy of five cervical cancer screening tests assessed in eleven studies in Africa and India. *Int J Cancer* 2008; **123**(1): 153–60.
- 41. Kamate B, Traore CB, Diallo D, et al. Extension of cervix cancer screening by visual methods to the community health centres in the district of Bamako. *Mali Med* 2008; **23**(4): 29–33.
- 42. Roblyer D, Richards-Kortum R, Park S, Adewole I, Follen M. Objective screening for cervical cancer in developing nations: lessons from Nigeria. *Gynecol Oncol* 2007; **107**(1 Suppl 1): S94–7.
- 43. Akinola O, Fabamwo A, Oshodi Y, et al. Efficacy of visual inspection of the cervix using acetic acid in cervical cancer screening: a comparison with cervical cytology. *J Obstet Gynaecol* 2007; **27**(7): 703–5.
- 44. Anorlu R, Ola E, Abudu O. Low cost methods for secondary prevention of cervical cancer in developing countries. *Niger Postgrad Med J* 2007; **14**(3): 242–6.
- 45. Kamal M, Sapkal R, Sarodey C, et al. Comparative study of four candidate strategies to detect cervical cancer in different health care settings. *J Obstet Gynaecol Res* 2007; **33**(4): 480–9.
- 46. Cronje H, Beyer E. Screening for cervical cancer in an African setting. *Int J Gynaecol Obstet* 2007; **98**(2): 168–71.
- 47. Sodhani P, Gupta S, Sharma J, et al. Test characteristics of various screening modalities for cervical cancer: a feasibility study to develop an alternative strategy for resource-limited settings. *Cytopathology* 2006; **17**(6): 348–52.
- 48. Cronje H. Screening for cervical cancer in the developing world. *Best Pract Res Clin Obstet Gynaecol* 2005; **19**(4): 517–29.
- 49. Denny L. The prevention of cervical cancer in developing countries. *BJOG* 2005; **112**(9): 1204–12.
- 50. Denny L, Wright T. Human papillomavirus testing and screening. *Best Pract Res Clin Obstet Gynaecol* 2005; **19**(4): 501–15.
- 51. Doh A, Nkele N, Achu P, Essimbi F, Essame O, Nkegoum B. Visual inspection with acetic acid and cytology as screening methods for cervical lesions in Cameroon. *Int J Gynaecol Obstet* 2005; **89**(2): 167–73.
- 52. Millogo F, Akotionga M, Lankoande J. Cervix cancer screening in a health district (Burkina Faso) by voluntary biopsies after the application of acetic acid and lugol. *Bull Soc Pathol Exot* 2004; **97**(2): 135–8.
- 53. El-Shalakany A, Hassan S, Ammar E, Ibrahim M, Salam M, Farid M. Direct visual inspection of the cervix for the detection of premalignant lesions. *J Low Genit Tract Dis* 2004; **8**(1): 16–20.
- 54. Bhatla N, Mukhopadhyay A, Joshi S, et al. Visual inspection for cervical cancer screening: evaluation by doctor versus paramedical worker. *Indian J Cancer* 2004; **41**(1): 32–6.
- 55. Okewole I, Fawole A, Omigbodun A, Adewole I. Does screening for cervical intra-epithelial neoplasm in developing countries prevent invasive cervical cancer? *Afr J Med Med Sci* 2003; **32**(3): 283–5.
- 56. Petry K, Scholz U, Hollwitz B, Von Wasielewski R, Meijer C. Human papillomavirus, coinfection with Schistosoma hematobium, and cervical neoplasia in rural Tanzania. *Int J Gynecol Cancer* 2003; **13**(4): 505–9.
- 57. Hawes S, Critchlow C, Faye Niang M, et al. Increased risk of high-grade cervical squamous intraepithelial lesions and invasive cervical cancer among African women with human immunodeficiency virus type 1 and 2 infections. *J Infect Dis* 2003; **188**(4): 555–63.
- 58. Claeys P, De Vuyst H, Gonzalez C, Garcia A, Bello R, Temmerman M. Performance of the acetic acid test when used in field conditions as a screening test for cervical cancer. *Trop Med Int Health* 2003; **8**(8): 704–9.
- 59. Cronje H, Parham G, Cooreman B, de Beer A, Divall P, Bam R. A comparison of four screening methods for cervical neoplasia in a developing country. *Am J Obstet Gynecol* 2003; **188**(2): 395–400.

- 60. Adanu R. Cervical cancer knowledge and screening in Accra, Ghana. *J Womens Health Gend Based Med* 2002; **11**(6): 487–8.
- 61. Blumenthal P, Gaffikin L, Chirenje Z, McGrath J, Womack S, Shah K. Adjunctive testing for cervical cancer in low resource settings with visual inspection, HPV, and the Pap smear. *Int J Gynaecol Obstet* 2001; **72**(1): 47–53.
- 62. Cronje H, Cooreman B, Beyer E, Bam R, Middlecote B, Divall P. Screening for cervical neoplasia in a developing country utilizing cytology, cervicography and the acetic acid test. *Int J Gynaecol Obstet* 2001; **72**(2): 151–7.
- 63. Singh V, Sehgal A, Parashari A, Sodhani P, Satyanarayana L. Early detection of cervical cancer through acetic acid application-an aided visual inspection. *Singapore Med J* 2001; **42**(8): 351–4.
- 64. Cronje H, van Rensburg E, Cooreman B, Niemand I, Beyer E. Speculoscopy vs. the acetic acid test for cervical neoplasia. *Int J Gynaecol Obstet* 2000; **69**(3): 249–53.
- 65. Denny L, Kuhn L, Risi L, et al. Two-stage cervical cancer screening: an alternative for resource-poor settings. *Am J Obstet Gynecol* 2000; **183**(2): 383–8.
- 66. Womack S, Chirenje Z, Gaffikin L, et al. HPV-based cervical cancer screening in a population at high risk for HIV infection. *Int J Cancer* 2000; **85**(2): 206–10.
- 67. Chirenje Z, Chipato T, Kasule J, Rusakaniko S. Visual inspection of the cervix as a primary means of cervical cancer screening: results of a pilot study. *Cent Afr J Med* 1999; **45**(2): 30–3.
- 68. Thistle P, Chirenje Z. Cervical cancer screening in a rural population of Zimbabwe. *Cent Afr J Med* 1997; **43**(9): 246–51.
- 69. Sitas F, Carrara H, Terblanche M, Madhoo J. Screening for cancer of the cervix in South Africa. *S Afr Med J* 1997; **87**(5): 620–2.
- 70. Megevand E, Denny L, Dehaeck K, Soeters R, Bloch B. Acetic acid visualization of the cervix: an alternative to cytologic screening. *Obstet Gynecol* 1996; **88**(3): 383–6.
- 71. Nolting W, Moodley J, Gouws E. Naked eye screening for cervical intra-epithelial abnormalities: a preliminary report. *Trop Doct* 1995; **25**(3): 130–1.
- 72. Moher D, Liberati A, Tetzlaff J, Altman D. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. *J Clin Epidemiol* 2009; **62**(10): 1006–12.
- 73. Composition of macro geographical (continental) regions, geographical sub-regions, and selected economic and other groupings.

  https://unstats.un.org/unsd/methods/m49/m49regnf.htm (accessed July 7, 2014).
- 74. Megevand E, Van Wyk W, Knight B, Bloch B. Can cervical cancer be prevented by a see, screen, and treat program? A pilot study. *Am J Obstet Gynecol* 1996; **174**(3): 923–8.
- 75. University of Zimbabwe. Visual inspection with acetic acid for cervical-cancer screening: test qualities in a primary-care setting. University of Zimbabwe/JHPIEGO Cervical Cancer Project. *Lancet* 1999; **353**(9156): 869–73.
- 76. Womack S, Chirenje Z, Blumenthal P, et al. Evaluation of a human papillomavirus assay in cervical screening in Zimbabwe. *BJOG* 2000; **107**(1): 33–8.
- 77. Wright T, Denny L, Kuhn L, Pollack A, Lorincz A. HPV DNA testing of self-collected vaginal samples compared with cytologic screening to detect cervical cancer. *JAMA* 2000; **283**(1): 81–6.
- 78. Kuhn L, Denny L, Pollack A, Lorincz A, Richart R, Wright T. Human papillomavirus DNA testing for cervical cancer screening in low-resource settings. *J Natl Cancer Inst* 2000; **92**(10): 818–25.
- 79. Denny L, Kuhn L, Pollack A, Wainwright H, Wright T. Evaluation of alternative methods of cervical cancer screening for resource-poor settings. *Cancer* 2000; **89**(4): 826–33.
- 80. Denny L, Kuhn L, Pollack A, Wright T. Direct visual inspection for cervical cancer screening: an analysis of factors influencing test performance. *Cancer* 2002; **94**(6): 1699–707.
- 81. Sankaranarayanan R, Chatterji R, Shastri S, et al. Accuracy of human papillomavirus testing in primary screening of cervical neoplasia: results from a multicenter study in India. *Int J Cancer* 2004; **112**(2): 341–7.

- 82. De Vuyst H, Claeys P, Njiru S, et al. Comparison of pap smear, visual inspection with acetic acid, human papillomavirus DNA-PCR testing and cervicography. *Int J Gynaecol Obstet* 2005; **89**(2): 120–6.
- 83. Sangwa-Lugoma G, Mahmud S, Nasr S, et al. Visual inspection as a cervical cancer screening method in a primary health care setting in Africa. *Int J Cancer* 2006; **119**(6): 1389–95.
- 84. Muwonge R, Manuel Mda G, Filipe A, Dumas J, Frank M, Sankaranarayanan R. Visual screening for early detection of cervical neoplasia in Angola. *Int J Gynaecol Obstet* 2010; **111**(1): 68–72.
- 85. Ngoma T, Muwonge R, Mwaiselage J, Kawegere J, Bukori P, Sankaranarayanan R. Evaluation of cervical visual inspection screening in Dar es Salaam, Tanzania. *Int J Gynaecol Obstet* 2010; **109**(2): 100–4.
- 86. Ibrahim A, Aro A, Rasch V, Pukkala E. Cervical cancer screening in primary health care setting in Sudan: a comparative study of visual inspection with acetic acid and Pap smear. *Int J Womens Health* 2012; **4**: 67–73.
- 87. Mahmud S, Sangwa-Lugoma G, Nasr S, et al. Comparison of human papillomavirus testing and cytology for cervical cancer screening in a primary health care setting in the Democratic Republic of the Congo. *Gynecol Oncol* 2012; **124**(2): 286–91.
- 88. Jeronimo J, Bansil P, Lim J, et al. A multicountry evaluation of careHPV testing, visual inspection with acetic acid, and papanicolaou testing for the detection of cervical cancer. *Int J Gynecol Cancer* 2014; **24**(3): 576–85.
- 89. Whiting P, Rutjes A, Westwood M, et al. QUADAS-2: a revised tool for the quality assessment of diagnostic accuracy studies. *Ann Intern Med* 2011; **155**(8): 529–36.