Supplementary Information

Characterization of the *Plasmodium* Interspersed Repeats (PIR) proteins of *Plasmodium chabaudi* indicates functional diversity

Xue Yan Yam¹⁺, Thibaut Brugat²⁺, Anthony Siau¹, Jennifer Lawton², Daniel. S.Wong¹, Abdirahman Farah^{1,3}, Jing Shun Twang¹, Xiaohong Gao¹, Jean Langhorne^{2*}, Peter R. Preiser^{1*}

¹: School of Biological Sciences, Nanyang Technological University, 637551, Singapore

²: Francis Crick Institute, Mill Hill Laboratory, London, UK

³: Karolinska Institutet, Stockholm, Sweden

Supplementary Data

Α

	Gene I.D.	CIR subfamily classification	Region of pro expressed (a	tein .a)	Theoret	ical M.W.	Theoretical p.l.	
	PCHAS_000730	S	24-238		2	26	5.77	
	PCHAS_140090	S	51-235		2	23	5.3	
	PCHAS_110020	S	23-233		2	26	7.6	
	PCHAS_000100	S	1-232		2	28	5.46	
	PCHAS_000950	L	54-306		3	80	6.03	
	PCHAS_040110	L	1-269		3	32	6.21	
в								
		Frequency in subfamilies						
	Mot	tifs	L	_	S	Peptide	s synthesised	
CIR-L	^a <u>K</u> KKřůKř		93.44		0.00	KNMKKVINLAYGK		
CIR-S	NRLOR OVLRE K		0.00	8	80.72	QYLREK	RKKAKRKVYNY	
C Variable region TM								
Conserved C-terminus								
Recombinar proteins	_;;							
CIR-L								
CIR-S								

Fig S1: Production of recombinant CIR proteins and subfamily-specific peptides

A) Table summarizing the characteristics of the His-tagged recombinant proteins expressed for this study. The gene I.D., CIR-subfamily, region of the protein expressed and the theoretical molecular weight (M.W.) and isoelectric point (p.I.) of the recombinant proteins are indicated. **B**) Conserved amino-acid motifs within the CIR repertoire were identified by MEME analysis by

Lawton et al [19] and shown here as Weblogo images. Blue: hydrophobic residues; Green: polar, non-charged residues; Purple: acidic residues; and Red: positively charged residues. The percentage of CIRs containing each motif within subfamilies L and S and the sequence of the peptides synthesized for antibody production are shown. C) Position of the regions selected for protein and peptide production on a typical CIR protein. TM: transmembrane domain (shown in grey).

PCHAS 000730 57	MLAFIEFDNILTEYCTEEEGGGKKECESYEVI. 32	,
PCHAS 110020 S7	MKOLFHYNTGSYPYCINGEGG-TOECESYEVI 31	
PCHAS 140090 57	SATTA 11	
PCHAS 000100 S7	MEDBSYKTEDVYNDTEKTSNYEDEDENDGTKTKYTNKATHNYCHYDDROEKDVCNDYYEM 60	
PCHAS_000050_12	MCDEDIRIEDVIRDIERIDRIEDEDERDOIRIRVIRRAIMICHIDDRQERDRCRDIIER 00	
DCHAS_000330_12_	MSKELCOCTNEN DURAUEDOUSESVUENDDI EVUVCENDICESNCLV 42	
FCHA5_040110_54_	MSRELCQGINERDANVYEDQASESIAENDIERVICENAACDSASEA 4/	
DCU1C 000720 C7		
PCHA5_000/30_5/_	ISSIFIKEEKIPMNEDD-LENDKRAVIAIEWFCRKENKNSQNGINNENDVINKIIKGNK 90	
PCHAS_110020_57_	VSSLFIKMVNHFMKGDDLKNNKLALIAILWLCIKLNKKKENEISNLNDFHNKIIKGNE 69	
PCHAS_140090_57_	VLNIFKNISVADGDDLEDDKIACIAILWLNRKLNQIFLIGVSNLDDFRNIIIKD 65	0
PCHAS_000100_57_	1556VIHLINNLKGKNVLDYDKLALIAILWLSYKLKIKENFIIKKLSVFYDSYIERNE 11	8
PCHAS_000950_L2_	LQQLFVNNEGDLELENKNDETSETAILWESNIIRHFNYKHIPHVQDFYIIFIKE 62	
PCHAS_040110_L4_	IGSAFTALLEYFKNIDNENSEDDKLIQTAILWFNSKLSQKPDIETERYTMYN 99	1
PCHAS 000730 57	FFFEKINDAGAYNSYFFFINKKODMUNUNIKDMSKEYFALOILCKMYTKIDESKSNCTKC 15	0
PCHAS 110020 S7	NHIDKINGAGSYNSYKNIINNYKYINTIDIKEMSKEYUDIKSICKIYTEYSKKKKNYTTC 14	0
PCHAS 140090 S7		3
PCHAS 000100 S7	VYNUNTNODEI TYVETTDVVVDI MOMDINETEVI FADENTI VYI VHOTVDVNSECSEV 17	6
PCHAS_000950_12	CNWVNDED VINVVNDIMUI OINOI VNI VEI I NII CVA ITVVDENDSNSSEC 11	4
PCHAS_000330_12_		4
FCHA5_040110_14_	Ibryinwroeboroierradindindindibradioeilinaa-onoribec 15	4
PCHAS 000730 57	LDDAOKFADEY-EKINSNS-ITGNDSYSOMLSNLEVDYDNLKNKCKNSSSLETIKKPO 20	6
PCHAS 110020 57	SODAODFAKHF-EDLNODSSITENISYBELLSSISIDYDDEKNECA-KNCSYCNDIPTIS 20	7
PCHAS 140090 S7	LEDSOKEVDEY-EKLNENY-DIKCUSYKKILSTLENDYDELRKKCNNEOSEPSMKITO 17	a
PCHAS 000100 S7	SDYANOFVOKE-FOLNNDPNNKEDSSYNKLISTISDDYNNLOKKCTNEPSLPVYPR 23	1
PCHAS 000950 L2	SNFANKWEERSKELVNKETKVEEDEHYCNALLNLKKAVEKETNGNNTNAEDKINETEKMN 17	4
PCHAS 040110 L4	IFSAKKCSGLYBACTI.HLDWDFTCNDYCNVLTNLKNDYDKTBENNBDKKLDFLTLDEGLY 21	4
PCHAD_040110_D4_	· · · · · · · · · · · · · · · · · · ·	-
PCHAS 000730 S7	NFEORFE 21	3
PCHAS 110020 S7	EIK 21	0
PCHAS 140090 S7	SYV0D 18	4
PCHAS 000100 S7		-
PCHAS 000950 L2	NCKKLCEKANRSWKVTHVEVKDVVDAKKVSENGESKKEGEEKTKESNPVETPGPVATLSN 23	4
PCHAS 040110 L4	DCKTECYKOFEGHKARVAALORSSDGSELDTTPG24	8
		-
PCHAS_000730_S7_	VTSLEHHHHHH 224	
PCHAS_110020_S7_	ТРЕЕННИНИ 220	
PCHAS_140090_S7_	SDLEHHHHHH 194	
PCHAS_000100_57_		
PCHAS_000950_L2_	TGDTSQNDAIKVRADESGENLEHHHHHH 262	
PCHAS_040110_L4_	ISLLDLSVPPTSINNGNKLPYIHHHHHH 276	

Fig S2: Alignment of the 6 His-tagged recombinant proteins

Alignment of protein sequences was performed using ClustalW. Colours determine the different amino acids groups. Highlighted box showed the highly conserved peptide motif.



Fig S3: Production of anti-CIR antibodies

A) Western blot experiments were performed to determine the specificity of CIR-specific polyclonal antibodies. The name of the recombinant proteins loaded on the SDS PAGE gel and antibodies used are indicated above, and the molecular weight on the side. Anti-His monoclonal antibody and rat pre-immune (P.I.) serum were used as loading and negative control respectively.
B) The titres of anti-peptide antibodies in rabbit sera before and after immunization ("Before Imm." and "After Imm." respectively) were determined by ELISA assays. Bars represent antibody titres in arbitrary unit (A.U.). C) Western blot experiments were performed to determine the specificity of anti-peptide antibodies in rabbit sera before ("pre-immune serum") and after immunization (CIR-L and CIR-S). Asterisks indicate the differences observed between

the anti-CIR-L and anti-CIR-S antisera. Anti-Ter119 (specific to red blood cells membrane) and anti-Aldolase (soluble protein) antibodies were used as loading controls. P.L.: parasite lysate; uRBC: uninfected red blood cells membranes; M.W.: molecular weight. **D**) IFA was performed on transfected CHO cells expressing PCHAS_040110 proteins. Ratio of Mean fluorescence intensity (MFI) of pooled anti-CIR antibodies/anti-C-myc antibodies was measured using fluorescence plate reader. Bars represent the mean \pm S.E.M. of three independent experiments. (T-test, ***: *P*<0.001).



Pooled anti- CIR



Fig S4: Immunofluorescence experiments using CIR-specific polyclonal antibodies Representative pictures of immunofluorescence assay on acetone/methanol fixed red blood cells (RBC) infected with *P.chabaudi* AS. Parasites nuclei were labeled with Hoechst 33342 (blue) and the mouse erythrocyte membranes with anti-Ter119 antibody (green). CIR proteins were labeled with either a single CIR-specific polyclonal antibodies or a pool of polyclonal anti-CIR antibodies (red). Scale bars represent 5 μm.



Fig S5: Live imaging of *P.chabaudi* parasites expressing CIRs-GFP tagged

Live cell imaging of transgenic *Plasmodium chabaudi* parasites expressing CIR proteins tagged with GFP (green). Representative pictures of the trophozoite (upper panel) and schizont (lower

panel) stages of each CIR member are shown. The subfamilies and gene accession number of the each CIR member are indicated along the side of both panels. Parasites nuclei were labeled with Hoechst 33342 (blue). The phase contrast image is shown in grey. Scale bars represent 5µm.



Fig S6: Comparison of cir transcriptional profile with CIR localization

A) The transcriptional graph of each *cir* gene is plotted using the raw microarray data from
Lawton *et al.*, *BMC genomics* 2012, with mRNA abundance against hours post parasite invasion.
First panel represent genes which are early transcribed (Ring/early trophozoites (ET) stage),

second panel are mid to late-trophozoites transcribed and last panel are late parasite stage transcribed. **B**) Representative pictures of immunofluorescence assay on acetone/methanol fixed red blood cells (RBC) infected with *P.chabaudi* AS. Parasites nuclei were labeled with Hoechst 33342 (blue) and the mouse erythrocyte membranes with an anti-Ter119 antibody (green). CIR proteins were labeled with individual polyclonal anti-CIR antibodies (red) in trophozoite (T) or schizont (S). First panel represent CIR member that is transcribed in early parasite stage (Ring/early trophozoites (ET)); second panel represent CIR member transcribed in trophozoite stage and last panel represent CIR members transcribed in the late parasite stage. **C**) Representative pictures of the trophozoite (T) and schizont (S) stages of each GFP-tagged CIR member are shown. Parasites nuclei were labeled with Hoechst 33342 (blue). The subfamilies and gene accession number of the each CIR member are indicated along the side of the pictures. First panel corresponds to the early transcribed genes; second panel to the mid to late trophozoites and last panel to the late parasite transcribed genes respectively to their transcription data as shown in (A). Scale bar represent 5 µm.



Fig S7: CIR proteins are recognized by the host immune system during infection

ELISA assays were performed to quantify the amount of CIR-specific Ig antibodies present in the plasma of C57BL/6 mice before (0) or after 1, 2 or 4 infections with *P. chabaudi* (n=10 mice per group). The name of the recombinant proteins used is indicated below (see also Fig S1A and S2). MSP1₂₁ recombinant protein was used as a positive control. Columns represent the median with interquartile range. Bars represent minimum and maximum values. (Mann-Whitney; *: P<0.05; **: P<0.01; ***: P<0.001)