

# **Electronic Supplementary Material**

**miR-20a regulates expression of the iron exporter ferroportin in lung cancer**

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## SUPPLEMENTARY MATERIAL:

Name	5' Sequence 3'
hs-ACTB_F	TTCCGCTGCCCTGAGGCACCTCT
hs-ACTB_R	TCTGCTGGAAGGTGGACAGCGA
hs-pre-ACTB_F	CTATTCTCGCAGCTCACCATG
hs-pre-ACTB_R	CAGCTCCCCTACCTGGTG
hs-FPN_F	TGCTGTTGCAGGCGTCATT
hs-FPN_R	TTGCAGCAACTGTGTCACAGTT
hs-pre-FPN_F	TACCCATTGGGAAGGGGAAT
hs-pre-FPN_R	CCAGGGTTTGGCTCAGTA
hs-HIF1A_F	CCCCAGATTCAAGGATCAGACA
hs-HIF1A_R	GGGACTATTAGGCTCAGGTGAACCT
hs-RPL19_F	TCGCCTCTAGTGTCCCTCCG
hs-RPL19_R	GCGGGCCAAGGTGTTTTC
hs-rnU6	GTGCTCGCTTCGGCAGCACATATAC
hs-miR-20a-5p	TAAAGTGCTTATAAGTGCAGGTAGG
hs-miR-21-5p	TAGCTTATCAGACTGATGTTGAGG

**Table S1: Primers used for qPCR analysis of mRNA and miRNA transcripts**

Name	5' Sequence 3'
h-FPN-F	AAAGCTAGCATGACCAGGGCGGGAG
h-FPN-R	AAACTCGAGAACAAACAGATGTATTGCTTGATT
Flag_F	[Phos]AATTCTAGACTACAAGGACGACGATGACAAGTGAGC
Flag_R	[Phos]GGCCGCTCACTGTCATCGTCGCTTGTAGTCTAG

**Table S2: Primers used to generate FPN expression constructs.** Sequences with 5'phosphate represented as [Phos] were annealed using oligo annealing buffer followed by direct ligating into the vector.

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Name	5' Sequence 3'
hs-FPN_F	GAGAGCTCGGAAGGAAAATCAAGCAAATACA
hs-FPN_R	GAGCTAGCTGGGAACAGAACATTCAAGGA
hs-HIF1A_F	GAGAGCTCACTCAGAGCTTGGATCAAGTT
hs-HIF1A_R	GAGCTAGGCCCTGGTCCACAGAACAGATGT
hs-RPL19_F	[Phos]CACACCTCCCACTTGTCTGTACATACTGGCCTCTGTGATTACA TAGATCAGCCATTAAAATAAAACAAGCCTTAATCTGCG
hs-RPL19_R	[Phos]CTAGCGCAGATTAAGGCTTGTGTTATTTAATGGCTGATCTAT GTAATCACAGAGGCCAGTATGTACAGACAAAGTGGAGGTTGAGC T
hs-miR-20a-5p(+)_F	[Phos]CCTACCTGCACATAAGCACTTTAG
hs-miR-20a-5p(+)_R	[Phos]CTAGCTAAAGTGCTTATAGTCAGGTAGGAGCT
hs-miR-20a-5p(-)_F	[Phos]CTAAAGTGCTTATAGTCAGGTAGGAGCT
hs-miR-20a-5p(-)_R	[Phos]CTAGCCTACCTGCACATAAGCACTTTAGAGCT

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**Table S3: Primers used for the amplification of 3'-UTR sequences.** Sequences with 5'phosphate represented as [Phos] were annealed using oligo annealing buffer followed by direct ligating into the vector.

Name	5' Sequence 3'
hs-FPN-MT1_F	TCAGTTGCAACATGTCTGTACCAAGATGGTAGAATGCCTTAACCG TTTATA
hs-FPN-MT1_R	TATAAACGGTTAACCGATTCTACCATCTGGTACAGACATGTTGCA AACTGA
hs-FPN-MT2_F	GATGGTAGAATGCCTAACCGTTATATGCAGAACATGGAGACTG CAATAC
hs-FPN-MT2_R	GTATTGCAGTCTCCATGATTCTGCATATAAACGGTTAACGCATTCT ACCATC
hs-FPN-MT3_F	GGAGACTGCAATACGTTGCTATGAGCAGAACATTTATCCTGGAGT TTAAT
hs-FPN-MT3_R	ATTAAAATCCAAGGATAAAAGATTCTGCTCATAGAACGTATTGCAG TCTCC
hs-HIF1A-MT1_F	TGGTATTAAACCATTGCATTGCAGTAGCATCAGAACAAAAAATGC ACCTTTTATTATTATTATTGG
hs-HIF1A-MT1_R	CCAAAAATAAATAAATAAAAGGTGCATTTTATTCTGATGCTAC TGCAATGCAATGGTTAAATACCA
hs-HIF1A-MT2_F	TACTGCTGGCAAAGCATTATTATTATGTATTCTGTGAAAAAAAAG GTGTAAAAATTTCAACTGCCT
hs-HIF1A-MT2_R	AGGCAGTTGAAAAATTTCACACCTTTTCACAGAACATAA ATAATAATGCTTGCCAGCAGTA
hs-HIF1A-MT3_F	TTTCTTTGAGCTGGCATTCTGACTATAGAACATCAGATGATTTC TCTGAATTG
hs-HIF1A-MT3_R	CAATTCAAGAGAAATCATCTGATGTTCTATAGTCAGAACGCCAGCT AAAAAGAAAA
hs-HIF1A-MT4_F	AGTATGTTAATTGCTAAAATACAATGTTGATTTCAGAACAT GTCGCTATTAAACATCCCTTTTT
hs-HIF1A-MT4_R	AAAAAAAGGATGTTAATAGCGACATTCTGCATAAAATCAAACATT GTATTTGAGCAAATTAACATACT

**Table S4: Primers used to mutagenize the miR-20a-5p seed sequences**

## Figure S1

A

Predicted site 1

HIF1A-WT	5'-(235)GCAUUGCAGUAGCAU <b>C</b> AUUUUAA(257)-3'
miR-20a-5p	3'-GAUGGACGUGAU <b>A</b> UUC <b>G</b> UGAAA <b>U</b> -5'
HIF1A-MUT	5'-(235)GCAUUGCAGUAGCAU <b>C</b> Agaa <b>U</b> A(257)-3'

Predicted site 2

HIF1A-WT	5'-(399)UUACACCUUUUUUUU <b>C</b> AUUUU <b>A</b> (417)-3'
miR-20a-5p	3'-GAUGGACGUGAU <b>A</b> UUC <b>G</b> UGAAA <b>U</b> -5'
HIF1A-MUT	5'-(399)UUACACCUUUUUUU <b>C</b> Agaa <b>U</b> A(417)-3'

Predicted site 3

HIF1A-WT	5'-(732)CUGAUGUUUCUAUAG <b>C</b> ACUU <b>U</b> <b>G</b> (754)-3'
miR-20a-5p	3'-GAUGGACGUGAU <b>A</b> UUC <b>G</b> UGAAA <b>U</b> -5'
HIF1A-MUT	5'-(732)CUGAUGUUUCUAUAG <b>C</b> Agaa <b>U</b> G(754)-3'

Predicted site 4

HIF1A-WT	5'-(943)AAUGUUUGAUUU <b>A</b> <b>G</b> <b>C</b> ACUU <b>U</b> <b>G</b> (965)-3'
miR-20a-5p	3'-GAUGGACGUGAU <b>A</b> UUC <b>G</b> UGAAA <b>U</b> -5'
HIF1A-MUT	5'-(943)AAUGUUUGAUUU <b>A</b> <b>G</b> <b>C</b> Agaa <b>U</b> G(965)-3'

B

Predicted site 1

FPN-WT	5'-(1093)GUCUGUACCUUGAU <b>G</b> <b>G</b> ACUU <b>U</b> <b>G</b> (1115)-3'
miR-20a-5p	3'-GAUGGACGUGAU <b>A</b> UUC <b>G</b> UGAAA <b>U</b> -5'
FPN-MUT	5'-(1093)GUCUGUACCUUGAU <b>G</b> <b>G</b> Agaa <b>U</b> G(1115)-3'

Predicted site 2

FPN-WT	5'-(1119)UAACCGUUUAU <b>A</b> <b>G</b> <b>C</b> ACUU <b>U</b> <b>C</b> (1139)-3'
miR-20a-5p	3'-GAUGGACGUGAU <b>A</b> UUC <b>G</b> UGAAA <b>U</b> -5'
FPN-MUT	5'-(1119)UAACCGUUUAU <b>A</b> <b>G</b> <b>C</b> Agaa <b>U</b> C(1139)-3'

Predicted site 3

FPN-WT	5'-(1152)AUACGUUGCUAUG <b>A</b> <b>G</b> <b>C</b> ACUU <b>U</b> <b>C</b> (1173)-3'
miR-20a-5p	3'-GAUGGACGUGAU <b>A</b> UUC <b>G</b> UGAAA <b>U</b> -5'
FPN-MUT	5'-(1152)AUACGUUGCUAUG <b>A</b> <b>G</b> <b>C</b> Agaa <b>U</b> C(1173)-3'

Figure S1. Putative miR-20a binding sites in the 3'UTRs of human (A) HIF1A and (B) FPN. The nucleotides that are mutated in the wild type 3'UTRs are indicated in bold and lower case.

Figure S2

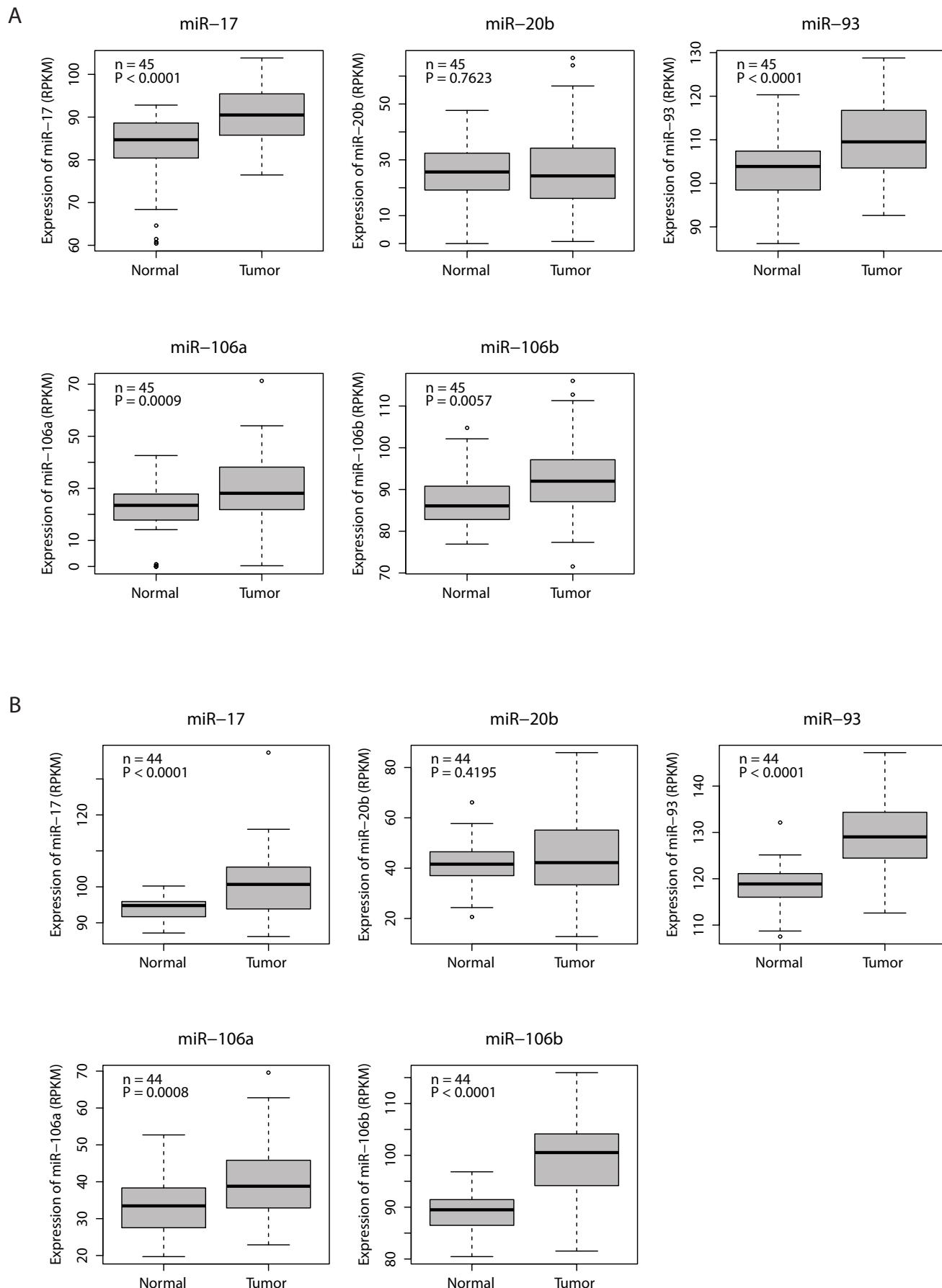


Figure S2. Boxplots indicating expression levels of the indicated member of the miR-17 seed family in the tumor and matching healthy tissue within TCGA datasets of (A) lung adenocarcinoma (LUAD) and (B) lung squamous cell carcinoma (LUSC) patients. 2-tailed Student's t test was used to calculate statistical significance.

Figure S3

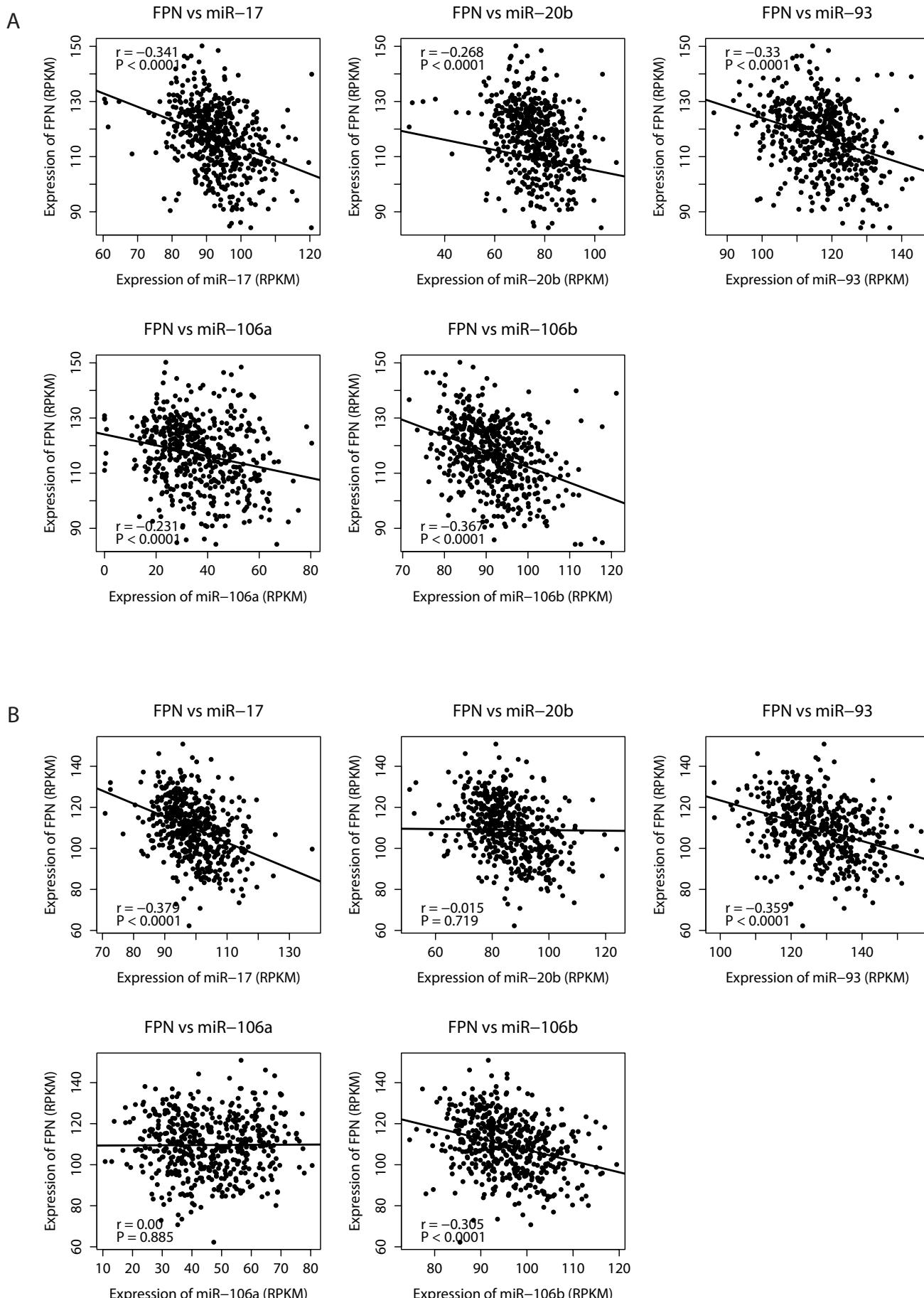
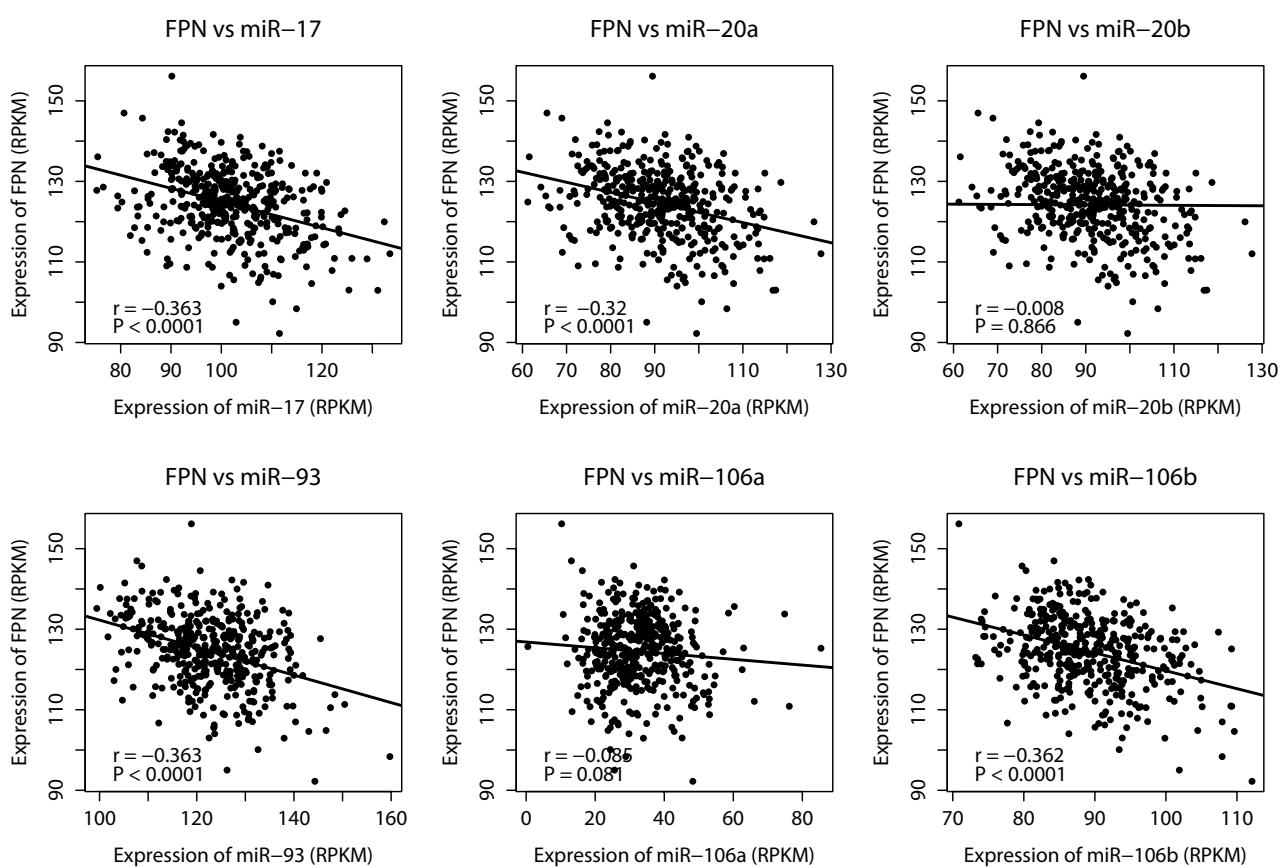


Figure S3. Scatter plots indicating correlations of FPN mRNA levels to the indicated member of the miR-17 seed family within TCGA datasets of (A) lung adenocarcinoma (LUAD) and (B) lung squamous cell carcinoma (LUSC) patients. Pearson's product moment correlation was used to calculate statistical significance.

Figure S4

A



B

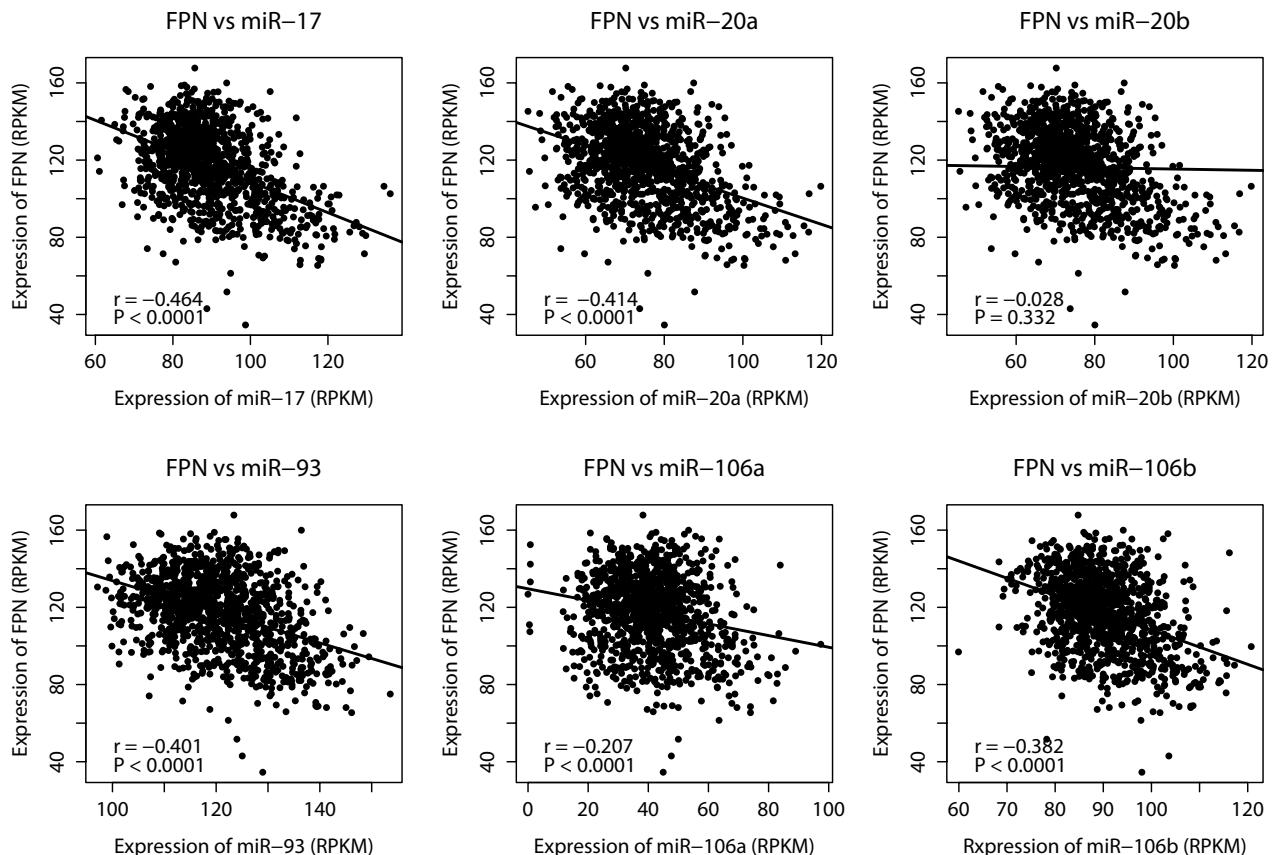
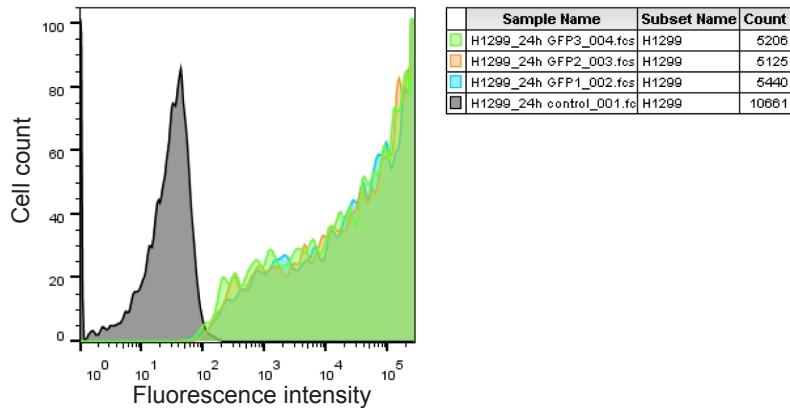


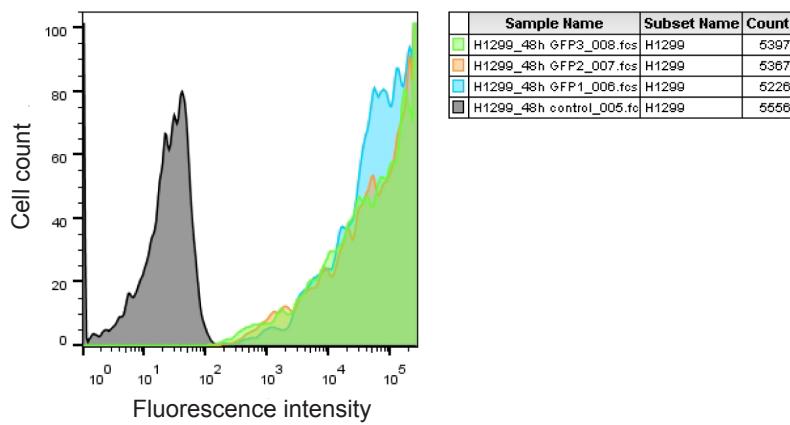
Figure S4. Scatter plots indicating correlation of FPN mRNA levels to the indicated member of the miR-17 seed family within TCGA datasets of (A) hepatocellular carcinoma (LIHC) and (B) breast cancer (BRCA) patients. Pearson's product moment correlation was used to calculate statistical significance.

Figure S5

A



B



C

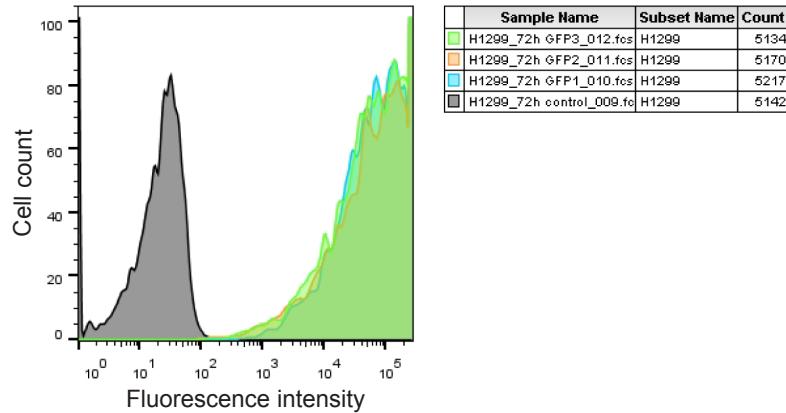


Figure S5. Chromatograms illustrating the transfection efficiency of the FPN expression construct (pcDNA FPN-EGFP) into H1299 cells. Green fluorescence intensity was analyzed by flow cytometry following transient transfection of pcDNA FPN-EGFP into H1299 cells after (A) 24 hrs (B) 48 hrs and (C) 72 hrs.

Figure S6

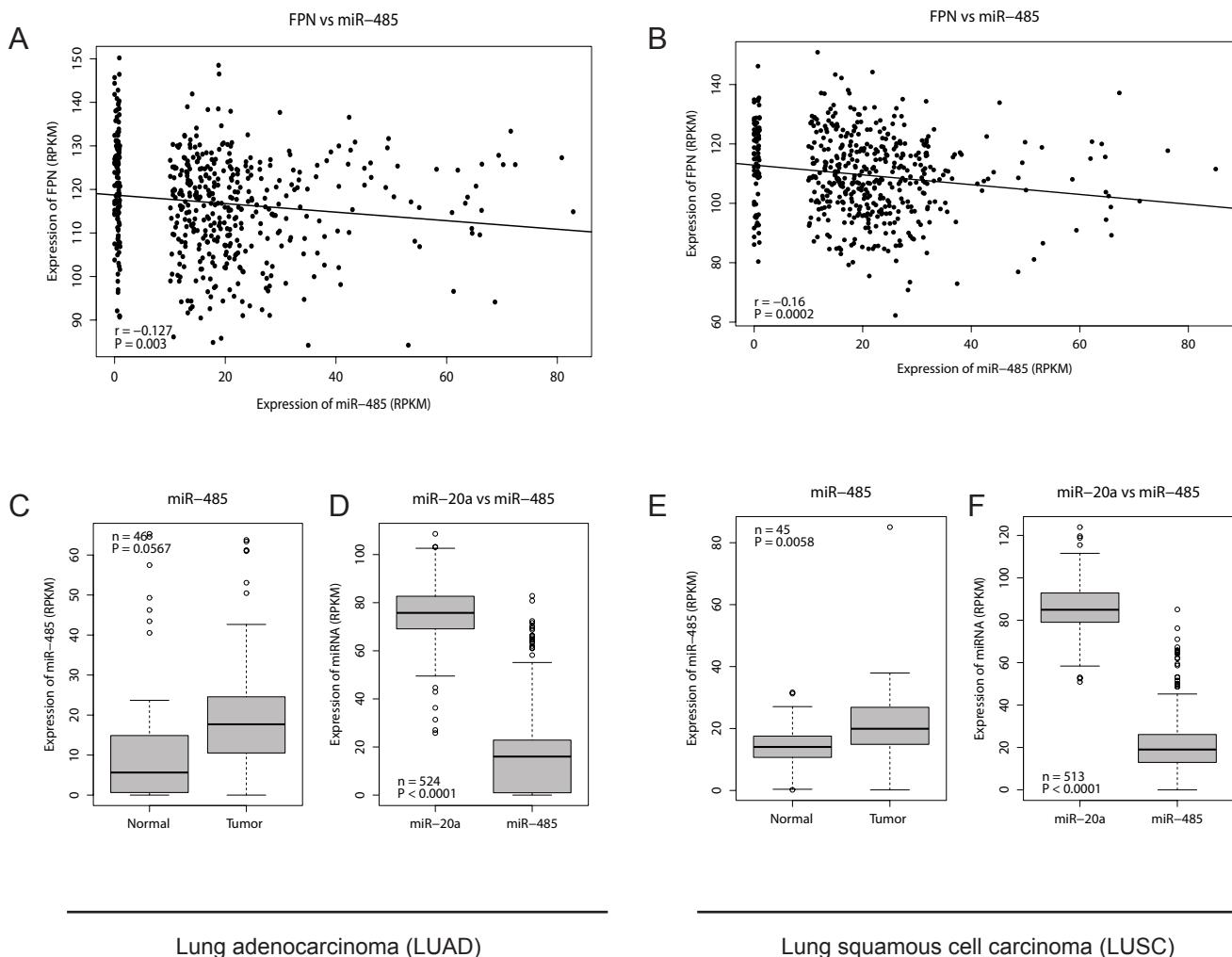


Figure S6. TCGA RNA-seq datasets available from Lung adenocarcinoma (LUAD) and Lung squamous cell carcinoma (LUSC) patients were downloaded from the UCSC cancer genomic browser (<https://genome-cancer.ucsc.edu>). Correlations between miR-485 and FPN mRNA expression levels are represented as scatter plots for (A) LUAD ( $n = 524$ ,  $r = -0.127$ ,  $P = 0.003$ ) and (B) LUSC ( $n = 513$ ,  $r = -0.16$ ,  $P = 0.0002$ ). Pearson's product moment correlation was used to calculate statistical significance. Boxplots illustrate that in patients with LUAD (C) miR-485 expression is not significantly elevated ( $n = 46$ ,  $P = 0.0567$ ) and that (D) miR-485 expression is significantly lower ( $n = 524$ ,  $P < 0.0001$ ) compared to miR-20a expression. In patients with LUSC (E) miR-485 is significantly increased compared to normal ( $n = 45$ ,  $P = 0.0058$ ) and (F) miR-485 expression is significantly lower ( $n = 513$ ,  $P < 0.0001$ ) compare to miR-20a levels. 2-tailed Student's t test was used to calculate statistical significance. RPKM = Reads Per Kilobase per Million mapped reads.

Figure S7

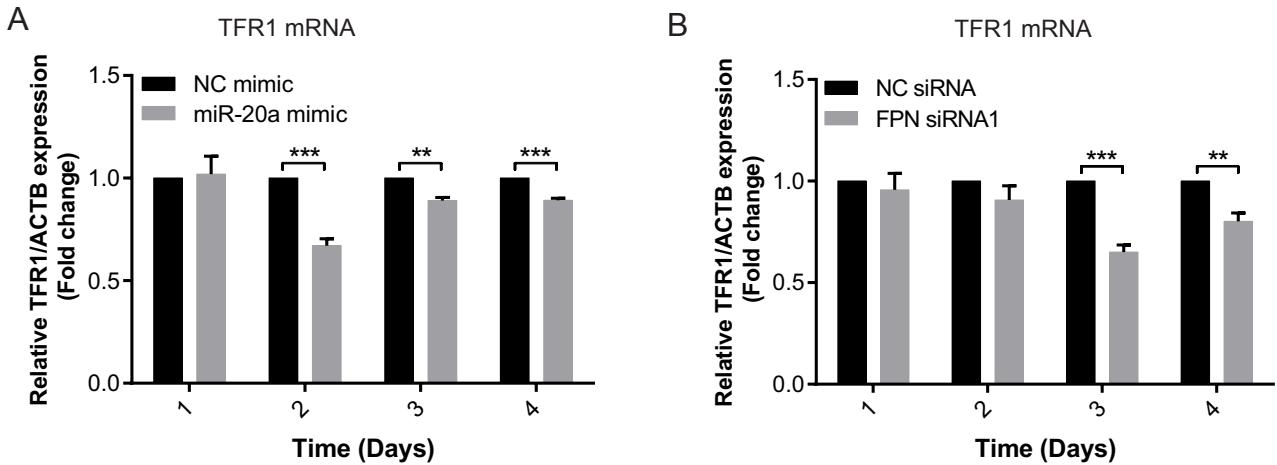


Figure S7. H1650 cells were transfected with 50 nM of (A) miR-20a mimic or (B) FPN siRNA1. After transfection TFR1 mRNA levels were analyzed by quantitative RT-PCR at an interval of 24 hrs for up to 4 days. Experiments were performed in triplicates and repeated at least three times. Data were normalized to ACTB. Data are represented as mean  $\pm$  SEM, and the values from negative control (NC) mimic were set to 1, \*\* $P < 0.001$ , \*\*\* $P < 0.001$ , 2-tailed Student's t test was used to calculate statistical significance.

Figure S8

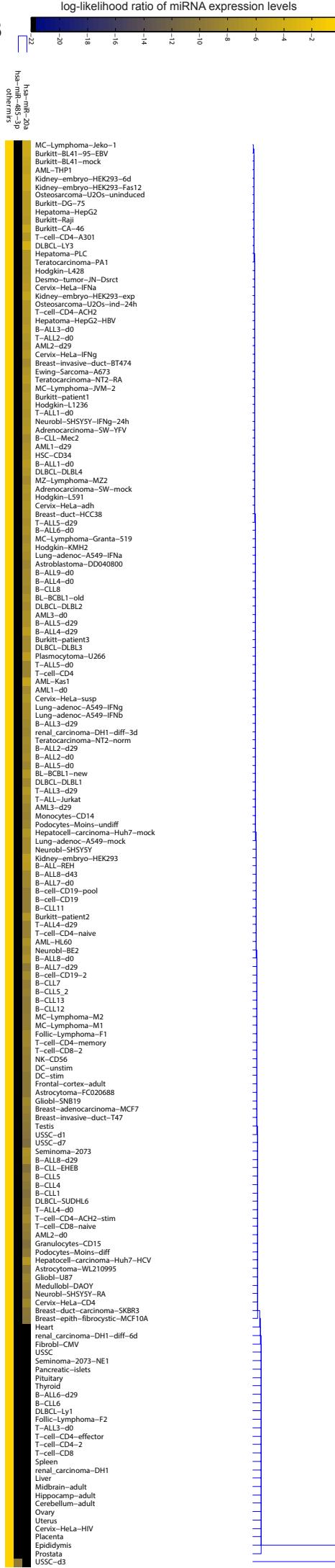


Figure S8. Heatmap illustrating comparative expression levels of miR-20a and miR-485-3p in various human tissues and cell lines.