

Online Supporting Information I

Modular synthesis of *N*-glycans and arrays for hetero-ligand binding analysis of HIV antibodies

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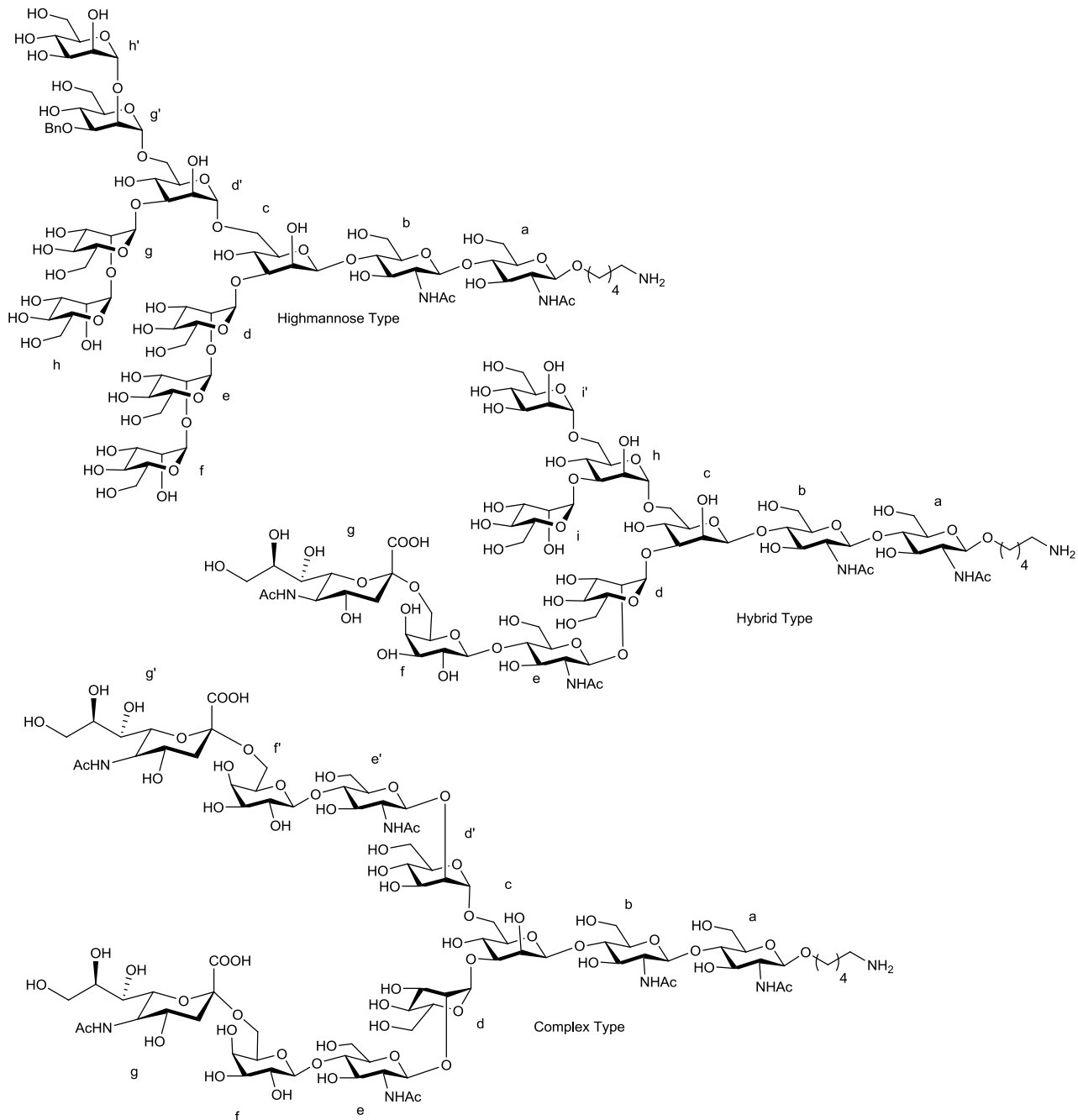
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1. Chemical Synthesis

i. Materials and Methods

All reagents were purchased from Sigma Aldrich, Across and used without further purification. Dry solvents were purchased from a commercial source without further distillation. Pulverized Molecular Sieves MS-4 Å (Aldrich) for glycosylation was activated by heating at 350 °C for 3 h. Reactions were monitored by analytical thin-layer chromatography (TLC) in EM silica gel 60 F254 plates and visualized under UV (254 nm) and/or by staining with acidic ceric ammonium molybdate or *p*-anisaldehyde. Flash chromatography was performed on silica gel (Merck) of 40-63 µm particle size. ¹H NMR spectra were recorded on a Bruker AVANCE 600 (600 MHz) spectrometer at 25 °C. All ¹H Chemical shifts (in ppm) were assigned according to CDCl₃ (δ = 7.24 ppm) and D₂O (δ = 4.80 ppm). ¹³C NMR spectra were obtained with Bruker AVANCE 600 spectrometer and were calibrated with CDCl₃ (δ = 77.00 ppm). Coupling constants (*J*) are reported in hertz (Hz). Splitting patterns are described by using the following abbreviations: s, singlet; brs, broad singlet; d, doublet; t, triplet; q, quartet; dd, doublet of doublet; m, multiplet. ¹H NMR spectra are reported in the following order: chemical shift, multiplicity, coupling constant(s) and number(s) of protons. All NMR signals were assigned on the basis of ¹H NMR, COSY, HSQC, HMQC, TOCSY, and ¹³C experiments. High resolution ESI mass spectra were recorded on a Bruker Daltonics spectrometer. NHS coated glass slides were purchased from SCHOTT (Nexterion H). Broadly neutralizing HIV antibodies PG9, PG16 and PGT141-145 were kindly gifted by Prof. Peter Kwong, NIH (PG9/PG16 also purchased from Polymun, Vienna Austria). PGT128 was kindly gifted by Prof. Dennis Burton, TSRI. Secondary antibody DyLight649-conjugated donkey anti-Human IgG was purchased from Jackson Immuno Research. Cytidine 5'-triphosphate (CTP), N-Acetylneuraminic acid (Neu5Ac), UDP galactose, L-fucose and Phospho (enol) pyruvic acid (PEP) were purchase from Sigma-Aldrich.

NMR Nomenclature. The individual sugar residues of highmannose, hybrid and complex type oligosaccharides have been labeled as shown bellow.



ii. General Procedures.

1. Enzyme expression, purification and reactions:

The functional domain of α -2,3 sialyltransferase (JTFAJ-16)¹ and α -2,6 sialyltransferase (JT-ISH-224)² were obtained according to our previous report³. Enzymes β -1, 4 galactosyl transferases from bovine milk was purchased from Sigma. Enzyme α -1, 2 fucosyl transferases (Human Fut2 derived from HEK-293 cells) was purchased from R&D SYSTEMS. The enzymes used in this work including *Bacteroides fragilis* L-fucokinase/GDP-fucose pyrophosphorylase (FKP), pyruvate kinase (PK), pyrophosphatase (PPA), cytidine monophosphate kinase (CMK), CMP-sialic acid synthetases (CSS) and α 1–3-fucosyltransferase from *Helicobacter pylori* (Hp1–3FT Δ 26695) are examples of suitable recombinant enzymes that are expressed and purified in our laboratory. Enzymatic reactions with cofactor regeneration were carried out according to the procedure reported previously from our group⁴.

2. Global deprotection:

Method 1 (for glycans with -Nphthalamide protection at all glucosamine residues): A mixture of protected glycans (50 mmol) and 10 mL of ethylene diamine: *n*BuOH (1:4) was stirred at 90 °C for overnight. Volatiles were then evaporated and the crude product was reacted with 10 mL Ac₂O/pyridine (1:2) for overnight. The solvents were removed using high vacuum and product was purified by flash column chromatography (acetone: toluene, 2/8, v/v). Product was de-acetylated using sodium methoxide in MeOH (10 mL) for overnight. The reaction mixture was neutralized by using IR-120, filtered and concentrated in *vacuo*. The residue was purified by flash column chromatography (acetone: toluene, 3/7, v/v). Product was dissolved in 10 mL MeOH: H₂O: HCOOH (6:3:1), Pd(OH)₂ (50% by weight) was added and the reaction mixture was hydrogenated for overnight. The reaction mixture was filtered through Celite and concentrated in *vacuo*. The residue was purified by Bio-Gel P-2 (BIO-RAD) column chromatography using water as eluent, and the product was the lyophilized to get desired oligosaccharides as a white color powder.

Method 2 (for glycans with -NHTroc protection at all glucosamine residues): A mixture of protected glycans (50 mmol) and lithium hydroxide (250 mmol) in 10 mL of 1, 4 dioxane: H₂O (4:1) was stirred at 90 °C for overnight. Volatiles were then evaporated and the crude product was reacted with 10 mL Ac₂O: pyridine (1:2) for overnight. The solvents were removed using high vacuum and product was purified by C18 gel column chromatography (MeOH: H₂O as an eluent). The product was de-acetylated using sodium methoxide in MeOH (10 mL) for overnight. The reaction mixture was neutralized by using IR-120, filtered and concentrated in *vacuo*. The residue was purified by C18 gel column chromatography (MeOH: H₂O as an eluent). The product was dissolved in 10 mL MeOH: H₂O: HCOOH (6:3:1), Pd(OH)₂ (50% by weight) was added and the mixture was hydrogenated for overnight. The reaction mixture was filtered through Celite and concentrated in *vacuo*. The residue was purified by Bio-Gel P-2 (BIO-RAD) column chromatography using water as eluent. The product was lyophilized to get desired oligosaccharides as a white color powder.

3. Enzymatic sialylation with cofactor regeneration: Glycans (5 μmol), Neu5Ac (10 μmol), ATP (0.05 μmol), CTP (1 μmol), phosphoenolpyruvate (10 μmol, monopotassium salt), cytidine monophosphate kinase (CMK, 80 units), CMP-sialic acid synthetases (CSS, 120 units), pyruvate kinase (PK, 40 units), pyrophosphatase (PPA, 40 units) and α 2,6/2,3 sialyltransferase (150 units) were dissolved in 50 μmol Tris buffer (25 mM, pH 7.5). The reaction was incubated at 37 °C with gentle agitation. Complete consumption of starting material was confirmed by mass spectrometric analysis. The reaction mixture was centrifuged and the supernatant subjected to gel filtration over P2-Biogel (eluent water). Fractions containing the product were combined and lyophilized to give the respective products as amorphous white solids.

4. Enzymatic β-1, 4-galactosylation:

Glycans (1 eq.) and UDP galactose (2 eq. per galactose) were dissolved in Tris buffer (25 mM, pH 7.5) containing MnCl₂ (10 mM). Enzyme β-1, 4-GalT-1 (150 units) was added to achieve a final concentration of glycan ranging from 2-5 mM. The resulting reaction mixture was incubated at 37 °C for 24 h. In di-galactosylation case, when TLC showed mono-galactosylated

intermediate, additional UDP-galactose (2eq), β -1,4-GalT (100 units) were added and incubated at 37 °C until complete consumption of intermediate. The reaction mixture was centrifuged and the supernatant was subjected to gel filtration over P2-Biogel (eluent water). Fractions containing the product were combined and lyophilized to give the respective products as amorphous white solids.

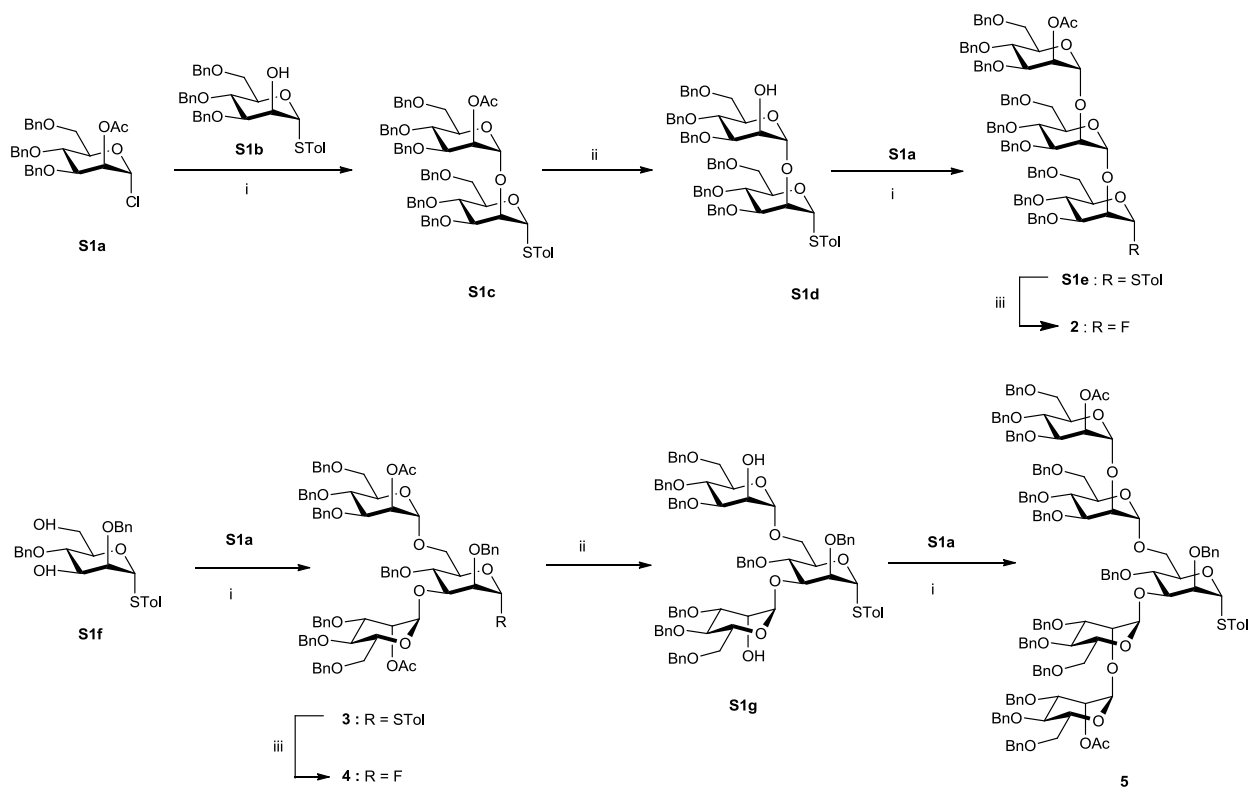
5. Enzymatic α -1,2/1,3-fucosylation : To a solution of glycans (5 μ mol), L-fucose (5 μ mol), ATP (0.5 μ mol), of GTP (0.5 μ mol), PEP (10 μ mol), and 10 mM MnCl_2 in a 25 mM Tris buffer (pH 7.4) was added L-fucokinase/GDP-fucose pyrophosphorylase (FKP, 200 units), PK (200 units), PPA (200 units), and α -1,2/1,3-fucosyltransferase (200 units), and the mixture was incubated at 37 °C for overnight. The reaction mixture was centrifuged and the supernatant was subjected to gel filtration over P2-Biogel (eluent water). Fractions containing the product were combined and lyophilized to give the respective products as amorphous white solids.

iii. Synthesis of building blocks 1-13.

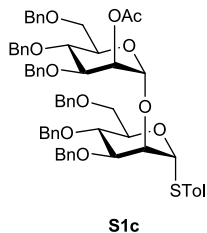
Synthesis of mono-, tri- and pentasaccharide building blocks 1-5.

The mannosyl trichloroacetimidate building block **1** was obtained by reported procedure⁵. The preparation of D1 arm trisaccharide **2** of $\text{Man}_9\text{GlcNAc}_2$ began by glycosylation of mannosyl chloride **S1a**⁶ and thioglycoside acceptor **S1b**⁷ under the treatment of 2,6-di-*tert*-butylpyridine (DTBP) and silver triflate (AgOTf) to obtain disaccharide **S1c** (Scheme S1). Zemplén deacetylation of **S1c** at 2-O position followed by glycosylation with **S1a** by employing DTBP and AgOTf gave desired trisaccharide **S1e** in 93% yield. The leaving group modification of **S1e** from thioglycoside to glycosyl fluoride **2** was performed under the action of *N*-bromosuccinimide (NBS) and diethylaminosulfur trifluoride (DAST). The fluoride transformation provided us better results in terms of α -selectivity and excellent yield during glycosylation with chitobiose trisaccharide. Under the activation of AgOTf, the condensation of donor **S1a**⁶ and acceptor **S1f**⁷ was performed to afford trimannose **3**, which was subsequently deacetylated and further di-glycosylated with **S1a** to afford D2/D3 arm pentasaccharide **5** in 71% yield. Compound **3** was next subjected to leaving group modification to trimannosyl fluoride **4**

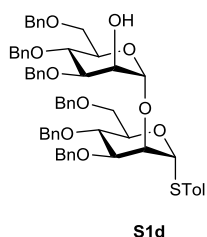
by using NBS and DAST in 68% yield. Careful observation of our strategy revealed that a single mannosyl chloride donor **S1a** and a unique DTBP/AgOTf mediated glycosylation condition was utilized to get D1, D2/D3 arm tri- and pentasaccharide intermediates in excellent yield.



Scheme S1 | Preparation of compound 1-5. **i**, DTBP, AgOTf, 4 Å MS, CH₂Cl₂, -30 °C to RT, overnight; **S1c**: 93%, **S1e**: 91%, **3**: 66%, **5**: 74%; **ii**, NaOMe, MeOH: CH₂Cl₂ = 1/1; **S1d**: 89%, **S1g**: 90%; **iii**, N-bromo succinimide (NBS), DAST, CH₂Cl₂, -30 °C to -10 °C; **2**: 58%, **4**: 61%. DTBP: 2, 6-di-tert-butylpyridine; DAST: Diethylaminosulfur trifluoride; AgOTf: Silver trifluoromethanesulfonate.

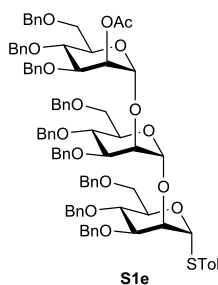


***p*-Tolyl-2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)-3,4,6-tri-O-benzyl-1-thio- α -D-mannopyranoside (S1c):** To a solution of acceptor **S1b** (1.70 g, 3.05 mmol) and donor **S1a** (2.34 g, 4.58 mmol) in 20 mL CH₂Cl₂ was added activated 4 Å molecular sieves and stirred for 1 h at RT. In a separate flask, AgOTf (1.19 g, 4.58 mmol) and DTBP (1.03 mL, 4.58 mmol) in 10 mL of CH₂Cl₂ were stirred with 4 Å MS for 1 h. The flask containing the AgOTf/DTBP was cooled to -30 °C and solution containing mixture of donor and acceptor was added over 5 min. The solution was stirred with gradual warming up to room temperature over 24 h. TLC (ethyl acetate: hexane, 2/8) indicated formation of product with consumption of starting material, the reaction was quenched with Et₃N, filtered through Celite, the filtrate was washed with aqueous NaHCO₃ (2 x 50 mL) and a brine (50 mL) solution. The organic layer was dried over Na₂SO₄, and concentrated in *vacuo*. The residue was purified by silica gel column chromatography (0% \rightarrow 15% EA in hexane) to afford **S1c** (2.90 g, 93%) as colorless foam. TLC (ethyl acetate: hexane = 2/8 v/v): R_f = 0.35; ¹H NMR (600 MHz, CDCl₃): δ 7.38-7.16 (m, 30H, Ar-H), 7.11-7.09 (m, 2H, Ar-H), 7.00 (d, *J* = 8.4 Hz, 2H, Ar-H), 5.44 (s, 1H, H-1^a), 5.51 (d, *J* = 2.4 Hz, 1H), 5.06 (s, 1H, H-1^b), 4.88 (d, *J* = 11.8 Hz, 1H), 4.81 (d, *J* = 11.2 Hz, 1H), 4.73-4.37 (m, 10H), 4.29 (t, *J* = 8.9 Hz, 1H), 4.22 (s, 1H), 3.96-3.88 (m, 4H), 3.83-3.71 (m, 2H), 3.69-3.63 (m, 2H), 3.55 (d, *J* = 12.1 Hz, 1H), 2.26 (s, 3H, -C(O)CH₃), 2.13 (s, 3H, -CH₃ of STol); ¹³C NMR (150 MHz, CDCl₃): δ 170.49, 138.75, 138.67, 138.62, 138.42, 138.32, 138.24, 137.87, 132.55, 130.51, 128.77, 128.64, 128.61, 128.56, 128.52, 128.39, 128.29, 128.20, 128.15, 128.03, 127.99, 127.94, 127.78, 127.63, 99.99, 87.78, 80.19, 78.35, 76.88, 75.48, 75.32, 75.03, 74.56, 73.46, 73.08, 72.47, 72.21, 72.18, 68.48, 69.02, 68.85, 21.42, 21.36; ESI-MS: *m/z* calcd for C₆₃H₆₆O₁₁S; 1030.4218; found 1053.4228 (*M* + Na)⁺.



***p*-Tolyl-3,4,6-tri-*O*-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)-3,4,6-tri-*O*-benzyl-1-thio- α -D-**

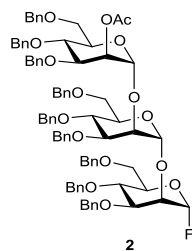
manno pyranoside (S1d): To a solution of compound **S1c** (1.01 g, 0.970 mmol) in 20 mL of methanol: CH₂Cl₂ (1/1) was added sodium methoxide (0.024 g, 0.42 mmol), stirred at RT until TLC (ethyl acetate: hexane, 3/7) indicated formation of a product with consumption of the starting material. The reaction mixture was neutralized with IR-120, filtered and concentrated *in vacuo* and the residue was purified by silica gel column chromatography (0% \rightarrow 30% EA in hexane) to afford **S1d** (0.859 g, 89%) as a colorless oil. TLC (ethyl acetate: hexane = 3/7 v/v): R_f = 0.29; ¹H NMR (600 MHz, CDCl₃): δ 7.33-7.10 (m, 32H, Ar-H), 6.96 (d, *J* = 7.8 Hz, 2H, Ar-H), 5.56 (s, 1H, H-1), 5.11 (s, 1H, H-1), 4.85 (d, *J* = 10.8 Hz, 1H), 4.75 (d, *J* = 10.8 Hz, 1H), 4.69-4.62 (m, 3H), 4.52-4.38 (m, 6H), 4.37 (d, *J* = 12.1 Hz, 1H), 4.25 (d, *J* = 12 Hz, 2H), 4.09 (s, 1H), 3.90-3.77 (m, 6H), 3.70 (d, *J* = 10.8 Hz, 1H), 3.62 (dd, *J* = 4.8, 8.2 Hz, 1H), 3.54 (d, *J* = 10.2 Hz, 1H), 2.23 (s, 3H, CH₃, STol); ¹³C NMR (150 MHz, CDCl₃): δ 138.61, 138.42, 138.33, 138.24, 137.97, 137.51, 132.18, 130.38, 129.73, 128.55, 128.47, 128.38, 128.29, 128.24, 127.93, 127.91, 127.89, 127.86, 127.85, 127.78, 127.68, 127.57, 127.48, 127.45, 127.33, 101.23, 87.62, 80.06, 79.99, 76.52, 75.18, 75.01, 74.95, 94.35, 73.21, 73.18, 72.87, 72.38, 72.18, 71.69, 69.28, 68.70, 68.56, 19.45; ESI-MS: *m/z* calcd for C₆₁H₆₄O₁₀S; 988.4112; found 1011.4125 (*M* + Na)⁺.



***p*-Tolyl-2-*O*-acetyl-3,4,6-tri-*O*-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)-3,4,6-tri-*O*-benzyl- α -D-**

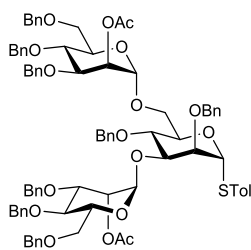
mannopyranosyl-(1 \rightarrow 2)-3,4,6-tri-*O*-benzyl-1-thio- α -D-mannopyranoside (S1e): To a solution of acceptor **S1d** (2.6 g, 2.62 mmol) and donor **S1a** (2.01 g, 3.94 mmol) in 10 mL CH₂Cl₂ was added activated 4 Å molecular sieves and stirred for 1 h at RT. In a separate flask,

AgOTf (1.02 g, 3.94 mmol) and DTBP (893 μ L, 3.94 mmol) in 30 mL of CH_2Cl_2 were stirred with activated 4 Å molecular sieves for 1 h. The mixture of AgOTf /DTBP was cooled to $-30\text{ }^\circ\text{C}$ and a solution of donor and acceptor was added over 5 min. The solution was stirred with gradual warming up to RT over 24 h until TLC (ethyl acetate: hexane, 2/8) indicated formation of product with consumption of starting material. The reaction was quenched with Et_3N , filtered through Celite. The filtrate was washed with aqueous NaHCO_3 (2 x 50 mL) and a brine (50 mL) solution. The organic layer was dried over Na_2SO_4 , filtered and concentrated in *vacuo*. The residue was purified by silica gel column chromatography (0% \rightarrow 20% EA in hexane) to afford **S1e** (3.50 g, 91%) as colorless foam. TLC (ethyl acetate: hexane = 2/8 v/v): $R_f = 0.41$; ^1H NMR (600 MHz, CDCl_3): δ 7.34-7.11 (m, 45H, Ar-H), 7.05-7.03 (m, 2H, Ar-H), 6.93 (d, $J = 7.2$ Hz, 2H, Ar-H), 5.65 (s, 1H, H-1), 5.50 (d, $J = 3.0$ Hz, 1H, H-1), 5.10 (d, $J = 3.2$ Hz, 1H, H-1), 4.82 (dd, $J = 3.1, 10.2$ Hz, 2H), 4.75 (d, $J = 12.1$ Hz, 1H), 4.68 (s, 2H), 4.60-4.54 (m, 8H), 4.35-4.19 (m, 9H), 3.90-3.84 (m, 4H), 3.78-3.72 (m, 2H), 3.66-3.63 (m, 3H), 3.55-3.52 (m, 1H), 3.46 (d, $J = 4.2$ Hz, 1H), 3.41 (dd, $J = 3.4, 9.2$ Hz, 1H), 3.31 (d, $J = 9.8$ Hz, 1H), 2.20 (s, 3H, $-\text{C}(\text{O})\text{CH}_3$), 2.05 (s, 3H, CH_3 , STol); ^{13}C NMR (150 MHz, CDCl_3): δ 170.84, 139.12, 138.85, 138.79, 138.65, 138.60, 138.52, 138.37, 138.25, 137.94, 137.60, 132.30, 130.75, 129.97, 129.90, 128.98, 128.63, 128.52, 128.33, 128.20, 128.03, 127.88, 127.64, 100.79, 96.52, 88.06, 80.83, 80.45, 76.45, 76.42, 76.04, 75.65, 75.21, 75.14, 74.69, 73.63, 73.05, 73.01, 72.95, 71.99, 71.87, 71.43, 69.66, 69.51, 69.35, 68.40, 68.34, 21.40, 21.36, 21.31; ESI-MS: m/z calcd for $\text{C}_{90}\text{H}_{94}\text{O}_{16}\text{S}$; 1462.6155; found 1485.6190 ($M + \text{Na}$) $^+$.



2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)-3,4,6-tri-O-benzyl- α -D-

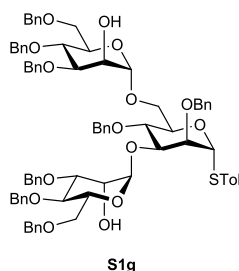
mannopyranosyl-(1 \rightarrow 2)-3,4,6-tri-O-benzyl-1- α -D-mannopyranosyl fluoride (2): To a solution of trisaccharide **S1e** (0.6 g, 0.410 mmol) in CH₂Cl₂ (10 mL) at -30 °C was added NBS (0.218 mg, 1.23 mmol), stirred for 10 minutes. DAST (324 μ L, 2.46 mmol) was added slowly and the resulting reaction mixture was stirred at -10 °C for 6 h. TLC (ethyl acetate: hexane, 3/7) indicated formation of product with consumption of starting material, the reaction was quenched with aq.NaHCO₃, and the filtrate was washed with aqueous NaHCO₃ (2 x 50 mL) and a brine (50 mL) solution. The organic layer was dried over Na₂SO₄, filtered and concentrated in *vacuo*. The residue was purified by silica gel column chromatography (0% \rightarrow 25% EA in hexane) to afford fluoride **2** (0.320 g, 58%) as white foam and 0.200 g alcohol (anomeric-OH) as side product. TLC (ethyl acetate: hexane = 3/7, v/v): R_f = 0.31; ¹H NMR (600 MHz, CHCl₃): δ 7.32-7.12 (m, 45H, Ar-H), 5.67 (d, *J* = 50.4 Hz, 1H, Ar-H), 5.49 (s, 1H, H-1), 5.16 (s, 1H, H-1), 4.99 (s, 1H, H-1), 4.82-4.77 (m, 3H), 4.65-4.38 (m, 15H), 4.30 (d, *J* = 12.1 Hz, 1H), 4.03 (s, 1H), 3.97-3.94 (m, 3H), 3.90-3.64 (m, 12H), 3.54 (d, *J* = 10.2 Hz, 1H), 2.11 (s, 3H, -C(O)CH₃); ¹³C NMR (150 MHz, CDCl₃): δ 170.44, 138.77, 138.64, 138.60, 138.54, 138.46, 138.41, 138.28, 128.76, 128.64, 128.61, 128.54, 128.44, 128.31, 128.19, 128.13, 128.09, 128.04, 128.01, 127.95, 127.90, 127.87, 127.84, 127.79, 127.73, 107.65, 106.19, 101.04, 99.72, 78.56, 78.31, 75.45, 75.41, 75.31, 74.86, 74.59, 74.31, 74.10, 73.86, 73.68, 73.65, 73.57, 72.69, 72.49, 72.38, 72.17, 69.51, 69.31, 69.01, 68.95, 21.44; ESI-MS: *m/z* calcd for C₈₃H₈₇FO₁₆; 1358.5870; found 1381.5891 (*M* + Na)⁺.



***p*-Tolyl-2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3)-2-O-acetyl-3,4,6-tri-O-**

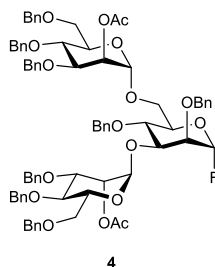
benzyl- α -D-mannopyranosyl-(1 \rightarrow 6)-2,4-di-O-benzyl-1-thio- α -D-mannopyranoside (3): To a

solution of acceptor **S1f** (0.54 g, 1.23 mmol) and donor **S1a** (1.57 g, 3.07 mmol) in CH₂Cl₂ (10 mL) was added 4 Å activated MS and stirred for 1 h at room temperature. In a separate flask, AgOTf (0.787 g, 3.07 mmol) and DTBP (690 µL, 3.07 mmol) in 10 mL of CH₂Cl₂ were stirred with MS 4 Å. After stirring for 1 h, the flask containing the AgOTf /DTBP was cooled to -30 °C and a solution containing a mixture of donor and acceptor was added over 5 min. The solution was stirred with gradual warming up to room temperature over 24 h. TLC (ethyl acetate: hexane, 2/8) indicated formation of product with consumption of starting material. The reaction was quenched with Et₃N, filtered through Celite, the filtrate was washed with aqueous NaHCO₃ (2 x 50 mL) and a brine (50 mL) solution. The organic layer was dried over Na₂SO₄, filtered and concentrated in *vacuo*. The residue was purified by silica gel column chromatography (0% → 20% EA in hexane) to afford **3** (1.10 g, 66%) as colorless foam. TLC (ethyl acetate: hexane = 2/8, v/v): R_f = 0.36; ¹H NMR (600 MHz, CDCl₃): δ 7.35-7.10 (m, 42H, Ar-H), 7.05 (d, *J* = 8.0 Hz, 2H, Ar-H), 5.51 (s, 1H), 5.47 (s, 1H, H-1), 5.45 (dd, *J* = 2.0, 2.8 Hz, 1H), 5.21 (s, 1H, H-1), 4.91 (d, *J* = 1.5 Hz, 1H, H-1), 4.86 (t, *J* = 10.2 Hz, 2H), 4.75 (d, *J* = 11.2 Hz, 1H), 4.67-4.58 (m, 5H), 4.54-4.36 (m, 8H), 4.23 (dd, *J* = 3.9, 9.6 Hz, 1H), 4.12-4.06 (m, 2H), 4.01 (dd, *J* = 3.2, 9.2 Hz, 1H), 3.97-3.86 (m, 5H), 3.82 (t, *J* = 9.2 Hz, 1H), 3.79-3.75 (m, 1H), 3.72 (dd, *J* = 3.9, 10.7 Hz, 1H), 3.70-3.56 (m, 4H), 2.17 (s, 3H, -C(O)CH₃), 2.13 (s, 3H, -C(O)CH₃), 2.09 (s, 3H, CH₃, STol); ¹³C NMR (150 MHz, CDCl₃): δ 170.27, 170.10, 138.57, 138.53, 138.18, 138.16, 137.81, 137.77, 137.42, 131.46, 130.85, 129.83, 128.44, 128.42, 128.36, 128.27, 128.21, 128.06, 127.81, 127.72, 127.68, 127.63, 127.53, 127.48, 99.83, 98.17, 85.23, 78.98, 78.08, 77.76, 75.15, 74.98, 74.89, 74.32, 74.15, 73.52, 73.30, 72.23, 72.05, 71.85, 71.48, 71.43, 71.36, 69.05, 68.72, 68.60, 68.41, 66.60, 60.39, 21.15, 21.00, 20.94; ESI-MS: *m/z* calcd for C₈₅H₉₀O₁₇S; 1414.5791; found 1437.5821 (*M* + Na)⁺.

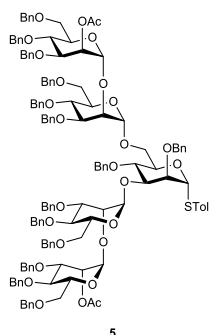


***p*-Tolyl-3,4,6-tri-*O*-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3)-3,4,6-tri-*O*-benzyl- α -D-mannopyranosyl-(1 \rightarrow 6)]-2,4-di-*O*-benzyl-1-thio- α -D-mannopyranoside (**S1g**):**

To a solution of trisaccharide **3** (1.30 g, 0.918 mmol) in 10 mL methanol: CH₂Cl₂ (1/1) was added sodium methoxide (0.024 g, 0.459 mmol), stirred at RT until TLC (ethyl acetate: hexane, 3/7) indicated formation of a product with consumption of the starting material. The reaction mixture was neutralized with IR-120, filtered and concentrated *in vacuo* and the residue was purified by silica gel column chromatography (0% \rightarrow 25% EA in hexane) to afford **S1g** (1.10 g, 90%) as a colorless oil. TLC (ethyl acetate : hexane = 3/7, v/v): R_f = 0.26; ¹H NMR (600 MHz, CDCl₃): δ 7.36-7.12 (m, 42H, Ar-H), 7.06 (d, *J* = 8.0 Hz, 2H, Ar-H), 5.46 (s, 1H, H-1), 5.24 (s, 1H, H-1), 5.01 (d, *J* = 1.1 Hz, 1H, H-1), 4.82 (t, *J* = 11.1 Hz, 2H), 4.69 (d, *J* = 11.6 Hz, 1H), 4.65-4.44 (m, 13H), 4.23 (dd, *J* = 4.1, 9.6 Hz, 1H), 4.16 (s, 1H), 4.07 (dd, *J* = 2.8, 9.4 Hz, 1H), 4.03 (d, *J* = 10.1 Hz, 1H), 3.96-3.88 (m, 4H), 3.87-3.81 (m, 3H), 3.80-3.75 (m, 1H), 3.73-3.64 (m, 4H), 3.62 (dd, *J* = 1.6, 10.8 Hz, 1H), 2.38 (s, 1H), 2.34 (s, 1H, -C(O)CH₃), 2.19 (s, 3H, CH₃, STol); ¹³C NMR (150 MHz, CDCl₃): δ 138.12, 138.11, 137.69, 131.75, 130.12, 128.82, 128.76, 128.73, 128.67, 128.63, 128.57, 128.54, 128.53, 128.41, 128.35, 128.29, 128.19, 128.12, 128.08, 128.03, 127.95, 128.92, 127.82, 124.43, 99.90, 85.55, 80.32, 80.00, 78.49, 79.46, 75.46, 75.41, 75.27, 75.19, 75.13, 74.66, 74.50, 74.39, 73.85, 73.81, 73.62, 72.32, 72.24, 71.85, 71.71, 71.40, 71.33, 69.48, 69.00, 68.96, 68.90, 68.32, 66.51, 21.15; ESI-MS: *m/z* calcd for C₈₁H₈₆O₁₅S; 1330.5580; found 1353.5614 (*M* + Na)⁺.



2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3)-2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 6)-2,4-di-O-benzyl- α -D-mannopyranosyl fluoride (4): To a solution of compound **3** (0.270 g, 0.197 mmol) in CH₂Cl₂ (10 mL) at -30 °C was added NBS (0.052 g, 0.296 mmol), stirred for 10 min. DAST (52 μ L, 0.395 mmol) was then added slowly and the resulting reaction mixture was stirred for 4 h at -10 °C. TLC (ethyl acetate: hexane, 3/7) indicated formation of product with consumption of starting material. The reaction mixture was quenched with aq.NaHCO₃, and the filtrate was washed with aqueous NaHCO₃ (2 x 50 mL) and a brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by silica gel column chromatography (0% \rightarrow 8% EA in toluene) to afford **4** (0.150 g, 61%) as white foam. TLC (ethyl acetate: toluene =1/9, v/v): R_f = 0.19; ¹H NMR (600 MHz, CHCl₃): δ 7.29-7.10 (m, 40H, Ar-H), 5.50 (s, 1H, H-1), 5.47 (d, *J* = 50 Hz, 1H, H-1), 5.46 (s, 1H, H-1), 5.18 (s, 1H), 4.92 (s, 1H), 4.86 (d, *J* = 11.2 Hz, 1H), 4.83 (d, *J* = 10.8 Hz, 1H), 4.72 (d, *J* = 10.0 Hz, 1H), 4.72-4.56 (m, 6H), 4.48-4.40 (m, 7H), 4.10 (d, *J* = 8.9 Hz, 1H), 4.00-3.92 (m, 4H), 4.90-3.77 (m, 5H), 3.71-3.64 (m, 5H), 3.57 (d, *J* = 7.2 Hz, 1H), 2.13 (s, 3H, -C(O)CH₃), 2.06 (s, 3H, -C(O)CH₃); ¹³C NMR (150 MHz, CHCl₃): δ 170.56, 170.36, 138.80, 138.74, 138.50, 138.39, 138.11, 138.04, 137.89, 137.88, 129.70, 128.81, 128.70, 128.68, 128.65, 128.58, 128.56, 128.54, 128.43, 128.27, 128.18, 128.12, 128.07, 128.04, 128.00, 127.97, 127.89, 127.82, 126.55, 106.66, 105.18, 100.02, 98.84, 98.81, 78.31, 77.70, 76.31, 76.03, 75.42, 75.37, 75.23, 74.62, 74.41, 74.27, 73.90, 73.80, 73.65, 73.16, 72.66, 72.14, 71.92, 71.58, 69.42, 68.96, 68.70, 68.61, 66.53, 42.20, 32.21, 29.98, 29.65, 22.98, 21.63, 21.44, 21.27, 14.71, 14.41; ESI-MS: *m/z* calcd for C₇₈H₈₃O₁₇F; 1310.5507; found 1333.5536 (*M* + Na)⁺.



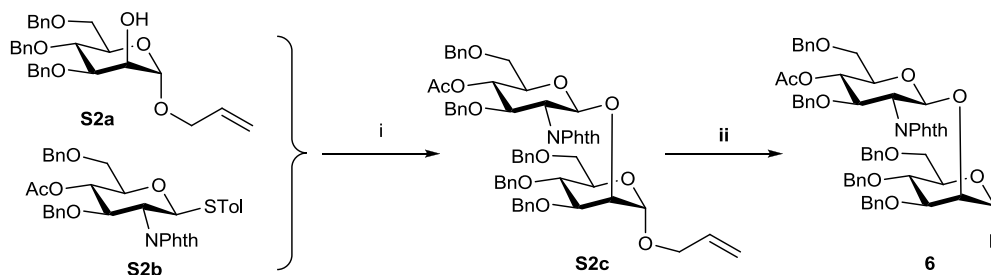
***p*-Tolyl-2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3)-[2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 6)]-2,4-di-O-benzyl-1-thio- α -D-mannopyranoside**

(5): To a solution of acceptor **S1g** (1.01 g, 0.739 mmol) and donor **S1a** (0.947 g, 1.84 mmol) in 20 mL CH₂Cl₂ was added 4 Å activated MS and stirred for 1 h at room temperature. In a separate flask, AgOTf (0.472 g, 1.85 mmol) and DTBP (415 μ L, 1.85 mmol) in 10 mL of CH₂Cl₂ were stirred with MS 4 Å. After stirring for 1 h, the flask containing the AgOTf /DTBP was cooled to -30 °C and a solution containing a mixture of donor and acceptor in CH₂Cl₂ was added over 5 min. The solution was stirred with gradual warming up to room temperature over 24 h. TLC (ethyl acetate: hexane, 3/7) indicated formation of product with consumption of starting material. The reaction was then quenched with Et₃N, filtered through Celite, and the filtrate was washed with aqueous NaHCO₃ (2 x 50 mL) and a brine (50 mL) solution. The organic layer was dried over Na₂SO₄, filtered and concentrated in *vacuo*. The residue was purified by silica gel column chromatography (0% \rightarrow 25% EA in hexane) to afford **5** (1.25 g, 74%) as white solid. TLC (ethyl acetate: hexane =3/7, v/v): R_f = 0.33; ¹H NMR (600 MHz, CDCl₃): δ 7.27-7.04 (m, 72H, Ar-H), 7.02 (d, *J* = 8.0 Hz, 2H, Ar-H), 5.52 (d, *J* = 1.5 Hz, 2H, 2 x H-1), 5.46 (s, 1H, H-1), 5.18 (s, 1H, H-1), 5.04 (d, *J* = 8.2 Hz, 1H), 4.85 (t, *J* = 11.0 Hz, 2H), 4.78 (t, *J* = 11.1 Hz, 2H), 4.72-4.44 (m, 18H), 4.41-4.30 (m, 8H), 4.27 (d, *J* = 12.3 Hz, 2H), 4.20 (s, 2H), 4.05-3.92 (m, 6H), 3.91-3.73 (m, 8H), 3.68-3.56 (m, 10H), 3.48 (d, *J* = 10.1 Hz, 1H), 3.40 (d, *J* = 10.3 Hz, 1H), 3.34 (d, *J* =

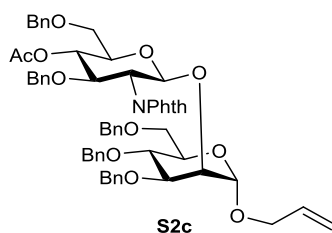
10.5 Hz, 1H), 2.10 (s, 6H, -C(O)CH₃), 2.01 (s, 3H, CH₃, STol); ¹³C NMR (150 MHz, CDCl₃): δ 170.22, 152.99, 152.78, 152.57, 138.43, 138.38, 138.24, 138.05, 138.02, 137.86, 137.81, 131.15, 129.80, 128.44, 128.30, 128.20, 128.05, 127.95, 127.85, 127.72, 127.70, 127.62, 127.55, 127.53, 127.47, 127.41, 127.38, 127.35, 127.23, 126.94, 101.16, 99.37, 98.86, 84.97, 79.24, 78.07, 78.02, 75.22, 74.99, 74.87, 74.77, 74.55, 74.47, 74.29, 74.17, 73.94, 73.82, 73.38, 73.24, 73.18, 73.01, 72.58, 71.93, 71.85, 71.77, 71.60, 71.55, 71.47, 70.99, 69.41, 68.64, 68.57, 68.46, 68.01, 29.69, 21.22, 20.91; ESI-MS: *m/z* calcd for C₁₃₉H₁₄₆O₂₇S; 2279.9698; found 2302.9747 (*M* + Na)⁺.

Synthesis of disaccharides building block 6.

Condensation of acceptor **S2a**⁸ with donor **S2b**⁹ mediated by NIS and TfOH supplied the disaccharide **S2c** with exclusively β-linkage (*J*_{1',2'} = 8.5 Hz) in 64% yield. Compound **S2c** was next modified to fluoride **6** via anomeric deallylation followed by conversion of free -OH to -F in presence of DAST (Scheme S2).

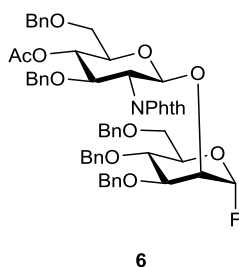


Scheme S2 | Preparation of compound 6. i, NIS, TfOH, CH₂Cl₂, 4 Å MS, -40 °C, 2h, 64%; **ii**, (1) PdCl₂, MeOH: CH₂Cl₂ = 1/1, (2) DAST, CH₂Cl₂, -30 °C to -10 °C, 3-5 h, 61%.



Allyl-O-4-O-acetyl-3,6-O-di-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl-(1 \rightarrow 2)-O-

3,4,6-tri-O-benzyl- α -D-mannopyranoside (S2c): A mixture of thioglycoside donor **S2b** (2.33 g, 3.65 mmol), acceptor **S2a** (1.50 g, 3.05 mmol) and activated 4 Å molecular sieves in 30 mL dry CH₂Cl₂ was stirred for 1 h at room temperature. NIS (1.35 g, 6.01 mmol) and TfOH (66.2 μ L, 0.75 mmol) were added slowly at -40 °C and stirred for 2 h until TLC (ethyl acetate: toluene, 1/9) indicated formation of product with consumption of starting material. The reaction mixture was quenched with Et₃N, filtered through Celite, and the filtrate was washed with aq. NaHCO₃ (2 x 50 mL), aq. Na₂S₂O₃ (2 x 50 mL) and finally with brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 10% EA in toluene) to afford **S2c** (2.10 g, 64%) as white solid. TLC (ethyl acetate: toluene = 1/9, v/v): R_f = 0.29; ¹H NMR (600 MHz, CDCl₃): δ 7.52-7.50 (m, 4H, Ar-H), 7.33-7.10 (m, 16H, Ar-H), 7.10-7.09 (m, 2H, Ar-H), 7.00-6.90 (m, 2H, Ar-H), 5.73-5.71 (m, 1H, -OCH₂-CH=CH₂), 5.26 (d, *J* = 8.4 Hz, 1H, H-1), 5.14-5.06 (m, 3H), 4.71 (t, *J* = 7.8 Hz, 2H), 4.58 (d, *J* = 12.1 Hz, 1H), 4.50-4.39 (m, 6H), 4.34 (d, *J* = 4.8 Hz, 1H), 4.32 (d, *J* = 6.6 Hz, 1H), 4.10 (t, *J* = 8.6 Hz, 1H), 4.07-4.00 (q, 2H), 3.78 (dd, *J* = 3.2, 6.1 Hz, 1H), 3.81-3.78 (m, 2H), 3.71-3.70 (dd, *J* = 3.2, 6.3 Hz, 1H), 3.65-3.60 (m, 1H), 3.58 (dd, *J* = 3.1, 6.5 Hz, 1H), 3.56 (t, *J* = 8.9 Hz, 1H), 3.48 (t, *J* = 10.2 Hz, 1H), 3.39 (d, *J* = 8.4 Hz, 1H), 2.97-2.96 (m, 1H), 1.93 (s, 3H, -C(O)CH₃); ¹³C NMR (150 MHz, CDCl₃): δ 169.9, 138.69, 138.6, 138.5, 137.9, 137.9, 134., 133.8, 133.7, 131.9, 130.3, 128.5, 128.4, 128.4, 128.3, 128.2, 128.0, 127.9, 127.7, 127.7, 127.6, 127.5, 127.5, 123.3, 117.5, 97.1, 96.4, 77.9, 76.9, 75.1, 74.8, 73.9, 73.8, 73.8, 73.05, 72.8, 71.9, 71.0, 70.4, 70.1, 68.1, 55.5, 28.7, 21.5, 21.1; ESI-MS: *m/z* calcd for C₆₀H₆₁NO₁₃; 1003.4035 found 1026.4043 (*M* + Na)⁺.

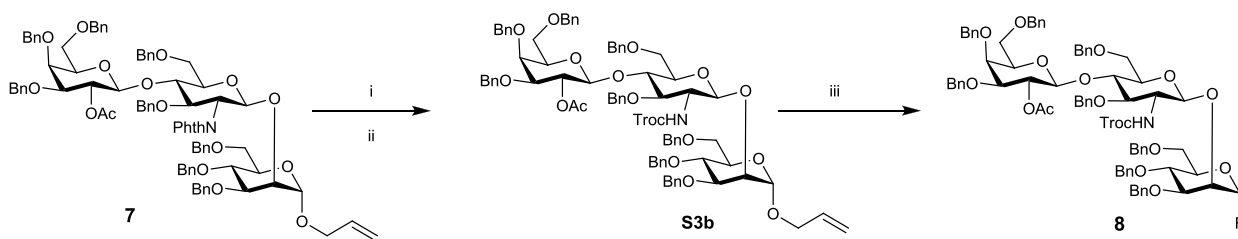


4-O-acetyl-3,6-O-di-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl-(1 \rightarrow 2)-O-3,4,6-tri-O-benzyl- α -D-mannopyranosyl fluoride (6): To a solution of **S2c** (0.750 g, 0.747 mmol) in 10 mL CH_2Cl_2 : MeOH (1/1) was added PdCl_2 (0.02 g) and stirred at rt for overnight until TLC (ethyl acetate: toluene, 2/8) indicated formation of a product with consumption of the starting material. The reaction mixture was then concentrated in *vacuo*, and the residue was purified by flash column chromatography to afford alcohol (0.6 g, 80%) as colorless foam. To a solution of alcohol (0.270 g, 0.197 mmol) in CH_2Cl_2 (10 mL) at $-30\text{ }^\circ\text{C}$ was DAST (52 μL , 0.395 mmol) and the resulting reaction mixture was stirred at $-10\text{ }^\circ\text{C}$ for 8 h until TLC indicated formation of product with consumption of starting material. The reaction was quenched with aq. NaHCO_3 , and the filtrate was washed with aqueous NaHCO_3 (2 x 50 mL) and a brine (50 mL) solution. The organic layer was dried over Na_2SO_4 and concentrated in *vacuo*. The residue was purified by silica gel column chromatography (0% \rightarrow 25% EA in hexane) to afford **6** (0.150 g, 61%) as white foam. TLC (ethyl acetate: hexane = 3/7, v/v): $R_f = 0.45$; $^1\text{H NMR}$ (600 MHz, CDCl_3): δ 7.57-7.55 (m, 4H, Ar-H), 7.35-7.21 (m, 11H, Ar-H), 7.07 (d, $J = 6.4$ Hz, 4H, Ar-H), 6.99 (d, $J = 7.2$ Hz, 2H, Ar-H), 6.88-6.82 (m, 3H, Ar-H), 5.30 (d, $J = 50.4$ Hz, 1H, H-1^{Man}), 5.30 (d, $J = 12.2$ Hz, 1H, H-1^{GlcNAc}), 4.88 (d, $J = 11.2$ Hz, 1H), 4.80-4.71 (m, 3H), 4.64 (d, $J = 11.3$ Hz, 1H), 4.49-4.46 (m, 2H), 4.43 (d, $J = 11.8$ Hz, 2H), 4.34-4.28 (m, 3H), 4.23 (dd, $J = 6.1, 11.3$ Hz, 1H), 4.16 (s, 1H), 4.06 (d, $J = 12.1$ Hz, 1H), 4.02 (d, $J = 12.3$ Hz, 1H), 3.76-3.57 (m, 5H), 3.32 (d, $J = 10.8$

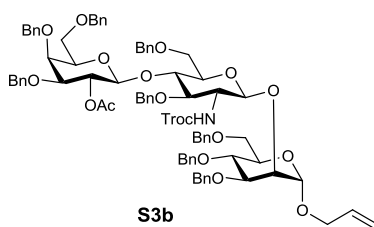
Hz, 1H), 3.02 (dd, $J = 4.2, 11.3$ Hz, 1H), 1.97 (s, 3H, $-\text{C}(\text{O})\text{CH}_3$); ^{13}C NMR (150 MHz, CDCl_3): δ 170.99, 138.40, 138.28, 138.16, 137.94, 137.70, 133.94, 131.82, 128.90, 128.63, 128.57, 128.47, 128.40, 128.31, 128.27, 128.04, 127.89, 127.89, 127.76, 123.54, 106.33, 104.85, 97.45, 79.55, 78.48, 75.43, 75.23, 74.09, 73.84, 73.70, 73.16, 72.78, 72.54, 71.36, 69.18, 63.40, 55.80, 31.21, 29.98, 21.11; ESI-MS: m/z calcd for $\text{C}_{57}\text{H}_{56}\text{FNO}_{12}$; 965.3679 found 988.3690 ($M + \text{Na}$) $^+$.

Synthesis of trisaccharide building blocks 7 and 8.

Building block **7** (Figure 2a) was obtained according to previous report³. The N-phthalamide protection at Glucosamine residues was modified to NH-Troc to prepare building block **8** (Scheme S3).

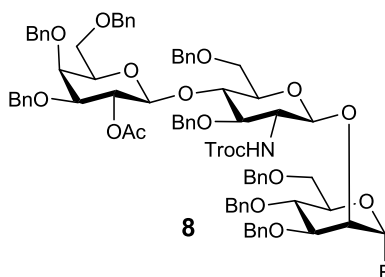


Scheme S3 | Preparation of compound 8. **i**, (1) EDA, n-BuOH, 90 °C, (2) Troc-Cl, NaHCO_3 , CH_2Cl_2 , **ii**, Ac_2O , pyridine, RT, overnight, 72% over 3 steps; **iii**, (1) PdCl_2 , $\text{MeOH}:\text{CH}_2\text{Cl}_2$, (2) DAST, CH_2Cl_2 , -30 °C, 66% over 2 steps.



Allyl-O-2-O-acetyl-3,4,6-O-tri-benzyl- β -D-galactopyranosyl-(1 \rightarrow 4)-O-3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino- β -D-glucopyranosyl-(1 \rightarrow 2)-O-3,4,6-tri-O-

benzyl- α -D-mannopyranoside (S3b): A mixture of compound **S3a** (1 g, 0.693 mmole) and 10 mL of ethylene diamine: *n*-BuOH (1:4) was stirred at 90 °C overnight. Volatiles were evaporated, and the crude product was dried using high vacuum. It was then dissolved in CH₂Cl₂ (20 mL), NaHCO₃ (0.376 g, 6.93 mmol) and 2,2,2-trichloro ethyl chloroformate (0.665 mL, 6.93 mmol) were added at 0 °C, allowed it to warm to rt and stirred for overnight. TLC (ethyl acetate: toluene, 2/8) indicated formation of product with consumption of starting material. The reaction mixture was diluted with CH₂Cl₂ (100 mL), washed with water (2 x 50 mL) and brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by silica gel column chromatography (0% → 15% EA in toluene). The product was then acetylated using 10 mL of pyridine/acetic anhydride (6:4) until TLC indicated (ethyl acetate: toluene, 2/8) complete consumption of starting material. The reaction mixture was then concentrated *in vacuo* and purified by silica gel column chromatography to afford **S3b** (0.760 g, 72%) as a white foam. TLC (ethyl acetate: toluene = 2/8, v/v): R_f = 0.64; ¹H NMR (600 MHz, CDCl₃): δ 7.34-7.13 (m, 40H, -Ph), 5.86-5.77 (m, 1H, allyl -CH), 5.30-5.28 (t, *J* = 8.2 Hz, 1H, H^{2gal}), 5.25 (bd, 1H, -NH*Troc*), 5.16 (d, *J* = 17.6 Hz, 1H, *Troc*), 5.10 (d, *J* = 10.2 Hz, 1H), 4.90 (t, 3H), 4.82-4.76 (m, 3H), 4.74 (s, 1H), 4.65-4.61 (dd, *J* = 8.4 & 2.8 Hz, 3H), 4.55-4.41 (m, 8H), 4.30 (d, *J* = 12.3 Hz, 2H), 4.20 (d, *J* = 12.2 Hz, 2H), 4.10-4.01 (m, 3H), 3.90-3.84 (m, 5H), 3.76-3.61 (m, 5H), 3.55-3.40 (m, 3H), 3.35-3.30 (m, 2H), 3.13 (d, *J* = 7.2 Hz, 1H), 1.92 (s, 3H, -C(O)CH₃); ¹³C NMR (150 MHz, CDCl₃): δ 170.23, 154.11, 139.17, 138.98, 138.75, 138.25, 138.20, 134.04, 127.41, 100.7, 97.5, 96.80, 80.55, 78.53, 75.36, 72.87, 73.55, 72.78, 72.49, 72.10, 71.96, 69.98, 68.90, 68.65, 57.64, 41.51, 21.30. ESI-MS: *m/z* calcd for C₈₂H₈₈Cl₃N₁O₁₈; 1458.4960 found 1504.4956 (*M* + Na)⁺

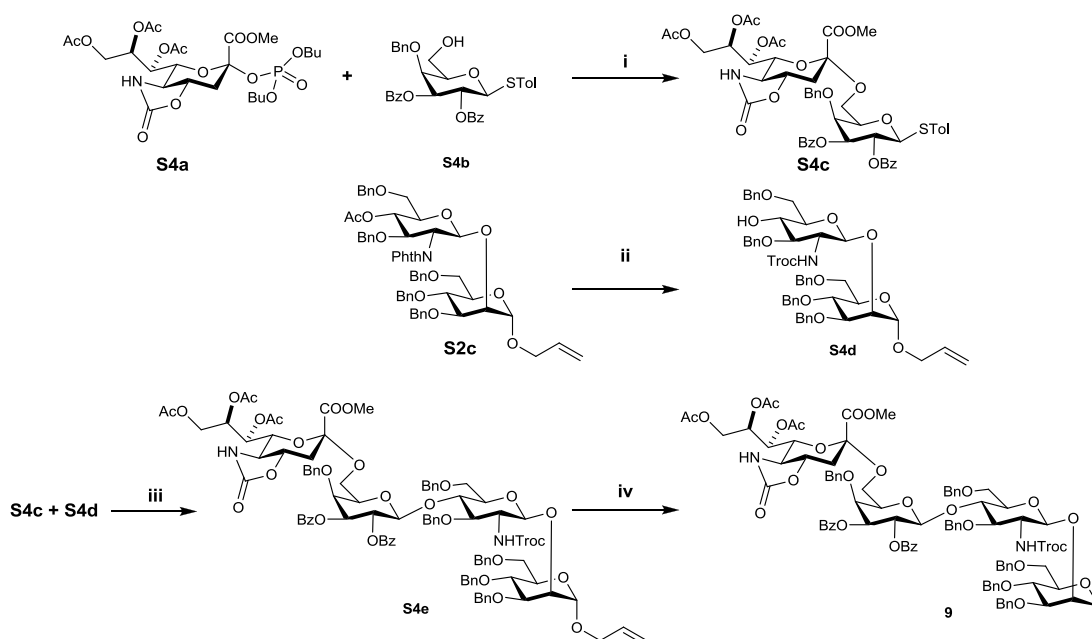


2-O-acetyl-3,4,6-O-tri-benzyl- β -D-galactopyranosyl-(1 \rightarrow 4)-O-3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino- β -D-glucopyranosyl-(1 \rightarrow 2)-O-3,4,6-tri-O-benzyl- α -D-mannopyranosyl fluoride (8**):** PdCl₂ (0.030 g) was added to a solution of **S3b** (1.3 g, 0.942 mmol) in 10 mL of CH₂Cl₂: MeOH (1:1). The reaction mixture was stirred at room temperature for 2 h until TLC (ethyl acetate: toluene, 2/8) indicated formation of product with consumption of the starting material. The reaction mixture was then filtered through Celite and concentrated in *vacuo*. The residue was purified by flash column chromatography to afford 1-OH compound (0.980 g) as white color foam. The residue (0.850 g, 0.608 mmol) was dissolved in CH₂Cl₂ (10 mL) at -30 °C, then DAST (160 μ L, 1.21 mmol) was added slowly. The resulting reaction mixture was stirred for 1 h. When TLC (ethyl acetate: toluene, 2/8) indicated formation of product with consumption of starting material, the reaction was quenched with aq. NaHCO₃. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL) and brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by silica gel column chromatography (0% \rightarrow 15% EA in toluene) to afford **8** (0.700 g, 66% over 2 steps) as white foam. TLC (ethyl acetate: toluene = 2/8, v/v): R_f = 0.64; ¹H NMR (600 MHz, CDCl₃): δ 7.35-7.10 (m, 40H), 4.57 (d, *J* = 51.4 Hz, 1H), 5.33 (t, *J* = 10.2 Hz, 1H), 5.22 (d, *J* = 8.2 Hz, 1H), 4.99-4.91 (m, 3H), 4.82 (d, *J* = 7.8 Hz, 1H), 4.75 (t, *J* = 10.3 Hz, 1H), 4.64-4.44 (m, 12H), 4.38 (dd, *J* = 3.4 & 10.2 Hz, 2H), 4.26 (s, 2H), 4.06-4.33 (m, 5H), 3.76-3.64 (m, 4H), 3.54-3.46 (m, 2H), 3.40-3.34 (m, 3H), 3.12 (d, *J* = 8.4 Hz, 1H), 1.65 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 169.5, 154.0, 139.0, 138.8, 138.7, 138.5, 138.4, 138.3, 138.1, 129.4, 128.7, 128.6, 128.4, 128.2, 128.0, 127.9, 127.7, 127.5, 127.4, 127.2, 127.1, 125.5, 107.6, 105.4, 100.7, 98.7, 95.8, 80.5, 77.4,

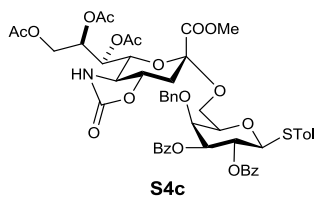
74.3, 73.6, 73.3, 73.1, 72.3, 72.1, 72.0, 71.9, 68.9, 68.7, 68.3, 57.4, 41.4, 21.7, 21.2; ESI-MS: m/z calcd for $C_{79}H_{83}Cl_3FN_1O_{17}$; 1443.4602 found 1466.4608 ($M + Na$)⁺.

Synthesis of complex type D1 arm tetrasaccharide **9**.

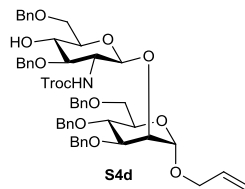
Preparation of sialylated D1 arm was achieved through coupling of two main building units **S4c** and **S4d**. Use of sialyl phosphate donor **S4a** for the α -2,6 glycosylation of Gal **S4b** resulted in complete α -selectivity¹⁰. The N-Phth protection at glucosamine of **S2c** was modified NHTroc, while doing so, the 4-OAc group was removed to afford the desired **S4d**. At last the coupling of **S4c** and **S4d** afforded the desired tetrasaccharide, which further underwent anomeric modification to get donor **9**.



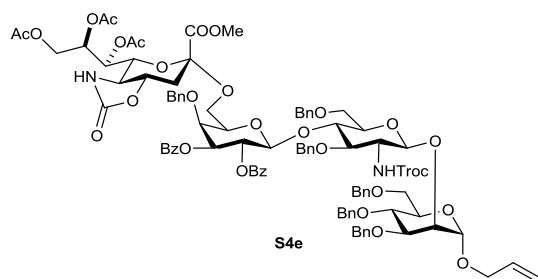
Scheme S4 | Preparation of compound **9.** **i**, TMSOTf, CH_2Cl_2 , $-50\text{ }^\circ C$, 64%; **ii**, (1) EDA, n -BuOH, $90\text{ }^\circ C$, (2) Troc-Cl, $NaHCO_3$, CH_2Cl_2 , 78% over 2 steps; **iii**, NIS, TfOH, CH_2Cl_2 , $-50\text{ }^\circ C$, 65%; **iv**, (1) $PdCl_2$, MeOH: CH_2Cl_2 , (2) DAST, CH_2Cl_2 , $-30\text{ }^\circ C$, 55% over 2 steps.



Methyl-5-acetamido-7,8,9-tri-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-non-2-uloypyranosylonate-(2 \rightarrow 6)-*p*-tolyl-4-O-benzyl-2,3-di-O-benzoyl-1-thio- β -D-glactopyranoside (S4c): A mixture of donor **S4a** (0.113 g, 0.178 mmol), acceptor **S4b** (0.220 g, 0.119 mmol) and activated 4 Å molecular sieves in dry CH₂Cl₂ (10 mL) was stirred at rt for 1 h. The reaction was cooled to -50 °C, trimethylsilyl triflate (12 μ L, 0.06 mmol) was added slowly and the resulting reaction mixture was stirred for 2 h. The reaction was quenched by adding Et₃N, diluted with CH₂Cl₂, filtered through Celite, extracted with saturated NaHCO₃, dried over sodium sulfate and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 25% EA in hexane) to afford **S4c** (0.190 g, 64%) as colorless foam. TLC: (ethyl acetate: hexane = 3/7, v/v): R_f = 0.46; ¹H NMR (600 MHz, CDCl₃): δ 7.93 (d, *J* = 7.8 Hz, 2H), 7.89 (d, *J* = 8.4 Hz, 2H), 7.50-7.45 (m, 2H), 7.42-7.30 (m, 6H), 7.24-7.13 (m, 5H), 7.04 (d, *J* = 8.2 Hz, 2H), 5.80 (t, *J* = 10.3 Hz, 1H), 5.47 (d, *J* = 8.9 Hz, 1H), 5.37 (s, 3H), 5.32 (dd, *J* = 4.2 & 7.8 Hz, 1H), 5.12 (dd, *J* = 4.1 & 7.2 Hz, 1H), 4.90 (d, *J* = 12.2 Hz, 1H), 4.65 (d, *J* = 12.1 Hz, 1H), 4.20 (dd, *J* = 3.2 & 7.8 Hz, 1H), 4.18 (d, *J* = 6.8 Hz, 1H), 3.94-3.91 (m, 3H), 3.74-3.72 (m, 3H), 3.71 (s, 3H), 3.65 (dd, *J* = 3.2 & 7.8 Hz, 1H), 3.06 (t, *J* = 10.2 Hz, 2H), 2.87 (dd, *J* = 3.6 & 8.6 Hz, 1H), 2.33 (s, 3H), 2.17 (s, 3H), 2.15 (t, *J* = 7.8 Hz, 1H), 2.13 (s, 3H), 2.10 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 171.9, 170.8, 169.8, 168.2, 166.8, 165.2, 159.2, 138.1, 133.7, 133.3, 130.0, 129.8, 129.4, 129.3, 129.2, 128.7, 128.6, 128.5, 128.4, 127.8, 127.7, 125.5, 100.5, 86.8, 76.8, 76.1, 74.7, 73.8, 68.9, 68.5, 68.3, 67.2, 63.7, 62.0, 58.1, 53.3, 37.7, 25.88, 21.7, 21.5, 21.3, 21.0, 20.9; ESI-MS: *m/z* calcd for C₅₁H₅₃NO₁₈S; 999.2876 found 1022.2882 (*M* + Na)⁺.

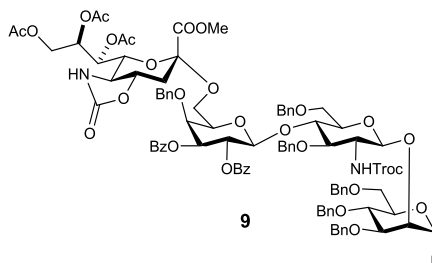


Allyl-O-3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino- β -D-glucopyranosyl-(1 \rightarrow 2)-O-3,4,6-tri-O-benzyl- α -D-mannopyranoside (S4d): A mixture of compound **S2c** (2 g, 1.99 mmole) and 20 mL of ethylene diamine: *n*-BuOH (1:4) was stirred at 90 °C overnight. Volatiles were evaporated, and the crude product was dried using high vacuum. It was then dissolved in CH₂Cl₂ (20 mL), NaHCO₃ (1.05 g, 19.9 mmol) and 2,2,2-trichloro ethyl chloroformate (1.9 mL, 19.9 mmol) were added at 0 °C, allowed it to warm to rt and stirred for overnight until TLC (ethyl acetate: toluene, 1.5/8.5) indicated formation of product with consumption of starting material. The reaction mixture was diluted with CH₂Cl₂ (100 mL), washed with water (2 x 50 mL) and brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by silica gel column chromatography (0% \rightarrow 10% EA in toluene) to afford **S4d** (1.5 g, 78%) as a white foam. TLC (ethyl acetate: toluene = 1.5/8.5, v/v): R_f = 0.44; ¹H NMR (600 MHz, CDCl₃): δ 7.30-7.15 (m, 25H), 5.87-7.78 (m, 1H), 5.33 (d, *J* = 5.2 Hz, 1H), 5.21 (d, *J* = 3.2 Hz, 1H), 5.14 (d, *J* = 7.2 Hz, 1H), 4.94 (d, *J* = 5.8 Hz, 1H), 4.80 (d, *J* = 8.5 Hz, 1H), 4.73-4.44 (m, 10H), 4.12-4.06 (m, 4H), 3.97-3.86 (m, 4H), 3.78-3.49 (m, 7H); ¹³C NMR (150 MHz, CDCl₃): δ 153.97, 138.45, 138.37, 138.28, 137.98, 137.45, 133.63, 128.94, 128.43, 128.40, 128.26, 128.16, 127.97, 127.91, 127.78, 127.65, 127.53, 127.41, 125.20, 117.23, 97.93, 96.88, 95.50, 79.33, 79.08, 75.08, 74.47, 74.37, 73.53, 73.19, 73.00, 71.98, 71.65, 70.83, 69.16, 67.91, 57.41, 44.53; ESI-MS: *m/z* calcd for C₅₃H₅₈Cl₃NO₁₂; 1005.3025 found 1006.3089 (*M* + H)⁺.



Allyl-[Methyl-5-acetamido-7,8,9-tri-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-non-2-ulopyranosylonate]-(2 \rightarrow 6)-4-O-benzyl-2,3-di-O-benzoyl-1- β -D-galactopyranosyl-(1 \rightarrow 4)-O-3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino- β -D-glucopyranosyl-(1 \rightarrow 2)-O-3,4,6-tri-O-benzyl- α -D-mannopyranoside (S4e): Activated 4 Å molecular sieves were added to a solution of acceptor **S4d** (0.208 g, 0.169 mmol) and donor **S4c** (0.310 g, 0.199 mmol) in anhydrous CH₂Cl₂ (10 mL). The mixture was stirred for 1 h at room temperature. The reaction mixture was cooled to -50 °C, NIS (0.076 g, 0.338 mmol) and TfOH (3.7 μ L, 0.042 mmol) were added slowly. The resulting reaction mixture was stirred for 2 h. When TLC (ethyl acetate: toluene, 1.5/8.5) indicated formation of product with consumption of starting material, the reaction was quenched by adding Et₃N and filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), aqueous Na₂S₂O₃ (2 x 50 mL), and brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 10% EA in toluene) to afford **S4e** (0.390 g, 65%) as clear foam. TLC (ethyl acetate: toluene = 1.5/8.5, v/v): R_f = 0.51; ¹H NMR (600 MHz, CDCl₃): δ 7.91-7.85 (m, 4H, Ph), 7.50-7.46 (m, 1H, Ph), 7.35-7.19 (m, 35H), 5.82-5.76 (m, 2H), 5.39-5.36 (m, 3H), 5.24 (d, *J* = 6.4 Hz, 1H), 5.22 (d, *J* = 7.2 Hz, 1H), 5.19 (d, *J* = 3.4 Hz, 1H), 5.15 (d, *J* = 3.4 Hz, 1H), 5.11-5.08 (dd, *J* = 6.3 & 3.1 Hz, 2H), 4.96 (d, *J* = 12.1 Hz, 2H), 4.83 (d, *J* = 12.1 Hz, 2H), 4.73-4.57 (m, 10H), 4.47-4.42 (m, 6H), 4.25-4.15 (m, 5H), 4.08-3.93 (m, 9H), 3.91 (s, 3H), 3.39 (d, *J* = 7.8 Hz, 1H), 3.21 (bd, 1H), 2.93 (t, *J* = 7.8 Hz, 1H), 2.78 (dd, *J* = 3.6 & 7.8 Hz, 1H), 2.11 (s, 3H), 2.01 (s, 3H), 1.66 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 171.83, 170.84, 169.91, 168.03, 166.15, 165.43, 159.57, 154.17, 139.14, 138.85, 138.36, 134.08, 133.78, 132.66, 13.54,

129.67, 128.78, 128.43, 128.23, 127.78, 127.67, 127.56, 127.43, 127.23, 127.10, 117.53, 100.67, 100.32, 98.94, 97.18, 75.90, 78.31, 75.32, 74.95, 7.81, 74.43, 73.99, 73.58, 73.45, 73.33, 73.29, 73.23, 73.12, 69.90, 69.56, 68.66, 67.56, 58.70, 53.43, 21.28, 21.07, 20.98; ESI-MS: m/z calcd for $C_{97}H_{103}Cl_3N_2O_{13}$; 1982.5562 found 1905.5558 ($M + Na$)⁺.

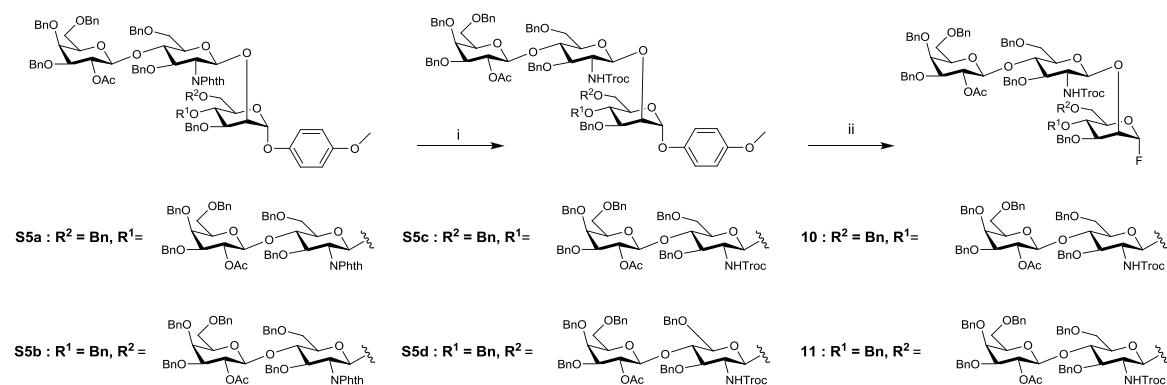


**Methyl-5-acetamido-7,8,9-tri-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-non-2-
ulopyranosylonate-(2 \rightarrow 6)-4-O-benzyl-2,3-di-O-benzoyl-1- β -D-galactopyranosyl-(1 \rightarrow 4)-O-
3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino- β -D-glucopyranosyl-
(1 \rightarrow 2)-O-3,4,6-tri-O-benzyl- α -D-mannopyranosyl fluoride (9):** PdCl₂ (0.050 g) was added to a solution of **S4e** (2.7 g, 1.42 mmol) in 20 mL of CH₂Cl₂: MeOH (1:1). The reaction mixture was stirred at room temperature for 5 h until TLC (ethyl acetate: toluene, 2/8) indicated formation of product with consumption of the starting material. The reaction mixture was then filtered through Celite and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 15% EA in toluene) to afford 1-OH compound (2.0 g) as white color foam. The residue (2 g, 1.07 mmol) was dissolved in CH₂Cl₂ (10 mL) at -30 °C, then DAST (284 μ L, 2.14 mmol) was added slowly. The resulting reaction mixture was stirred for 1 h. When TLC (ethyl acetate: toluene, 2/8) indicated formation of product with consumption of starting material, the reaction was quenched with aq. NaHCO₃. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL) and brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by silica gel column chromatography (0% \rightarrow 15% EA in toluene) to afford **9** (1.45 g, 55% over 2 steps) as white foam. TLC (ethyl acetate: toluene = 2/8, v/v): R_f = 0.64; ¹H NMR (600 MHz, CDCl₃): δ 7.96 (m, 4H), 7.55 (m, 2H), 7.40-7.17 (m, 34H), 5.86 (t, J =

10.2Hz, 1H), 5.50 (d, $J = 52$ Hz, 1H), 5.44-5.40 (m, 2H), 5.29 (dd, $J = 1.6$ & 7.8 Hz, 1H), 5.15 (dd, $J = 1.7$ & 7.2 Hz, 1H), 5.05-4.99 (m, 2H), 4.84-4.44 (m, 10H), 4.33-4.20 (m, 3H), 4.27-4.20 (m, 4H), 4.03-3.90 (m, 3H), 3.72-3.65 (s, 3H), 3.70-3.06 (m, 12H), 3.46-3.30 (m, 1H), 3.00 (t, $J = 10.2$ Hz, 1H), 2.86 (dd, $J = 3.2$ & 12.3 Hz, 1H), 2.19 (s, 3H), 2.16 (s, 3H), 2.12 (t, $J = 8.4$ Hz, 1H), 2.0 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3): δ 170.8, 170.0, 168.0, 166.1, 165.2, 159.5, 154.1, 138.9, 138.4, 138.2, 138.1, 133.7, 133.6, 133.4, 130.0, 129.9, 129.6, 129.3, 128.8, 128.6, 128.5, 128.4, 128.3, 128.1, 127.9, 127.5, 127.2, 125.5, 107.7, 100.7, 100.1, 99.2, 95.8, 78.1, 77.1, 76.6, 75.3, 74.9, 74.6, 74.4, 74.1, 73.9, 73.7, 73.5, 73.3, 73.0, 72.6, 72.5, 69.2, 69.0, 67.3, 63.2, 61.8, 58.1, 57.3, 53.4, 37.3, 21.7, 21.3, 21.0, 20.9 ; ESI-MS: m/z calcd for $\text{C}_{94}\text{H}_{98}\text{Cl}_3\text{FN}_2\text{O}_{29}$; 1844.5204 found 1867.5192 ($M + \text{Na}$) $^+$.

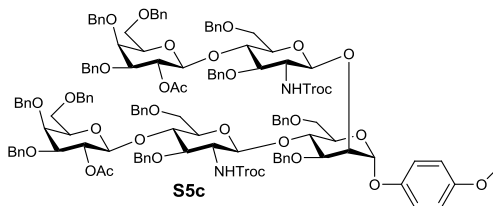
Synthesis of building blocks 10 and 11.

Pentasaccharides **S5a** and **S5b** were obtained according to our previous report³, in which the functional group was further modified from N-phthalamide to N-Troc. Finally the anomeric *p*-methoxy phenyl group was removed and resulting -OH was changed to fluoride in presence of DAST.



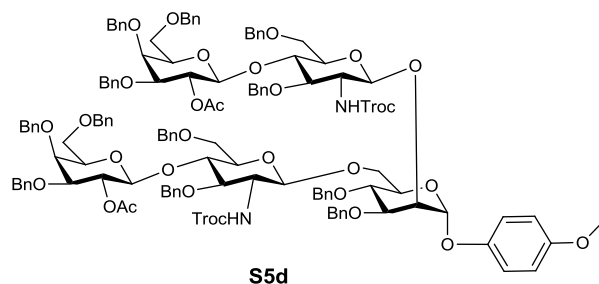
Scheme S5 | Preparation of compound 10 and 11. i, (1) EDA, n-BuOH, 90 °C; (2) Troc-Cl, NaHCO₃, CH₂Cl₂; (3) Ac₂O, pyridine; ii, (1) CAN, ACN: Toluene: H₂O; (2) DAST, CH₂Cl₂, -30 °C.

General procedure for step i: A mixture of compound **S5a** and **S5b** (0.500 g, 0.212 mmol) and 10 mL of ethylene diamine: *n*-BuOH (1:4) was stirred at 90 °C overnight. Volatiles were evaporated, and the crude product was dried using high vacuum. It was then dissolved in CH₂Cl₂ (20 mL), NaHCO₃ (0.114 g, 2.12 mmol) and 2,2,2-trichloro ethyl chloroformate (0.2 mL, 2.12 mmol) were added at 0 °C, allowed it to warm to RT and stirred for overnight. TLC (ethyl acetate: toluene, 2/8) indicated formation of product with consumption of starting material. The reaction mixture was diluted with CH₂Cl₂ (100 mL), washed with water (2 x 50 mL) and brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by silica gel column chromatography (0% → 15% EA in toluene). The product was then acetylated using 10 mL of pyridine/acetic anhydride (6:4) until TLC indicated (ethyl acetate: toluene, 2/8) complete consumption of starting material. The reaction mixture was then concentrated *in vacuo* and purified by silica gel column chromatography (0% → 15% EA in toluene) to afford **S5c** (0.332 g 65%) and **S5d** (0.340 g, 68%) as a colorless foam.



***p*-methoxyphenyl-O-2-O-acetyl-3,4,6-O-tri-benzyl- β -D-galactopyranosyl-(1→4)-O-3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino- β -D-glucopyranosyl-(1→2)-[2-O-acetyl-3,4,6-O-tri-benzyl- β -D-galactopyranosyl-(1→4)-O-3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino- β -D-glucopyranosyl-(1→4)]-3,6-O-di-benzyl- α -D-mannopyranoside (**S5c**):** Compounds **S5c** was prepared according to the above mentioned general procedure. TLC (ethyl acetate: toluene = 2/8, v/v): R_f = 0.64; ¹H NMR (600 MHz, CDCl₃): δ 7.40-7.08 (m, 50H, Ar-H), 6.92-6.72 (m, 10H, Ar-H), 6.63-6.53 (m, 2H, PMP-H), 5.31-5.24 (m, 2H), 5.22 (d, *J* = 7.8 Hz, 1H), 5.15 (d, *J* = 8.4 Hz, 1H), 4.94 (d, *J* = 3 Hz, 2H),

4.88 (d, $J = 3.6$ Hz, 1H), 4.86 (d, $J = 3.6$ Hz, 1H), 4.80-4.71 (m, 7H), 4.64-4.56 (m, 3H), 4.49-4.44 (m, 10H), 4.39-4.10 (m, 14H), 4.15-4.05 (m, 2H), 4.03 (t, $J = 8.4$ Hz, 1H), 3.98-3.86 (m, 3H), 3.79-3.72 (m, 1H), 3.69 (s, 3H), 3.58-3.56 (m, 1H), 3.49-3.25 (m, 10H), 3.08 (d, $J = 8.4$ Hz, 1H), 3.05 (d, $J = 8.9$ Hz, 1H), 2.90 (bs, 1H), 1.93 (s, 3H), 1.92 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3): δ 169.5, 169.4, 168.3, 167.8, 154.8, 150.3, 139.2, 139.1, 139.0, 138.9, 138.8, 138.5, 138.3, 138.2, 138.0, 134.0, 133.6, 132.1, 131.9, 131.7, 129.2, 128.6, 128.5, 128.3, 128.2, 128.1, 128.0, 127.9, 127.9, 127.8, 127.7, 127.6, 127.5, 127.4, 127.4, 127.3, 127.1, 127.0, 126.8, 123.5, 123.4, 123.3, 118.1, 114.4, 108.9, 100.9, 100.7, 98.5, 97.6, 97.1, 80.5, 50.5, 79.7, 77.9, 77.8, 77.7, 77.4, 77.2, 75.4, 74.9, 74.6, 74.5, 73.7, 73.6, 73.6, 73.4, 73.3, 73.0, 72.8, 72.7, 72.2, 71.9, 71.8, 71.7, 71.2, 69.2, 68.5, 68.3, 68.2, 67.6, 67.3, 56.4, 55.9, 55.7, 21.3, 21.2, 21.2; ESI-MS: m/z calcd for $\text{C}_{131}\text{H}_{138}\text{Cl}_6\text{N}_2\text{O}_{31}$; 2449.7492; found 2449.7272 ($M + \text{H}$) $^+$.

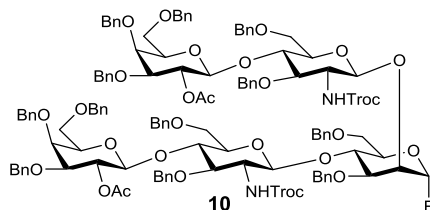


***p*-methoxyphenyl-O-2-O-acetyl-3,4,6-O-tri-benzyl- β -D-galactopyranosyl-(1 \rightarrow 4)-O-3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino- β -D-glucopyranosyl-(1 \rightarrow 2)-[2-O-acetyl-3,4,6-O-tri-benzyl- β -D-galactopyranosyl-(1 \rightarrow 4)-O-3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino- β -D-glucopyranosyl-(1 \rightarrow 6)]-3,6-O-di-benzyl- α -D-**

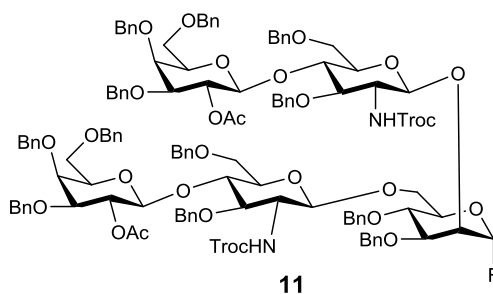
mannopyranoside (S5d): Compounds **S5d** was prepared according to the above mentioned general procedure. TLC (ethyl acetate: toluene = 2/8, v/v): $R_f = 0.61$; ^1H NMR (600 MHz, CDCl_3): δ 7.42-7.05 (m, 60H), 7.02 (d, $J = 9.2$ Hz, 2H), 6.85 (d, $J = 9.0$ Hz, 2H), 5.59 (s, 1H), 5.42 (bs, 1H), 5.39-5.29 (m, 2H), 5.15 (d, $J = 3.8$ Hz, 1H), 5.00-4.84 (m, 7H), 4.80-4.25 (m,

20H), 4.10 (dd, $J = 3.8$ & 7.8 Hz, 2H), 4.00-3.83 (m, 6H), 3.76 (s, 3H), 3.62-3.20 (m, 23H), 1.99 (s, 3H), 1.96 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3): δ 169.9, 169.5, 168.5, 155.0, 154.6, 150.1, 138.9, 138.4, 138.2, 138.1, 134.3, 132.0, 129.2, 128.6, 128.4, 128.3, 128.2, 128.1, 128.0, 127.9, 127.8, 127.6, 127.5, 127.3, 125.5, 123.6, 117.8, 117.2, 114.9, 106.1, 101.5, 100.9, 100.7, 98.3, 96.4, 95.7, 80.3, 80.1, 79.4, 78.3, 77.9, 78.3, 75.6, 74.8, 74.5, 74.4, 74.3, 73.6, 73.4, 73.2, 73.0, 72.6, 72.3, 71.8, 68.9, 68.1, 67.9, 67.5, 57.9, 56.7, 55.8, 41.3, 40.4, 37.7, 29.9, 21.6, 21.2, 21.1; ESI-MS: m/z calcd for $\text{C}_{131}\text{H}_{138}\text{Cl}_6\text{N}_2\text{O}_{31}$; 2448.7492 found 2449.7414 ($M + \text{H}$)⁺.

General procedure for step ii: Cerium ammonium nitrate (0.616 g, 0.725 mmol) was added to a solution of compound **S5c** or **S5d** (0.350 g, 0.145 mmol) in 10 mL of acetonitrile: toluene: H_2O (4:2:1). The resulting reaction mixture was stirred at 0 °C for 2 h. The reaction was diluted with EtOAc (100 mL) and washed with H_2O (30 x 2 mL) and brine (30 mL). The organic layer was dried over Na_2SO_4 and concentrated in *vacuo*. The product was purified by flash column chromatography (0% → 15% EA in toluene) to afford **1-OH** compound (0.180 g) as clear foam. The residue (0.180 g, 0.078 mmol) was dissolved in CH_2Cl_2 (10 mL) at -30 °C. Then, DAST (30 μL , 0.234 mmol) was added slowly, and the resulting reaction mixture was stirred for 1 h. When TLC (ethyl acetate: toluene, 1/9) indicated formation of product with consumption of starting material, the reaction was quenched with aq. NaHCO_3 . The filtrate was washed with aqueous NaHCO_3 (2 x 50 mL) and brine (50 mL) solution. The organic layer was dried over Na_2SO_4 and concentrated in *vacuo*. The residue was purified by silica gel column chromatography (0% → 8% EA in toluene) to afford **10** (0.120 g, 34% over 2 steps) and **11** (0.150 g, 42 % over 2 steps).



2-O-acetyl-3,4,6-O-tri-benzyl-β-D-galactopyranosyl-(1→4)-O-3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino-β-D-glucopyranosyl-(1→2)-[2-O-acetyl-3,4,6-O-tri-benzyl-β-D-galacto-pyranosyl-(1→4)-O-3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy) carbonylamino-β-D-glucopyranosyl-(1→4)]-3,6-O-di-benzyl-α-D-mannopyranosyl fluoride (10): TLC (ethyl acetate: toluene =1/9, v/v): $R_f = 0.34$; $^1\text{H NMR}$ (600 MHz, CDCl_3): δ 7.34-7.13 (m, 60H), 5.41 (s, 1H), 5.50 (d, $J = 51.1\text{Hz}$, 1H), 5.30-5.25 (m, 2H), 5.12 (d, $J = 8.4\text{ Hz}$, 1H), 4.92-4.85 (m, 5H), 4.69-4.38 (m, 20H), 4.32-4.14 (m, 8H), 3.91-3.77 (m, 7H), 3.70 (dd, $J = 3.8$ and 7.8 Hz , 1H), 3.60-3.56 (m, 3H), 3.49-3.24 (m, 14H), 2.03 (s, 3H), 1.98 (s, 3H); $^{13}\text{C NMR}$ (150 MHz, CHCl_3): δ 169.68, 169.58, 154.31, 154.09, 139.06, 138.97, 138.93, 138.32, 138.29, 138.27, 138.20, 138.17, 129.32, 128.99, 128.69, 128.63, 128.50, 128.49, 128.45, 128.27, 128.24, 128.16, 128.15, 128.07, 128.05, 128.00, 127.97, 127.94, 127.91, 127.82, 127.72, 127.60, 127.57, 127.44, 125.58, 100.64, 100.45, 95.86, 80.54, 80.51, 76.81, 76.70, 76.03, 75.34, 75.08, 86, 74.77, 74.66, 74.57, 74.47, 73.74, 73.70, 73.67, 73.64, 73.58, 73.46, 73.42, 73.27, 72.90, 72.19, 72.17, 71.99, 71.95, 68.49, 68.27, 68.14, 68.01, 57.53, 21.40, 21.27; ESI-MS: m/z calcd for $\text{C}_{124}\text{H}_{131}\text{Cl}_6\text{FN}_2\text{O}_{29}$; 2340.6953 found 2363.7200 ($M + \text{Na}$) $^+$.

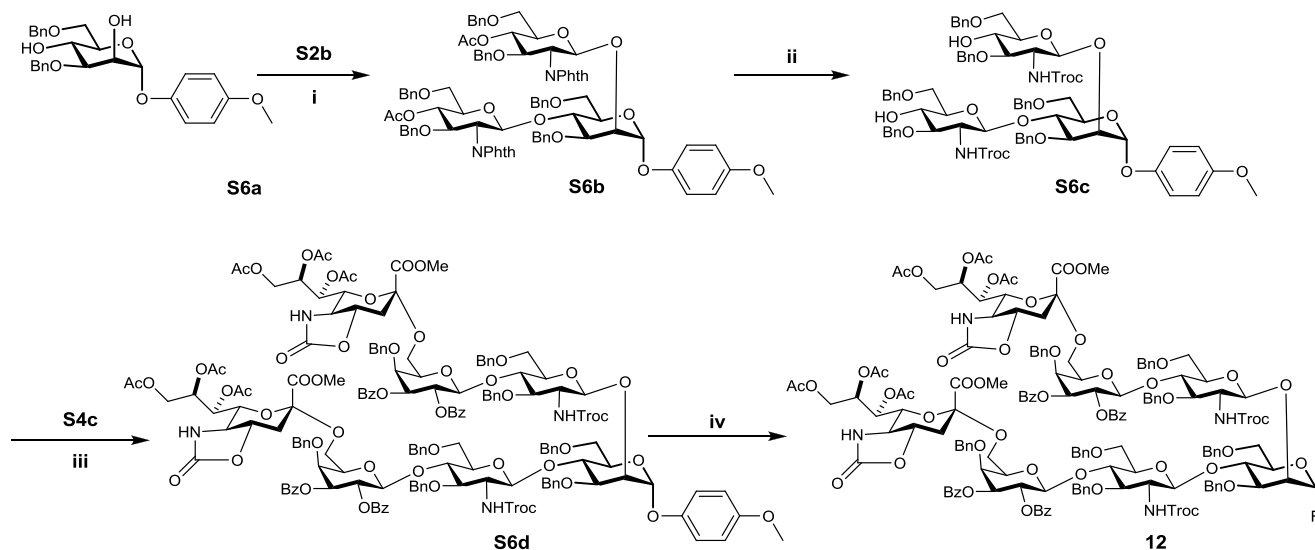


2-O-acetyl-3,4,6-O-tri-benzyl-β-D-galactopyranosyl-(1→4)-O-3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino-β-D-glucopyranosyl-(1→2)-[2-O-acetyl-3,4,6-O-tri-benzyl-β-D-galacto-pyranosyl-(1→4)-O-3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy) carbonylamino-β-D-glucopyranosyl-(1→4)]-3,6-O-di-benzyl-α-D-mannopyranosyl fluoride

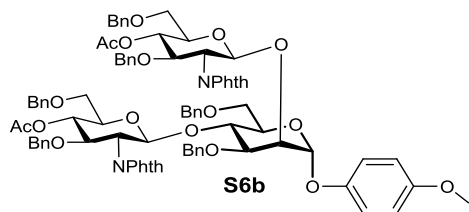
(11): TLC (ethyl acetate: toluene =1/9, v/v): R_f = 0.38; ^1H NMR (600 MHz, CDCl_3): δ 7.33-7.09 (m, 60H), 5.60 (d, J = 52.1 Hz, 1H), 5.32- 5.28 (m, 2H), 5.10 (d, J = 2.1 Hz, 1H), 4.93-4.80 (m, 5H), 4.78 (d, J = 8.4 Hz, 1H), 4.74 (d, J = 8.2 Hz, 1H), 4.65-4.40 (m, 28H), 3.95 (dd, J = 8.4 Hz, 1H), 3.79-3.69 (m, 10H), 3.50-3.40 (m, 5H), 3.39-3.20 (m, 8H), 1.91 (s, 6H) ; ^{13}C NMR (150 MHz, CHCl_3): δ 169.69, 169.56, 154.72, 154.66, 139.55, 139.02, 139.00, 138.43, 138.34, 138.27, 138.20, 138.19, 128.93, 128.80, 128.76, 128.72, 128.65, 128.45, 128.41, 128.30, 128.28, 128.21, 127.67, 127.34, 127.10, 106.45, 105.33, 101.82, 101.21, 100.00, 97.86, 97.46, 95.60, 75.46, 75.27, 74.86, 74.58, 74.26, 74.10, 73.50, 73.47, 73.10, 72.90, 72.88, 72.37, 72.30, 71.40, 71.37, 71.09, 70.57, 70.35, 70.45, 69.78, 69.46, 29.99, 25.78, 22.87, 21.36, 21.29, 15.57, 14.49, 144.42; ESI-MS: m/z calcd for $\text{C}_{124}\text{H}_{131}\text{Cl}_6\text{FN}_2\text{O}_{29}$; 2340.6953 found 2362.7225 ($M + \text{Na}$) $^+$.

Synthesis of building blocks 12.

Preparation of disialylated antennae was commenced with β -1, 2 and β -1, 4 glycosylation of mannosyl acceptor **S6a** with donor **S2b**. Compound **S6b** was then modified to **S6c** and subsequently glycosylated with **S4c** to get desired heptasaccharide **S6d**. At last, the reducing end was modified to get donor **12**.



Scheme S6 | Preparation of compound 12. i, NIS, TfOH, CH₂Cl₂, -50 °C, 76%; **ii**, (1) EDA, n-BuOH, 90 °C, (2) Troc-Cl, NaHCO₃, CH₂Cl₂, 72% over 2 steps; **iii**, NIS, TfOH, CH₂Cl₂, -50 °C, 86%; **iv**, (1) CAN, ACN: Toluene: H₂O, (2) DAST, CH₂Cl₂, -30 °C, 40% over 2 steps.

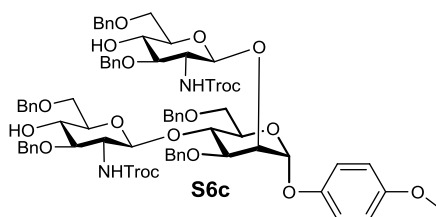


***p*-methoxyphenyl-O-4-O-acetyl-3,6-O-di-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl-(1→2)-[4-O-acetyl-3,6-O-di-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl-**

(1→4)]-3,6-O-di-benzyl-α-D-mannopyranoside (S6b):

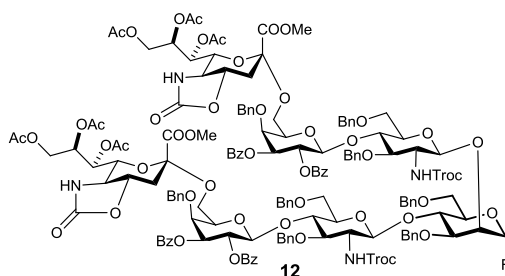
Activated 4 Å molecular sieves were added to a solution of acceptor **S6a** (0.560 g, 1.19 mmol) and donor **S2b** (1.74 g, 2.74 mmol) in anhydrous CH₂Cl₂ (10 mL). The reaction mixture was stirred for 1 h at room temperature then cooled to -50 °C. NIS (1.07 g, 4.75 mmol) and TfOH (52 μL, 0.595 mmol) were added slowly, and the resulting reaction mixture was stirred for 2 h. When TLC (ethyl acetate: toluene, 1.5/8.5) indicated formation of product with consumption of starting material, the reaction was quenched by adding Et₃N then filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), aqueous Na₂S₂O₃ (2 x 50 mL), and brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 10% EA in toluene) to afford **S6b** (1.27 g, 76%) as clear foam. TLC (ethyl acetate : toluene = 1.5/8.5, v/v): R_f=0.57; ¹H NMR (600 MHz, CDCl₃): δ 7.70-7.68 (m, 8H), 7.66-7.16 (m, 18H), 6.97 (m, 12H), 6.66-6.49 (m, 4H), 5.25 (t, *J* = 10 Hz, 2H), 5.08-5.00 (dd, *J* = 9.6 & 3.4 Hz, 2H), 4.90 (d, *J* = 4.6 Hz, 1H), 4.69 (d, *J* = 12 Hz, 1H), 4.57 (t, 3H), 4.42-4.21 (m, 10H), 4.12 (dd, *J* = 3.2 & 8.4 Hz, 2H), 3.90 (dd, *J* = 3.2 & 7.8 Hz, 1H), 3.84-3.60 (m, 7H), 3.66 (s, 3H), 3.54-3.31 (m, 3H), 3.01 (d, *J* = 7.8 Hz, 1H), 1.84 (s, 3H), 1.80 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 169.61, 169.12, 154.49, 150.07, 138.69, 138.21, 137.99, 137.77, 137.60, 133.52, 131.29, 128.28, 128.19, 127.99, 127.94, 127.91, 127.83, 127.68, 127.59, 127.47,

127.25, 127.08, 127.94, 123.17, 117.55, 114.08, 98.10, 97.37, 96.54, 78.89, 74.56, 73.56, 73.27, 72.98, 72.42, 72.32, 71.91, 71.43, 71.02, 69.91, 69.25, 68.90, 55.73, 55.45, 55.26, 20.80; ESI-MS: m/z calcd for $C_{77}H_{82}Cl_6N_2O_{21}$; 1585.3589 found 1586.5687 ($M + H$)⁺.



***p*-methoxyphenyl-O-3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy) carbonylamino- β -D-glucopyranosyl-(1 \rightarrow 2)-[3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino- β -D-glucopyranosyl-(1 \rightarrow 4)]-3,6-O-di-benzyl- α -D-mannopyranoside (S6c):** A mixture of compound **S6b** (1.9 g, 1.27 mmol) and 20 mL of ethylene diamine: *n*-BuOH (1:4) was stirred at 90 °C overnight. Volatiles were evaporated, and the crude product was dried using high vacuum. It was then dissolved in CH_2Cl_2 (20 mL), $NaHCO_3$ (0.687 g, 12.7 mmol) and 2,2,2-trichloro ethyl chloroformate (1.75 mL, 12.7 mmol) were added at 0 °C, allowed it to warm to rt and stirred for overnight until TLC (acetone: toluene, 1/9) indicated formation of product with consumption of starting material. The reaction mixture was diluted with CH_2Cl_2 (100 mL), washed with water (2 x 50 mL) and brine (50 mL) solution. The organic layer was dried over Na_2SO_4 and concentrated in *vacuo*. The residue was purified by silica gel column chromatography (0% \rightarrow 8% EA in toluene) to afford **S6c** as a colorless solid (1.35 g, 72%). TLC (acetone: toluene = 1/9, v/v): R_f = 0.54; 1H NMR (600 MHz, $CDCl_3$): δ 7.35-7.14 (m, 30H), 6.90 (d, J = 8.4 Hz, 2H), 6.79 (d, J = 8.6 Hz, 2H), 5.32 (d, J = 3.2 Hz, 1H), 5.27 (d, J = 7.2 Hz, 1H), 4.93 (d, J = 8.4 Hz, 1H), 4.73-4.58 (m, 10H), 4.48-4.43 (m, 7H), 4.19 (d, J = 7.2 Hz, 1H), 4.05-3.96 (m, 3H), 3.68 (s, 3H), 3.62-3.56 (m, 5H), 3.26 (q, 1H), 3.26-3.15 (m, 3H), 2.96 (s, 1H), 2.12 (s, 1H); ^{13}C NMR (150

chromatography (0% → 15% EA in toluene) to afford **S6d** (0.950 g, 86%) as clear foam. TLC (acetone: toluene = 2/8, v/v): R_f = 0.48; $^1\text{H NMR}$ (600 MHz, CDCl_3): δ 7.90-7.87 (m, 7H), 7.81-7.79 (m, 3H), 7.50-7.07 (m, 50H), 6.98 (d, 2H), 6.40 (d, 2H), 5.78-5.73 (m, 2H), 5.36-5.26 (m, 4H), 5.20 (d, J = 8.6 Hz, 1H), 5.19-5.15 (dd, J = 3.2 & 8.4 Hz, 2H), 5.05 (dd, J = 4.3 & 8.4 Hz, 2H), 4.94-4.78 (m, 4H), 4.68-4.15 (m, 30H), 3.96-3.86 (m, 6H), 3.71 (s, 6H), 3.70-3.59 (m, 11H), 3.57 (s, 3H), 3.34-3.26 (m, 5H), 2.98-2.76 (m, 4H), 2.15 (s, 3H), 2.09 (s, 3H), 2.00 (s, 3H), 1.98 (s, 3H), 1.89 (s, 3H), 1.89 (s, 3H); $^{13}\text{C NMR}$ (150 MHz, CDCl_3): δ 171.56, 170.89, 169.56, 167.20, 165.28, 165.00, 159.11, 154.79, 138.78, 138.56, 138.21, 137.29, 137.00, 133.67, 133.24, 130.79, 129.70, 129.60, 129.26, 128.78, 128.56, 128.42, 128.32, 128.11, 127.69, 127.56, 127.44, 127.32, 127.14, 117.78, 100.89, 100.56, 99.87, 99.68, 95.41, 79.13, 75.56, 74.18, 73.48, 73.26, 73.12, 71.00, 72.90, 72.78, 71.76, 68.78, 68.40, 68.42, 61.78, 57.78, 57.33, 55.89, 53.80, 38.90, 38.7, 30.98, 30.76, 28.87, 23.89, 22.65, 20.87, 20.65, 20.54, 14.08, 10.86; ESI-MS: m/z calcd for $\text{C}_{161}\text{H}_{168}\text{Cl}_6\text{N}_4\text{O}_{55}$; 3251.8695 found 3252.8657 (M)⁺.

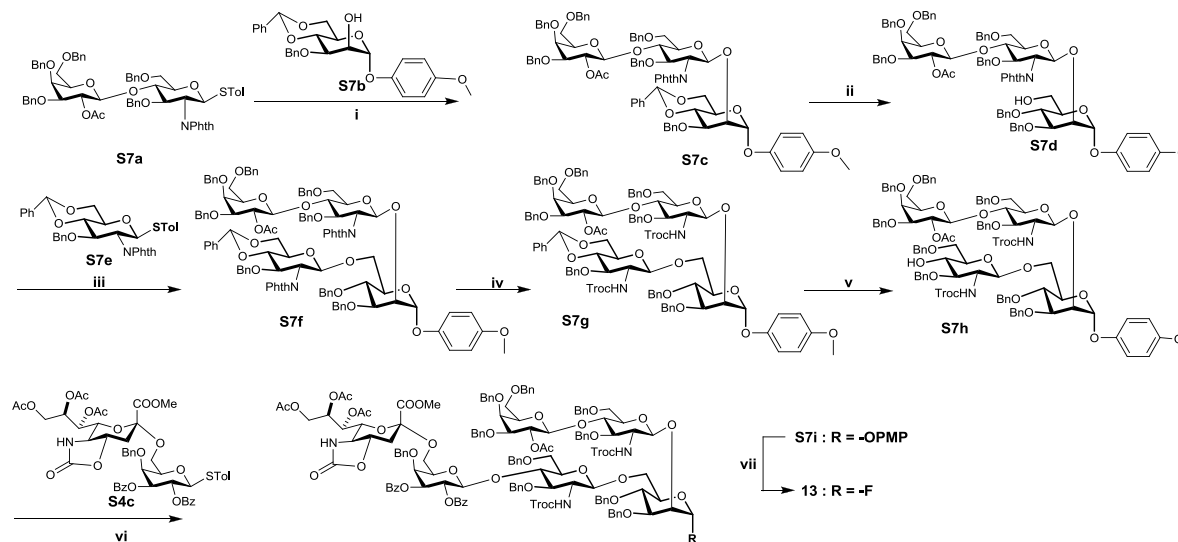


[Methyl-5-acetamido-7,8,9-tri-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-non-2-ulopyranosylate]-(2→6)-4-O-benzyl-2,3-di-O-benzoyl-1- β -D-galacto pyranosyl-(1→4)-O-3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino- β -D-glucoopyranosyl-(1→2)-[Methyl-5-acetamido-7,8,9-tri-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-non-2-ulopyranosylate]-(2→6)-4-O-benzyl-2,3-di-O-benzoyl-1- β -D-galactopyranosyl-(1→4)-O-3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino- β -D-glucoopyranosyl-(1→4)-O-3,4,6-tri-O-benzyl- α -D-mannopyranosyl fluoride (12**): Cerium ammonium nitrate**

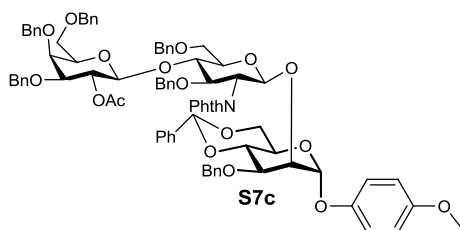
(0.470 g, 0.554 mmol) was added to a solution of compound **S6d** (0.300 g, 0.092 mmol) in 10 mL of acetonitrile: toluene: H₂O (4:2:1). The resulting reaction mixture was stirred at rt for 3 h. The reaction was diluted with EtOAc (100 mL) and washed with H₂O (30 x 2 mL) and brine (30 mL). The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The product was purified by flash column chromatography (0% → 20% EA in toluene) to afford **1-OH** compound (0.180 g) as a foam. The residue (0.180 g, 0.057 mmol) was dissolved in CH₂Cl₂ (10 mL) at -30 °C. Then, DAST (22 μL, 0.171 mmol) was added slowly, and the resulting reaction mixture was stirred for 1 h. When TLC (acetone: toluene, 2/8) indicated formation of product with consumption of starting material, the reaction was quenched with aq. NaHCO₃. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL) and brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by silica gel column chromatography (0% → 15% EA in toluene) to afford **13** (0.120 g, 41% over 2 steps) as white solid. TLC (acetone: toluene =2/8, v/v): R_f = 0.24; ¹H NMR (600 MHz, CDCl₃): δ 7.92-7.83 (m, 10H), 7.54-7.47 (m, 5H), 7.40-6.93 (m, 45H), 5.83 (m, 2H), 5.40-5.35 (m, 6H), 5.2 (dd, *J* = 3.2 & 7.8 Hz, 2H), 5.11 (dd, *J* = 4.3 & 8.4 Hz, 2H), 4.98-4.89 (m, 4H), 4.76-4.56 (m, 15H), 4.28-4.10 (m, 11H), 4.00-3.89 (m, 6H), 3.79-3.72 (m, 6H), 3.65 (s, 3H), 3.67 (s, 3H), 3.62-3.43 (m, 7H), 3.36-3.21 (m, 6H), 3.02-2.98 (m, 4H), 2.16 (s, 3H), 2.14 (s, 3H), 2.11 (s, 3H), 2.06 (s, 3H), 2.01 (s, 3H), 1.98 (s, 3H), 1.89-1.76 (m, 2H); ¹³C NMR (150 MHz, CDCl₃): δ 171.80, 170.85, 170.06, 168.03, 166.12, 165.41, 159.56, 154.16, 138.75, 138.34, 133.73, 130.11, 129.94, 129.57, 129.19, 128.79, 128.72, 128.50, 128.38, 128.03, 127.75, 127.56, 100.69, 100.00, 96.67, 95, 78, 95.35, 95.12, 74.91, 74.65, 74.29, 73.00, 70.91, 69.01, 67.35, 61.87, 58.15, 57.60, 53.42, 37.17, 21.28, 20.97; ESI-MS: *m/z* calcd for C₁₅₄H₁₆₁Cl₆FN₄O₅₃; 3147.6554 found 3169.8022 (*M* + Na)⁺.

Synthesis of building blocks **13**.

Total chemical synthesis of partially sialylated antennae **13** is shown in scheme S7.

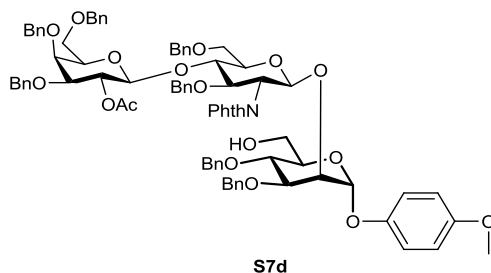


Scheme S7 | Preparation of compound 13. **i**, NIS, TfOH, CH₂Cl₂, -50 °C, 86%; **ii**, BH₃.THF, Bu₂BOTf, CH₂Cl₂, 65%; **iii**, NIS, TfOH, CH₂Cl₂, -50 °C, 76%; **iv**, (1) EDA, n-BuOH, 90 °C, (2) Troc-Cl, NaHCO₃, CH₂Cl₂; (3) Ac₂O, pyridine, 65% over 3 steps; **v**, Triethyl silane, TFA, CH₂Cl₂, 58%; **vi**, NIS, TfOH, CH₂Cl₂, -50 °C, 73%; **vii**, (1) CAN, ACN: Toluene: H₂O, (2) DAST, CH₂Cl₂, -30 °C, 30% over 2 steps.



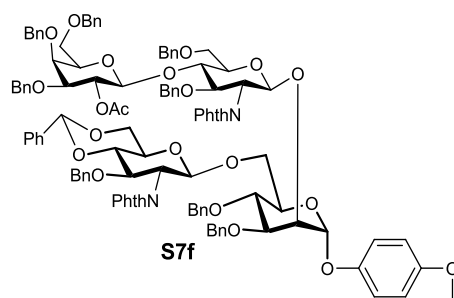
***p*-methoxyphenyl-O-2-O-acetyl-3,4,6-O-tri-benzyl-β-D-galactopyranosyl-(1→4)-O-3,6-O-di-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl-(1→2)-O-3,4-O-benzyl-α-D-mannopyranoside (S7c):** Activated 4 Å molecular sieves were added to a solution of acceptor **S7a** (0.500 g, 1.07 mmol) and donor **S7b** (1.21 g, 1.28 mmol) in anhydrous CH₂Cl₂ (10 mL). The reaction mixture was stirred for 1 h at room temperature then cooled to -50 °C. NIS (0.481 g, 2.14 mmol) and TMSOTf (48 μL, 0.267 mmol) were added slowly, and the resulting reaction

mixture was stirred for 3 h. When TLC (ethyl acetate: toluene, 1/9) indicated formation of product with consumption of starting material, the reaction was quenched by adding Et₃N then filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), aqueous Na₂S₂O₃ (2 x 50 mL), and brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 10% EA in toluene) to afford **S7c** (1.30 g, 86%) as a pale yellow solid. TLC (ethyl acetate: toluene = 1/9, v/v): R_f = 0.59; ¹H NMR (600 MHz, CDCl₃): δ 7.80-7.77 (m, 4H), 7.75-7.65 (m, 5H), 7.37-7.15 (m, 30H), 6.98 (d, *J* = 8.1 Hz, 2H), 6.82 (d, *J* = 8.4 Hz, 2H), 6.70 (d, *J* = 8.4 Hz, 2H), 6.60 (d, *J* = 9.2 Hz, 2H), 5.39 (s, 1H), 5.34 (t, *J* = 10.2 Hz, 1H), 5.22 (dd, *J* = 7.2 & 2.8 Hz, 1H), 5.06 (d, *J* = 3.2 Hz, 1H), 4.89 (d, *J* = 11.4 Hz, 1H), 4.82 (d, *J* = 12.1 Hz, 1H), 4.75 (d, *J* = 11.8 Hz, 1H), 4.66-4.62 (m, 3H), 4.54-4.38 (m, 6H), 4.32 (d, *J* = 11.2 Hz, 1H), 4.29 (dd, *J* = 3.1 & 7.2 Hz, 2H), 4.27 (d, *J* = 10.8 Hz, 1H), 4.19 (t, *J* = 8.3 Hz, 1H), 4.00-3.89 (m, 4H), 3.72 (s, 3H), 3.58-3.33 (m, 4H), 3.0 (t, *J* = 8.7 Hz, 1H), 2.07 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 169.3, 154.9, 149.6, 138.8, 138.7, 138.6, 138.5, 138.1, 138.0, 137.9, 137.5, 133.6, 131.9, 128.7, 128.4, 128.4, 128.3, 128.2, 128.1, 128.1, 128.1, 128.0, 127.8, 127.8, 127.7, 127.7, 127.7, 127.6, 127.6, 127.5, 127.4, 127.3, 127.2, 126.8, 126.0, 123.0, 117.03, 114.5, 101.4, 100.9, 97.0, 96.2, 96.2, 80.3, 78.1, 78.0, 75.6, 74.9, 74.5, 74.4, 73.7, 73.4, 73.3, 72.6, 72.6, 72.0, 71.7, 71.3, 68.4, 68.1, 68.1, 64.2, 55.6, 55.6, 55.6, 21.0; ESI-MS: *m/z* calcd for C₈₄H₈₃NO₁₉; 1409.5559 found 1432.5499 (*M* + Na)⁺.



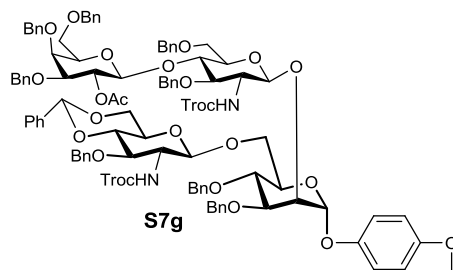
***p*-methoxyphenyl-O-2-O-acetyl-3,4,6-O-tri-benzyl-β-D-galactopyranosyl-(1→4)-O-3,6-O-di-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl-(1→2)-O-3,4-O-dibenzyl-α-D-**

mannopyranoside (S7d): To a mixture of compound **S7c** (1.30 g, 1.02 mmol) and activated molecular sieves in anhydrous CH₂Cl₂ (15 mL) was added borane.THF complex (0.978 mL of a 1M solution in THF, 10.2 mmol) and Bu₂BOTf (0.439 mL of a 1M solution in CH₂Cl₂, 10.2 mmol) were added at 0 °C. The reaction mixture was allowed to stirred at room temperature for 4 h. TLC (acetone: toluene, 1/9) indicated formation of a product with consumption of the starting material. Triethyl amine was added to the reaction mixture followed by slow addition of methanol at 0 °C. When no more hydrogen was produced, the reaction mixture was filtered through Celite, The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), and brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 7% EA in toluene) to afford **S7d** (0.850 g, 65%) as clear foam. TLC (acetone: toluene = 1/9, v/v): R_f = 0.31; ¹H NMR (600 MHz, CDCl₃): δ 7.98-7.89 (m, 4H), 7.84-7.66 (m, 5H), 7.42-7.02 (m, 30H), 7.03 (d, *J* = 8.4 Hz, 2H), 6.87 (t, *J* = 7.8 Hz, 2H), 6.67 (d, *J* = 12.1 Hz, 2H), 6.48 (d, *J* = 12.1 Hz, 2H), 5.39 (t, *J* = 10.2 Hz, 1H), 5.17 (d, *J* = 8.4 Hz, 1H), 4.93 (d, *J* = 12 Hz, 2H), 4.88 (d, *J* = 7.2 Hz, 1H), 4.85 (d, *J* = 12.1 Hz, 2H), 4.67 (dd, *J* = 4.3 & 8.4 Hz, 2H), 4.59-4.30 (m, 9H), 4.18 (t, *J* = 7.4 Hz, 1H), 4.01-3.96 (m, 3H), 3.84-3.78 (m, 3H), 3.76 (s, 3H), 3.68 (dd, *J* = 3.2 & 7.8 Hz, 1H), 3.54-3.34 (m, 3H), 3.25-3.20 (m, 1H), 2.01 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 169.3, 167.9, 154.7, 149.6, 138.9, 138.7, 138.4, 138.2, 138.0, 138.0, 137.9, 134.1, 133.6, 131.9, 131.5, 129.0, 128.4, 128.4, 128.4, 128.3, 128.2, 128.2, 128.1, 128.1, 127.9, 127.8, 127.8, 127.7, 127.7, 127.7, 127.6, 127.6, 127.5, 127.3, 127.3, 126.8, 123.2, 123.0, 116.8, 114.4, 100.9, 97.6, 95.7, 80.3, 78.0, 76.9, 75.2, 75.1, 74.4, 74.4, 73.9, 73.6, 73.4, 73.3, 72.5, 72.3, 72.0, 71.6, 70.7, 68.6, 68.1, 62.2, 55.9, 55.6, 21.0; ESI-MS: *m/z* calcd for C₈₄H₈₅NO₁₉; 1411.5716 found 1412.5726 (*M* + H)⁺.



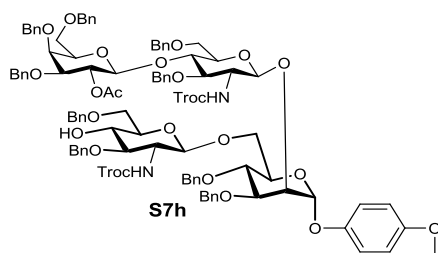
***p*-methoxyphenyl-O-[2-O-acetyl-3,4,6-O-tri-benzyl- β -D-galactopyranosyl-(1 \rightarrow 4)-O-3,6-O-di-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl]-(1 \rightarrow 2)-O-[3-O-benzyl-4,6-O-benzylidene-2-deoxy-2-phthalimido- β -D-glucopyranosyl]-(1 \rightarrow 6)-3,4-O-dibenzyl- α -D-mannopyranoside (S7f):** Activated 4 Å molecular sieves were added to a solution of acceptor S7d (0.600 g, 0.470 mmol) and donor S7e (0.557 g, 0.940 mmol) in anhydrous CH₂Cl₂ (10 mL). The reaction mixture was stirred for 1 h at room temperature then cooled to -50 °C. NIS (0.211 g, 0.940 mmol) and TfOH (10.4 μL, 0.117 mmol) were added slowly, and the resulting reaction mixture was stirred for 2 h. When TLC (ethyl acetate: toluene, 2/8) indicated formation of product with consumption of starting material, the reaction was quenched by adding Et₃N then filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), aqueous Na₂S₂O₃ (2 x 50 mL), and brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 15% EA in toluene) to afford S7f (0.610g, 76%) as clear foam. TLC (ethyl acetate: toluene = 2/8, v/v): R_f = 0.49; ¹H NMR (600 MHz, CDCl₃): δ 7.66 (m, 8H), 7.38-7.15 (m, 30H), 7.10 (d, *J* = 8.4 Hz, 2H), 6.95-6.91 (m, 5H), 6.88-6.78 (m, 8H), 6.63 (d, *J* = 8.4 Hz, 2H), 6.50 (d, *J* = 8.8 Hz, 2H), 5.41 (s, 1H), 5.32 (t, *J* = 10.2 Hz, 1H), 5.11 (d, *J* = 8.4 Hz, 1H), 4.89 (d, *J* = 4.3 Hz, 1H), 4.87 (s, 1H), 4.80 (t, *J* = 8.8 Hz, 2H), 4.73 (s, 1H), 4.70 (s, 1H), 4.60 (t, *J* = 10.8 Hz, 3H), 4.57-4.40 (m, 5H), 4.35-4.16 (m, 8H), 4.08 (t, *J* = 10.2 Hz, 1H), 4.00-3.89 (m, 4H), 3.85 (dd, *J* = 3.6 & 7.8 Hz, 1H), 3.78 (s, 3H), 3.70-3.61 (m, 3H), 3.52-3.41 (m, 4H), 3.40-3.28 (m, 4H), 3.20 (t, *J* = 10.2 Hz, 2H), 2.98 (m, 1H), 1.98 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 168.5, 164.8, 158.0, 153.0, 152.8, 151.6, 151.5, 150.6, 150.5, 149.2, 139.6, 138.9, 138.4, 138.1, 138.0, 137.4, 129.3, 129.2,

128.9, 128.9, 128.8, 128.9, 128.7, 128.5, 128.4, 128.2, 128.1, 128.0, 128.0, 127.9, 127.9, 127.7, 127.5, 127.4, 126.2, 125.5, 117.3, 114.9, 114.2, 102.2, 101.4, 100.9, 98.8, 96.5, 95.8, 95.7, 95.5, 95.3, 95.3, 95.0, 82.3, 80.6, 78.3, 76.2, 76.0, 75.8, 75.5, 75.4, 75.2, 74.9, 74.8, 74.7, 74.7, 74.2, 74.1, 73.8, 73.7, 73.5, 72.9, 72.3, 71.9, 68.9, 68.8, 68.2, 68.0, 66.4, 58.0, 57.1, 55.9, 44.3, 42.9, 40.8, 40.4, 40.3, 40.0, 21.7, 21.2; ESI-MS: m/z calcd for $C_{111}H_{108}N_2O_{24}$; 1882.7247 found 1882.7343.



***p*-methoxyphenyl-O-[2-O-acetyl-3,4,6-O-tri-benzyl- β -D-galactopyranosyl-(1 \rightarrow 4)-O-3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino- β -D-glucopyranosyl]-(1 \rightarrow 2)-O-[3-O-benzyl-4,6-O-benzylidene-2-(2,2,2-trichloroethoxy)carbonylamino- β -D-glucopyranosyl]-(1 \rightarrow 6)-3,4-O-dibenzyl- α -D-mannopyranoside (**S7g**):** A mixture of compound **S7f** (0.950 g, 0.545 mmol) and 10 mL of ethylene diamine: *n*-BuOH (2:8) was stirred at 90 °C overnight. Volatiles were evaporated, and the crude product was dried using high vacuum. It was then dissolved in CH_2Cl_2 (20 mL), $NaHCO_3$ (0.363 g, 5.45 mmol) and 2,2,2-trichloro ethyl chloroformate (0.86 mL, 5.45 mmol) were added at 0 °C, allowed it to warm to rt and stirred for overnight. TLC (ethyl acetate: toluene, 2/8) indicated formation of product with consumption of starting material. The reaction mixture was diluted with CH_2Cl_2 (100 mL), washed with water (2 x 50 mL) and brine (50 mL) solution. The organic layer was dried over Na_2SO_4 and concentrated *in vacuo*. The residue was purified by silica gel column chromatography (0% \rightarrow 10% EA in toluene). The product was then acetylated using 10 mL of pyridine/acetic anhydride (6:4) until TLC indicated (ethyl acetate: toluene, 2/8) complete consumption of starting material. The reaction mixture was then concentrated *in vacuo* and purified by silica gel column chromatography (0% \rightarrow 10% EA in toluene) to afford **S7g** as a white foam (0.650 g, 65%). TLC

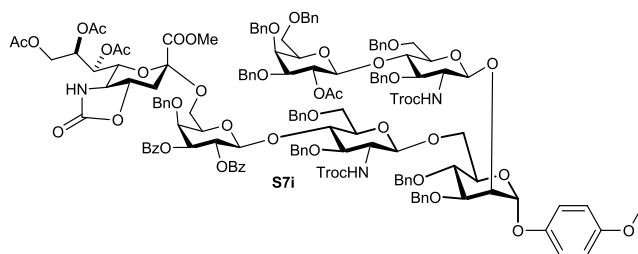
(ethyl acetate: toluene = 2/8, v/v): R_f = 0.64; ^1H NMR (600 MHz, CDCl_3): δ 7.48 (d, J = 7.8 Hz, 2H), 7.39 (d, J = 8.2 Hz, 2H), 7.35-7.12 (m, 41H), 6.91 (d, J = 10.2 Hz, 2H), 6.87 (d, J = 10.3 Hz, 2H), 5.55 (s, 1H), 5.50 (s, 1H), 5.32 (t, J = 10.2 Hz, 1H), 4.94-4.89 (m, 8H), 4.83-4.67 (m, 10H), 4.64-4.66 (m, 5H), 4.54-4.50 (m, 3H), 4.36-4.22 (m, 6H), 4.06-3.87 (m, 6H), 3.83-3.46 (m, 6H), 3.38-3.34 (m, 3H), 3.20-3.10 (m, 2H), 2.33 (s, 3H); ^{13}C NMR (150 MHz, CDCl_3): δ 169.5, 162.8, 155.0, 155.0, 154.8, 154.6, 154.51, 150.6, 150.5, 150.2, 139.6, 138.9, 138.4, 138.3, 138.3, 138.2, 138.1, 137.5, 129.3, 129.2, 128.7, 128.6, 128.6, 128.5, 128.5, 128.4, 128.3, 128.2, 128.1, 128.0, 128.0, 127.9, 127.9, 127.7, 127.5, 127.4, 126.2, 125.5, 117.3, 114.9, 114.2, 102.2, 101.4, 100.8, 98.3, 96.5, 95.8, 95.7, 95.5, 95.3, 95.3, 95.0, 82.3, 80.6, 78.3, 76.2, 76.0, 75.8, 75.5, 75.4, 75.2, 74.9, 74.8, 74.7, 74.7, 74.2, 74.1, 73.8, 73.7, 73.5, 72.9, 72.3, 71.9, 68.9, 68.8, 68.2, 68.0, 66.4, 58.0, 57.1, 55.9, 44.3, 42.9, 40.8, 40.4, 40.3, 40.0, 21.7, 21.2; ESI-MS: m/z calcd for $\text{C}_{102}\text{H}_{106}\text{Cl}_6\text{N}_2\text{O}_{25}$; 1972.6590 found 1973.5278 ($M + \text{H}$) $^+$.



***p*-methoxyphenyl-O-[2-O-acetyl-3,4,6-O-tri-benzyl- β -D-galactopyranosyl-(1 \rightarrow 4)-O-3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino- β -D-glucopyranosyl]-(1 \rightarrow 2)-O-[3,6-O-dibenzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino- β -D-glucopyranosyl]-**

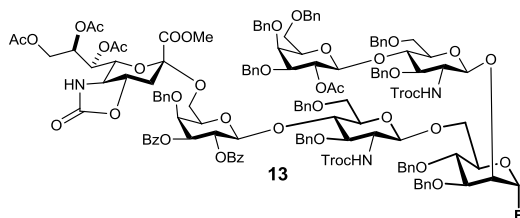
(1 \rightarrow 6)-3,4-O-dibenzyl- α -D-mannopyranoside (S7h): To a solution of S7g (0.600 g, 0.328 mmol) in anhydrous CH_2Cl_2 (70 mL) was added triethyl silane (2.10 mL, 13.2 mmol) followed by trifluoroacetic acid (0.953 mL, 13.2 mmol) at 0 $^\circ\text{C}$. The resulting reaction mixture was stirred for 2 h. After 2 h, TLC (ethyl acetate: toluene, 1.5/8.5 v/v) indicated product formation with consumption of starting material. The reaction mixture was washed with sat. NaHCO_3 (2 x 50 mL). The aqueous layer was further extracted with CH_2Cl_2 (3 x 30 mL), and the combined

organic layer were washed with brine solution (100 mL), dried over MgSO₄, filtered and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 10% EA in toluene) to afford **S7h** (0.350 g, 58%) as clear oil. TLC (ethyl acetate: toluene = 1.5/8.5, v/v): R_f = 0.35; ¹H NMR (600 MHz, CDCl₃): δ 7.33-7.04 (m, 45H), 6.92 (d, *J* = 8.2 Hz, 2H), 6.80 (d, *J* = 8.3 Hz, 2H), 5.55 (s, 1H), 5.31 (t, *J* = 10.1 Hz, 1H), 5.06 (d, *J* = 3.2 Hz, 1H), 4.93-4.80 (m, 4H), 4.47-4.35 (m, 13H), 4.35-4.18 (m, 8H), 4.05 (dd, *J* = 3.2 & 7.8 Hz, 2H), 3.82 (d, *J* = 8.4 Hz, 2H), 3.80-3.62 (m, 14H), 3.49-3.20 (m, 5H), 3.10 (bs, 2H), 2.08 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 169.5, 168.0, 155.3, 155.0, 154.7, 150.2, 138.9, 138.4, 138.3, 138.2, 137.8, 132.7, 131.1, 129.0, 128.7, 128.7, 128.6, 128.6, 128.6, 128.5, 128.4, 128.3, 128.2, 128.1, 128.1, 128.0, 127.9, 127.8, 127.7, 127.5, 127.4, 117.3, 114.9, 101.5, 100.8, 95.7, 80.6, 78.5, 75.8, 75.3, 74.9, 74.8, 74.6, 74.1, 74.0, 73.8, 73.7, 73.64, 73.5, 72.8, 72.3, 71.9, 71.1, 68.9, 68.8, 68.4, 68.2, 67.6, 58.0, 56.5, 55.9, 41.5, 39.0, 37.3, 33.7, 32.0, 31.4, 30.6, 29.9, 29.20, 26.6, 24.0, 23.2, 22.9, 21.2, 20.1, 14.6, 14.4, 14.3, 14.3, 11.2; ESI-MS: *m/z* calcd for C₁₀₂H₁₀₈Cl₆N₂O₂₅; 1974.6750 found 1975.5438 (*M* + H)⁺.



***p*-methoxyphenyl-O-[2-O-acetyl-3,4,6-O-benzyl-β-D-galactopyranosyl-(1→4)-O-3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino-β-D-glucopyranosyl-(1→2)-[Methyl-5-acetamido-7,8,9-tri-O-acetyl-3,5-dideoxy-D-glycero-α-D-galacto-non-2-ulopyranosylonate]-(2→6)-4-O-benzyl-2,3-di-O-benzoyl-1-β-D-galactopyranosyl-(1→4)-O-3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino-β-D-glucopyranosyl-(1→6)-O-3,4-di-O-benzyl-α-D-mannopyranoside (S7i)**: A mixture of donor **S4c** (0.187 g, 0.182 mmol), acceptor **S7h** (0.300 g, 0.151 mmol) and activated 4 Å molecular sieves in dry

CH₂Cl₂ (10 mL) was stirred at room temperature for 1 h. The reaction mixture was cooled to -50 °C, NIS (0.067 g, 0.302 mmol) and TfOH (4 μL, 0.037 mmol) were added slowly and resulting reaction mixture was stirred for 2 h. The reaction was quenched by adding Et₃N, diluted with CH₂Cl₂, filtered through Celite, extracted with saturated Na₂S₂O₃ followed by NaHCO₃, dried over sodium sulfate and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 10% EA in toluene) to afford **S7i** (0.317 g, 73%) as colorless foam. TLC: (ethyl acetate: toluene = 1.5/8.5, v/v): R_f = 0.61; ¹H NMR (600 MHz, CDCl₃): δ 7.90 (m, 4H), 5.52-7.45 (m, 2H), 7.37-7.08 (m, 54H), 6.90 (d, *J* = 8.3 Hz, 2H), 6.80 (d, *J* = 8.3 Hz, 2H), 5.72 (t, *J* = 10.2 Hz, 1H), 5.53 (s, 1H), 5.35-5.32 (m, 3H), 5.20 (dd, *J* = 3.2 & 8.3 Hz, 1H), 5.08 (dd, *J* = 2.8 & 8.4 Hz, 2H), 4.99-4.79 (m, 7H), 4.70-4.18 (m, 25H), 4.06 (dd, *J* = 3.2 & 7.8 Hz, 1H), 3.94 (m, 2H), 3.90-3.84 (m, 16H), 3.74 (s, 3H), 2.60 (s, 3H), 2.59-2.57 (m, 3H), 3.45-3.32 (m, 6H), 3.20 (t, *J* = 9.8 Hz, 1H), 2.90 (t, *J* = 10.2 Hz, 2H), 2.83 (dd, *J* = 3.2 & 7.8 Hz, 1H), 2.08 (s, 3H), 2.06 (s, 3H), 2.02 (t, *J* = 10.1 Hz, 1H), 1.99 (s, 3H), 1.91 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 171.4, 170.5, 169.7, 169.1, 167.4, 165.7, 165.1, 159.1, 154.6, 159.8, 138.5, 138.2, 138.1, 137.9, 137.4, 133.3, 133.1, 132.3, 130.2, 129.8, 129.6, 129.4, 128.7, 128.5, 128.4, 128.1, 127.9, 127.5, 127.4, 127.1, 127.08, 116.9, 114.5, 101.3, 100.6, 100.4, 100.0, 95.3, 80.2, 79.9, 77.4, 75.5, 75.3, 74.5, 74.3, 74.1, 73.7, 73.4, 73.2, 73.1, 72.5, 72.1, 71.5, 71.2, 70.8, 68.6, 68.5, 68.3, 67.8, 67.5, 67.3, 63.1, 61.5, 57.8, 56.6, 55.5, 53.1, 38.6, 37.0, 36.8, 33.3, 32.0, 31.0, 30.9, 28.8, 26.2, 25.5, 23.6, 22.8, 22.5, 20.8, 20.6, 19.7, 14.3, 13.9, 10.8; ESI-MS: *m/z* calcd for C₁₄₆H₁₅₃Cl₆N₃O₄₃; 2850.5080 found 2850.8248.

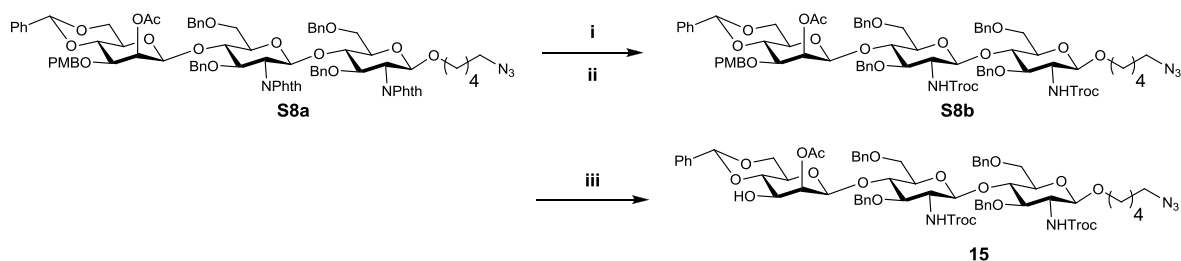


[2-O-acetyl-3,4,6-O-benzyl- β -D-galactopyranosyl-(1 \rightarrow 4)-O-3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino- β -D-glucopyranosyl-(1 \rightarrow 2)-[Methyl-5-acetamido-7,8,9-tri-O-acetyl-3,5-dideoxy-D-glycero- α -D-galacto-non-2-ulopyranosylonate]-(2 \rightarrow 6)-4-O-benzyl-2,3-di-O-benzoyl-1- β -D-galactopyranosyl-(1 \rightarrow 4)-O-3,6-O-di-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy) carbonylamino- β -D-glucopyranosyl-(1 \rightarrow 6)-O-3,4,-di-O-benzyl- α -D-mannopyranosyl fluoride (13): To a solution of compound **S7i** (0.430 g, 0.150 mmol) in 10 mL of acetonitrile: toluene: H₂O (4:2:1) was added cerium ammonium nitrate (0.641 g, 0.754 mmol) and the resulting reaction mixture was stirred at 0 °C for 2 h. The reaction was diluted with EtOAc (100 mL) and washed with H₂O (30 x 2 mL) and brine (30 mL). The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The product was purified by flash column chromatography (0% \rightarrow 20% EA in toluene) to afford **1-OH** compound (0.240 g, 57%) as clear foam. The residue (0.200 g, 0.072 mmol) was dissolved in CH₂Cl₂ (10 mL) at -30 °C. Then, DAST (29 μ L, 0.216 mmol) was added slowly, and the resulting reaction mixture was stirred for 2 h. When TLC (ethyl acetate: toluene, 2/8) indicated formation of product with consumption of starting material, the reaction was quenched with aq. NaHCO₃. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL) and brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by silica gel column chromatography (0% \rightarrow 10% EA in toluene) to afford **13** (0.104 g, 50%) as white foam. TLC (ethyl acetate: toluene =2/8, v/v): R_f = 0.44; ¹H NMR (600 MHz, CDCl₃): δ 7.95-7.90 (m, 4H), 7.58-7.49 (m, 2H), 7.39-7.18 (m, 54H), 5.82 (t, *J* = 10.2Hz, 1H), 5.65 (d, *J* = 51 Hz, 1H), 5.34 (t, *J* = 10.2 Hz, 2H), 5.21 (dd, *J* = 3.6 & 7.8 Hz, 1H), 5.13-4.19 (m, 30H), 4.10-4.09 (m, 5H), 4.02-3.74 (m, 12H), 3.65 (s, 3H), 3.64-3.27 (m, 14H), 3.02 (t, *J* = 10.2 Hz, 2H), 2.87 (dd, *J* = 3.2 & 7.8 Hz, 1H), 2.12 (s, 3H), 2.10 (s, 3H), 2.09 (t, *J* = 10.2 Hz, 1H), 2.00 (s, 3H), 1.97 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 171.4, 170.2, 169.7, 169.1, 167.4, 165.6, 165.1, 159.1, 154.4, 139.0, 138.8, 138.2, 138.1, 138.0, 137.7, 133.3, 129.8, 129.7, 129.2, 128.7, 128.4, 128.1, 127.9, 127.7, 127.6, 127.5, 127.4, 127.3, 127, 1, 127.0, 125.2, 101.4, 100.7, 100.4, 9.9, 97.3, 95.180.2, 79.6, 78.6, 75.5, 74.5, 74.3, 73.8, 73.5, 73.3, 73.1, 72.8, 71.3, 71.3, 70.7, 70.1, 68.6, 67.8, 66.9, 63.1,

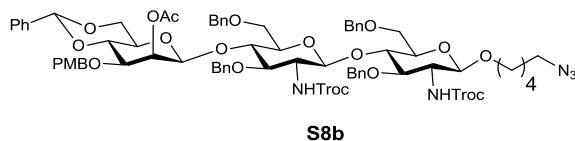
61.5, 57.8, 53.1, 37.1, 25.5, 21.3, 20.9, 20.5; ESI-MS: m/z calcd for $C_{139}H_{146}Cl_6FN_3O_{41}$; 2746.3454 found 2746.7604.

iv. Preparation of core trisaccharides 14-15

The reducing end trisaccharide (Man- β -1,4-GlcNAc- β -1,4-GlcNAc- β -linker) **14** with N-phthalamide protections was obtained according to our previous report³. Compound **14** was next modified to **15** as shown in scheme S8.



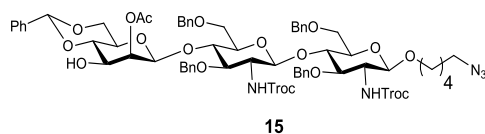
Scheme S8 | Preparation of compound 15. **i**, (1) EDA, *n*-BuOH, 90 °C, (2) Troc-Cl, NaHCO₃, CH₂Cl₂, 72% over 2 steps; **ii**, Ac₂O, pyridine, 79%; **iii**, DDQ, CH₂Cl₂ H₂O, 58%.



5-Azidopentyl-O-2-O-acetyl-3-O-*p*-methoxy-benzyl-4,6-O-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-O-(3,6-di-O-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino- β -D-glucopyranosyl)-(1 \rightarrow 4)-O-3,6-di-O-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)

carbonylamino- β -D-glucopyranoside (S8b): A mixture of compound **S8a** (0.950 g, 0.545 mmol) and 10 mL of ethylene diamine: *n*-BuOH (2:8) was stirred at 90 °C overnight. Volatiles were evaporated, and the crude mixture was dried using high vacuum. It was then dissolved in CH₂Cl₂ (20 mL), NaHCO₃ (0.363 g, 5.45 mmol) and 2,2,2-trichloro ethyl chloroformate (0.86 mL, 5.45 mmol) were added at 0 °C, allowed it to warm to rt and stirred for overnight. TLC (ethyl acetate: toluene, 2/8) indicated formation of product with consumption of starting material.

Reaction was diluted with CH₂Cl₂ (100 mL), washed with water (2 x 50 mL) and brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by silica gel column chromatography (0% → 10% EA in toluene). The product was then acetylated using 10 mL of pyridine/acetic anhydride (6:4) until TLC indicated (ethyl acetate: toluene, 2/8) complete consumption of starting material. The reaction mixture was then concentrated *in vacuo* and purified by silica gel column chromatography (0% → 10% EA in toluene) to afford **S8b** as a white foam (0.650 g, 65%). TLC (ethyl acetate: toluene = 2/8, v/v): R_f = 0.64; ¹H NMR (600MHz, CDCl₃): δ 7.62-7.20 (m, 30H), 5.89 (s, 1H), 4.56 (d, *J* = 3.2 Hz, 1H), 4.23 (d, *J* = 8.9 Hz, 2H), 4.20-4.10 (m, 12H), 4.00-3.86 (m, 5H), 3.69-3.30 (m, 11H), 3.29 (s, 3H), 3.22-3.20 (m, 4H), 3.20-3.00 (m, 1H), 2.66 (s, 3H, -C(O)CH₃), 1.56-1.38 (m, 4H, -CCH₂C-, linker), 1.21-1.23(m, 2H, -CCH₂C-, linker) ; ¹³C NMR (150 MHz, CDCl₃): δ 177.7, 169.6, 167.0, 138.5, 138.4, 138.3, 138.2, 137.2, 137.8, 137.5, 134.2, 133.2, 133.6, 131.7, 131.1, 131.45, 129.8, 129.06, 128.9, 128.2, 128.4, 128.2, 128.4, 128.7, 127.8, 127.87, 127.83, 127.4, 127.4, 127.0, 127.2, 127.29, 127.6, 127.20, 126.88, 126.24, 125.1, 123.6, 123.3, 101.5, 102.7, 99.7, 98.0, 97.1, 78.13, 78.55, 78.05, 75.82, 74.57, 75.38, 75.27, 74.25, 74.30, 72.78, 72.26, 70.89, 66.85, 68.43, 68.25, 67.75, 66.87, 55.57, 53.71, 52.10, 28.97, 28.87, 23.80, 22.77, 20.12; ESI-MS: *m/z* calcd for C₇₄H₈₃Cl₆N₅O₂₀: 1575.1930 ; found 1576.235 (*M* + H)⁺.



5-Azidopentyl-O-2-O-acetyl-4,6-O-benzylidene-β-D-mannopyranosyl-(1→4)-O-(3,6-di-O-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino-β-D-glucopyranosyl)-(1→4)-O-3,6-di-O-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino-β-D-glucopyranoside (15): To a solution of **S8a** (1.0 g, 0.673 mmol) in 10 mL (CH₂Cl₂ : H₂O, 10/1) was added DDQ (0.183 g, 0.808 mmol) at 0 °C and the resulting reaction mixture was stirred until TLC (ethyl acetate: toluene, 2/8) indicated formation of a product with consumption of the starting material. The

reaction mixture was then filtered and the organic layer washed with H₂O (2 x 30 mL). The aqueous layer was further extracted with CH₂Cl₂ (2 x 50 mL). The combined organic layers were washed with brine solution (40 mL), dried over Na₂SO₄, and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 15% EA in toluene) to afford **15** (0.652 g, 70%) as colorless foam. TLC (ethyl acetate: toluene =2/8 v/v): R_f = 0.24; ¹H NMR (600 MHz, CDCl₃): δ 7.44-7.16 (m, 25H), 5.44 (s, 1H), 5.15 (d, *J* = 3.2 Hz, 1H), 4.88 (t, *J* = 8.9 Hz, 2H), 4.54-4.40 (m, 12H), 4.08-3.58 (m, 3H), 3.56-3.40 (m, 14H), 3.25-3.20 (m, 4H), 3.02-3.00 (m, 1H), 2.16 (s, 3H, -C(O)CH₃), 1.51-1.47 (m, 4H, -CCH₂C-, linker), 1.30-0.99 (m, 2H, -CCH₂C-, linker); ¹³C NMR (150 MHz, CDCl₃): δ 171.5, 168.6, 167.0, 138.7, 138.4, 138.38, 138.2, 137.2, 137.8, 137.5, 134.2, 133.2, 133.6, 131.7, 131.1, 131.45, 129.8, 129.06, 128.9, 128.2, 128.4, 128.2, 128.4, 128.7, 127.8, 127.87, 127.83, 127.4, 127.4, 127.0, 127.2, 127.29, 127.6, 127.20, 126.88, 126.24, 125.1, 123.6, 123.3, 101.5, 102.7, 99.7, 98.0, 97.1, 78.13, 78.55, 78.05, 75.82, 74.57, 74.37, 74.27, 73.25, 73.30, 72.78, 71.26, 69.89, 68.85, 69.43, 68.25, 67.73, 66.67, 56.57, 55.71, 51.10, 28.67, 28.27, 23.00, 21.47, 21.02 ; ESI-MS: *m/z* calcd for C₆₆H₇₅Cl₆N₅O₁₉: 1455.0420; found 1478.3082 (*M* + Na)⁺.

v. Synthesis of high-mannose type oligosaccharides (G1-G6)

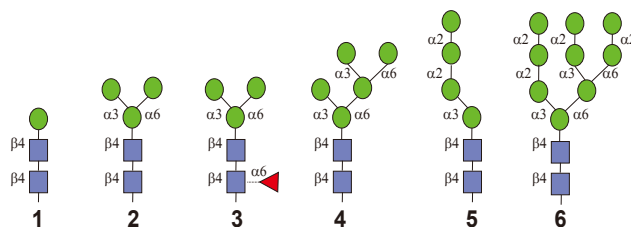
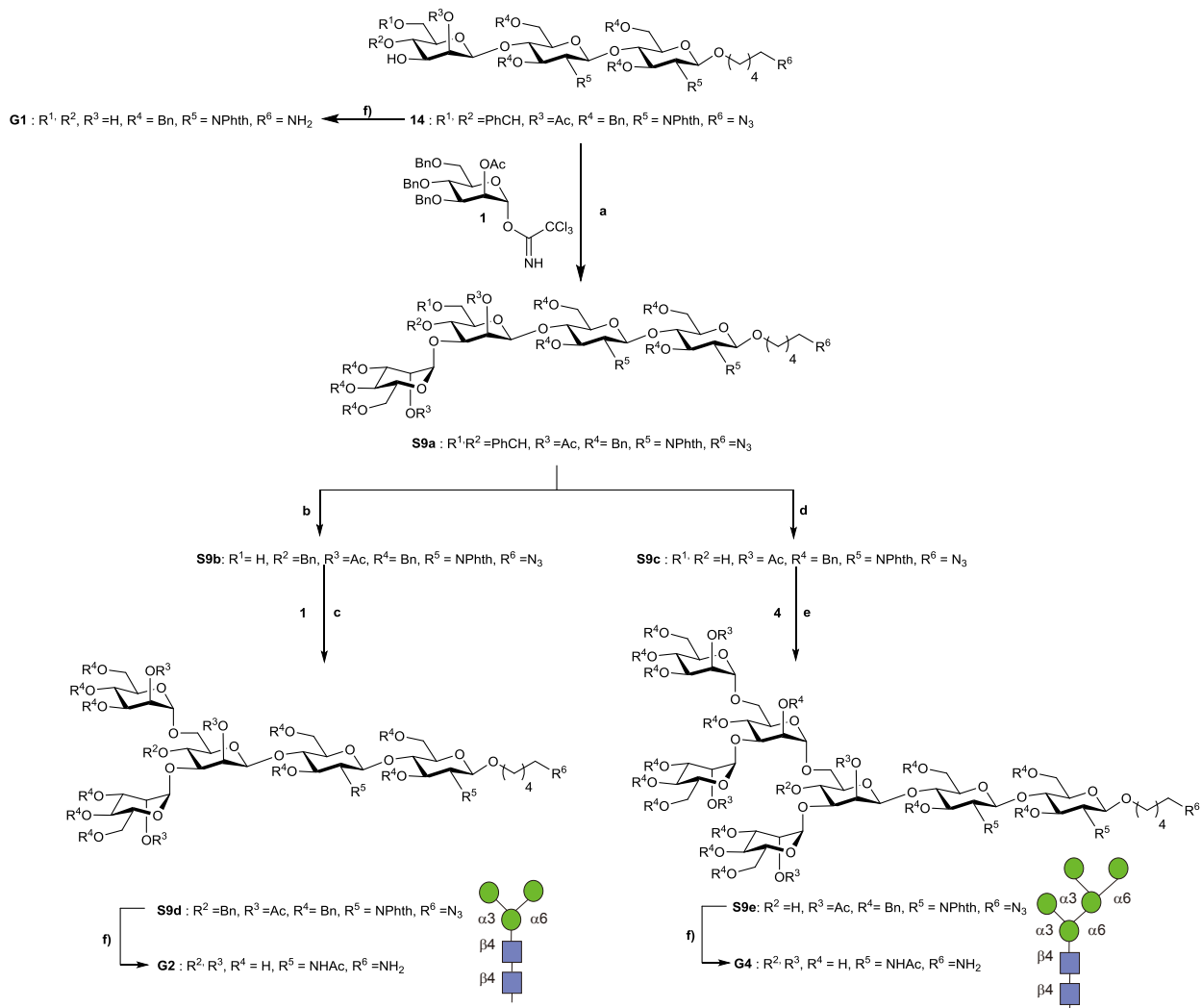


Figure S1 | Structures of high mannose type glycans and their fragments

With the donors **1-5** in hand, we investigated their glycosylation with core trisaccharide acceptor **14** and the coupling products (Scheme S9). The glycosylation of **14** with known imidate donor **1** to give tetrasaccharide **S9a** in 70% yield¹¹. After selective benzylidene ring opening of **S9a**, the resulting 6''-OH **S9b** was glycosylated with **1** to afford the desired fully protected Man₃GlcNAc₂

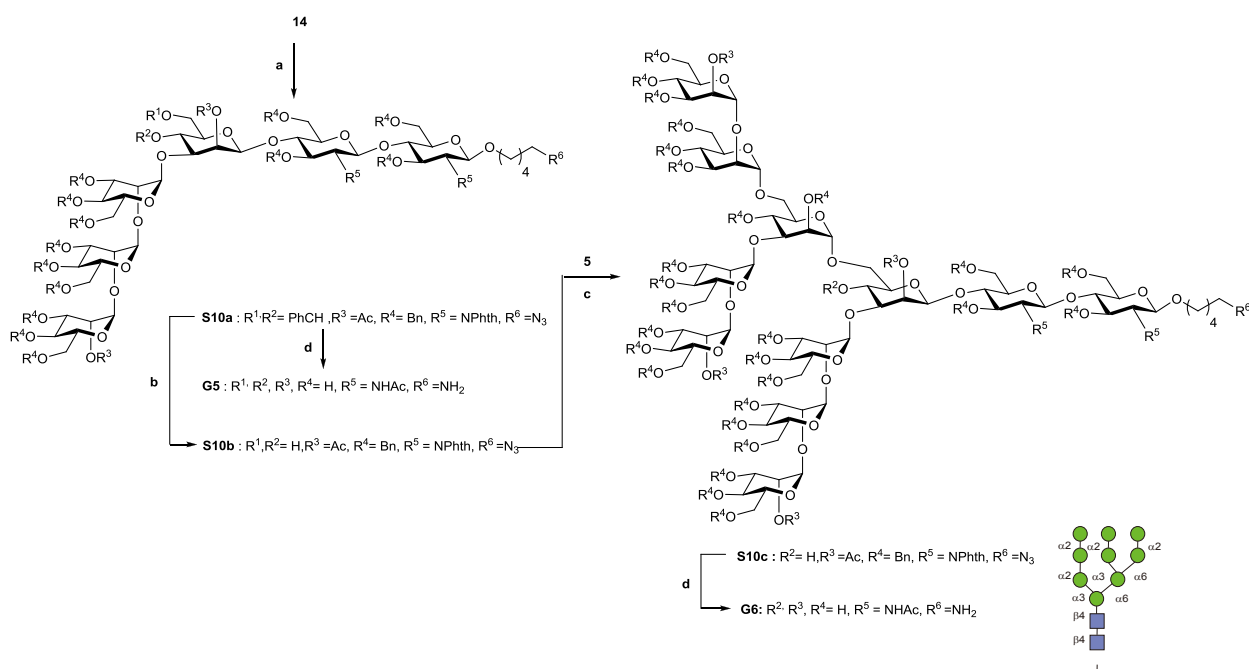
pentasaccharide **S9d**, an early intermediate in *N*-Glycan biosynthesis and thus a conserved motif in all *N*-glycans, in 60% yield. In another pathway, opening of the benzylidene group **S9a** resulted in diol **S9c**, which was further condensed with trimannosyl fluoride **4** by using Cp_2HfCl_2 and AgOTf to obtain heptasaccharide **S9e**, an intermediate in GlcNAc mediated branching in the Golgi apparatus, in 52% yield¹¹.



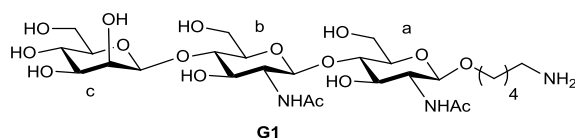
Scheme S9 | Preparation of Man_3 and $\text{Man}_5\text{GlcNAc}_2$. **a**, $\text{BF}_3 \cdot \text{OEt}_2$, CH_2Cl_2 , 4 Å MS, -40°C , 2 h, 70%; **b**, Triethyl silane, PhBCl_2 , CH_2Cl_2 , 4 Å MS, -78°C , 1 h, 82%; **c**, **1**, $\text{BF}_3 \cdot \text{OEt}_2$, CH_2Cl_2 , 4 Å MS, -60°C to -20°C , 2 h, 60%; **d**, *p*TsOH, CH_3CN , 2 h, 66%; **e**, **4**, AgOTf , Cp_2HfCl_2 , Toluene, 4 Å MS, -40°C , 2 h, 52%; **f**, (1) $\text{NH}_2\text{CH}_2\text{CH}_2\text{NH}_2$, *n*BuOH, 90°C , overnight; (2) Ac_2O ,

pyridine, overnight; (3) NaOMe, MeOH, overnight; (4) Pd(OH)₂, MeOH: H₂O: HCOOH (5:3:2), H₂; **G1**: 65%; **G2**: 52%; **G4**: 29%; Cp₂HfCl₂: Bis (cyclopentadienyl) hafnium Dichloride.

To synthesize Man₉GlcNAc₂, a major glycoform found on HIV-1 gp120 surface and an important component of epitope recognized by broadly neutralizing antibodies, compound **14** was subjected to 3"-O glycosylation with fluoride **2** under the promotion of Cp₂HfCl₂/AgOTf to get hexasaccharide **S10a** in 65% yield. The *p*-toluene sulfonic acid mediated ring opening of **S10a** provided diol **S10b**, which was further glycosylated with **5** to afford undecasaccharide **S10c** (Scheme S10).

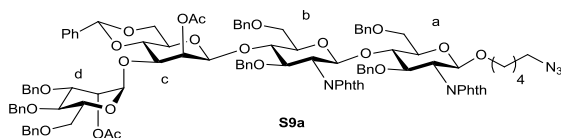


Scheme S10 | Preparation of Man₄ (G5) and Man₉GlcNAc₂ (G6). **a**, **2**, AgOTf, Cp₂HfCl₂, Toluene, 4 Å MS, -40 °C, 2 h, 58%; **b**, *p*-TsOH, CH₃CN, 2 h, 60%; **c**, **5**, (BrC₆H₄)₃NSbCl₆, CH₃CN, 4 Å MS, -10 °C to RT, 4 h, 52%; **d**, (1) NH₂CH₂CH₂NH₂, *n*BuOH, 90 °C, overnight; (2) Ac₂O, pyridine, overnight; (3) NaOMe, MeOH, overnight; (4) Pd(OH)₂, MeOH: H₂O: HCOOH (5:3:2), H₂; **G5**:42%; **SG6**: 26%; (BrC₆H₄)₃NSbCl₆: Tris (4-bromophenyl) ammoniumyl hexachloroantimonate.



5-Aminopentyl- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-

(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranoside (G1): Compound **14** (0.150 g, 0.110 mmol) was deprotected by following general procedure 2 (Method 1) to yield desired trisaccharide **G1** (0.045 g, 65%) as a white solid. ^1H NMR (600 MHz, D_2O): δ 4.72 (s, 1H, overlapped with D_2O , H-1^c), 4.60 (d, $J = 7.8$ Hz, 1H, H-1^a), 4.49 (d, $J = 7.8$ Hz, 1H, H-1^b), 4.06 (d, $J = 3$ Hz, 1H), 3.94-3.89 (m, 4H), 3.86-3.58 (m, 14H), 3.50-3.49 (m, 1H), 3.42-3.42 (m, 1H), 2.97 (t, $J = 10.8$ Hz, 2H, $-\text{CH}_2\text{NH}_2$, linker), 2.07 (s, 3H, $-\text{C}(\text{O})\text{CH}_3$), 2.03 (s, 3H, $-\text{C}(\text{O})\text{CH}_3$), 1.68-1.65 (m, 2H, $-\text{CCH}_2\text{C}-$, linker), 1.60-1.58 (m, 2H, $-\text{CCH}_2\text{C}-$, linker), 1.40-1.39 (m, 2H, $-\text{CCH}_2\text{C}-$, linker); ^{13}C NMR (150 MHz, D_2O): δ 177.32, 177.17, 173.76, 104.12 (C-1^a, $^1J_{\text{C,H}} = 163.1$), 103.80 (C-1^b, $^1J_{\text{C,H}} = 162.2$), 102.80 (C-1^c, $^1J_{\text{C,H}} = 160.2$), 82.07, 81.35, 79.15, 77.31, 77.25, 75.49, 75.13, 74.68, 73.23, 72.84, 69.34, 63.65, 62.83, 62.77, 57.75, 57.72, 42.05, 30.78, 29.10, 24.86, 24.84, 24.83; ESI-MS: m/z calcd for $\text{C}_{27}\text{H}_{49}\text{N}_3\text{O}_{16}$: 671.3005; found 694.3115 ($M + \text{Na}$)⁺.

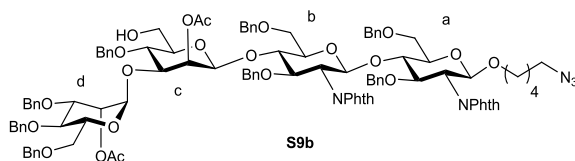


5-Azidopentyl-O-(2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3)-2-O-acetyl-

4,6-O-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido-

β -D-glucopyranosyl)-(1 \rightarrow 4)-O-3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranoside (S91): A mixture of trichloroacetimidate **1** (0.278 g, 0.437 mmol), chitobiose acceptor **14** (0.300 g, 0.219 mmol) and activated 4 Å molecular sieves in dry CH_2Cl_2 (10 mL) was stirred at room temperature for 1 h. The reaction was cooled to -40 °C, boron trifluoride ethyl etherate (12 μL , 0.109 mmol) was then added slowly and the resulting reaction mixture was stirred for 2 h. The reaction was quenched by adding Et_3N , diluted with CH_2Cl_2 , filtered through

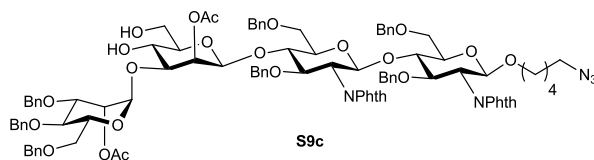
Celite and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 15% EA in toluene) to afford **S9a** (0.310 g, 70%) as white foam. TLC (ethyl acetate: toluene = 2/8, v/v): $R_f = 0.46$; $^1\text{H NMR}$ (600 MHz, CDCl_3): δ 7.84-7.65 (m, 8H, Ar-H), 7.38-7.31 (m, 2H, Ar-H), 7.30-7.18 (m, 28H, Ar-H), 6.98-6.90 (m, 7H, Ar-H), 6.75-6.71 (m, 3H, Ar-H), 5.45 (s, 1H, Ph-CH, benzylidene), 5.43-5.43 (d, $J = 3.4$ Hz, 1H, H-2^d), 5.34 (d, $J = 3.1$ Hz, 1H, H-2^c), 5.21 (d, $J = 8.1$ Hz, 1H, H-1^a), 5.19 (d, $J = 2.1$ Hz, 1H, H-1^d), 4.90 (d, $J = 8.1$ Hz, 1H, H-1^a), 4.83-4.81 (m, 3H), 4.67 (s, 1H), 4.67-4.61 (m, 2H), 4.53-4.43 (m, 7H), 4.38-4.23 (q, 2H), 4.22-4.16 (m, 1H), 4.13-4.06 (m, 6H), 3.87-3.75 (m, 6H), 3.66-3.60 (m, 3H), 3.53-3.45 (m, 3H), 3.38 (dd, $J = 4.2$ & 12.0 Hz, 1H), 3.26-3.17 (m, 3H), 3.09-3.07 (m, 1H), 2.87-2.82 (m, 2H), 2.06 (s, 3H, -C(O)CH₃), 2.03 (s, 3H, -C(O)CH₃), 1.35-1.23 (m, 4H, -CCH₂C-, linker), 1.07-1.01 (m, 2H, -CCH₂C-, linker); $^{13}\text{C NMR}$ (150 MHz, CDCl_3): δ 169.87, 168.75, 167.83, 138.97, 138.47, 138.17, 137.36, 134.30, 134.10, 131.72, 129.12, 128.90, 128.64, 128.55, 128.51, 128.32, 128.23, 128.09, 127.84, 127.46, 128.14, 126.24, 123.40, 101.44, 99.38, 98.90, 98.37 (C-1^d, $^1J_{\text{C,H}} = 172$ Hz), 97.31, 79.05, 78.84, 77.89, 76.14, 75.02, 74.78, 73.70, 73.40, 73.06, 73.04, 72.37, 72.03, 71.00, 69.11, 69.85, 69.11, 68.85, 68.59, 68.50, 67.84, 66.70, 56.85, 55.99, 51.38, 28.95, 28.55, 23.28, 21.33, 21.16; ESI-MS: m/z calcd for $\text{C}_{105}\text{H}_{107}\text{N}_5\text{O}_{25}$: 1838.7180; found 1861.7223 ($M + \text{Na}$)⁺.



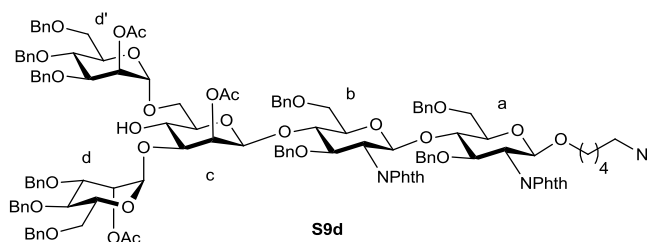
5-Azidopentyl-O-(2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1→3)-2-O-acetyl-4-O-benzyl- β -D-mannopyranosyl-(1→4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)-(1→4)-O-3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranoside

(S9b): A mixture of tetrasaccharide **S9a** (0.280 g, 0.152 mmol) and 4 Å activated molecular

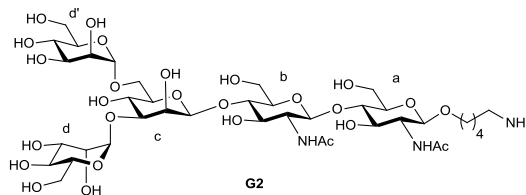
sieves in CH₂Cl₂ (10 mL) was stirred at room temperature for 1 h. Reaction was cooled to -78 °C, triethyl silane (73 μL, 0.456 mmol) and dichlorophenyl borane (69 μL, 0.532 mmol) were added and stirred for 1 h. The reaction was quenched by adding Et₃N, filtered through Celite and concentrated in *vacuo*. The residue was co-distilled with methanol 2-3 times before being purified by flash column chromatography (0% → 15% EA in toluene) to afford 6"-OH **S9b** (0.230 g, 82%). TLC: (ethyl acetate: toluene =2/8, v/v): R_f = 0.36; ¹H NMR (600 MHz, CDCl₃): δ 7.85-7.52 (m, 8H, Ar-H), 7.52-7.12 (m, 30H, Ar-H), 7.00-6.88 (m, 8H, Ar-H), 6.74-6.70 (m, 2H, Ar-H), 5.39 (d, *J* = 1.8 Hz, 1H, H-2^d), 5.32 (d, *J* = 3.6 Hz, 1H, H-2^c), 5.21 (d, *J* = 8.4 Hz, 1H, H-1^a), 5.10 (s, 1H, H-1^d), 4.90-4.81 (m, 4H), 4.67-4.61 (m, 2H, overlapped H-2^b), 4.59 (s, 1H, H-1^c), 4.59-4.44 (m, 9H), 4.37(d, *J* = 12 Hz, 2H), 4.24-4.12 (m, 3H), 4.10-4.05 (m, 3H), 3.90-3.77 (m, 3H), 3.69-3.58 (m, 7H), 3.49 (t, *J* = 11.5 Hz, 2H), 3.41-3.36 (dd, *J* = 6.1, 11.2 Hz, 2H), 3.26 (dd, *J* = 6.0, 9.2 Hz, 1H), 3.26-3.18 (m, 2H), 3.06-2.98 (m, 1H), 2.88-2.81 (m, 2H, linker), 2.06 (s, 3H, -C(O)CH₃), 2.04 (s, 3H, -C(O)CH₃), 1.37-1.22 (m, 4H, linker), 1.07 -1.00 (m, 2H, linker); ¹³C NMR (150 MHz, CDCl₃): δ 170.45, 169.88, 168.78, 168.24, 167.86, 138.95, 138.92, 138.77, 138.58, 138.21, 138.11, 137.95, 137.37, 134.35, 134.16, 133.89, 132.09, 131.70, 128.90, 128.76, 128.71, 128.64, 128.58, 128.51, 128.41, 128.31, 128.08, 127.96, 127.81, 127.64, 127.13, 123.97, 123.46, 100.14, 98.40, 98.36, 97.36, 76.24, 75.52, 75.30, 74.96, 74.71, 74.51, 74.29, 73.41, 73.09, 72.55, 72.12, 69.41, 69.20, 68.85, 68.48, 67.93, 67.71, 61.87, 56.78, 56.00, 51.38, 28.96, 28.55, 23.28, 21.37; ESI-MS: *m/z* calcd for C₁₀₅H₁₀₉N₅O₂₅: 1840.7337; found 1863.7383 (*M* + Na)⁺.



5-Azidopentyl-O-(2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3)-2-O-acetyl- β -D-mannopyranosyl-(1 \rightarrow 4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)-(1 \rightarrow 4)-O-3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranoside (S9c): *p*-Toluenesulfonic acid monohydrate (0.045 g, 0.239 mmol) was added to a stirred solution of tetrasaccharide **S9a** (0.220 g, 0.119 mmol) in acetonitrile (10 mL) at room temperature. After 8 h, the reaction mixture was quenched with Et₃N and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 15% EA in toluene) to give the title diol **S9c** (0.140 g, 66%) as white solid. TLC: (ethyl acetate: toluene = 2/8, v/v): R_f = 0.26; ¹H NMR (600 MHz, CDCl₃): δ 7.85-7.52 (m, 8H, Ar-H), 7.32-7.17 (m, 25H, Ar-H), 6.98-6.91 (m, 7H, Ar-H), 6.75-6.71 (m, 3H, Ar-H), 5.30 (d, *J* = 3.6 Hz, 1H,), 5.21(d, *J* = 8.6 Hz, 1H, H-1^a), 5.20 (s, 2H, overlapped H-1^d), 4.89 (d, *J* = 8.4 Hz, 1H, H-1^b), 4.85-4.82 (m, 3H), 4.64-4.60 (m, 3H), 4.54-4.46 (m, 7H), 4.38 (d, *J* = 12.0 Hz, 1H), 4.34 (d, *J* = 12.2 Hz, 1H), 4.21-4.04 (m, 5H), 3.92 (m, 1H), 3.87-3.79 (m, 3H), 3.70-3.59 (m, 5H), 3.52-3.43 (m, 4H), 3.39 (dd, *J* = 3.2, 8.3 Hz, 1H), 3.28 (dd, *J* = 3.2, 8.2 Hz, 1H), 3.29-3.19 (m, 2H), 2.29 (m, 1H), 2.86-2.83 (m, 3H), 2.11 (s, 3H, -C(O)CH₃), 2.03 (s, 3H, -C(O)CH₃), 1.35-1.23 (m, 4H, -CCH₂C-, linker), 1.06-1.01 (m, 2H, -CCH₂C-, linker); ¹³C NMR (150 MHz, CDCl₃): δ 170.76, 169.88, 138.92, 138.70, 138.62, 138.52, 138.12, 138.06, 134.27, 134.07, 133.81, 131.95, 131.64, 128.82, 128.64, 128.59, 128.52, 128.50, 128.39, 138.33, 128.20, 128.09, 128.06, 128.03, 128.01, 127.94, 127.81, 127.75, 127.54, 127.42, 127.29, 127.06, 123.90, 123.36, 98.51, 98.30, 98.24(C-1^d, ¹*J*_{C,H} = 173 Hz), 97.30, 79.07, 78.38, 77.60, 76.10, 75.52, 75.03, 74.79, 74.71, 74.59, 74.51, 73.39, 73.03, 72.07, 72.01, 71.30, 69.53, 69.27, 69.05, 68.44, 67.70, 67.17, 62.44, 56.74, 55.94, 51.32, 28.89, 28.49, 23.22, 21.31, 21.22; ESI-MS: *m/z* calcd for C₉₈H₁₀₃N₅O₂₅: 1750.6868; found 1773.6913 (*M* + Na)⁺.

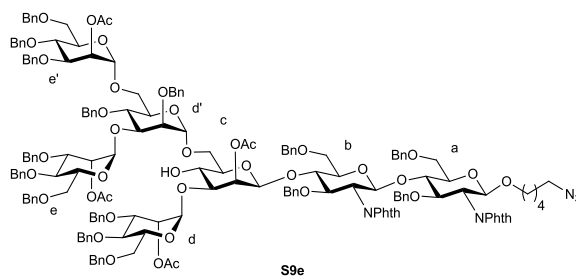


5-Azidopentyl-O-di-(2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3), (1 \rightarrow 6)-2-O-acetyl-4-O-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)-(1 \rightarrow 4)-O-3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranoside (S9d): A mixture of donor **1** (0.113 g, 0.178mmol), acceptor **S9b**(0.220 g, 0.119 mmol) and activated 4 Å molecular sieves in dry CH₂Cl₂ (10 mL) was stirred at room temperature for 1 h. The reaction was cooled to -60 °C, boron trifluoride ethyl etherate (12 μ L, 0.109 mmol) was added slowly and the resulting reaction mixture was stirred for 2 h at -20 °C. The reaction was quenched by adding Et₃N, diluted with CH₂Cl₂, filtered through Celite and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 20% EA in hexane) to afford **S9d** (0.170 g, 60%) as colorless foam. TLC: (ethyl acetate: hexane = 3/7, v/v): R_f = 0.46; ¹H NMR (600 MHz, CDCl₃): δ 7.77-7.59 (m, 8H, Ar-H), 7.32-7.16 (m, 37H, Ar-H), 7.14-7.08 (m, 2H, Ar-H), 6.91-6.89 (m, 5H, Ar-H), 6.73-6.67 (m, 6H, Ar-H), 5.45 (s, 1H, H-2^d), 5.34 (d, *J* = 3.1 Hz, 1H, H-2^c), 5.32 (s, 1H, H-2^d), 5.17 (d, *J* = 8.4 Hz, 1H, H-1^a), 5.09 (s, 1H, H-1^d), 5.0 (s, 1H, H-1^d), 4.87 (d, *J* = 8.4 Hz, 1H, H-1^b), 4.83-4.78 (q, 2H), 4.74-4.71 (q, 2H), 4.65-4.61 (m, 4H), 4.56-4.42 (m, 15H), 4.31 (d, *J* = 9.2 Hz, 1H), 4.12-4.03 (m, 5H), 4.03-3.76 (m, 7H), 3.68-3.43 (m, 9H), 3.30 (dd, *J* = 3.2, 9.1 Hz, 1H), 3.21-3.15 (m, 2H), 3.13 (d, *J* = 9.2 Hz, 1H), 3.09 (d, *J* = 8.4 Hz, 1H), 2.89-2.79 (m, 2H, -CH₂N₃-, linker), 2.12 (s, 3H, -C(O)CH₃), 2.07 (s, 3H, -C(O)CH₃), 1.69 (s, 3H, -C(O)CH₃), 1.37-1.12 (m, 4H, -CCH₂C-, linker), 1.07-1.01 (m, 2H, -CCH₂C-, linker); ¹³C NMR (150 MHz, CDCl₃): δ 170.45, 170.22, 170.18, 168.21, 167.71, 138.93, 138.72, 138.68, 138.50, 138.24, 138.22, 138.07, 133.83, 133.75, 128.87, 128.79, 128.67, 128.63, 128.58, 128.42, 128.21, 128.06, 127.99, 127.96, 127.81, 127.76, 127.70, 127.46, 127.08, 123.36, 100.13 (C-1^a, ¹*J*_{C,H} = 159.3 Hz), 99.39 (C-1^d, ¹*J*_{C,H} = 171.5 Hz), 98.35 (C-1^b, ¹*J*_{C,H} = 162.3 Hz), 98.31(C-1^d, ¹*J*_{C,H} = 170 Hz), 97.16, 76.20, 76.05, 75.58, 75.21, 74.97, 74.89, 74.70, 74.47, 74.28, 73.94, 73.72, 73.00, 72.25,72.20, 71.70,71.50, 69.15,69.07, 69.04,68.99, 68.50, 68.45, 67.80, 65.45, 56.83, 51.37, 29.94, 23.27, 21.31, 21.28, 20.83; ESI-MS: *m/z* calcd for C₁₃₄H₁₃₉N₅O₃₁: 2314.9379; found 2337.9461 (*M* + Na)⁺.



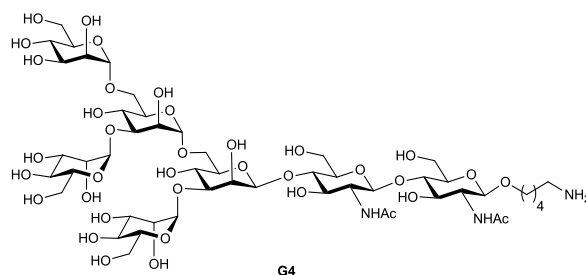
5-Aminopentyl-di-(α -D-mannopyranosyl)-(1 \rightarrow 3),(1 \rightarrow 6)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-gluco-pyranoside

(G2): Pentasaccharide **S9d** (0.090 g, 0.038 mmol) was deprotected by following the general procedure 2 (Method 1) to afford compound **G2** (0.020 g, 52%) as a white powder. ^1H NMR (600 MHz, D_2O): δ 5.10 (d, $J = 1.3$ Hz, 1H, H-1 $^{\text{d}}$), 5.09 (d, $J = 1.5$ Hz, 1H, H-1 $^{\text{d}}$), 4.78 (s, 1H, overlapped with D_2O , H-1 $^{\text{c}}$), 4.59 (d, $J = 8.4$ Hz, 1H, H-1 $^{\text{a}}$), 4.49 (d, $J = 7.8$ Hz, 1H, H-1 $^{\text{b}}$), 4.25 (s, 1H), 4.01 (d, $J = 1.5$ Hz, 1H), 4.00 (d, $J = 1.3$ Hz, 1H), 3.97-3.77 (m, 8H), 3.75-3.65 (m, 19H), 3.52-3.48 (m, 2H), 2.98 (t, $J = 11.2$ Hz, 2H), 2.06 (s, 3H), 2.02 (s, 3H), 1.68-1.65 (m, 2H), 1.60-1.57 (m, 2H), 1.49-1.40 (m, 2H); ^{13}C NMR (150 MHz, D_2O): δ 181.17, 174.69, 174.42, 102.51 (C-1 $^{\text{d}}$, $^1J_{\text{C,H}} = 171.5$ Hz), 101.34, 101.04, 100.35 (C-1 $^{\text{d}}$, $^1J_{\text{C,H}} = 173.5$ Hz), 99.60, 80.48, 79.61, 79.30, 74.49, 74.36, 74.16, 73.43, 72.66, 72.37, 71.93, 70.39, 70.31, 70.13, 70.09, 69.98, 69.85, 66.84, 66.76, 65.80, 61.12, 90.93, 60.07, 59.97, 54.97, 54.85, 39.30, 28.03, 26.35, 23.07, 22.17, 22.07; ESI-MS: m/z calcd for $\text{C}_{39}\text{H}_{69}\text{N}_3\text{O}_{26}$: 995.4067; found 1018.4106 ($M + \text{Na}$) $^+$.



5-Azidopentyl-O-(2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3))-2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3)}-[2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 6)]-2,4-di-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 6)}-2-O-acetyl- β -D-mannopyranosyl-(1 \rightarrow 4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)-

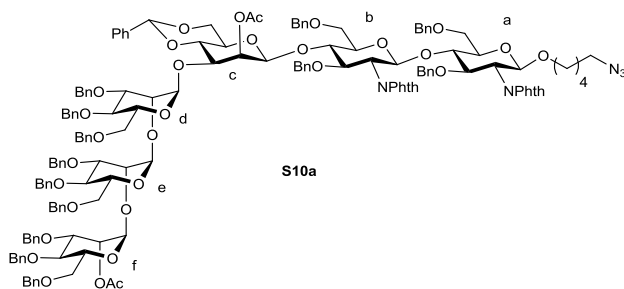
(1→4)-O-3,6-di-O-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranoside (S9e): A mixture of Silver triflate (0.127 g, 0.497 mmol), Bis(cyclopentadienyl) hafnium dichloride (0.121 g, 0.319 mmol) and 4 Å activated molecular sieves in dry toluene (10 mL) was stirred at room temperature for 1 h. The reaction mixture was then cooled to -40 °C, a solution of donor **4** (0.103 g, 0.085 mmol) and acceptor **S9c** (0.125 g, 0.071 mmol) in 5 mL toluene was added. The mixture was stirred for 2 h, quenched with Et₃N, diluted with CH₂Cl₂ and filtered through celite. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), and a brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 15% EA in toluene) to afford **S9e** (0.110 g, 52%) as colorless foam. TLC: (acetone: toluene = 2/8, v/v): R_f = 0.36; ¹H NMR (600 MHz, CDCl₃): δ 7.77-7.40 (m, 8H, Ar-H), 7.32-7.07 (m, 65H, Ar-H), 6.99-6.88 (m, 4H, Ar-H), 6.79-6.68 (m, 6H, Ar-H), 5.44 (s, 1H, H-2^d), 5.43 (s, 1H, H-2^e), 5.41 (s, 1H, H-2^{e'}), 5.29 (d, *J* = 3.4 Hz, 1H, H-2^c), 5.18 (s, 2H, H-1^{d,d'}), 5.16 (s, 1H, H-1^e), 5.09 (s, 1H, H-1^{e'}), 4.92 (s, 1H, H-1^c), 4.91 (d, *J* = 8.5 Hz, 1H, H-1^a), 4.83-4.78 (q, 1H), 4.84-4.73 (m, 6H), 4.65-4.61 (m, 2H), 4.60-4.57 (m, 5H), 4.52-4.35 (m, 17H), 4.27 (d, *J* = 12.3 Hz, 1H), 4.20-4.19 (m, 1H), 4.10-3.99 (m, 6H), 3.97-3.70 (m, 13H), 3.69-3.57 (m, 6H), 3.56-3.31 (m, 6H), 3.30 (dd, *J* = 3.2, 9.2 Hz, 1H), 3.30-3.18 (m, 5H), 2.97-2.96 (m, 1H), 2.89-2.79 (m, 2H, -CCH₂C-, linker), 2.12 (s, 3H, -C(O)CH₃), 2.07 (s, 3H, -C(O)CH₃), 2.01 (s, 6H, -C(O)CH₃), 1.33-1.23 (m, 4H, -CCH₂C-, linker), 1.06-0.99 (m, 2H, -CCH₂C-, linker); ¹³C NMR (150 MHz, CDCl₃): δ 170.56, 170.42, 170.26, 170.02, 168.45, 168.69, 138.95, 138.93, 138.72, 138.68, 138.58, 138.24, 138.22, 138.07, 133.83, 132.09, 131.76, 128.87, 128.79, 128.67, 128.63, 128.61, 128.58, 128.52, 128.42, 128.13, 128.06, 127.99, 127.92, 127.81, 127.76, 127.74, 127.46, 127.08, 123.71, 123.36, 100.13 (C-1^{e,e'}, ¹*J*_{C,H} = 169.2 Hz), 99.39 (C-1^b, ¹*J*_{C,H} = 162.2 Hz), 98.35 (C-1^d, ¹*J*_{C,H} = 171.5 Hz), 98.31 (C-1^a, ¹*J*_{C,H} = 160.2 Hz), 97.16 (C-1^{d'}, ¹*J*_{C,H} = 170.5 Hz), 76.20, 76.05, 75.42, 74.97, 74.89, 74.80, 74.70, 74.65, 74.47, 74.28, 73.94, 73.68, 73.26, 72.60, 72.25, 72.20, 71.70, 71.50, 69.15, 69.07, 69.04, 68.99, 68.50, 68.45, 67.80, 65.54, 56.73, 55.90, 51.29, 31.14, 29.91, 28.87, 28.47, 23.19, 21.33, 21.27, 21.19; HRMS (MALDI-TOF): *m/z* calcd for C₁₇₆H₁₈₅N₅O₄₂: 3042.2400; found 3065.2210 (*M* + Na)⁺.



5-Aminopentyl- α -D-mannopyranosyl(1 \rightarrow 3),[di-(α -D-mannopyranosyl)-(1 \rightarrow 3), (1 \rightarrow 6)- α -D-mannopyranosyl](1 \rightarrow 6)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-

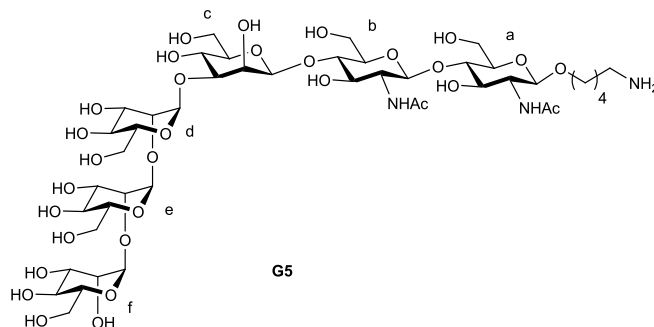
glucopyranosyl -(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranoside (G4):- Heptasaccharide

S9e (0.140 g, 0.024 mmol), was deprotected by following the general procedure 2 (Method 1) to obtain title compound **G4** (0.039 g, 29%) as a white solid. ^1H NMR (600 MHz, D_2O): δ 5.09 (s, 1H, H-1^d), 4.89 (s, 1H, H-1^{d'}), 4.86 (d, $J = 1.5$ Hz, 1H, H-1^c), 4.75 (d, $J = 1.5$ Hz, 1H, H-1^e), 4.76 (s, 1H, overlapped with D_2O , H-1^c), 4.50 (d, $J = 7.8$ Hz, 1H, H-1^a), 4.35 (d, $J = 7.9$ Hz, 1H, H-1^b), 4.17 (d, $J = 1.7$ Hz, 1H), 4.07 – 4.05 (m, 1H), 4.00-3.97 (m, 2H), 3.94-3.37 (m, 40H), 2.90-2.85 (m, 2H, $-\text{CH}_2\text{NH}_2$, linker), 1.98 (s, 3H, $-\text{C}(\text{O})\text{CH}_3$), 1.94 (s, 3H, $-\text{C}(\text{O})\text{CH}_3$), 1.61-1.46 (m, 4H, $-\text{CCH}_2\text{C}-$, linker), 1.35-1.26 (m, 2H, $-\text{CCH}_2\text{C}-$, linker); ^{13}C NMR (150 MHz, D_2O): δ 177.2, 172.3, 102.44, 101.44, 101.01, 100.37, 99.85, 99.25, 80.54, 79.49, 79.28, 78.58, 74.45, 74.43, 72.50, 71.93, 70.64, 70.24, 70.15, 70.04, 70.01, 69.87, 69.46, 69.39, 66.69, 66.66, 65.94, 65.91, 65.56, 65.13, 65.12, 60.93, 60.91, 60.43, 60.13, 60.04, 59.97, 54.90, 54.86, 39.28, 27.98, 26.48, 22.08, 22.02, 21.99; ESI-MS: m/z calcd for $\text{C}_{51}\text{H}_{89}\text{N}_3\text{O}_{36}$: 1319.5123; found 1342.5165 ($M + \text{Na}$)⁺.

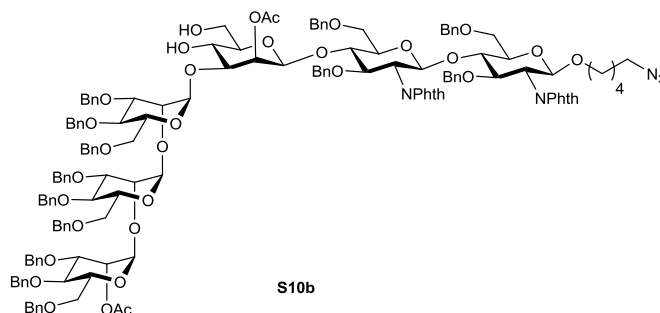


5-Azidopentyl-O-(2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3)-2-O-acetyl-4,6-O-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)-(1 \rightarrow 4)-O-3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranoside (S10a):

A mixture of Silver triflate (0.328 g, 1.28 mmol), Bis(cyclopentadienyl) hafnium dichloride (0.340 g, 0.897 mmol) and 4 Å activated molecular sieves in dry toluene (10 mL) was stirred at rt for 1 h. The reaction mixture was then cooled to -40 °C, a solution of donor **2** (0.383 g, 0.282 mmol) and acceptor **14** (0.350 g, 0.256 mmol) in 5 mL toluene was added. The mixture was stirred for 2 h, quenched with Et₃N, diluted with CH₂Cl₂ and filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), and a brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 15% EA in toluene) to afford **S10a** (0.405 g, 58%) as colorless foam. TLC: (ethyl acetate: toluene = 2/8, v/v): R_f = 0.46; ¹H NMR (600 MHz, CDCl₃): δ 7.89-7.35 (m, 8H, Ar-H), 7.38-7.12 (m, 56H, Ar-H), 7.12-7.08 (m, 3H, Ar-H), 7.03-6.88 (m, 7H, Ar-H), 6.78-6.69 (m, 4H, Ar-H), 5.50 (s, 1H, H-1^f), 5.37 (d, *J* = 1.4 Hz, 1H, H-2^c), 5.25 (s, 1H, Ph-CH-, benzylidene), 5.23-5.17 (m, 3H, overlapped, H-1^{d,e,f}), 4.98 (s, 1H, H-1^c), 4.95 (d, *J* = 7.5 Hz, 1H, H-1^a), 4.91-4.53 (m, 5H), 4.69-4.58 (m, 3H), 4.55-4.28 (m, 16H), 4.25-4.00 (m, 13H), 3.95-3.91 (m, 1H), 3.91-3.60 (m, 10H), 3.58 (d, *J* = 12 Hz, 1H), 3.51-3.45 (m, 4H), 3.42-3.38 (m, 3H), 3.29-3.10 (m, 3H), 2.98-2.78 (m, 3H), 2.08 (s, 3H, -C(O)CH₃), 1.99 (s, 3H, -C(O)CH₃), 1.35-1.23 (m, 4H, -CCH₂C-, linker), 1.07-1.00 (m, 2H, -CCH₂C-, linker); ¹³C NMR (150 MHz, CDCl₃): δ 170.38, 169.81, 168.76, 168.26, 139.06, 138.95, 138.92, 138.87, 138.60, 137.36, 128.85, 128.66, 128.64, 128.58, 128.55, 128.48, 128.43, 128.39, 128.35, 128.26, 128.10, 128.02, 127.82, 127.71, 101.48, 100.08, 99.75, 99.39, 98.36, 97.27, 77.80, 78.75, 76.10, 75.38, 74.93, 74.82, 74.70, 73.58, 73.28, 73.21, 73.06, 72.79, 72.65, 72.57, 72.17, 71.01, 69.60, 69.10, 68.96, 68.63, 66.74, 56.85, 55.97, 51.37, 28.95, 28.54, 23.27, 21.43, 21.09; ESI-MS: *m/z* calcd for C₁₅₉H₁₆₃N₅O₃₅: 2703.1054; found 2726.1214 (*M* + Na)⁺.

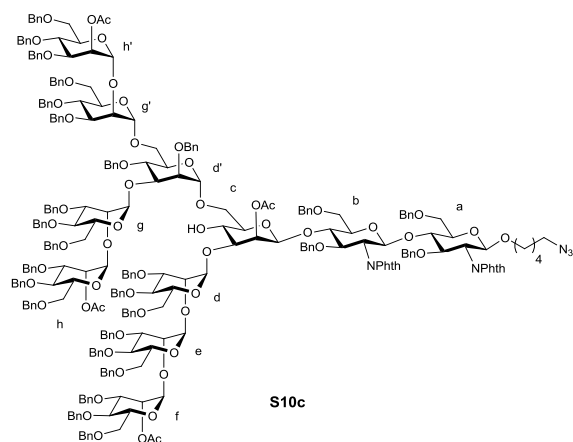


5-Aminopentyl- α -D-mannopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl-(1 \rightarrow 3)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranoside (G5): Hexasaccharide **S10a** (0.140 g, 0.051 mmol), was deprotected by following general procedure 2 (Method 1) to obtain the title compound **G5** (0.025 g, 42%) as white solid. ^1H NMR (600 MHz, D_2O): δ 5.32 (s, 1H, H-1^d), 5.27 (s, 1H, H-1^c), 5.01 (s, 1H, H-1^f), 4.74 (s, 1H, H-1^e), 4.56 (d, $J = 7.8$ Hz, 1H, H-1^a), 4.63 (d, $J = 7.8$ Hz, 1H, H-1^b), 4.18 (d, $J = 3.3$ Hz, 1H), 4.07 (s, 1H), 4.04 (d, $J = 3.2$ Hz, 1H), 4.03 (d, $J = 3.1$ Hz, 1H), 3.32-3.33 (m, 35H), 2.05 (t, $J = 11.2$ Hz, 2H, $-\text{CH}_2\text{NH}_2$, linker), 2.03 (s, 3H, $-\text{C}(\text{O})\text{CH}_3$), 2.00 (s, 3H, $-\text{C}(\text{O})\text{CH}_3$), 1.64 (m, 2H, $-\text{CCH}_2\text{C}-$, linker), 1.50 (m, 2H, $-\text{CCH}_2\text{C}-$, linker), 1.36 (m, 2H, $-\text{CCH}_2\text{C}-$, linker); ^{13}C NMR (150 MHz, D_2O): δ 174.54, 174.38, 102.17 (C-1^e, $^1J_{\text{C,H}} = 170.2$ Hz), 101.34 (C-1^b, $^1J_{\text{C,H}} = 161.2$ Hz), 101.014 (C-1^d, $^1J_{\text{C,H}} = 169.8$ Hz), 100.70 (C-1^f, $^1J_{\text{C,H}} = 172.2$ Hz), 100.56 (C-1^a, $^1J_{\text{C,H}} = 160.2$ Hz), 99.85, 80.50, 79.25, 78.66, 78.55, 78.47, 76.12, 74.48, 73.37, 73.13, 72.33, 71.88, 70.24, 70.17, 70.05, 69.95, 69.89, 66.95, 66.81, 66.71, 65.81, 61.01, 60.97, 60.88, 60.76, 60.02, 59.92, 54.92, 39.25, 27.99, 26.32, 23.19, 22.06, 22.04; ESI-MS: m/z calcd for $\text{C}_{45}\text{H}_{79}\text{N}_3\text{O}_{31}$: 1157.4590; found 1180.4591 ($M + \text{Na}$)⁺.



5-Azidopentyl-O-(2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3)-2-O-acetyl- β -D-mannopyranosyl-(1 \rightarrow 4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)-(1 \rightarrow 4)-O-3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranoside

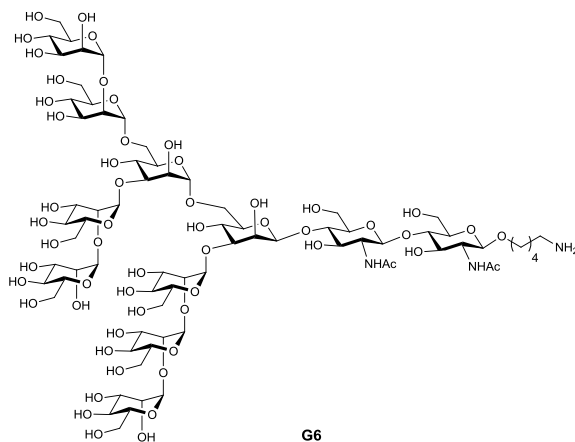
(S10b):- To a solution of **S10a** (0.205 g, 0.075 mmol) in 10 mL acetonitrile was added *p*-toluene sulfonic acid monohydrate (0.020 g, 0.113 mmol), stirred at room temperature for 9 h. The reaction was quenched with Et₃N and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 20% EA in toluene) to give diol **S10b** (0.120 g, 60%). TLC: (acetone: toluene = 2/8, v/v): R_f = 0.26; ¹H NMR (600 MHz, CDCl₃): δ 7.83-7.62 (m, 8H, Ar-H), 7.41-7.06 (m, 57H, Ar-H), 6.95-6.89 (m, 6H, Ar-H), 6.73-6.71 (m, 2H, Ar-H), 5.50 (d, *J* = 4.8 Hz, 1H, H-2^f), 5.23 (d, *J* = 1.8 Hz, 1H, H-1^d), 5.22 (d, *J* = 3.8 Hz, 1H, H-1^c), 5.19 (d, *J* = 3.2 Hz, 1H, H-1^e), 5.18 (s, 1H, H-1^f), 5.03 (d, *J* = 7.8 Hz, 1H, H-1^a), 4.88 (t, *J* = 7.8 Hz, 1H), 4.82-4.74 (m, 5H), 4.68-4.38 (m, 21H), 4.33 (d, *J* = 7.8 Hz, 1H), 4.29 (d, *J* = 7.9 Hz, 1H), 4.20-4.00 (m, 7H), 3.95-3.71 (m, 9H), 3.69-3.53 (m, 6H), 3.52-3.33 (m, 8H), 3.29-3.15 (m, 5H), 2.90-2.79 (m, 2H), 2.08 (s, 3H, -C(O)CH₃), 1.92 (s, 3H, -C(O)CH₃), 1.35-1.20 (m, 4H, -CCH₂C-, linker), 1.08-1.01 (m, 2H, -CCH₂C-, linker); ¹³C NMR (150 MHz, CDCl₃): δ 170.40, 169.88, 168.68, 138.20, 138.94, 138.78, 138.71, 138.56, 138.48, 138.38, 138.21, 138.02, 134.30, 133.84, 132.02, 128.78, 128.65, 128.63, 128.43, 128.40, 128.38, 128.25, 128.15, 128.06, 127.94, 127.88, 127.84, 127.77, 127.69, 127.65, 127.56, 127.10, 123.90, 123.41, 100.98, 99.83, 98.34, 97.32, 78.31, 78.17, 78.11, 78.00, 76.22, 75.52, 75.28, 75.05, 74.82, 74.40, 73.98, 73.67, 73.58, 73.36, 73.06, 72.59, 72.48, 72.42, 72.09, 71.32, 70.52, 69.46, 69.09, 68.83, 68.72, 68.42, 68.20, 67.61, 62.57, 56.77, 55.98, 51.37, 29.99, 29.65, 28.94, 28.54, 23.27, 21.44, 21.13, 14.41; ESI-MS: *m/z* calcd for C₁₅₂H₁₅₉N₅O₃₅: 1615.0741; found 2638.0754 (*M* + Na)⁺.



5-Azidopentyl-O-(2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2))-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2))-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3))-2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2))-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3))-2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2))-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 6))-2,4-di-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 6))-2-O-acetyl- β -D-mannopyranosyl-(1 \rightarrow 4))-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)-(1 \rightarrow 4))-O-3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranoside (S10c):

A mixture of hexasaccharide acceptor **S10a** (0.1 g, 0.038 mmol), thiomannoside donor **5** (0.104 g, 0.045 mmol) and activated 4 Å molecular sieves in CH₃CN (10 mL) was stirred at rt for 1 h. The resulting mixture was cooled to -10 °C, tris(4-bromophenyl)aminium hexachloroantimonate (0.096 g, 0.114 mmol) was added and stirred at rt for 6 h. TLC (ethyl acetate: toluene, 2/8) indicated formation of product with consumption of starting material, reaction was quenched by Et₃N. The reaction mixture was diluted with CH₂Cl₂ and filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), and a brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 10% EA in toluene) to afford **S10c** (0.095 g, 52%) as colorless foam. TLC: (acetone: toluene = 2/8, v/v): R_f = 0.41; ¹H NMR (600 MHz, CDCl₃): δ 7.77-7.50 (m, 8H, Ar-H), 7.29-6.98 (m, 126H, Ar-H), 6.95-6.89 (m, 4H, Ar-H), 6.74-6.64 (m, 5H, Ar-H), 5.56 (s, 1H, H-2^f), 5.53 (s, 1H, H-2^h), 5.51 (s, 1H^{h'}), 5.34 (s, 1H, H-2^c), 5.32 (d, *J* = 12.2 Hz, 2H), 5.18 (d,

$J = 7.8$ Hz, 1H), 5.12 (s, 1H), 5.08 (s, 1H), 5.04 (s, 2H), 4.59-3.75 (m, 116H), 3.26-3.16 (m, 4H), 3.08-3.02 (m, 2H), 2.93-2.83 (m, 2H), 2.12 (s, 3H), 2.06 (s, 3H), 2.06 (s, 3H), 1.97 (s, 3H), 1.39-1.26 (m, 4H), 1.07-1.05 (m, 2H); ^{13}C NMR (150 MHz, CDCl_3): δ 170.12, 170.03, 169.76, 168.14, 167.36, 138.64, 138.54, 138.45, 138.41, 138.27, 138.07, 133.68, 133.57, 131.74, 129.05, 128.62, 128.57, 128.51, 128.33, 128.23, 128.14, 128.06, 127.99, 127.94, 127.81, 127.78, 127.73, 127.45, 101.24, 99.81, 99.59, 99.48, 99.34, 99.24, 99.05, 98.02, 96.95, 79.82, 79.23, 78.33, 78.24, 78.14, 75.86, 75.22, 75.12, 75.02, 74.93, 74.77, 74.57, 74.47, 74.38, 74.30, 74.17, 73.29, 73.11, 72.50, 72.20, 71.94, 71.35, 71.25, 69.13, 69.03, 68.79, 68.53, 68.48, 68.35, 68.17, 67.40, 56.56, 55.69, 51.09, 28.66, 28.26, 22.99, 21.17, 21.12; HRMS (MALDI-TOF): m/z calcd for $\text{C}_{284}\text{H}_{297}\text{N}_5\text{O}_{62}$: 4772.0232; found 4794.9783 ($M + \text{Na}$) $^+$.



5-Aminopentyl- α -D-mannopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl-(1 \rightarrow 3)-{ α -D-mannopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl-(1 \rightarrow 3)-[α -D-mannopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl-(1 \rightarrow 6)]- α -D-mannopyranosyl-(1 \rightarrow 6)}- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranoside (G6): Compound **S10c** (0.120 g, 0.024 mmol) was deprotected by following general procedure 2 (Method 1) to get $\text{Man}_9\text{GlcNAc}_2$ **G6** (0.012 g, 26%) as a white solid. ^1H NMR (600 MHz, D_2O): δ 5.40 (s, 1H, H-1 d), 5.33 (s, 1H, H-1 d), 5.30 (s, 1H, H-1 e), 5.14 (s, 1H, H-1), 5.05 (s, 1H, H-1), 5.03 (d, $J = 2.4$ Hz, 2H, H-1), 4.86 (s, 1H, H-1), 4.58 (d, $J = 7.8$ Hz, 1H,

H-1^a), 4.48 (d, $J = 7.8$ Hz, 1H, H-1b), 4.22 (d, $J = 2.4$ Hz, 1H), 4.14 (s, 1H), 4.14-3.95 (m, 12H), 3.94-3.59 (m, 54H), 3.50-3.48 (m, 1H), 3.02 (t, $J = 9.2$ Hz, 2H), 2.06 (s, 3H, -C(O)CH₃), 2.02 (s, 3H, -C(O)CH₃), 1.68-1.63 (m, 2H, -CCH₂C-, linker), 1.59-1.56 (m, 2H, -CCH₂C-, linker), 1.41-1.37 (m, 2H, -CCH₂C-, linker); ¹³C NMR (150 MHz, D₂O): δ 174.47, 171.01, 102.24 (C-1^d, $^1J_{C,H} = 171.8$ Hz), 102.21 (C-1^{d'}, $^1J_{C,H} = 169.0$ Hz), 102.19 (C-1^e, $^1J_{C,H} = 173.8$ Hz), 101.43, 101.03, 100.82, 100.59 (C-1^a, $^1J_{C,H} = 162.8$ Hz), 100.21, 99.61, 97.97 (C-1^b, $^1J_{C,H} = 159.8$ Hz), 81.86, 79.32, 78.88, 78.63, 78.45, 74.50, 74.48, 74.14, 73.39, 73.27, 73.20, 73.17, 72.69, 72.36, 71.90, 71.61, 70.21, 70.03, 69.93, 69.37, 67.01, 66.94, 66.90, 66.85, 66.81, 66.76, 65.42, 65.40, 65.07, 64.90, 61.11, 61.06, 61.01, 60.91, 60.06, 59.94, 54.95, 39.29, 38.61, 28.02, 26.34, 22.16, 22.10, 22.06; ESI-MS: m/z calcd for C₇₅H₁₂₉N₃O₅₆: 1967.7231; found 1990.7279 ($M + Na$)⁺.

vi. Synthesis of hybrid type oligosaccharides (G7-G14).

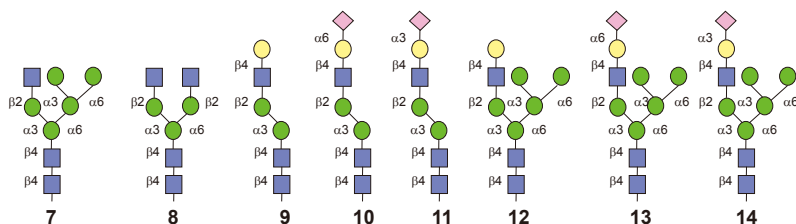
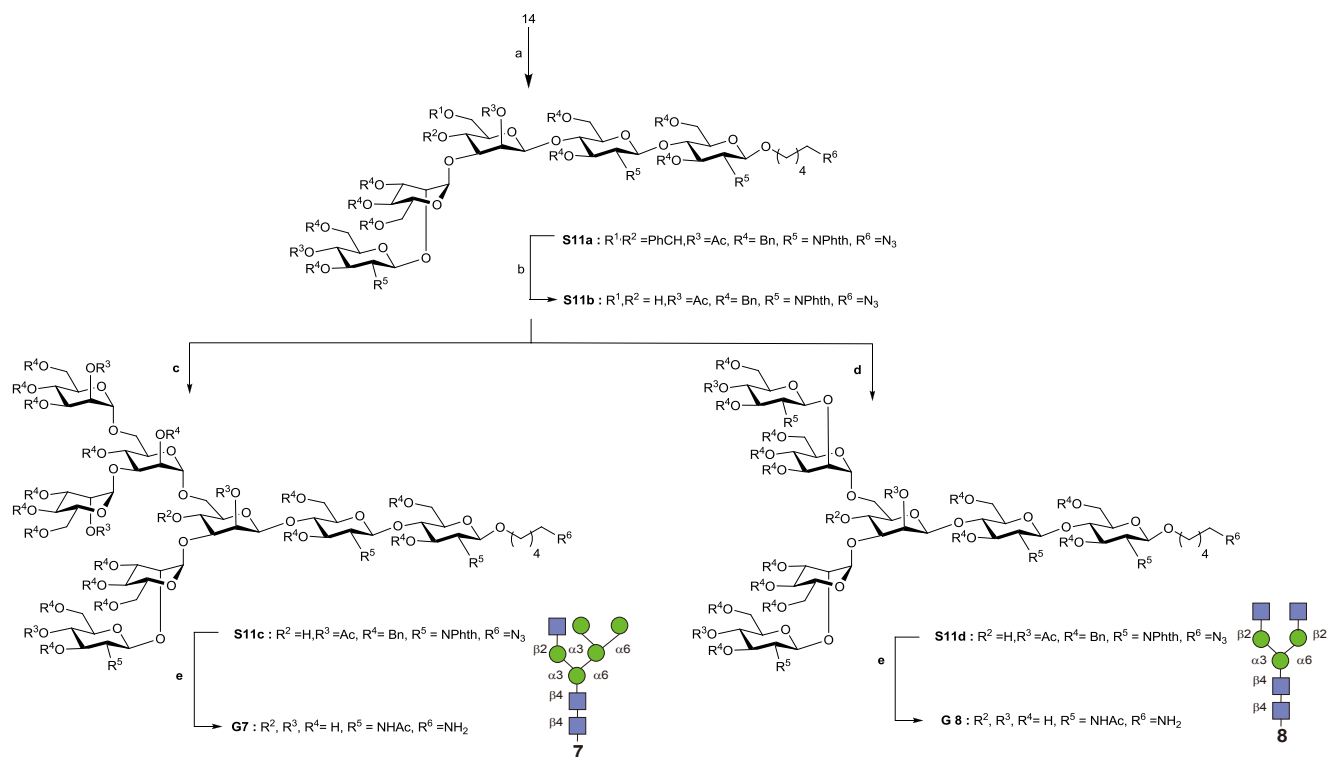
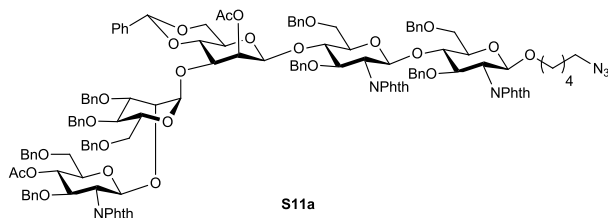


Figure S2 | Structures of hybrid type glycans and their fragments

Synthesis of hybrid type sugars commenced with condensation of **14** with **6** catalyzed by AgOTf/CP₂HfCl₂ to give pentasaccharide **S11a** in 63% yield (Scheme S11). The benzilidene opening resulted in diol **S11b**, which was further glycosylated at 6''-O with trimannosyl thioglycoside **3** to afford **S11c** in 55% yield. Cp₂HfCl₂ /AgOTf mediated condensation of acceptor **S11b** with **6** resulted in the formation of **S11d** in 58% yield.

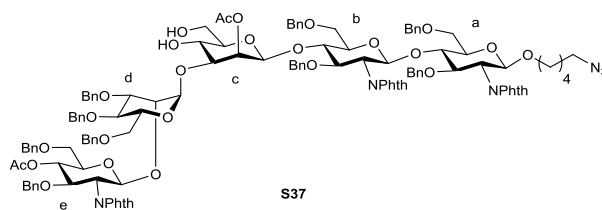


Scheme S11 | Preparation of octasaccharide G7 and heptasaccharide G8. **a**, **6**, AgOTf, Cp₂HfCl₂, Toluene, 4 Å MS, -40 °C, 4 h, 63%; **b**, *p*TsOH, CH₃CN, 5 h, 78%; **c**, **3**, (BrC₆H₄)₃NSbCl₆, CH₃CN, 4 Å MS, -10 °C to RT, 4 h, 55%; **d**, **6**, AgOTf, Cp₂HfCl₂, Toluene, 4 Å MS, -40 °C, 4 h, 58%; **e**, (1) NH₂CH₂CH₂NH₂, *n*BuOH, 90 °C, overnight; (2) Ac₂O, pyridine, overnight; (3) NaOMe, MeOH, overnight; (4) Pd(OH)₂, MeOH: H₂O: HCOOH (5:3:2), H₂; **G7**:26; **G8**:32%.

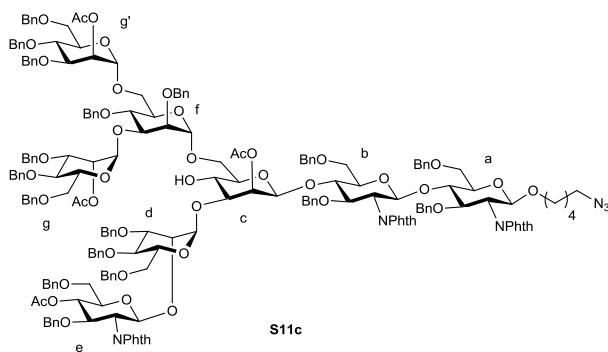


5-Azidopentyl-O-2-O-acetyl-3,6-O-di-benzyl-2-deoxy-2-phthalimido-β-D-gluco-pyranosyl-(1→2)-O-(3,4,6-tri-O-benzyl-α-D-mannopyranosyl-(1→3)-2-O-acetyl-4,6-O-benzylidene-β-D-mannopyranosyl-(1→4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)

-(1→4)-O-3,6-di-O-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranoside (S11a): A mixture of Silver triflate (0.080 g, 0.315 mmol), Bis(cyclopentadienyl)hafnium dichloride (0.084 g, 0.220 mmol) and 4 Å activated molecular sieves in dry toluene (10 mL) was stirred at rt for 1 h. The reaction mixture was then cooled to -40 °C, a solution of donor **6** (0.341 g, 0.308 mmol), chitobiose acceptor **14** (0.350 g, 0.256 mmol) in 5 mL toluene was added. The mixture was stirred for 4 h, quenched with Et₃N, diluted with CH₂Cl₂ and filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), and a brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 10% EA in toluene) to afford **S11a** (0.360 g, 63%) as white foam. TLC: (ethyl acetate: toluene = 2/8, v/v): R_f = 0.56; ¹H NMR (600 MHz, CDCl₃): δ 7.84-7.55 (m, 12H, Ar-H), 7.45-7.10 (m, 28H, Ar-H), 7.06-6.90 (m, 18H, Ar-H), 6.75-6.71 (m, 4H, Ar-H), 5.37 (d, *J* = 8.4 Hz, 1H, H-1), 5.21 (d, *J* = 8.4 Hz, 1H, H-1), 5.19 (s, 1H, Ph-CH₂, benzylidene), 5.09 (t, *J* = 9.0 Hz, 1H), 4.93-4.76 (m, 6H), 4.57 (s, 1H), 4.54 (d, *J* = 12.0 Hz, 1H), 4.45-4.07 (m, 20H), 3.69-3.15 (m, 22H), 3.02-3.00 (m, 1H), 2.86-2.81 (m, 2H, linker), 2.25 (s, 3H, -C(O)CH₃), 1.84 (s, 3H, -C(O)CH₃), 1.35-1.23 (m, 4H, -CCH₂C-, linker), 1.07-1.02 (m, 2H, -CCH₂C-, linker); ¹³C NMR (150 MHz, CDCl₃): δ 170.68, 170.05, 167.88, 138.98, 138.89, 138.85, 138.71, 138.63, 138.14, 138.07, 137.78, 134.08, 133.90, 133.46, 132.06, 131.81, 129.32, 128.90, 128.83, 128.67, 128.55, 128.50, 128.34, 128.27, 127.95, 127.79, 127.70, 127.60, 101.00, 99.41, 98.38, 97.91, 97.29, 80.38, 79.02, 76.82, 76.76, 76.52, 76.14, 75.25, 74.90, 74.83, 74.67, 74.51, 74.44, 73.68, 73.52, 73.38, 73.06, 73.02, 72.93, 71.46, 70.65, 70.40, 70.03, 69.66, 69.14, 68.52, 67.56, 56.83, 55.98, 55.85, 51.37, 29.99, 28.95, 28.55, 23.28, 21.67, 21.19; ESI-MS: *m/z* calcd for C₁₃₃H₁₃₂N₆O₃₁: 2309.8862; found 2332.8861 (*M* + Na)⁺.

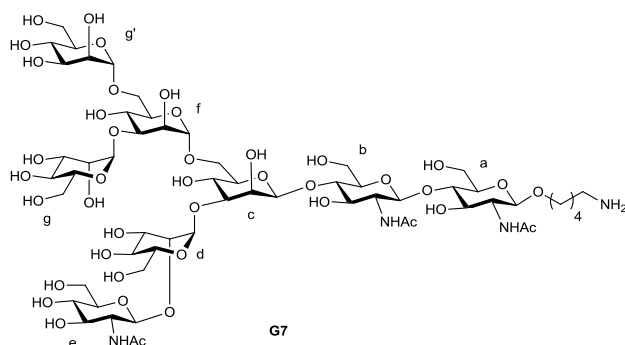


5-Azidopentyl-O-2-O-acetyl-3,6-O-di-benzyl-2-deoxy-2-phthalimido-β-D-gluco-pyranosyl-(1→2)-O-(3,4,6-tri-O-benzyl-α-D-mannopyranosyl-(1→3)-2-O-acetyl-β-D-mannopyranosyl-(1→4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido-β-D-gluco-pyranosyl)-(1→4)-O-3,6-di-O-benzyl-2-deoxy-2-phthalimido-β-D-gluco-pyranoside (S11b): *p*-Toluene sulfonic acid monohydrate (0.020 g, 0.143 mmol) was added to a solution of **S11a** (0.220 g, 0.095 mmol) in acetonitrile (20 mL) and the resulting reaction mixture was stirred at rt for 5 h. The reaction was quenched by adding Et₃N and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 15% EA in toluene) to give diol **S11b** (0.165 g, 78%). TLC: (ethyl acetate: toluene = 2/8, v/v): R_f = 0.32; ¹H NMR (600 MHz, CDCl₃): δ 7.89-7.43 (m, 12H, Ar-H), 7.35-7.16 (m, 28H, Ar-H), 7.09-7.05 (m, 5H, Ar-H), 7.03-6.89 (m, 9H, Ar-H), 6.75-6.72 (m, 3H, Ar-H), 5.30 (d, *J* = 8.4 Hz, 1H, H-1^a), 5.23 (d, *J* = 8.4 Hz, 1H, H-1^b), 5.12 (d, *J* = 9.0 Hz, 1H, Ar-H), 5.09 (t, *J* = 9.8 Hz, 1H), 4.95 (d, *J* = 12.0 Hz, 1H), 4.90 (d, *J* = 8.4 Hz, 1H, H-1^c), 4.87-4.80 (m, 2H), 4.58-4.55 (m, 2H), 4.50-4.41 (m, 10H, overlapped, H-1^d), 4.37-4.34 (m, 3H), 4.27-4.24 (m, 2H), 4.18-4.15 (m, 2H), 4.10-3.92 (m, 5H), 3.73-3.70 (m, 2H), 3.65-3.61 (m, 2H), 3.59-3.51 (m, 6H), 3.47-3.38 (m, 6H), 3.23-3.17 (m, 4H), 3.04 (dd, *J* = 3.6, 9.0 Hz, 1H), 3.01-3.98 (m, 1H), 2.83-2.78 (m, 3H), 2.38 (s, 3H, -C(O)CH₃), 1.90 (s, 3H, -C(O)CH₃), 1.36-1.22 (m, 4H, -CCH₂C-, linker), 1.06-1.01 (m, 2H, -CCH₂C-, linker); ¹³C NMR (150 MHz, CDCl₃): δ 170.93, 170.05, 168.77, 167.86, 167.69, 138.90, 138.83, 138.65, 138.47, 138.23, 138.14, 138.07, 138.04, 134.35, 134.13, 133.91, 132.04, 131.84, 131.72, 128.81, 128.65, 128.64, 128.62, 128.54, 128.53, 128.48, 128.46, 128.36, 128.17, 128.09, 128.02, 127.99, 127.97, 127.92, 127.91, 127.83, 127.77, 127.74, 127.63, 127.13, 124.73, 123.97, 123.43, 123.08, 118.73, 100.25 (C-1^e), 98.37 (C-1^d), 98.21 (C-1^c), 97.86 (C-1^b), 97.30 (C-1^a), 82.86, 80.05, 78.38, 76.92, 76.15, 75.72, 75.60, 75.45, 74.86, 74.83, 74.70, 74.42, 74.24, 73.75, 73.42, 73.38, 72.82, 70.81, 70.60, 70.27, 69.99, 69.50, 69.15, 69.13, 68.53, 67.88, 66.28, 62.98, 56.78, 55.99, 55.88, 51.37, 32.21, 29.98, 28.94, 28.54, 23.27, 21.69, 21.18; ESI-MS: *m/z* calcd for C₁₂₆H₁₂₈N₆O₃₁: 2221.8549; found 2244.8529 (*M* + Na)⁺.



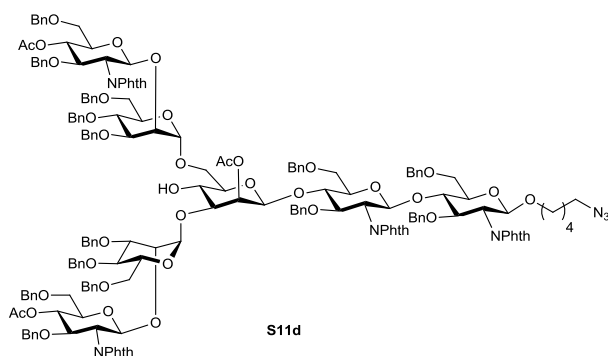
5-Azidopentyl-O-{4-O-acetyl-3,6-O-di-benzyl-2-deoxy-2-phthalimido- β -D-gluco-pyranosyl-(1 \rightarrow 2)}-O-{3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3)}-{2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3)}-[2-O-acetyl-3,4,6-tri-O-benzyl- α -D-manno-pyranosyl-(1 \rightarrow 6)]-2,4-di-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 6)}-2-O-acetyl- β -D-manno-pyranosyl-(1 \rightarrow 4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)-(1 \rightarrow 4)-O-3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranoside (S11c): A mixture of pentasaccharide acceptor **S11b** (0.150 g, 0.067 mmol), thiomannoside donor **3** (0.110 g, 0.080 mmol) and activated 4 Å molecular sieves in CH₃CN (10 mL) was stirred at rt for 1 h. The resulting mixture was cooled to -10 °C, tris (4-bromophenyl) aminium hexachloroantimonate (0.170 g, 0.201 mmol) was added and resulting reaction was stirred at room temperature for 4 h. TLC (ethyl acetate: toluene, 2/8) indicated formation of product with consumption of starting material, the reaction was quenched by Et₃N. The reaction mixture was diluted with CH₂Cl₂ and filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), and a brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 10% EA in toluene) to afford **S11c** (0.130 g, 55%) as colorless foam. TLC: (ethyl acetate: toluene= 1/9, v/v): R_f = 0.42; ¹H NMR (600 MHz, CDCl₃): δ 7.71-7.30 (m, 12H, Ar-H), 7.28-7.10 (m, 70H, Ar-H), 7.97-6.86 (m, 12H, Ar-H), 6.69 (m, 3H, Ar-H), 5.47 (d, *J* = 8.4 Hz, 2H), 5.44 (s, 1H), 5.32 (d, *J* = 8.4 Hz, 1H, H-1^a), 5.15 (d, *J* = 8.4 Hz, 1H, H-1^b), 5.13-5.10 (m, 2H), 5.00 (s, 1H, H-1^d), 4.95 (s, 1H, H-1^f), 4.91 (s, 2H, H-1^e, H-1^g), 4.87-4.73 (m, 8H), 4.62 (d, *J* = 8.7 Hz, 1H), 4.57-4.28 (m, 26H), 4.15-3.70 (m, 20H), 3.68-

3.40 (m, 13H), 3.38-3.25 (m, 6H), 3.24-3.09 (m, 5H), 2.98 (dd, $J = 2.3, 7.8$ Hz, 1H), 2.89-2.79 (m, 2H), 2.73-2.71 (m, 1H), 2.27 (s, 3H, -C(O)CH₃), 2.05 (s, 3H, -C(O)CH₃), 1.99 (s, 3H, -C(O)CH₃), 1.88 (s, 3H, -C(O)CH₃), 1.32-1.28 (m, 4H, -CCH₂C-, linker), 1.07-0.98 (m, 2H, -CCH₂C-, linker); ¹³C NMR (150 MHz, CDCl₃): δ 170.26, 168.89, 138.38, 128.47, 128.37, 128.32, 128.30, 128.26, 128.24, 128.19, 128.08, 128.04, 127.91, 127.86, 127.73, 127.56, 127.49, 127.39, 127.26, 100.35, 100.23, 99.89, 99.56, 99.23, 98.54, 98.33, 98.23, 78.43, 76.09, 75.37, 75.07, 74.08, 74.44, 74.04, 73.47, 73.31, 72.63, 68.81, 51.08, 29.71, 29.37, 28.66, 28.25, 22.98; HRMS (MALDI-TOF): m/z calcd for C₂₀₄H₂₁₀N₆O₄₈; 3513.4134 found 3536.4050 ($M + Na$)⁺.



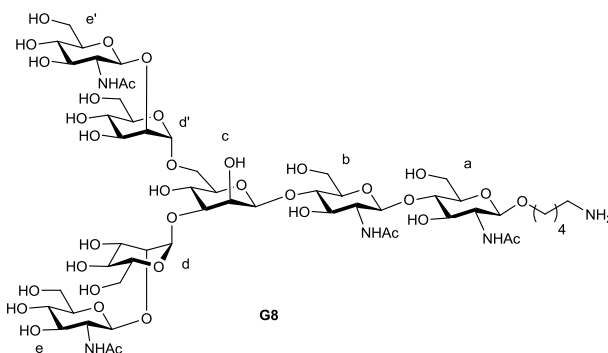
5-Aminopentyl-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl (1 \rightarrow 3),[di-(α -D-mannopyranosyl)-(1 \rightarrow 3),(1 \rightarrow 6)- α -D-mannopyranosyl](1 \rightarrow 6)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranoside (G7): Compound **S11c** (0.105 g, 0.030 mmol) was deprotected by following general procedure 2 (Method 1) to get the title compound **G7** (0.012 g, 26%) as a white solid. ¹H NMR (600 MHz, D₂O): δ 5.09 (d, $J = 1.2$ Hz, 1H, H-1^d), 5.02 (d, $J = 8.4$ Hz, 1H, H-1^a), 4.95 (s, 1H, H-1^f), 4.91 (s, 1H, H-1^g), 4.85 (s, 1H, H-1^{g'}), 4.78 (s, 1H, H-1^c), 4.61 (d, $J = 7.8$ Hz, 1H, H-1^e), 4.51 (d, $J = 7.8$ Hz, 1H, H-1^b), 4.22 (t, $J = 10.2$ Hz, 2H), 4.18 (dd, $J = 1.8, 3.0$ Hz, 1H), 4.00 (dd, $J = 1.8, 3.1$ Hz, 1H), 4.08-3.38 (m, 46H), 3.03 (t, $J = 8.2$ Hz, 2H, -NCH₂-, linker), 2.09 (s, 3H, -C(O)CH₃), 2.08 (s, 3H, -C(O)CH₃), 2.01 (s, 3H, -C(O)CH₃), 1.71-1.66 (m, 2H, -CCH₂C-, linker), 1.62-1.59 (m, 2H, -CCH₂C-, linker), 1.44-1.40 (m, 2H, -CCH₂C-, linker); ¹³C NMR (150 MHz, D₂O): δ 174.91, 174.42, 171.02, 110.64 (C-1^d, ¹ $J_{C,H} = 174$ Hz), 102.32, 101.48, 100.51

(C-1^a, ¹J_{C,H} = 161 Hz), 100.44, 100.15, 99.26, 97.58 (C-1^g, ¹J_{C,H} = 169.6 Hz), 79.48, 78.86, 78.72, 76.64, 76.04, 75.72, 74.65, 74.51, 74.42, 73.31, 72.69, 72.35, 72.09, 71.96, 70.86, 70.57, 70.34, 70.12, 70.09, 69.94, 69.42, 67.39, 66.70, 66.36, 65.54, 65.14, 61.68, 61.02, 60.93, 39.31, 38.62, 28.03, 26.39, 22.58, 22.20, 22.11, 22.08, 21.82; ESI-MS: *m/z* calcd for C₅₉H₁₀₂N₄O₄₁; 1522.6092 found 1523.6148 (*M* + H)⁺.



5-Azidopentyl-O-di-[[4-O-acetyl-3,6-O-di-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl-(1→2)]-O-[3,4,6-tri-O-benzyl-α-D-mannopyranosyl]]-(1→3),(1→6)-2-O-acetyl-β-D-mannopyranosyl-(1→4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranosyl)-(1→4)-O-3,6-di-O-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranoside (S11d):- A mixture of Silver triflate (0.080 g, 0.315 mmol), Bis(cyclopentadienyl)hafnium dichloride (0.084 g, 0.220 mmol) and 4 Å activated molecular sieves in dry toluene (10 mL) was stirred at rt for 1 h. The reaction mixture was then cooled to -40 °C, a solution of donor **6** (0.091 g, 0.094 mmol) and acceptor **S11b** (0.140 g, 0.063 mmol) in 5 mL toluene was added. The mixture was stirred for 4 h, quenched with Et₃N, diluted with CH₂Cl₂ and filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), and a brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 15% EA in toluene) to afford **S11d** (0.110 g, 58%) as colorless foam. TLC: (acetone: toluene = 2/8, v/v): R_f = 0.39; ¹H NMR (600 MHz, CDCl₃): δ 7.62-7.32 (m, 16H, Ar-H), 7.27-6.87 (m, 41H, Ar-H), 6.85-6.59 (m, 29H, Ar-H), 5.27 (d, *J* = 8.4 Hz, 1H), 5.16 (s,

1H), 5.14 (d, $J = 3.6$ Hz, 1H), 5.10 (d, $J = 1.8$ Hz, 1H), 4.98 (d, $J = 3.6$ Hz, 1H), 4.89-4.68 (m, 10H), 4.59-4.21 (m, 21H), 4.19-3.87 (m, 7H), 3.80 (dd, $J = 3.2, 1.8$ Hz, 1H), 3.70-3.52 (m, 7H), 3.51-3.28 (m, 8H), 3.27-3.18 (m, 5H), 3.15-3.03 (m, 3H), 3.01 (dd, $J = 3.2, 1.8$ Hz, 1H), 2.91-2.75 (m, 5H), 2.73-2.69 (m, 1H), 2.27 (s, 3H), 1.98 (s, 3H), 1.86 (s, 3H), 1.32-1.23 (m, 4H), 1.03-0.99 (m, 2H); ^{13}C NMR (150 MHz, CDCl_3): δ 171.09, 171.04, 170.094, 167.91, 167.57, 139.19, 138.86, 138.63, 138.54, 138.42, 138.31, 138.21, 138.14, 137.93, 133.84, 133.71, 132.04, 131.80, 131.69, 128.80, 128.64, 128.39, 128.32, 128.13, 128.08, 127.79, 127.70, 127.60, 100.18, 100.00, 98.36, 97.88, 97.61, 97.30, 97.22, 80.14, 79.97, 79.31, 75.85, 75.56, 75.39, 75.30, 75.04, 74.71, 74.59, 73.95, 73.64, 73.24, 72.86, 72.14, 71.55, 71.37, 71.04, 70.53, 70.40, 70.15, 69.71, 69.12, 68.12, 65.40, 63.29, 56.34, 55.92, 51.45, 51.37, 51.27, 28.95, 28.54, 23.26, 21.13, 21.02; HRMS (MALDI-TOF): m/z calcd for $\text{C}_{183}\text{H}_{183}\text{N}_7\text{O}_{43}$; 3168.2306 found 3191.2375 ($M + \text{Na}$) $^+$.

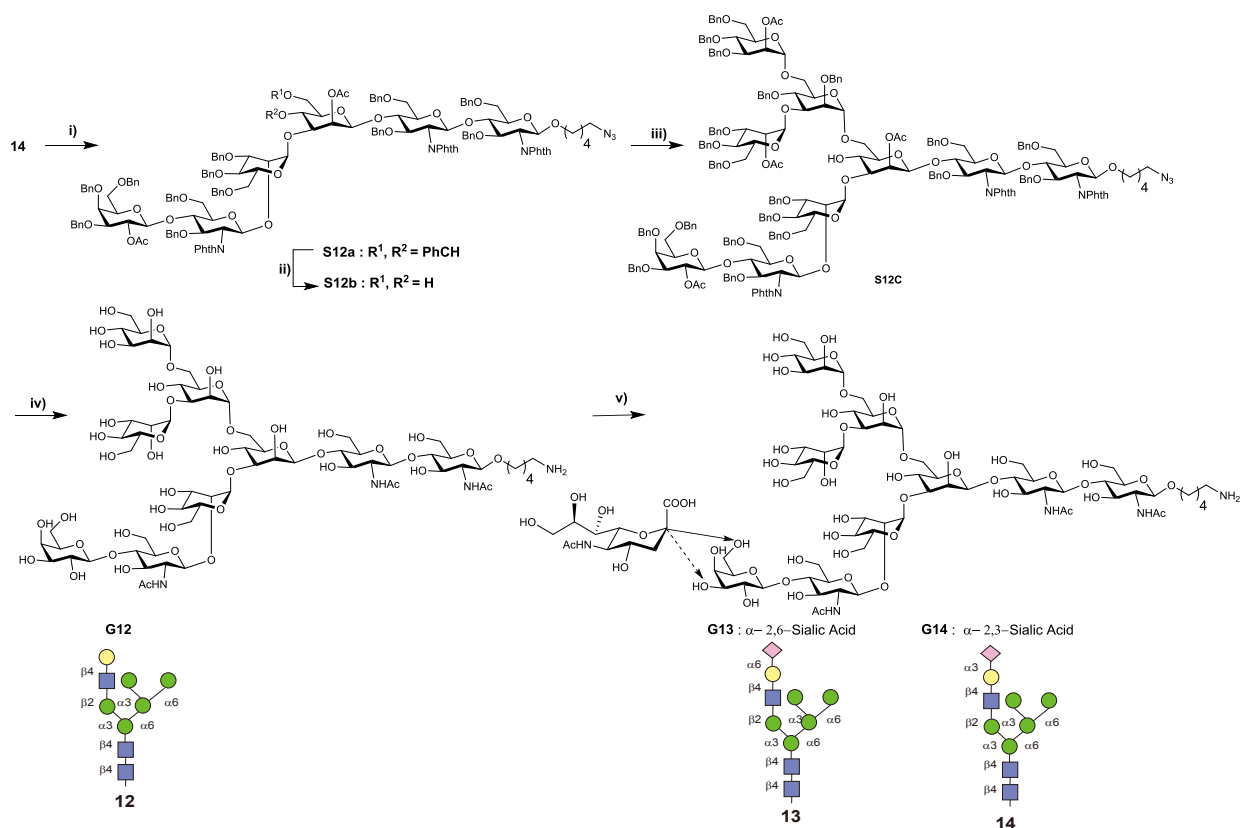


5-Aminopentyl-di-[2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl]-(1 \rightarrow 3),(1 \rightarrow 6)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranoside (G8**):** Compound **S11d** (0.130 g, 0.041 mmol) was deprotected by following general procedure 2 (Method 1) to get the desired heptasaccharide **G8** (0.015 g, 32%) as a white solid. ^1H NMR (600 MHz, D_2O): δ 5.99 (d, $J = 8.5$ Hz, 1H, H-1 a), 4.93 (s, 1H, H-1 d), 4.83 (s, 1H, H-1 d), 4.76 (s, 1H, H-1 c), 4.60 (d, $J = 7.8$ Hz, 1H, H-1 b), 4.56 (d, $J = 7.9$ Hz, 1H, H-1 e), 4.49 (d, $J = 7.9$ Hz, 1H, H-1 e), 4.27 (s, 2H), 4.11 (d, $J = 1.8$ Hz, 1H), 3.96-3.41 (m, 41H), 2.98 (t, $J = 8.9$ Hz, 2H, $-\text{NCH}_2-$, linker), 2.13 (s, 3H, -

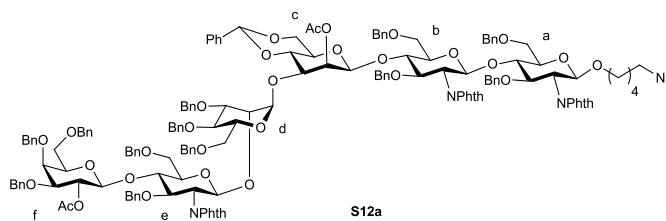
C(O)CH₃), 2.09 (s, 3H, -C(O)CH₃), 2.06 (s, 3H, -C(O)CH₃), 2.01 (s, 3H, -C(O)CH₃), 1.69-1.64 (m, 2H, -CCH₂C-, linker), 1.60-1.57 (m, 2H, -CCH₂C-, linker), 1.42-1.38 (m, 2H, -CCH₂C-, linker); ¹³C NMR (150 MHz, D₂O): δ 174.88, 174.78, 174.65, 174.42, 171.0, 101.39, 101.05, 100.61 (C-1^c), 100.39 (C-1^a), 99.55 (C-1^e), 97.60 (C-1^d), 97.20 (C-1^d), 79.41, 79.28, 78.92, 76.63, 76.25, 75.79, 74.50, 73.64, 72.84, 71.95, 70.09, 69.89, 69.48, 67.39, 67.35, 66.20, 65.39, 61.66, 61.61, 60.75, 60.61, 59.96, 55.68, 54.98, 54.94, 39.30, 28.04, 26.35, 22.55, 22.31, 22.21, 22.12, 22.08; ESI-MS: *m/z* calcd for C₅₅H₉₅N₅O₃₆; 1402.5830; found 1403.5862 (*M* + H)⁺.

Preparation of glycans G12-G14

Stereoselective installation of trisaccharide antenna **7** at the 3-O position of the core trisaccharide acceptor **14** was performed under the promotion of Cp₂HfCl₂/AgOTf to afford hexasaccharide **S12a**. The *p*-toluene sulfonic acid-mediated ring opening of **S12a** provided diol **S12b**. Considering the higher reactivity of the primary hydroxyl, **S12b** was further glycosylated at the 6-O position with trimannosyl thioglycoside **3** and, then, activated by the stable radical cation tris(4-bromophenyl) ammoniumyl hexachloroantimonate in a one electron transfer reaction to afford nonasaccharide **S12c**. With compound **S12c** in hand, a series of functional group transformations were carried out to afford the desired fully deprotected glycan **G12**. The precursor oligosaccharide **G12**, is an appropriate starting material for sialylation by α-2,6 or 2,3 sialyltransferase derived from marine bacteria, known for their broader acceptor specificity and no intrinsic sialidase activity. Sialyl transferase (SiaT)-mediated enzymatic terminal sialylation¹⁻⁴ of **G12** efficiently permitted access to α-2,6 and α-2,3 sialylated glycans **G13** and **G14**, respectively (Scheme S12).



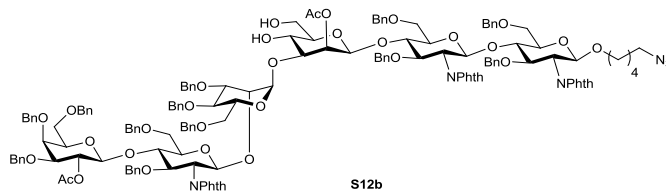
Scheme S12 | Preparation of G12-G14. i) **7**, Cp_2HfCl_2 , AgOTf , 4 Å MS, $-40\text{ }^\circ\text{C}$, 2 h, 63%; ii) *p*-TsOH, CH_3CN , 5 h, 78%; iii) **3**, $(\text{BrC}_6\text{H}_4)_3\text{NSbCl}_6$, CH_3CN , 4 Å MS, $-10\text{ }^\circ\text{C}$ to RT, 4 h, 55%; iv) (1) $\text{NH}_2\text{CH}_2\text{CH}_2\text{NH}_2$, *n*-BuOH, $90\text{ }^\circ\text{C}$, overnight; (2) Ac_2O , pyridine, overnight; (3) NaOMe, MeOH, overnight; (4) $\text{Pd}(\text{OH})_2$, MeOH: H_2O : HCOOH (5:3:2), H_2 ; 62%; v) CMP- β -D-Sialic acid, α -2,6- or 2,3-sialyltransferase, G13: 60%; G14: 56%; Cp_2HfCl_2 : Bis(cyclopentadienyl) hafnium dichloride; AgOTf : Silver trifluoromethanesulfonate; $(\text{BrC}_6\text{H}_4)_3\text{NSbCl}_6$: Tris (4-bromophenyl) ammonium hexachloroantimonate.



5-Azidopentyl-O-2-O-acetyl-3,4,6-O-tri-benzyl- β -D-galactopyranosyl-(1 \rightarrow 4)-O-3,6-O-di-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl-(1 \rightarrow 2)-O-(3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3)-2-O-acetyl-4,6-O-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)-(1 \rightarrow 4)-O-3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranoside (S12a):

A mixture of silver triflate (0.327 g, 1.28 mmol), bis (cyclopentadienyl) hafnium dichloride (0.339 g, 0.896 mmol) and 4 Å activated molecular sieves in dry toluene (10 mL) was stirred at rt for 1 h. The reaction mixture was then cooled to -40 °C, a solution of donor **7** (0.430 g, 0.307 mmol) and acceptor chitobiose trisaccharide **14** (0.35 g, 0.256 mmol) in 5 mL toluene was added. The mixture was stirred for 2 h, quenched with Et₃N, diluted with EtOAc and filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), and a brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 15% EA in toluene) to afford **S12a** (0.500 g, 71%) as white foam. TLC: (ethyl acetate: toluene = 2/8, v/v): R_f = 0.62; ¹H NMR (600 MHz, CDCl₃): δ 7.85-7.49 (m, 12H, Ar-H), 7.02-7.39 (m, 45H, Ar-H), 7.19-7.02 (m, 10H, Ar-H), 6.99-6.90 (m, 6H, Ar-H), 6.83-6.78 (m, 4H, Ar-H), 5.310 (t, J = 7.5 Hz, 1H, H-2^f), 5.27 (d, J = 8.1 Hz, 1H, H-1^a), 5.25 (d, J = 8.4 Hz, 1H, H-1^c), 5.15 (d, J = 2.3 Hz, 1H), 5.01 (s, 1H, Ph-CH, benzylidene), 4.90-4.80 (m, 7H), 4.62 (d, J = 8.5 Hz, 1H), 4.58-4.53 (m, 3H), 4.52-4.38 (m, 10H), 4.32-4.13 (m, 10H), 4.12-4.05 (m, 5H), 4.02 (m, 1H), 3.89 (d, J = 3.2 Hz, 1H), 3.75-3.73 (m, 2H), 3.72-3.49 (m, 8H), 3.48-3.37 (m, 5H), 3.34-3.29 (m, 4H), 3.29-3.18 (m, 4H), 2.98 (m, 1H), 2.89-2.79 (m, 2H, linker), 2.29 (s, 3H, -C(O)CH₃), 1.99 (s, 3H, -C(O)CH₃), 1.49-1.30 (m, 4H, -CCH₂C-), 1.10-1.00 (m, 2H, -CCH₂C-); ¹³C NMR (150 MHz, CDCl₃): δ 170.8, 169.4, 168.7, 167.7, 139.1, 139.9, 138.9, 138.8, 138.8, 138.5, 138.2, 138.2, 138.1, 137.9, 137.6, 134.2, 133.8, 133.1, 131.9, 131.8, 131.6, 129.1, 128.8, 128.6, 128.5, 128.4, 128.4, 128.4, 128.3, 128.3, 128.2, 128.1, 128.0, 128.0, 127.9, 127.8, 127.8, 127.7, 127.7, 127.5, 127.5, 127.4, 127.3, 127.0, 126.7, 126.1, 123.8, 123.3, 122.8, 100.88, 100.83, 99.94, 99.38, 98.43, 98.30, 97.26, 80.31, 79.00, 78.05, 77.61, 76.42, 76.31, 76.1, 75.1, 74.8, 74.7, 74.6, 74.6, 74.4, 74.4, 74.2, 73.6, 73.4, 73.3, 72.9, 72.8, 72.2, 71.8, 71.7, 70.4, 70.0, 69.8, 69.0,

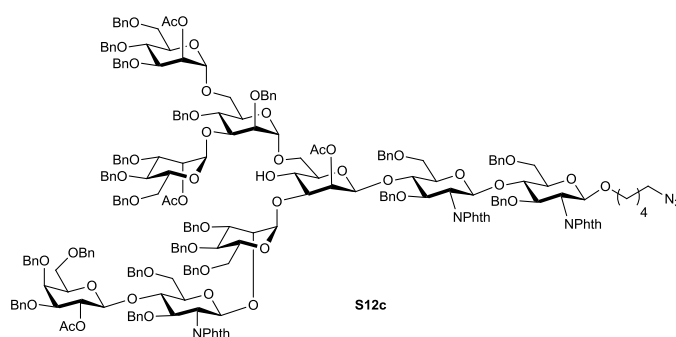
68.8, 68.4, 68.2, 67.7, 67.6, 56.7 56.0, 55.9, 51.3, 28.8, 28.4, 23.2, 21.7, 21.2; ESI-MS: m/z calcd for $C_{160}H_{160}N_6O_{36}$; 2742.0799 found 2765.0952 ($M + Na$)⁺.



5-Azidopentyl-O-2-O-acetyl-3,4,6-O-tri-benzyl- β -D-galactopyranosyl-(1 \rightarrow 4)-O-3,6-O-di-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl-(1 \rightarrow 2)-O-(3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3)-2-O-acetyl- β -D-mannopyranosyl-(1 \rightarrow 4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)-(1 \rightarrow 4)-O-3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranoside (S12b):

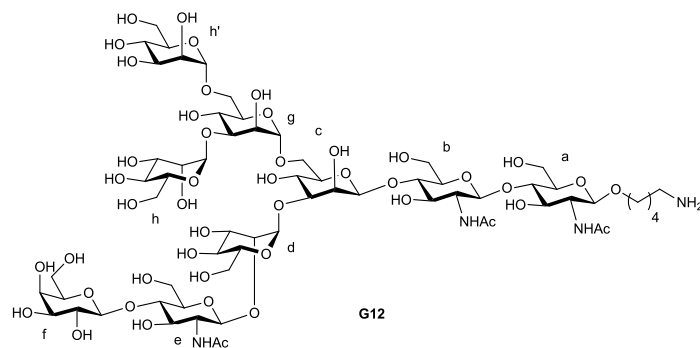
To a solution of **S12a** (0.601 g, 0.219 mmol) in acetonitrile (10 mL) was added *p*-toluene sulfonic acid monohydrate (0.046 g, 0.328 mmol), stirred at rt for 5 h. The reaction was quenched with Et₃N and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 15% EA in toluene) to afford diol **S12b** (0.460 g, 78%). TLC: (acetone: toluene = 2/8, v/v): R_f = 0.32; ¹H NMR (600 MHz, CDCl₃): δ 7.91-7.42 (m, 12H, Ar-H), 7.39-7.12 (m, 41H, Ar-H), 7.10-6.93 (m, 13H, Ar-H), 6.85-6.81 (m, 3H, Ar-H), 6.78-6.73 (m, 3H, Ar-H), 5.33 (t, J = 7.8 Hz, 1H, H-2^f), 5.23 (t, J = 8.4 Hz, 2H, H-1), 5.08 (d, J = 6.2 Hz, 1H), 4.97 (d, J = 8.9 Hz, 1H, H-1), 4.94-4.80 (m, 5H), 4.64-4.60 (m, 2H), 4.49-4.21 (m, 21H), 4.19-4.15 (m, 2H), 4.15-3.89 (m, 8H), 3.89 (d, J = 6.2 Hz, 1H), 3.78 (d, J = 5.8 Hz, 1H), 3.69 (dd, J = 6.3, 12.1 Hz, 1H), 3.65-3.64 (m, 1H), 3.56-3.32 (m, 14H), 3.29 (dd, J = 6.6, 12.5 Hz, 1H), 3.29-3.23 (m, 3H), 3.00-2.99 (m, 2H), 2.87-2.82 (m, 2H), 2.74 (t, J = 9.5 Hz, 1H), 2.28 (s, 3H, -C(O)CH₃), 1.98 (s, 3H, -C(O)CH₃), 1.37-1.22 (m, 4H, -CCH₂C-), 1.07-1.00 (m, 2H, -CCH₂C-); ¹³C NMR (150 MHz, CDCl₃): δ 171.29, 169.57, 168.78, 168.58, 168.03, 167.90, 139.01, 138.92, 138.84, 138.68, 138.45, 138.39, 138.36, 138.22, 138.14, 137.98, 134.35, 134.15, 134.02, 133.89, 132.04, 131.95, 131.68, 130.10, 129.54, 129.46, 128.80, 128.70, 128.66, 128.62, 128.60, 128.52, 128.47, 128.46, 128.39, 128.30, 128.15, 128.07, 128.01, 127.98, 127.94, 127.91,

127.83, 127.72, 127.60, 127.39, 127.20, 126.96, 123.90, 123.46, 122.88, 101.11, 100.87, 98.87, 98.33, 83.75, 80.57, 80.14, 78.58, 78.29, 76.15, 75.78, 75.45, 74.11, 74.96, 74.83, 74.70, 74.61, 74.44, 74.10, 73.77, 73.73, 73.59, 73.43, 73.38, 73.03, 72.30, 71.97, 70.61, 70.29, 69.82, 69.14, 69.08, 68.54, 68.38, 67.94, 66.94, 66.47, 63.21, 56.80, 56.09, 56.01, 51.37, 31.71, 31.21, 30.97, 30.59, 29.99, 29.65, 28.55, 27.87, 27.29, 26.96, 26.24, 25.59, 25.36, 24.26, 23.27, 22.98, 21.86, 21.32, 20.80, 20.49, 18.94, 16.76, 15.95, 15.44, 15.10, 14.40, 14.07, 13.67, 13.47; ESI-MS: m/z calcd for $C_{153}H_{156}N_6O_{36}$; 1327.0189; found 1350.0207 ($M + Na$)⁺.



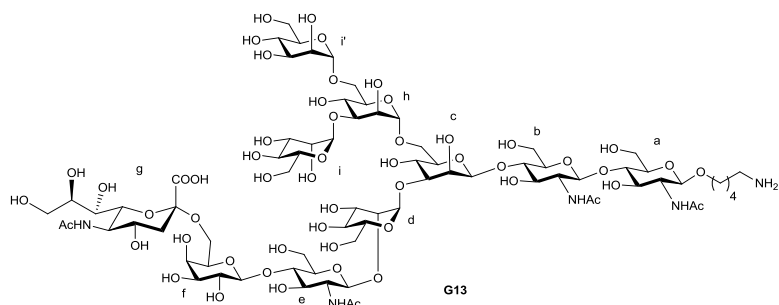
5-Azidopentyl-O-{2-O-acetyl-3,4,6-O-tri-benzyl- β -D-galactopyranosyl-(1 \rightarrow 4)}-O-{3,6-O-di-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl-(1 \rightarrow 2)}-O-{3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3)}-{2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3)}-[2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 6)]-2,4-di-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 6)}-2-O-acetyl- β -D-mannopyranosyl-(1 \rightarrow 4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl)-(1 \rightarrow 4)-O-3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranoside (S12c): A mixture of hexasaccharide acceptor **S12b** (0.40 g, 0.15 mmol), thiomannoside donor **3** (0.229 g, 0.226 mmol) and activated 4 Å molecular sieves in CH_3CN (10 mL) was stirred at rt for 1 h. The resulting mixture was cooled to -10 °C, tris (4-bromophenyl)aminium hexachloroantimonate (0.254 g, 0.30 mmol) was added and stirred at rt for 4 h. TLC indicated formation of product with consumption of starting material, the reaction was then quenched by Et_3N . The reaction mixture was diluted with CH_2Cl_2 and filtered through celite. The filtrate was washed with aqueous $NaHCO_3$ (2 x 50 mL), and a brine (50 mL) solution.

The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 10% EA in toluene) to afford **S12c** (0.335 g, 55%) as colorless foam and acceptor **9** (0.103 g). TLC: (ethyl acetate: toluene = 1/9, v/v): R_f = 0.46; ¹H NMR (600 MHz, CDCl₃): δ 7.72-7.45 (m, 12H, Ar-H), 7.37-7.28 (m, 5H, Ar-H), 7.28-7.12 (m, 67H, Ar-H), 7.09-7.02 (m, 11H, Ar-H), 6.94-6.90 (m, 7H, Ar-H), 6.80-6.78 (m, 4H, Ar-H), 6.69-6.68 (m, 3H, Ar-H), 6.62-6.61 (m, 3H, Ar-H), 5.49 (s, 1H), 5.48 (s, 1H), 5.35 (t, *J* = 8.4 Hz, 1H), 5.28 (d, *J* = 8.4 Hz, 1H, H-1), 5.17 (d, *J* = 8.4 Hz, 1H, H-1), 5.09 (d, *J* = 6.2 Hz, 1H), 5.00 (s, 1H, H-1), 4.92 (d, *J* = 3.1 Hz, 1H), 4.90-4.72 (m, 12H), 4.69-4.72 (m, 23H), 4.26-4.17 (m, 5H), 4.13-3.97 (m, 12H), 3.89-3.82 (m, 12H), 3.72 (t, 1H), 3.68-3.62 (m, 1H), 3.65-3.55 (m, 7H), 3.54-3.45 (m, 4H), 3.43-3.40 (m, 4H), 3.40-3.29 (m, 9H), 3.23-3.19 (m, 3H), 3.17-3.09 (m, 2H), 2.91-2.79 (m, 3H), 2.68 (s, 1H), 2.28 (s, 3H, -C(O)CH₃), 2.05 (s, 3H, -C(O)CH₃), 2.01 (s, 3H, -C(O)CH₃), 1.99 (s, 3H, -C(O)CH₃), 1.34-1.22 (m, 4H, linker), 1.06-1.00 (m, 2H, linker); ¹³C NMR (150 MHz, CDCl₃): δ 171.37, 170.38, 170.29, 169.54, 168.49, 168.23, 167.97, 167.53, 139.28, 139.11, 139.03, 138.97, 138.94, 138.67, 138.63, 138.58, 138.53, 138.41, 138.38, 138.25, 138.10, 128.74, 128.70, 128.67, 128.63, 128.58, 128.55, 128.53, 128.51, 128.48, 128.44, 128.40, 128.37, 128.34, 128.30, 128.15, 128.03, 128.00, 127.95, 127.92, 127.84, 127.77, 127.75, 127.70, 127.64, 127.61, 101.17, 100.92, 98.84, 98.28, 97.71, 97.45, 75.49, 75.36, 75.26, 75.17, 75.06, 74.96, 74.82, 74.75, 74.65, 74.60, 74.38, 74.30, 73.99, 73.77, 73.73, 73.59, 73.25, 73.25, 73.06, 72.99, 72.91, 72.26, 72.18, 71.98, 71.82, 71.57, 71.45, 70.52, 70.03, 69.92, 69.56, 69.10, 68.90, 68.85, 68.57, 68.48, 68.37, 66.64, 66.06, 56.75, 56.09, 55.99, 51.37, 34.25, 29.99, 28.94, 28.54, 25.90, 23.27, 21.86, 21.42, 21.33; HRMS (MALDI-TOF): *m/z* calcd for C₂₃₁H₂₃₈N₆O₅₃; 3945.6071; found 3968.6006 (*M* + Na)⁺.

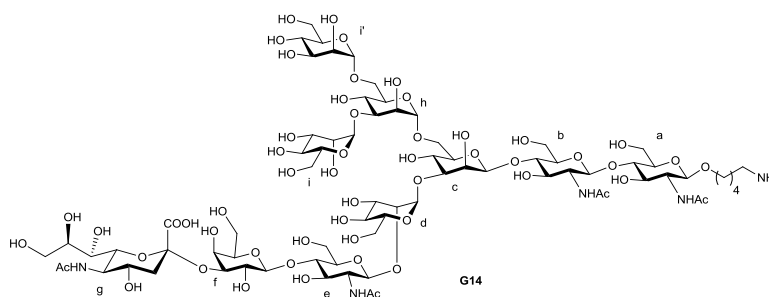


5-Aminopentyl- β -D-galactopyranosyl-(1 \rightarrow 4)-[2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl-(1 \rightarrow 3),[di-(α -D-mannopyranosyl)-(1 \rightarrow 3),(1 \rightarrow 6)- α -D-mannopyranosyl](1 \rightarrow 6)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranoside (G12):

Compound **S12c** (0.16 g, 0.048 mmol) was deprotected by following general procedure 2 (Method 1) to afford desired the nonasaccharide **G12** (0.052 g, 62%). ^1H NMR (600 MHz, D_2O): δ 5.10 (s, 1H, H-1^d), 5.07 (d, J = 9.5 Hz, 1H, H-1^a), 4.91 (s, 2H, H-1^{h,h'}), 4.84 (s, 1H, H-1^f), 4.61(s, 1H, H-1^c), 4.50 (d, J = 9.2 Hz, 1H, H-1^b), 4.49 (d, J = 8.1 Hz, 1H, H-1^e), 4.31 (s, 2H), 4.19 (bs, 1H), 4.07 (bs, 1H), 4.02-3.51 (m, 53H), 3.40-3.30 (m, 2H) 2.99 (t, J = 11.2 Hz, 2H, -NCH₂-, linker), 2.08 (s, 3H, -C(O)CH₃), 2.07 (s, 3H, -C(O)CH₃), 2.04 (s, 3H, -C(O)CH₃), 1.70–1.65 (m, 2H, -CCH₂C-, linker), 1.62-1.58 (m, 2H, -CCH₂C-, linker), 1.48–1.38 (m, 2H, -CCH₂C-, linker); ^{13}C NMR (150 MHz, D_2O): δ 177.60, 177.17, 105.68 (C-1^b), 105.05 (C-1^d), 104.23, 104.04, 103.79, 103.37, 103.18, 102.88, 102.23 (C-1^h), 102.03 (C-1^h), 100.24 (C-1^e), 82.20, 82.06, 82.01, 81.58, 81.51, 81.40, 79.38, 78.70, 78.08, 77.43, 77.37, 77.18, 77.03, 76.07, 75.66, 75.43, 75.23, 75.11, 75.06, 74.96, 74.83, 74.71, 73.72, 73.65, 73.33, 73.20, 73.11, 72.95, 72.89, 72.84, 72.69, 72.18, 71.28, 70.15, 68.50, 69.46, 69.36, 69.01, 68.31, 67.90, 67.86, 64.45, 63.75, 63.69, 63.61, 62.87, 62.82, 62.75, 57.94, 57.72, 42.06, 30.78, 29.10, 25.35, 25.30, 24.96, 24.87, 24.83; ESI-MS: m/z calcd for $\text{C}_{65}\text{H}_{112}\text{N}_4\text{O}_{46}$; 1684.6620 found 1685.6968 ($M + \text{H}$)⁺.



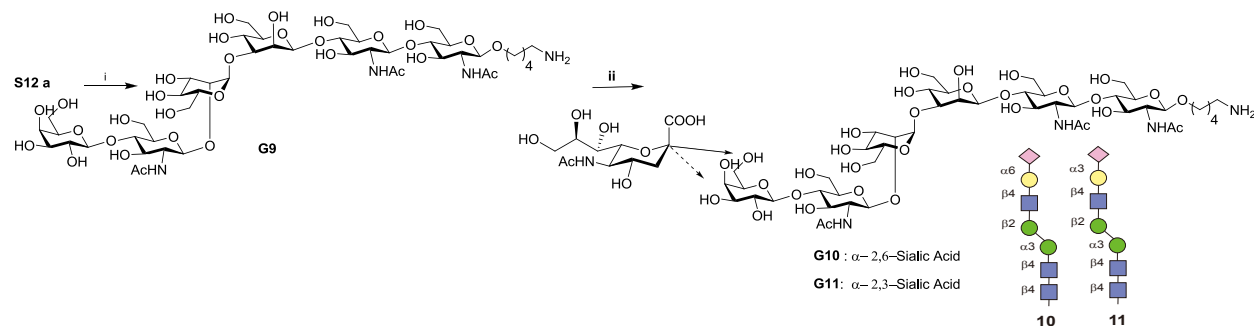
5-Aminopentyl-5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyrano-sylonate-(2 \rightarrow 6)- β -D-galactopyranosyl-(1 \rightarrow 4)-[2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl-(1 \rightarrow 3),[di-(α -D-mannopyranosyl)-(1 \rightarrow 3),(1 \rightarrow 6)- α -D-manno-pyranosyl] (1 \rightarrow 6)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranoside (G13): Compound G12 (5 mg, 0.0029 mmol) was sialylated with α -2,6-sialyltransferase for 2 days by following general procedure 3 to get the desired title compound G13 (3.5 mg, 60%) as white solid. ^1H NMR (600 MHz, D_2O): δ 5.07 (s, 1H, H-1^d), 5.06 (d, $J = 8.5$ Hz, 1H, H-1^a), 4.92 (s, 1H, H-1^h), 4.90 (s, 1Hⁱ), 4.89 (s, 1H, H-1ⁱ), 4.78 (s, 1H, H-1^c), 4.58 (d, $J = 7.2$ Hz, 1H, H-1^f), 4.46 (d, $J = 7.8$ Hz, 1H, H-1^b), 4.41 (d, $J = 8.4$ Hz, 1H, H-1^e), 4.25-4.24 (m, 2H), 4.12 (s, 1H), 4.04 (d, $J = 4.7$ Hz, 1H), 4.04-3.42 (m, 55H), 3.40 (t, $J = 10.8$ Hz, 1H), 3.39-3.35 (m, 1H), 2.95 (t, $J = 10.9$ Hz, 2H), 2.64 (dd, $J = 4.2, 12.2$ Hz, 1H, H-3^{equi. g}), 2.04 (s, 3H, -C(O)CH₃), 2.04 (s, 3H, -C(O)CH₃), 2.00 (s, 3H, -C(O)CH₃), 2.00 (s, 3H, -C(O)CH₃), 1.71-1.61 (m, 3H, -CCH₂C-, linker and H-3^{axial g}), 1.58-1.54 (m, 2H, -CCH₂C-, linker), 1.39-1.35 (m, 2H, -CCH₂C-, linker); ^{13}C NMR (150 MHz, D_2O): δ 174.90, 174.79, 174.38, 173.47, 103.42, 102.29 (C-1^d, $^1J_{\text{C,H}} = 170.9$ Hz), 101.48, 101.01, 100.41, 100.18 (C-1^a, $^1J_{\text{C,H}} = 162.9$ Hz), 100.15, 99.24 ((C-1^h, $^1J_{\text{C,H}} = 171.3$ Hz)), 97.87, 97.27, 80.78, 79.59, 79.20, 78.72, 78.36, 76.61, 75.81, 74.79, 74.48, 74.36, 73.63, 73.28, 72.63, 72.52, 72.35, 72.30, 72.09, 71.94, 71.65, 70.82, 70.69, 70.53, 70.31, 70.12, 70.05, 69.89, 69.41, 68.33, 68.17, 67.32, 67.17, 66.66, 65.47, 65.09, 63.30, 62.62, 61.66, 60.98, 60.89, 60.32, 60.02, 59.94, 57.95, 54.91, 51.81, 40.01, 39.27, 28.00, 26.37, 23.62, 23.19, 22.59, 22.17, 22.05, 22.00; ESI-MS (negative mode): m/z calcd for C₇₆H₁₂₉N₅O₅₄; 1975.7418 found 1974.7600 (M -H)⁻.



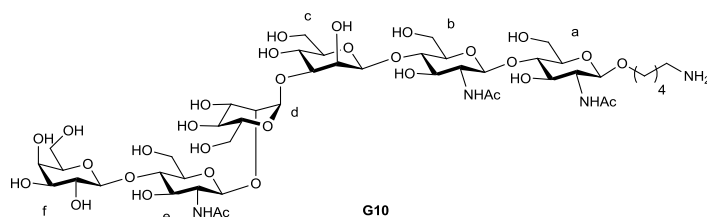
5-Aminopentyl-5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylate-(2 \rightarrow 3)- β -D-galactopyranosyl-(1 \rightarrow 4)-[2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl-(1 \rightarrow 3),[di-(α -D-mannopyranosyl)-(1 \rightarrow 3),(1 \rightarrow 6)- α -D-manno-pyranosyl] (1 \rightarrow 6)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranoside (G14): Compound G12 (5 mg, 0.0029 mmol) was sialylated with α -2,3-sialyltransferase for 3 d by following general procedure 3 to get the desired title compound G14 (3.1 mg, 56%) as white solid. ^1H NMR (600 MHz, D_2O): δ 5.01 (s, 1H, H-1^d), 4.97 (dd, $J = 3.2, 7.2$ Hz, 1H, H-1^a), 4.84 (s, 1H, H-1^h), 4.81 (s, 1H, H-1ⁱ), 4.75 (s, 1H, H-1ⁱ), 4.62 (s, 1H, H-1^c), 4.52 (d, $J = 7.8$ Hz, 1H, H-1^f), 4.97 (d, $J = 9.2$ Hz, 1H, H-1^b), 4.42 (d, $J = 8.3$ Hz, 1H, H-1^c), 4.39 (d, $J = 8.3$ Hz, 1H), 4.22 (t, $J = 7.8$ Hz, 2H), 4.09 (s, 1H), 4.06 (d, $J = 7.4$ Hz, 1H), 4.03 (d, $J = 7.3$ Hz, 1H), 4.00 (d, $J = 3.2$ Hz, 1H), 3.96-3.85 (m, 16H), 3.85-3.71 (m, 19H), 3.72-3.52 (m, 26H), 3.51-3.48 (m, 2H), 3.38-3.370 (m, 2H), 2.90 (t, $J = 7.9$ Hz, 2H, -NCH₂-, linker), 2.68 (dd, $J = 4.8, 12.0$ Hz, 1H, H-3^{equi. g}), 2.01 (s, 3H, -C(O)CH₃), 1.99 (s, 3H, -C(O)CH₃), 1.96 (s, 3H, -C(O)CH₃), 1.95 (s, 3H, -C(O)CH₃), 1.73 (t, $J = 8.6$ Hz, 1H, H-3^{axial g}), 1.61-1.56 (m, 2H, -CCH₂C-, linker), 1.54-1.46 (m, 2H, -CCH₂C-, linker), 1.34-1.33 (m, 2H, -CCH₂C-, linker); ^{13}C NMR (150 MHz, D_2O): δ 171.94, 171.81, 171.37, 170.80, 113.44, 99.86, 99.53, 99.27, 98.42, 97.99, 97.38, 97.08, 96.74, 96.21, 75.74, 75.66, 75.52, 73.56, 72.85, 72.41, 72.10, 71.62, 71.45, 71.36, 70.27, 69.81, 69.62, 69.28, 69.14, 69.02, 68.90, 68.69, 67.91, 67.82, 67.50, 67.20, 67.08, 66.87, 66.36, 65.46, 65.29, 65.01, 64.39, 64.32, 63.68, 62.47, 62.09, 61.02, 59.50, 59.39, 58.64, 57.96, 57.87, 57.10, 56.91, 52.12, 51.91, 48.60, 42.85, 40.32, 36.54, 36.25,

35.04, 25.27, 24.98, 23.34, 20.60, 19.53, 19.13, 19.03, 18.95, 13.70; ESI-MS (negative mode): m/z calcd for $C_{76}H_{129}N_5O_{54}$; 1975.7418 found 1974.7671 ($M - H$)⁻.

Preparation of glycans G9-G11

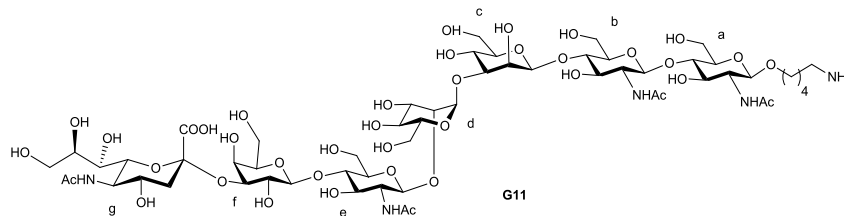


Scheme S13 | Preparation of G9-G11. i) (1) $NH_2CH_2CH_2NH_2$, n-BuOH, 90 °C, overnight; (2) Ac_2O , pyridine, overnight; (3) NaOMe, MeOH, overnight; (4) $Pd(OH)_2$, MeOH: H_2O : $HCOOH$ (5:3:2), H_2 ; 51%; ii) CMP- β -D-Sialic acid, α -2,6/2,3- sialyltransferase, **G10**: 77%; **G11**: 52%



5-Aminopentyl- β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl (1 \rightarrow 2)- α -D-mannopyranosyl]-(1 \rightarrow 3)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranoside (G10): Hexasaccharide **S12a** (0.110 g, 0.041 mmol) was deprotected following general procedure 2 (Method 1) to get the desired compound **G10** (0.025 g, 51%) as white solid. 1H NMR (600 MHz, D_2O): δ 5.04 (d, $J = 8.4$ Hz, 1H, H-1^a), 4.84 (s, 1H, H-1^d), 4.78 (s, 1H, H-1^c), 4.60 (d, $J = 8.2$ Hz, 1H, H-1^e), 4.51 (d, $J = 8.1$ Hz, 1H, H-1^b), 4.46 (d, $J = 7.8$ Hz, 1H, H-1^f), 4.28 (d, $J = 3.3$ Hz, 1H), 4.26 (d, $J = 3.6$ Hz, 1H), 4.10-3.82 (m, 9H), 3.82-3.56 (m, 22H), 3.52-3.38 (m, 5H), 2.97 (t, $J = 7.8$ Hz, 2H, -NCH₂-, linker), 2.07 (s, 3H, -C(O)CH₃), 2.05 (s, 3H, -C(O)CH₃), 2.03 (s, 3H, -C(O)CH₃), 1.70-

54.87, 51.85, 40.02, 39.30, 28.01, 26.36, 22.56, 22.10, 22.08, 21.99; ESI-MS (negative mode): m/z calcd for $C_{58}H_{99}N_5O_{39}$; 1489.5833 found 1488.5949 ($M - H$)⁻.



5-Aminopentyl-[5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyrano-sylonate-(2 \rightarrow 3)- β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl]-(1 \rightarrow 3)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranoside (G11): Compound **G9** (7 mg, 0.0058 mmol) was sialylated with α -2,3-sialyltransferase for 3 days by following general procedure 3 to get desired the title compound **G11** (4.6 mg, 52%) as white solid. ¹H NMR (600 MHz, D₂O): δ 5.06 (d, $J = 8.4$ Hz, 1H, H-1^a), 4.79 (s, 1H, H-1^d), 4.75 (s, 1H, H-1^c), 4.62 (d, $J = 6.6$ Hz, 1H, H-1^e), 4.56 (d, $J = 7.8$ Hz, 1H, H-1^b), 4.51 (d, $J = 7.8$ Hz, 1H, H-1^f), 4.28 (d, $J = 8.4$ Hz, 2H), 4.13 (d, $J = 9.6$ Hz, 1H), 3.99-3.83 (m, 12H), 3.82-3.57 (m, 25H), 3.53-3.39 (m, 4H), 2.99 (t, $J = 7.8$ Hz, 2H), 2.77 (dd, $J = 4.8, 2.0$ Hz, 1H, H-3^{equi. g}), 2.12 (s, 3H, -C(O)CH₃) 2.09 (s, 3H, -C(O)CH₃), 2.05 (s, 3H, -C(O)CH₃), 2.03 (s, 3H, -C(O)CH₃), 1.80 (t, $J = 7.8$ Hz, 1H, H-3^{axial g}), 1.71-1.66 (m, 2H, -CCH₂C-, linker), 1.62-1.59 (m, 2H, -CCH₂C-, linker), 1.44-1.41 (m, 2H, -CCH₂C-, linker); ¹³C NMR (150 MHz, D₂O): δ 174.88, 174.53, 174.46, 174.40, 102.50, 102.37, 101.65, 101.31, 100.12, 99.86, 98.31, 80.32, 79.67, 78.78, 78.56, 76.66, 75.32, 75.07, 74.51, 74.48, 74.38, 73.67, 72.52, 72.38, 72.36, 72.08, 71.90, 71.66, 70.70, 70.06, 68.37, 68.34, 67.33, 63.35, 62.63, 61.67, 60.88, 60.09, 59.98, 54.99, 54.87, 51.85, 40.02, 39.30, 28.01, 26.36, 22.56, 22.10, 22.08, 21.27 20.27; ESI-MS (negative mode): m/z calcd for $C_{58}H_{99}N_5O_{39}$; 1489.5833 found 1488.5944($M - H$)⁻.

vii. Synthesis of complex type oligosaccharides

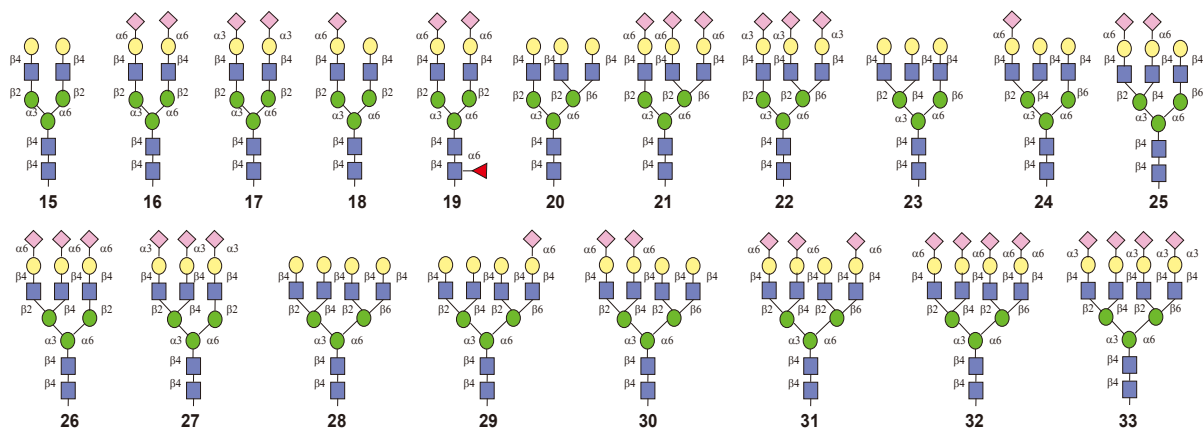
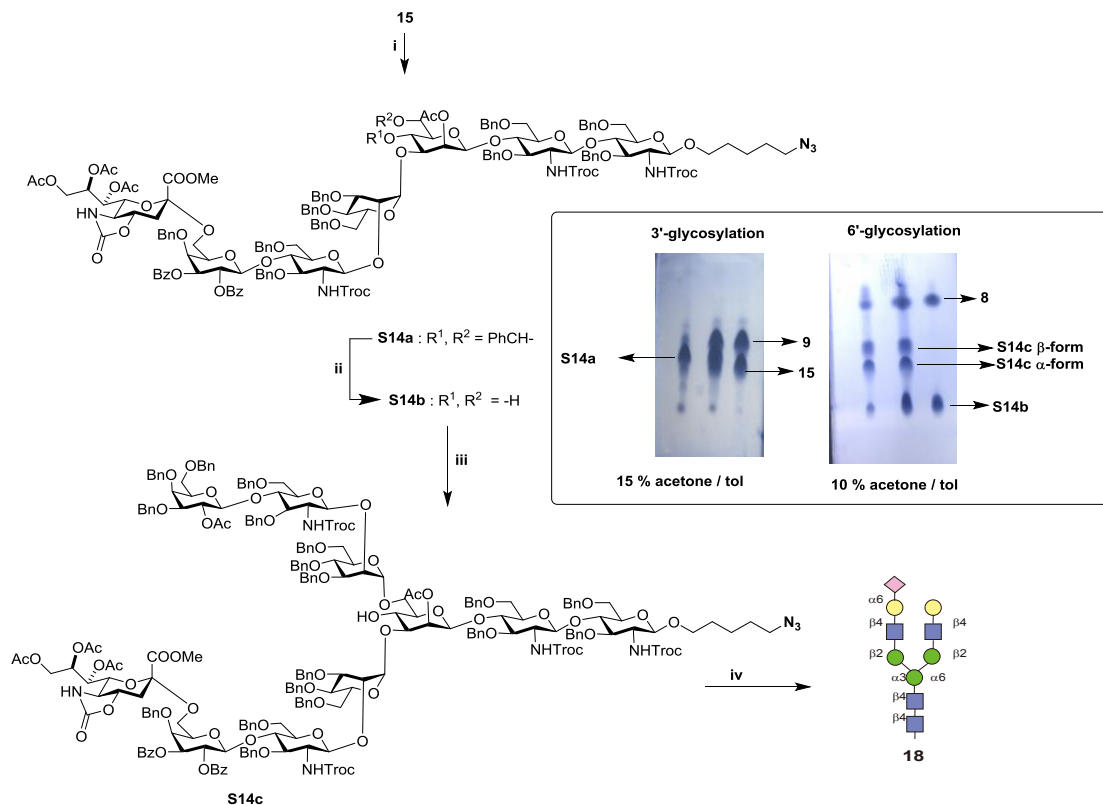


Figure S3 | Structures of complex type glycans.

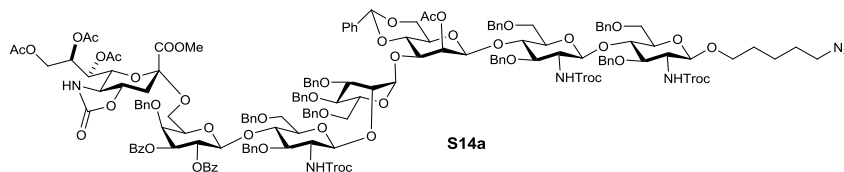
Previously, we have developed an efficient chemo-enzymatic strategy for the rapid production of bi-(G15, G16, and G17), tri- (G20-G23, G26, and G27) and tetraantennary (G28, G32, and G33) complex type *N*-glycans, with and without terminal *N*-acetylneuraminic acid residues connected via the α -2,6 or α -2,3 linkages. In addition, we have developed a total chemical synthesis strategy which utilized a modular set of glycosyl donors (1-13) for stereo- and region-selective glycosylation to core trisaccharide **15**. However, the synthesis of asymmetrically sialylated glycans by enzyme is complicated by their specificity. For this reason, we demonstrated the utility of glycosyl fluoride to prepare asymmetric complex type glycans. Initially, we started with phtallamide protections at C2-amine of all glucosamine residues of both antennae and core trisaccharides (**14**), however, in presence of preinstalled sialic acid, the process of global deprotection was found to be complicated. To overcome this difficulty, the phtallamide protections at C2-amine of all glucosamine residues were replaced with trichloroethyl carbonate (troc).

Synthesis of glycan G18.

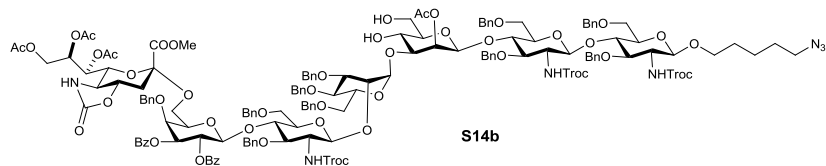
Glycosylation of sialylated antennae **9** with core **15** at 3-O site provided desired heptasaccharide **S14a** in 63% yield. Benzylidene was removed in the presence of *p*-TSA to get 4,6-diol **S14b**, which was further alpha glycosylated at 6-O position with **8** to afford desired **S14c** as mixture of α and β forms (7/3 isolated yield). The major α isomer was isolated by column chromatography and finally deprotected to get **G18** as a white solid.



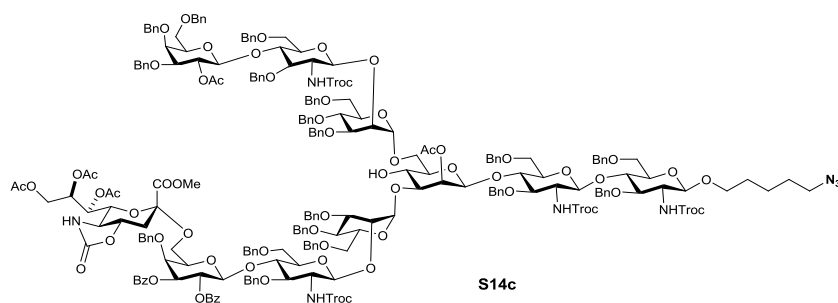
Scheme S14 | Preparation of G18. i, **9**, AgOTf, Cp₂HfCl₂, toluene, -20 °C to 0 °C, 63%; ii, *p*TSA, acetonitrile, 59%; iii, **8**, AgOTf, Cp₂HfCl₂, toluene, -40 °C to -20 °C, 58%; iv, (1) LiOH, 1, 4-dioxane: H₂O, 90 °C, (2) Ac₂O, pyridine, (3) NaOMe, MeOH, (4) Pd (OH)₂, MeOH: H₂O, H₂, 25%.



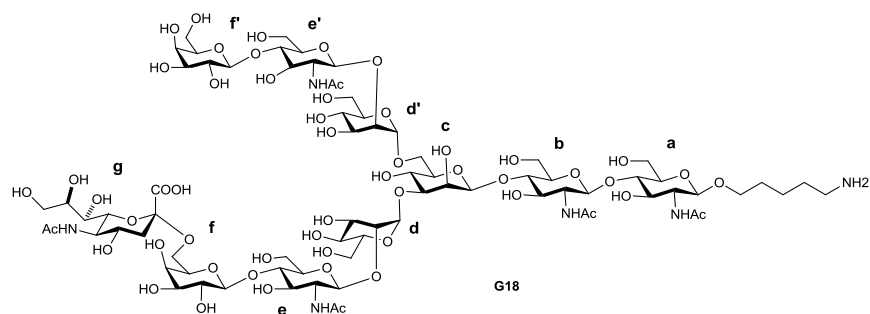
Compound S14a: A mixture of silver triflate (0.087 g, 0.34 mmol), bis (cyclopentadienyl) hafnium dichloride (0.090 g, 0.23 mmol) and 4 Å activated molecular sieves in dry toluene (10 mL) was stirred at rt for 1 h. The reaction mixture was then cooled to -20 °C, a solution of donor **9** (0.153 g, 0.082 mmol) and acceptor chitobiose trisaccharide **15** (0.100 g, 0.068 mmol) in 5 mL toluene was added. The mixture was stirred for 2 h at 0 °C, quenched with Et₃N, diluted with EtOAc and filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), and a brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 10% acetone in toluene) to afford **S14a** (0.140 g, 63%) as white foam. TLC: (acetone: toluene = 1.5/8.5, v/v): R_f = 0.52; ¹H NMR (600 MHz, CDCl₃): δ 7.89 (d, *J* = 7.8 Hz, 4H), 7.49-7.46 (m, 2H), 7.37-7.06 (m, 59H), 5.77 (t, *J* = 10.8 Hz, 1H), 5.35-5.37 (m, 2H), 5.26 (s, 1H), 5.18 (dd, *J* = 3.8 & 7.7 Hz, 1H), 5.08 (d, *J* = 3.2 Hz, 1H), 5.07 (dd, *J* = 3.2 & 8.4 Hz, 1H), 4.97 (d, *J* = 8.4 Hz, 1H), 4.91 (d, *J* = 9.1 Hz, 1H), 4.86 (d, *J* = 8.7 Hz, 3H), 4.73-4.13 (m, 32H), 4.02-3.30 (m, 30H), 3.20 (t, *J* = 10.8 Hz, 2H), 3.10 (d, *J* = 8.7 Hz, 1H), 2.98 (t, *J* = 9.2 Hz, 1H), 2.87-2.85 (m, 1H), 2.80 (dd, *J* = 3.2 & 7.8 Hz, 1H), 2.10 (s, 6H), 2.08 (t, *J* = 9.3 Hz, 1H), 2.00 (s, 6H), 1.40-1.17 (m, 4H), 0.87-0.82 (m, 2H); ¹³C NMR (150 MHz, CDCl₃): δ 171.9, 170.8, 170.2, 170.0, 167.9, 166.1, 165.5, 159.5, 154.5, 154.3, 154.1, 138.2, 130.1, 129.8, 129.6, 129.4, 128.9, 128.8, 128.7, 128.6, 128.5, 128.4, 128.3, 128.2, 128.1, 127.9, 127.8, 127.6, 127.0, 100.8, 100.5, 98.0, 95.8, 76.8, 76.7, 75.5, 74.8, 74.6, 74.4, 74.3, 74.1, 73.8, 73.7, 73.4, 73.2, 69.9, 69.7, 69.5, 68.7, 67.5, 66.7, 61.9, 58.9, 57.6, 53.4, 37.4, 29.9, 29.2, 28.8, 23.4, 21.3, 21.2, 21.0, 20.9, 14.3; ESI-MS: *m/z* calcd for C₁₆₀H₁₇₂Cl₉N₇O₄₈; 3278.8343 found 3278.8127.



Compound S14b: *p*-Toluene sulfonic acid monohydrate (0.004 g, 0.022 mmol) was added to a solution of **S14a** (0.150 g, 0.045 mmol) in acetonitrile (5 mL) and the resulting reaction mixture was stirred at room temperature for 5h. The reaction was quenched by adding Et₃N and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 15% EA in toluene) to give diol **S14b** (0.085 g, 58%). TLC: (acetone: toluene = 2/8, v/v): R_f = 0.32; ¹H NMR (600 MHz, CDCl₃): δ 7.89-7.86 (m, 4H), 7.49-7.37 (m, 2H), 7.30-7.09 (m, 54H), 5.78 (t, *J* = 10.2 Hz, 1H), 5.51 (d, *J* = 7.8 Hz, 1H), 5.37 (dt, *J* = 2.4 & 7.2 Hz, 1H), 5.26 (s, 1H), 5.19 (dd, *J* = 2.4 & 10.8 Hz, 1H), 5.12 (d, *J* = 3.6 Hz, 1H), 4.95-4.85 (m, 5H), 4.75-4.12 (m, 25H), 4.08-4.03 (m, 3H), 3.96-3.89 (m, 7H), 3.74-3.67 (m, 8H), 3.63-3.30 (m, 20H), 3.20 (t, *J* = 10.2 Hz, 2H), 3.10 (d, *J* = 8.9 Hz, 2H), 2.89 (s, 1H), 2.76 (dd, *J* = 3.6 & 7.9 Hz, 1H), 2.08 (s, 3H), 2.05 (s, 3H), 1.99 (s, 3H), 1.95 (s, 3H), 1.41-1.31 (m, 4H), 1.27-1.17 (m, 2H); ¹³C NMR (150 MHz, CDCl₃): δ 172.4, 172.3, 171.3, 171.2, 170.7, 170.3, 170.0, 168.4, 168.3, 166.1, 165.5, 154.5, 154.3, 154.1, 153.9, 138.5, 138.4, 138.1, 138.0, 133.7, 133.3, 130.1, 129.8, 129.7, 129.5, 129.3, 128.9, 128.8, 128.7, 128.6, 128.6, 128.5, 128.4, 128.3, 128.2, 128.1, 128.0, 127.9, 101.1, 100.8, 100.6, 100.4, 99.7, 99.2, 96.0, 95.8, 76.3, 76.1, 75.7, 75.5, 74.9, 74.8, 74.5, 74.3, 74.0, 73.7, 73.4, 73.3, 73.2, 73.1, 73.0, 72.6, 72.3, 72.1, 71.3, 71.0, 69.9, 69.6, 69.4, 69.3, 68.4, 67.8, 66.1, 66.2, 62.8, 62.2, 59.2, 57.7, 53.4, 51.5, 36.9, 29.9, 29.2, 28.8, 25.0, 23.4, 21.4, 21.3, 21.2, 21.1, 15.5, 14.4; ESI-MS (negative mode): *m/z* calcd for C₁₅₃H₁₆₈Cl₉N₇O₄₈; 3191.8105 found 3236.8033 (*M* + 2Na).



Compound S14c: A mixture of silver triflate (0.067 g, 0.27 mmol), bis (cyclopentadienyl) hafnium dichloride (0.071 g, 0.18 mmol) and 4 Å activated molecular sieves in dry toluene (10 mL) was stirred at rt for 1 h. The reaction mixture was then cooled to -40 °C, a solution of donor **8** (0.097 g, 0.068 mmol) and acceptor **S14b** (0.175 g, 0.054 mmol) in 5 mL toluene was added. The mixture was stirred for 2 h at -20 °C, quenched with Et₃N, diluted with EtOAc and filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), and a brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 10% acetone in toluene) to separate mixture of α and β isomers to afford **S14c** (0.149 g, 58%) as white foam. TLC: (acetone: toluene = 1.5/8.5, v/v): R_f = 0.52; ¹H NMR (600 MHz, CDCl₃): δ 7.90 (d, *J* = 7.2 Hz, 2H), 7.83 (d, *J* = 7.8 Hz, 2H), 7.51-7.41 (m, 4H), 7.34-7.06 (m, 92H), 5.83 (t, *J* = 10.2 Hz, 1H), 5.48-5.03 (m, 3H), 5.00-4.75 (m, 13H), 4.60-4.08 (m, 41H), 3.99-3.07 (m, 53H), 2.96 (t, *J* = 10.2 Hz, 2H), 2.80 (dd, *J* = 3.8 & 7.9 Hz, 1H), 2.15 (s, 3H), 2.10 (s, 3H), 2.07 (s, 3H), 2.00 (s, 3H), 1.93 (s, 3H), 1.40-1.17 (m, 5H), 0.91-0.81 (m, 2H); ¹³C NMR (150 MHz, CDCl₃): δ 170.9, 170.6, 170.4, 169.9, 169.4, 166.5, 159.5, 154.7, 154.4, 139.8, 138.8, 138.7, 138.6, 138.4, 138.2, 133.7, 130.9, 130.7, 129.7, 129.6, 129.4, 129.2, 129.0, 128.7, 128.6, 128.6, 128.4, 128.3, 128.2, 127.9, 127.6, 127.3, 100.4, 100.2, 99.8, 99.4, 74.9, 74.7, 74.5, 73.3, 73.2, 66.1, 60.6, 51.5, 32.1, 29.8, 29.7, 29.6, 29.3, 28.7, 23.7, 22.8, 22.4, 22.0, 21.6, 21.4, 21.2, 20.9, 15.4, 14.3, 14.2, 14.0, 11.3, 10.2; ESI-MS: *m/z* calcd for C₂₃₂H₂₅₀Cl₁₂N₈O₆₅; 4607.2765 found 2330.6261 (*M* + Na)²⁺.



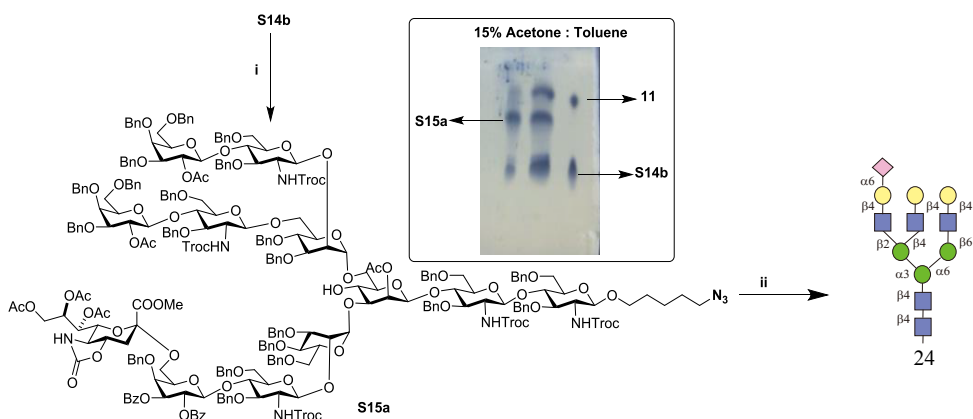
5-Aminopentyl-[5-Acetamido-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyrano-sylonate-(2→6)-β-D-galactopyranosyl-(1→4)-2-acetamido-2-deoxy-β-D-glucopyranosyl-(1→2)-α-D-

mannopyranosyl)-(1→3),-[β-D-galactopyranosyl-(1→4)-2-acetamido-2-deoxy-β-D-glucopyranosyl-(1→2)-α-D-mannopyranosyl-(1→6)-β-D-mannopyranosyl-(1→4)-2-acetamido-2-deoxy-β-D-glucopyranosyl-(1→4)-2-acetamido-2-deoxy-β-D-glucopyranoside

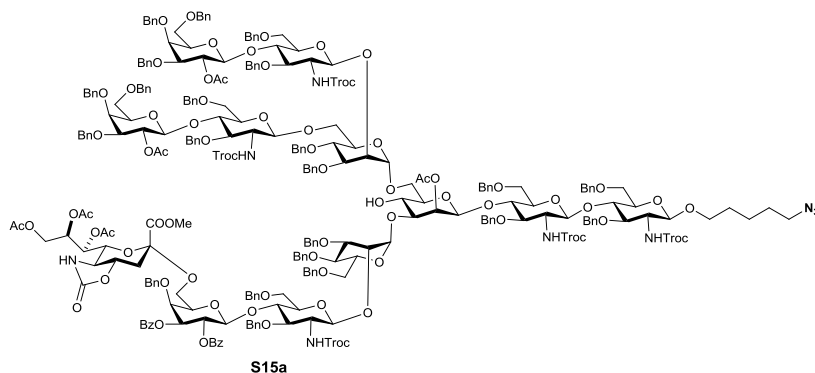
G18: Compound **S14c** (0.135 g, 0.029 mmol) was deprotected by following general procedure 2 (method 2) to afford the desired glycan **G18** (0.015 g, 25%). ¹H NMR (600 MHz, D₂O): δ 5.11 (s, 1H, H-1^d), 4.89 (s, 1H, H-1^d), 4.73 (s, 1H, H-1^c), 4.57 (dt, *J* = 10.2 Hz, 2H, H-1^{f,f}), 4.47 (d, *J* = 7.8 Hz, 2H, H-1^{a,b}), 4.47 (d, *J* = 8.4 Hz, 2H, H-1^{e,e'}), 4.23 (s, 1H), 4.17 (s, 1H), 4.09 (d, *J* = 1.3 Hz, 1H), 4.03-3.48 (m, 59H), 2.98 (t, *J* = 10.2 Hz, 2H, -CH₂- linker), 2.65 (dd, *J* = 4.8 & 12.6 Hz, 1H, H-3^{equi. g}), 2.10 (s, 3H), 2.09 (s, 3H), 2.07 (s, 3H), 2.02 (s, 6H), 1.72-1.64 (m, 3H, H-3^{axial g}, linker -CH₂-), 1.63-1.54 (m, 2H), 1.39-1.35 (m, 2H); ¹³C NMR (150 MHz, D₂O): δ 177.7, 177.5, 177.4, 177.1, 176.2, 176.1, 106.2, 106.1, 104.1, 103.7, 103.5, 103.1, 102.9, 102.9, 99.5, 83.6, 83.1, 82.2, 82.1, 81.7, 79.4, 79.1, 78.9, 77.3, 77.2, 76.8, 76.4, 75.3, 75.1, 74.8, 74.5, 73.5, 73.4, 72.8, 71.2, 70.9, 70.0, 67.6, 66.0, 65.6, 65.4, 64.5, 62.8, 57.7, 57.3, 55.0, 54.6, 54.0, 42.8, 42.3, 42.0, 41.78, 30.7, 30.5; ESI-MS (negative mode): *m/z* calcd for C₇₈H₁₃₂N₆O₅₄ 2016.7767; found 1007.3799 (*M* - H)²⁻.

Synthesis of glycan **G24**.

Compound **S14b** acts as common acceptor for both **G18** and **G24**. Glycosylation of **S14b** at 6-O site provided the desired **S15a** with 10% acceptor recovery. At last, global deprotection yielded the desired glycan **G24**.

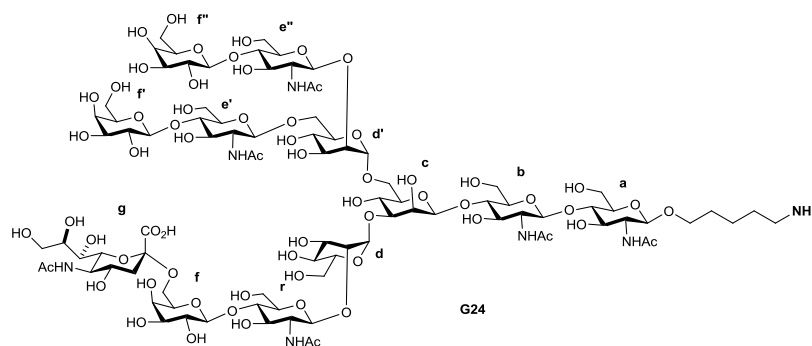


Scheme S15 | Preparation of G24. i, **11**, AgOTf, Cp₂HfCl₂, toluene, -40 °C, 63%; ii, (1) LiOH, 1, 4-dioxane, 90 °C, (2) Ac₂O, pyridine, (3) NaOMe, MeOH, (4) Pd(OH)₂, MeOH: H₂O, H₂, 42%.



Compound S15a: A mixture of silver triflate (0.058 g, 0.23 mmol), bis (cyclopentadienyl) hafnium dichloride (0.061 g, 0.23 mmol) and 4 Å activated molecular sieves in dry toluene (10 mL) was stirred at rt for 1 h. The reaction mixture was then cooled to -40 °C, a solution of donor **11** (0.135 g, 0.056 mmol) and acceptor **S14b** (0.150 g, 0.046 mmol) in 5 mL toluene was added. The mixture was stirred for 2 h at -20 °C, quenched with Et₃N, diluted with EtOAc and filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), and a brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 10% acetone in toluene) to afford **S15a** (0.165 g, 63%) as white foam. TLC: (acetone: toluene = 1.5/8.5, v/v): R_f = 0.58; ¹H NMR (600 MHz,

CDCl₃): δ 8.00-7.80 (m, 4H), 7.50-6.80 (m, 116H), 5.80 (t, *J* = 10.2 Hz, 1H), 5.40-5.30 (m, 7H), 5.10-4.08 (m, 65H), 4.00-3.05 (m, 65H), 2.98 (dd, *J* = 3.2 & 8.5 Hz, 1H), 2.15 (s, 6H), 2.10 (s, 6H), 2.00 (s, 6H), 1.37-1.17 (m, 5H), 0.87-0.84 (m, 2H); ¹³C NMR (150 MHz, CDCl₃): δ 170.8, 167.9 154.1, 139.1, 139.0, 138.9, 138.8, 138.6, 138.3, 138.1, 138.0, 129.8, 129.6, 129.3, 128.9, 128.8, 128.7, 128.7, 128.6, 128.5, 128.4, 128.2, 128.0, 127.7, 127.4, 127.3, 127.2, 127.1, 127.0, 100.8, 100.0, 74.9, 74.8, 74.5, 74.4, 73.6, 73.3, 72.6, 53.5, 29.9, 29.2, 23.4, 21.3, 21.2, 21.0, 20.9, 15.5, 14.4; ESI-MS (negative mode): *m/z*. calcd for C₂₇₇H₂₉₈Cl₁₅N₉O₇₇; 5517.1670 found 2803.7442 (*M* + 2Na)²⁺.

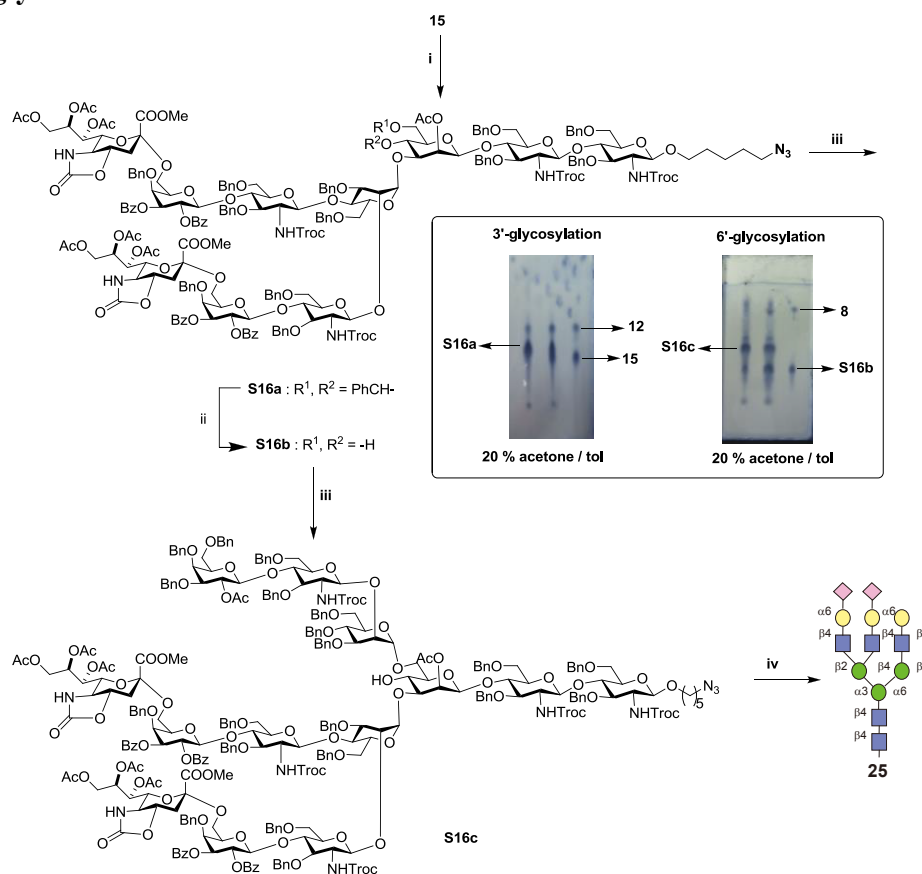


5-Aminopentyl-[5-Acetamido-3,5-dideoxy-D-glycero-α-D-galacto-2-nonulopyrano-sylonate-(2→6)-β-D-galactopyranosyl-(1→4)-2-acetamido-2-deoxy-β-D-glucopyranosyl-(1→2)-α-D-mannopyranosyl]-(1→3),-[β-D-galactopyranosyl-(1→4)-2-acetamido-2-deoxy-β-D-glucopyranosyl-(1→2), (1→6)-α-D-mannopyranosyl]-(1→6)-β-D-mannopyranosyl-(1→4)-2-acetamido-2-deoxy-β-D-glucopyranosyl-(1→4)-2-acetamido-2-deoxy-β-D-

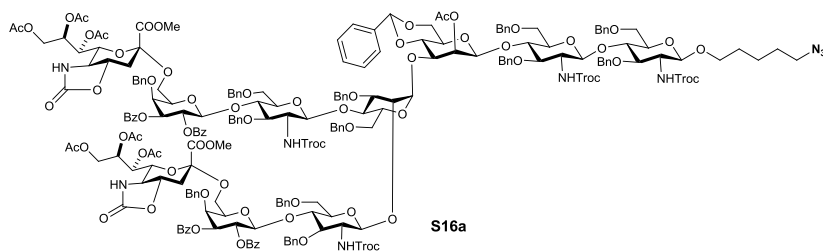
glucopyranoside G24: Compound **S15a** (0.090 g, 0.016 mmol) was deprotected by following general procedure 2 (method 2) to afford desired glycan **G24** (0.016 g, 42%) as a white solid. ¹H NMR (600 MHz, D₂O): δ 5.14 (s, 1H, H-1^d), 4.88 (s, 1H, H-1^{d'}), 4.60 (t, *J* = 10.2 Hz, 3H, H-1^{e,f,e'}), 4.55 (d, *J* = 8.4 Hz, 1H), 4.48 (dd, *J* = 3.2 & 8.6 Hz, 4H), 4.45 (d, *J* = 7.8 Hz, 1H), 4.26 (s, 1H), 4.20 (s, 1H), 4.09 (s, 1H), 4.01-3.50 (m, 71H), 2.97 (t, *J* = 10.2 Hz, 2H, linker -CH₂-), 2.68 (dd, *J* = 3.8 & 8.3 Hz, 1H, H-3^{equi. g}), 2.08 (s, 3H, -C(O)CH₃), 2.07 (s, 3H, -C(O)CH₃), 2.04 (s,

3H, -C(O)CH₃), 2.03 (s, 6H, -C(O)CH₃), 1.72 (t, *J* = 10.2 Hz, 1H, H-3^{axial}g), 1.67-1.65 (m, 2H), 1.60-1.56 (m, 2H, linker -CH₂-), 1.40-1.39 (m, 2H, linker -CH₂-); ¹³C NMR (150 MHz, D₂O): δ 174.9, 174.7, 174.6, 174.5, 174.4, 174.1, 173.5, 103.6, 103.5, 102.9, 102.8, 101.5, 101.4, 101.0, 100.3, 100.1, 99.5, 99.3, 80.7, 80.4, 79.5, 79.3, 79.2, 78.5, 76.5, 75.4, 75.2, 74.7, 74.5, 74.4, 74.3, 73.7, 73.4, 72.5, 72.4, 72.3, 72.1, 71.9, 71.6, 71.3, 70.9, 70.8, 70.5, 70.3, 69.5, 69.4, 68.5, 68.3, 68.2, 67.5, 67.3, 65.4, 63.3, 62.5, 61.7, 61.0, 60.2, 60.1, 59.8, 55.08, 54.9, 54.8, 54.6, 51.0, 40.9, 39.4, 28.0, 26.7, 22.5, 22.4, 22.3, 22.2, 22.1, 22.0, 21.09 ; ESI-MS (negative mode): *m/z* calcd for C₉₂H₁₅₅N₇O₆₄; 2383.2370 found 1213.4483 (*M* + Na)²⁻.

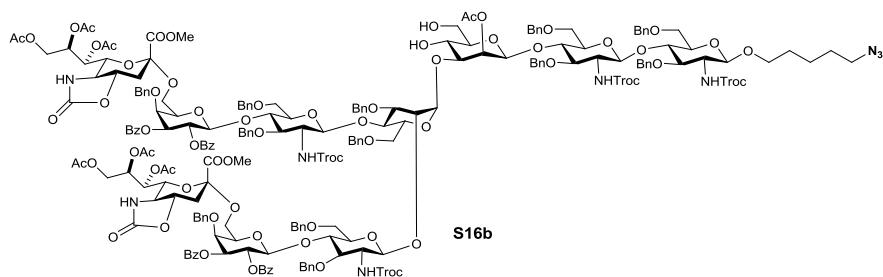
Synthesis of glycan G25.



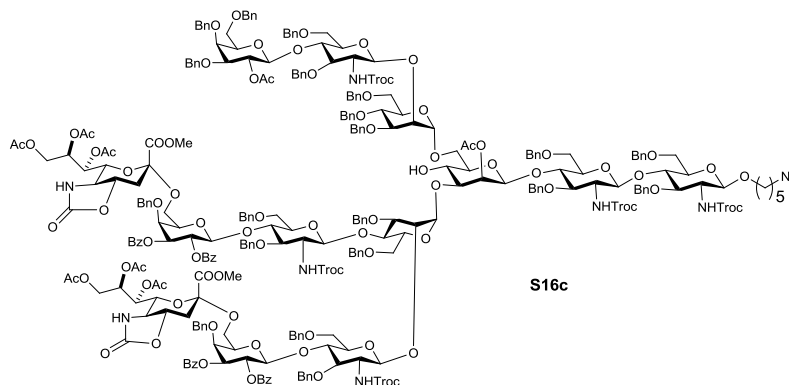
Scheme S16 | Preparation of G25. i, **12**, AgOTf, Cp₂HfCl₂, toluene, -40 °C, 64%; ii, *p*TSA, acetonitrile, 56%; iii, **8**, AgOTf, Cp₂HfCl₂, toluene, -78 °C, 49%; iv, (1) LiOH, 1,4-dioxane, 90 °C, (2) Ac₂O, pyridine, (3) NaOMe, MeOH, (4) Pd(OH)₂, MeOH: H₂O, H₂, 38%.



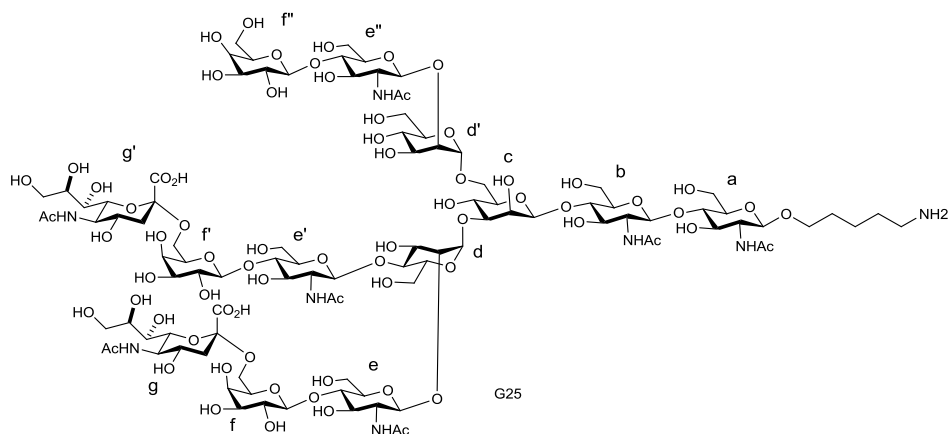
Compound 16a: A mixture of silver triflate (0.087 g, 0.34 mmol), bis (cyclopentadienyl) hafnium dichloride (0.090 g, 0.23 mmol) and 4 Å activated molecular sieves in dry toluene (10 mL) was stirred at rt for 1 h. The reaction mixture was then cooled to -40 °C, a solution of donor **12** (0.259 g, 0.082 mmol) and acceptor **15** (0.100 g, 0.068 mmol) in 5 mL toluene was added. The mixture was stirred for 3 h at -10 °C, quenched with Et₃N, diluted with EtOAc and filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), and a brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 20% acetone in toluene) to afford **S16a** (0.201 g, 64%) as white foam. TLC: (acetone: toluene = 2.5/7.5, v/v): R_f = 0.50; ¹H NMR (600 MHz, CDCl₃): δ 7.92-7.83 (m, 9H), 7.69-7.67 (m, 2H), 7.52-7.44 (m, 6H), 7.37-6.96 (m, 68H), 5.34-5.32 (m, 2H), 5.26-5.24 (m, 4H), 5.20-5.10 (m, 3H), 5.08 (d, *J* = 9.8 Hz, 3H), 4.59-4.40 (m, 16H), 4.75-4.50 (m, 28H), 4.49-4.00 (m, 20H), 3.98-3.60 (m, 16H), 3.48-3.06 (m, 16H), 2.98 (t, *J* = 10.7 Hz, 2H), 2.76 (dd, *J* = 3.8 & 7.8 Hz, 2H), 2.08 (s, 3H), 2.07 (s, 3H), 2.06 (s, 6H), 1.99 (s, 9H), 1.37-1.17 (m, 6H), 0.89-0.84 (m, 2H); ¹³C NMR (150 MHz, CDCl₃): δ 171.8, 171.7, 171.6, 169.8, 169.4, 168.0, 167.6, 166.6, 166.4, 166.2, 165.4, 165.3, 159.7, 159.4, 154.3, 154.2, 139.8, 139.7, 139.6, 139.4, 139.2, 138.7, 138.6, 138.5, 138.4, 138.3, 138.2, 133.9, 132.7, 131.9, 131.2, 130.6, 129.8, 129.6, 129.4, 129.3, 129.2, 129.0, 128.8, 128.7, 128.6, 128.6, 128.4, 128.2, 128.1, 128.0, 127.9, 127.8, 127.7, 127.6, 127.5, 127.4, 127.3, 127.2, 127.0, 102.7, 102.4, 101.9, 101.8, 100.8, 100.7, 100.4, 100.2, 100.0, 99.2, 99.4, 98.4, 96.5, 95.9, 74.3, 74.1, 74.0, 73.8, 73.7, 73.5, 73.4, 73.2, 73.1, 72.7, 72.5, 72.1, 71.3, 70.9, 70.4, 69.5, 69.4, 68.5, 68.4, 68.3, 68.1, 67.9, 67.4, 53.4, 51.6, 37.5, 33.9, 32.2, 31.8, 29.9, 28.8, 26.5, 23.4, 22.9, 21.2, 21.0, 20.0, 14.6, 14.3; ESI-MS: *m/z* calcd for C₂₂₀H₂₃₅Cl₁₂N₉O₇₂; 4582.6910 found 915.6684 (*M* + H)⁵⁺.



Compound 16b: *p*-Toluene sulfonic acid monohydrate (0.002 g, 0.008 mmol) was added to a solution of **S14a** (0.200 g, 0.043 mmol) in acetonitrile (10 mL) and resulting reaction mixture was stirred at rt for overnight. Reaction was quenched by adding Et₃N and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 20% acetone in toluene) to give diol **S16b** (0.110 g, 56%). TLC: (acetone: toluene = 2.5/7.5, v/v): R_f = 0.32; ¹H NMR (600 MHz, CDCl₃): δ 7.90-7.80 (m, 8H), 7.79 (d, *J* = 8.4 Hz, 2H), 7.69-7.46 (m, 5H), 7.41-7.04 (m, 65H), 5.78 (t, *J* = 10.1 Hz, 1H), 5.73 (t, *J* = 10.7 Hz, 1H), 5.35-5.25 (m, 7H), 5.20 (dd, *J* = 3.2 & 7.9 Hz, 1H), 5.15 (dd, *J* = 3.8 & 8.4 Hz, 1H), 5.08 (t, *J* = 10.8 Hz, 2H), 4.90-4.80 (m, 5H), 4.79-4.50 (m, 15H), 4.48-4.30 (m, 18H), 4.20-4.10 (m, 25H), 3.92-3.20 (m, 35H), 3.2 (t, *J* = 8.8 Hz, 2H), 3.18-3.05 (m, 3H), 3.0-2.95 (m, 3H), 2.81-2.70 (m, 3H), 2.08 (s, 6H), 2.07 (s, 6H), 2.03 (s, 6H), 1.92 (s, 6H), 1.32-1.17 (m, 6H), 0.89-0.83 (m, 2H); ¹³C NMR (150 MHz, CDCl₃): δ 171.8, 170.7, 168.0, 166.1, 159.6, 138.1, 133.7, 132.7, 131.1, 130.1, 129.9, 129.7, 129.4, 128.9, 128.8, 128.7, 128.6, 128.4, 128.3, 128.2, 127.8, 127.7, 127.6, 74.9, 74.7, 74.6, 73.9, 68.8, 66.1, 53.4, 51.6, 37.3, 32.4, 31.4, 29.9, 29.8, 29.7, 28.7, 28.4, 23.4, 22.3, 20.9, 20.7, 20.4, 15.5, 14.6, 14.2 ; ESI-MS: *m/z* calcd for C₂₁₃H₂₃₁Cl₁₂N₉O₇₂; 4494.5820 found 681.2953 (*M* + *K*)⁷⁺.



Compound 16c: A mixture of silver triflate (0.068 g, 0.34 mmol), bis (cyclopentadienyl) hafnium dichloride (0.065 g, 0.17 mmol) and 4 Å activated molecular sieves in dry toluene (10 mL) was stirred at rt for 1 h. The reaction mixture was then cooled to -78 °C, a solution of donor **8** (0.109 g, 0.076 mmol) and acceptor **16b** (0.230 g, 0.051 mmol) in 5 mL toluene was added. The mixture was stirred for 3 h at -20 °C, quenched with Et₃N, diluted with EtOAc and filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), and a brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 20% acetone in toluene) to afford **S16c** (0.148 g, 49%) as white foam. TLC: (acetone: toluene = 2.5/7.5, v/v): R_f = 0.50; ¹H NMR (600 MHz, CDCl₃): δ 7.90 (m, 10H), 7.52-6.80 (m, 110H), 5.79 (t, *J* = 10.2 Hz, 2H), 5.48 (dd, *J* = 4.2 & 8.7 Hz, 2H), 5.34-5.14 (m, 10H), 4.98-3.10 (m, 123H), 2.85 (t, *J* = 10.2 Hz, 2H), 2.30 (dd, *J* = 3.4 & 8.2 Hz, 2H), 2.01 (s, 3H), 2.00 (s, 6H), 1.95 (s, 6H), 1.94 (s, 6H), 1.91 (s, 3H), 1.39-1.29 (m, 6H), 1.26-1.91 (m, 2H); ¹³C NMR (150 MHz, CDCl₃): δ 172.1, 170.8, 170.3, 170.1, 170.0, 169.3, 169.2, 168.0, 167.3, 166.5, 165.8, 165.1, 165.0, 154.0, 153.8, 153.7, 153.7, 139.0, 138.9, 138.8, 138.7, 138.6, 138.3, 138.2, 138.1, 137.9, 137.4, 133.4, 133.2, 132.0, 130.3, 129.8, 129.7, 129.4, 129.3, 129.2, 129.1, 128.9, 128.8, 128.5, 128.3, 128.2, 128.1, 128.0, 127.9, 100.4, 100.2, 99.3, 75.8, 75.6, 75.4, 75.3, 75.2, 75.1, 74.8, 74.7, 74.6, 74.5, 74.4, 74.3, 74.2, 74.2, 74.1, 73.5, 73.4, 73.3, 73.2, 73.1, 73.0, 72.9, 72.6, 72.2, 71.9, 71.7, 71.6, 71.3, 71.1, 70.1, 69.9, 69.8, 68.5, 68.3, 68.2, 63.4, 58.8, 58.3, 58.0, 53.4, 51.3, 31.9, 30.3, 30.0, 29.8, 29.7, 29.5, 28.7, 28.5, 28.3, 27.1, 24.5, 24.1, 23.8, 22.7, 22.1, 21.7, 20.9, 20.7, 20.6, 14.1, 14.0, 10.9 ; ESI-MS: *m/z* calcd for C₂₉₂H₃₁₃Cl₁₅N₁₀O₈₉; 5918.4470 found 1486.4746 (*M* + 6H)⁴⁺.

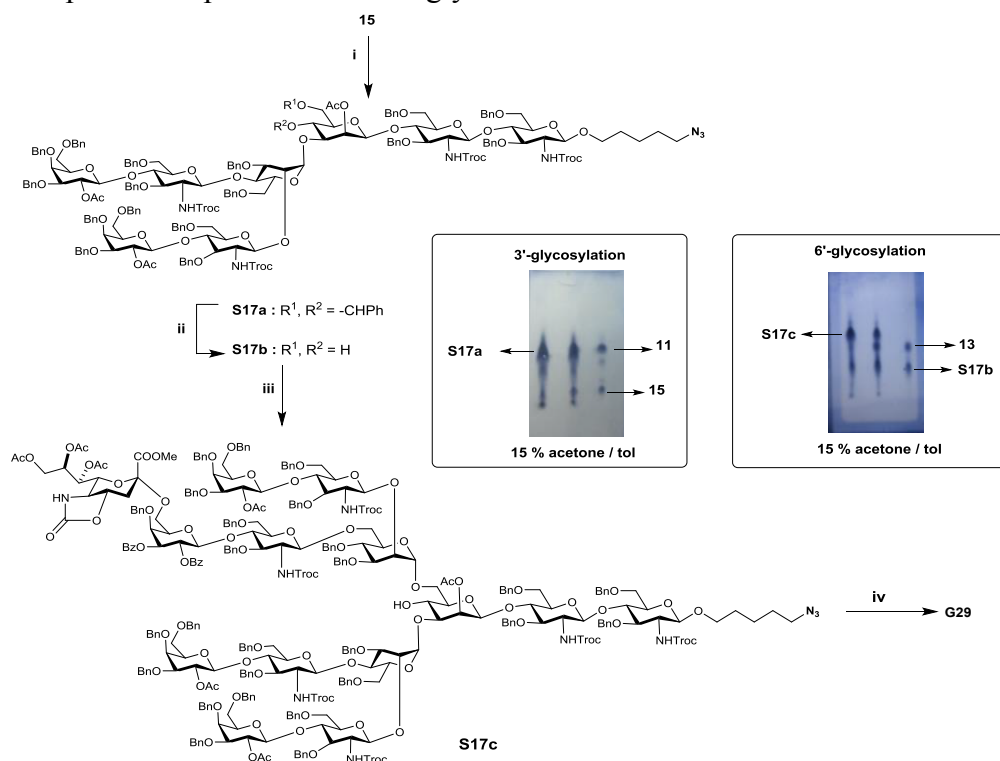


5-Aminopentyl-di-[5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate-(2 \rightarrow 6)- β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 2),(1 \rightarrow 4)- α -D-mannopyranosyl]-(1 \rightarrow 3)-[β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl]-(1 \rightarrow 6)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-

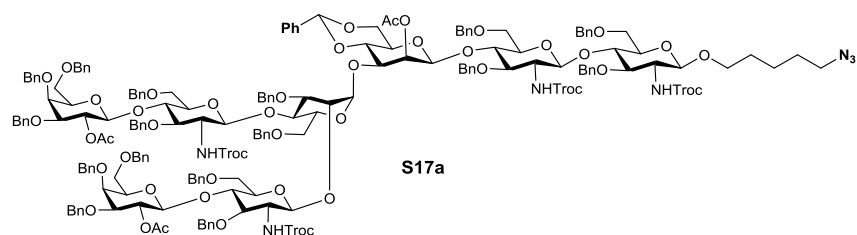
glucopyranoside G25: Compound **S16c** (0.115 g, 0.019 mmol) was deprotected by following general procedure 2 (method 2) to afford the desired glycan **G25** (0.020 g, 38%). ^1H NMR (600 MHz, D_2O): δ 5.15 (d, 1H, H-1^d), 4.94 (s, 1H, H-1^{d'}), 4.63 (d, $J = 8.4$ Hz, 2H, H-1^{a,b}), 4.58 (d, $J = 7.8$ Hz, 2H, H-1^{e,e'}), 4.52 (d, $J = 7.8$ Hz, 2H, H-1^{e'',f'}), 4.50 (s, 1H, H-1^c), 4.42 (dd, $J = 3.2$ & 10.2 Hz, 2H, H-1^{f,f''}), 4.23 (s, 2H), 4.08 (s, 1H), 3.92 (dd, $J = 3.2$ & 7.8 Hz, 2H), 3.90-3.45 (m, 75H), 3.01 (t, $J = 10.2$ Hz, 2H, linker CH_2), 2.69 (dd, $J = 3.6$ & 12.1 Hz, 2H, H-3_{equi.}^{g,g'}), 2.12 (s, 3H), 2.10 (s, 3H), 2.09 (s, 3H), 2.07 (s, 3H), 2.05 (s, 9H), 1.77-1.66 (m, 4H, H-3_{axial.}^{g,g'} and linker CH_2), 1.62-1.60 (m, 2H), 1.42-1.40 (m, 2H); ^{13}C NMR (150 MHz, D_2O): δ 177.6, 177.4, 177.3, 177.1, 176.1, 106.3, 105.6, 104.2, 104.1, 103.7, 103.1, 102.8, 102.1, 101.8, 99.8, 78.7, 78.0, 77.4, 77.2, 77.1, 76.4, 75.5, 75.2, 75.1, 75.0, 74.8, 74.4, 74.0, 73.7, 73.4, 73.0, 72.8, 72.1, 71.5, 70.9, 70.8, 70.0, 68.4, 66.0, 65.4, 64.3, 64.0, 63.7, 63.1, 63.8, 62.6, 57.7, 57.6, 57.5, 57.3, 54.6, 42.7, 42.0, 30.7, 29.0, 25.3, 25.0, 24.9, 24.8, 24.7; ESI-MS (negative mode): m/z calcd for $\text{C}_{103}\text{H}_{172}\text{N}_8\text{O}_{72}$; 2674.4930 found 1335.9948 ($M - 2\text{H}$)²⁻.

Synthesis of glycan G29.

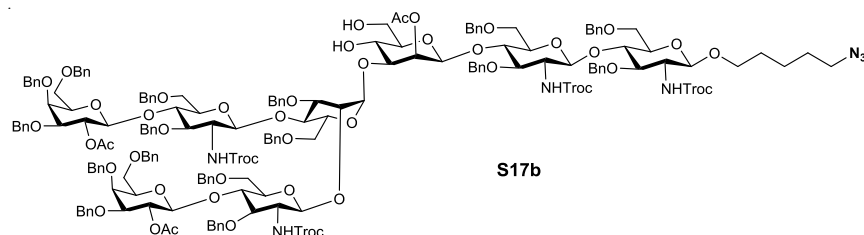
Glycan **G29** is mono sialylated tetraantennary complex type structure. The synthesis began with construction of D1 arm by glycosylation of donot **11** at 3-O position of **15** to afford compound **S17a**. The reaction was progress was checked by TLC as shown in Scheme S17. Removal benzylidene followed by 6-O glycosylation afforded desired tetra-decasaccharide **S17c**. Finally global deprotection provided desired glycan **G29**.



Scheme S17 | Preparation of G29. i, **11**, AgOTf, Cp_2HfCl_2 , toluene, $-40\text{ }^\circ\text{C}$, 77%; ii, *p*TSA, acetonitrile, 73%; iii, **13**, AgOTf, Cp_2HfCl_2 , toluene, $-40\text{ }^\circ\text{C}$, 57%; iv, (1) LiOH, 1,4-dioxane, $90\text{ }^\circ\text{C}$, (2) Ac_2O , pyridine, (3) NaOMe, MeOH, (4) $Pd(OH)_2$, MeOH:H₂O, H₂, 34%.

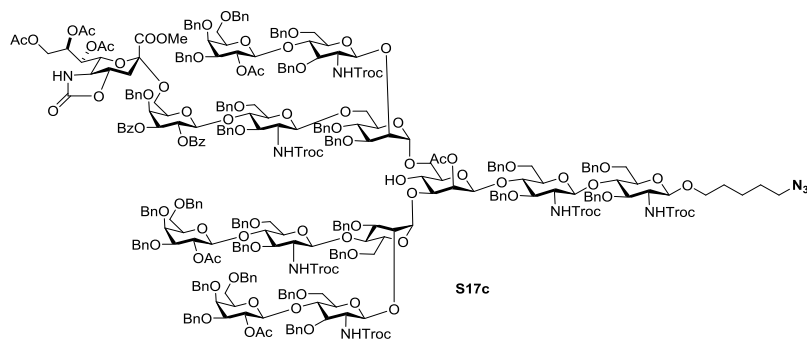


Compound S17a: A mixture of silver triflate (0.173 g, 0.675 mmol), bis (cyclopentadienyl) hafnium dichloride (0.177 g, 0.469 mmol) and 4 Å activated molecular sieves in dry toluene (10 mL) was stirred at room temperature for 1 h. The reaction mixture was then cooled to -40 °C, a solution of donor **11** (0.345 g, 0.470 mmol) and acceptor **15** (0.195 g, 0.134 mmol) in 5 mL toluene was added. The mixture was stirred for 2 h at -10 °C, quenched with Et₃N, diluted with EtOAc and filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), and a brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 10% acetone in toluene) to afford **S17a** (0.380 g, 77%) as white foam. TLC: (acetone: toluene = 1/9, v/v): R_f = 0.60; ¹H NMR (600 MHz, CDCl₃): δ 7.30-7.12 (m, 60H), 7.08-6.72 (m, 25H), 5.39 (t, *J* = 10.2 Hz, 1H), 5.29 (t, *J* = 10.3 Hz, 1H), 5.16 (d, *J* = 4.2 Hz, 1H), 5.09 (d, *J* = 8.4 Hz, 1H), 4.91-4.72 (m, 18H, overlapped), 4.63-4.56 (m, 5H), 4.01-4.80 (m, 30H), 3.80-3.95 (m, 10H), 3.60-3.75 (m, 4H), 3.52-3.48 (m, 6H), 3.41-3.15 (m, 20H), 2.98 (d, *J* = 9.6 Hz, 1H), 2.90-2.76 (m, 3H), 2.66-2.62 (m, 1H), 1.95 (s, 3H), 1.93 (s, 3H), 1.87 (s, 3H), 1.38-1.23 (m, 4H, -CCH₂C-), 1.08-1.01 (m, 2H, -CCH₂C-); ¹³C NMR (150 MHz, CDCl₃): δ 172.1, 171.9, 170.5, 156.7, 156.6, 142.0, 141.8, 141.6, 141.5, 141.4, 141.3, 141.3, 141.1, 140.8, 140.6, 140.5, 139.8, 135.2, 133.6, 133.2, 131.8, 131.5, 131.3, 131.1, 130.6, 130.5, 130.2, 130.1, 129.8, 129.0, 127.3, 104.8, 104.8, 103.2, 102.9, 77.7, 77.5, 77.3, 77.2, 77.1, 76.9, 76.2, 76.1, 76.0, 75.9, 75.5, 75.4, 74.8, 74.6, 74.5, 74.4, 74.3, 71.0, 70.8, 70.6, 54.0, 31.7, 31.3, 25.9, 23.9, 23.8, 23.7, 23.6, 17.2, 17.1, 16.8, 16.7, 13.7, 13.6; HRMS: *m/z* calcd for C₁₉₀H₂₀₅Cl₁₂N₇O₄₈; 3778.9981 found 3824.9969 (*M* + 2Na).



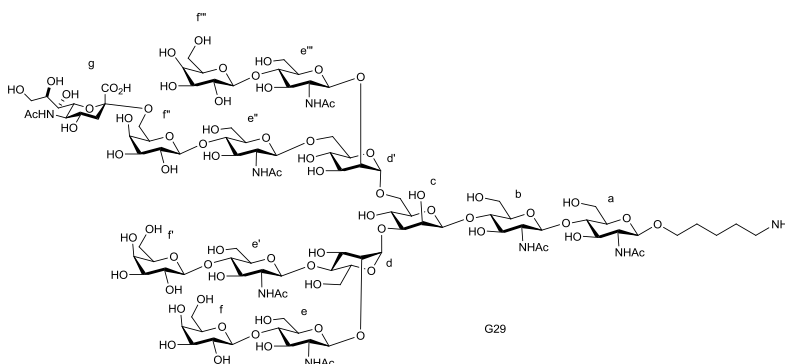
Compound S17b: *p*-Toluene sulfonic acid monohydrate (0.004 g, 0.02 mmol) was added to a solution of **S17a** (0.377 g, 0.102 mmol) in acetonitrile (10 mL) and the resulting reaction mixture

was stirred at rt for overnight. The reaction was quenched by adding Et₃N and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 10% acetone in toluene) to give diol **S17b** (0.270 g, 73%). TLC: (acetone: toluene = 1.5/8.5, v/v): R_f = 0.32; ¹H NMR (600 MHz, CDCl₃): δ 7.34-7.11 (m, 80H), 5.37 (s, 1H), 5.29 (t, *J* = 10.2 Hz, 1H), 5.22 (t, *J* = 9.8 Hz, 1H), 5.02 (d, *J* = 3.6 Hz, 2H), 4.92-4.74 (m, 7H), 4.70-4.21 (m, 34H), 3.93-3.51 (m, 17H), 3.65-3.20 (m, 34H), 3.20 (t, *J* = 10.2 Hz, 4H), 2.98 (d, *J* = 3.8 Hz, 2H), 2.89 (s, 1H), 2.11 (s, 3H), 1.94 (s, 3H), 1.90 (s, 3H), 1.40-1.17 (m, 4H), 0.91-0.84 (m, 2H); ¹³C NMR (150 MHz, CDCl₃): δ 170.1, 170.0, 169.6, 168.0, 154.3, 154.1, 139.6, 139.1, 139.0, 138.9, 138.7, 138.5, 138.3, 138.1, 138.0, 132.7, 131.1, 129.6, 129.4, 129.3, 129.0, 128.7, 128.6, 128.5, 128.4, 128.3, 128.2, 128.1, 128.0, 127.9, 127.6, 127.5, 127.3, 127.1, 127.0, 100.9, 100.6, 100.5, 98.6, 95.9, 95.8, 76.75, 76.3, 75.6, 74.8, 74.6, 74.5, 74.3, 74.2, 73.8, 73.7, 73.5, 73.2, 72.8, 72.5, 72.1, 69.6, 68.8, 68.5, 68.4, 68.2, 68.1, 62.8, 62.1, 57.6, 39.0, 37.3, 33.7, 32.5, 31.4, 30.6, 29.9, 29.2, 28.8, 26.6, 24.0, 23.4, 23.2, 22.9, 21.5, 21.3, 20.1, 15.5, 14.6, 14.4, 14.3, 11.2; HRMS: *m/z* calcd for C₁₈₃H₂₀₁Cl₁₂N₇O₄₈; 3692.0222 found 3737.9645 (*M* + 2Na).



Compound S17c: A mixture of silver triflate (0.046 g, 0.180 mmol), bis (cyclopentadienyl) hafnium dichloride (0.048 g, 0.126 mmol) and 4 Å activated molecular sieves in dry toluene (5 mL) was stirred at rt for 1 h. The reaction mixture was then cooled to -40 °C, a solution of donor **13** (0.100 g, 0.36 mmol) and acceptor **S17b** (0.118 g, 0.032 mmol) in 5 mL toluene was added. The mixture was stirred for 3 h at -10 °C, quenched with Et₃N, diluted with EtOAc and filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), and a brine (50 mL)

solution. The organic layer was dried over Na_2SO_4 and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 10% acetone in toluene) to afford **S17c** (0.120 g, 57%) as white foam. TLC: (acetone: toluene = 1/9, v/v): $R_f = 0.60$; ^1H NMR (600 MHz, CDCl_3): δ 7.58-7.54 (m, 6H), 7.50-6.98 (m, 134H), 5.57 (t, $J = 10.3$ Hz, 4H), 5.42-5.30 (m, 24H), 5.01-4.70 (m, 35H), 4.65-4.10 (m, 50H), 4.0-2.90 (m, 54H), 2.80 (d, $J = 9.8$ Hz, 2H), 2.15 (s, 3H), 2.09 (s, 3H), 2.07 (s, 3H), 1.96 (s, 3H), 1.93 (s, 3H), 1.88 (s, 3H), 1.86 (s, 3H), 1.34-1.17 (m, 5H), 0.89-0.81 (m, 2H); ^{13}C NMR (150 MHz, CDCl_3): δ 139.1, 139.0, 138.3, 138.2, 138.1, 130.1, 129.9, 129.1, 129.0, 128.9, 128.8, 128.7, 128.6, 128.4, 128.3, 128.1, 128.0, 127.9, 127.8, 127.6, 127.4, 74.8, 74.6, 73.7, 73.6, 71.9 ; HRMS: m/z calcd for $\text{C}_{322}\text{H}_{346}\text{Cl}_{18}\text{N}_{10}\text{O}_{89}$; 6418.3910 found 3253.3555 ($M + 2\text{Na}$) $^{2+}$.



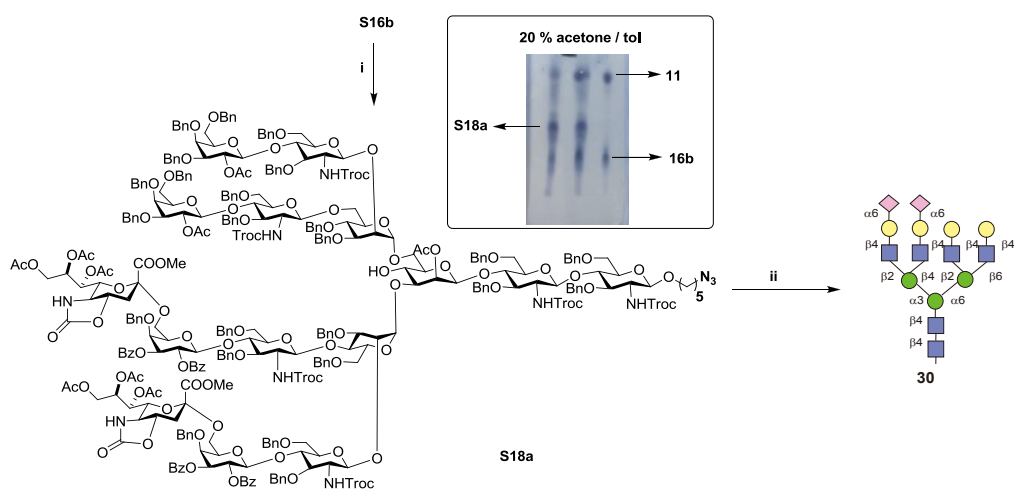
5-Aminopentyl-di-[β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 2),(1 \rightarrow 4)- α -D-mannopyranosyl]-(1 \rightarrow 3),-[5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate-(2 \rightarrow 6)- β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 6)- β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl]-(1 \rightarrow 6)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranoside

G29: Compound **S17c** (0.095 g, 0.015 mmol) was deprotected by following general procedure 2 (method 2) to afford desired glycan **G29** (0.014 g, 34%) as a white powder. ^1H NMR (600 MHz, D_2O): δ 4.99 (s, 1H, H-1 d), 4.75 (s, 1H, H-1 $^{d'}$), 4.72 (d, $J = 8.4$ Hz, 1H, H-1 e), 4.62 (d, 1H, $J = 7.8$ Hz, 1H, H-1 $^{e'}$), 4.63 (s, 1H, H-1 c), 4.45 (dd, $J = 3.6$ & 7.8 Hz, 4H, H-1 $^{e''}, e'''$, f, f'), 4.36-4.31 (m,

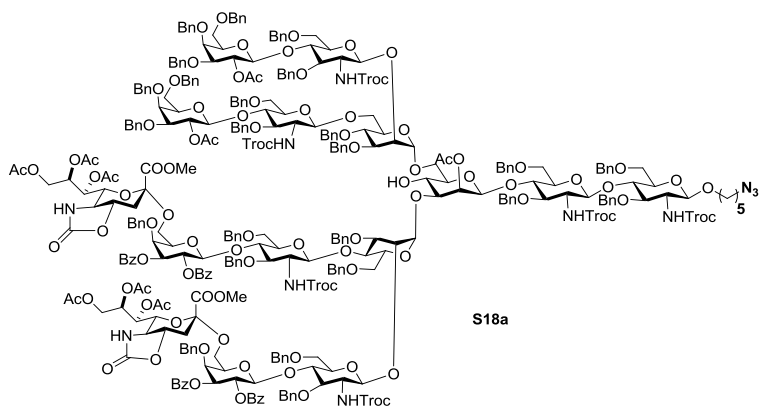
4H), 4.09 (s, 3H), 3.98-3.04 (m, 82H), 2.83 (t, $J = 10.2$ Hz, 2H, linker), 2.56 (dd, $J = 3.2$ & 8.4 Hz, 1H, H-3^{equi.g'}), 1.95 (s, 3H, -C(O)CH₃), 1.94 (s, 3H, -C(O)CH₃), 1.93 (s, 3H, -C(O)CH₃), 1.92 (s, 3H, -C(O)CH₃), 1.92 (s, 3H, -C(O)CH₃), 1.90 (s, 3H, -C(O)CH₃), 1.89 (s, 3H, -C(O)CH₃), 1.61-1.42 (m, 5H, H-3^{axial.g'} and linker CH₂), 1.30-1.24 (m, 2H, linker); ¹³C NMR (150 MHz, D₂O): δ 186.6, 174.7, 174.3, 174.2, 103.3, 102.7, 101.8, 100.8, 99.9, 98.5, 97.3, 78.9, 78.1, 77.8, 75.1, 74.4, 74.2, 74.1, 72.2, 71.7, 71.5, 70.7, 69.8, 68.3, 68.1, 63.1, 62.4, 60.8, 60.2, 59.7, 54.9, 54.6, 51.6, 39.9, 39.1, 27.8, 26.1, 22.2, 21.8, 17.6; ESI-MS (negative mode): m/z calcd for C₁₀₆H₁₇₈N₈O₇₄; 2747.0411 found 1396.0177 ($M + Na$)²⁻.

Synthesis of glycan G30.

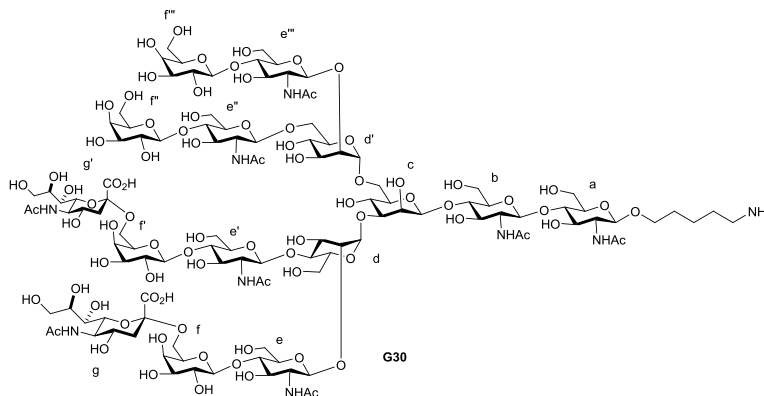
Depicted in Scheme S18, precursor S16b from scheme S16 was used as acceptor for 6-O glycosylation with donor **11** under the promotion of silver triflate and hafnocene dichloride to obtain the desired **S18a**. The final deprotection provided the disialylated tetra-antennary complex glycan G30.



Scheme S18 | Preparation of G30. i, **11**, AgOTf, Cp₂HfCl₂, toluene, -40 °C, 37%; ii, (1) LiOH, 1, 4-dioxane, 90 °C, (2) Ac₂O, pyridine, (3) NaOMe, MeOH, (4) Pd(OH)₂, MeOH:H₂O, H₂, 45%



Compound S18a: A mixture of silver triflate (0.051 g, 0.200 mmol), bis (cyclopentadienyl) hafnium dichloride (0.053 g, 0.140 mmol) and 4 Å activated molecular sieves in dry toluene (5 mL) was stirred at room temperature for 1 h. The reaction mixture was then cooled to $-50\text{ }^{\circ}\text{C}$, a solution of donor **11** (0.144 g, 0.060 mmol) and acceptor **S16b** (0.180 g, 0.040 mmol) in 5 mL toluene was added. The mixture was stirred for 3 h at $-10\text{ }^{\circ}\text{C}$, quenched with Et_3N , diluted with EtOAc and filtered through Celite. The filtrate was washed with aqueous NaHCO_3 (2 x 50 mL), and a brine (50 mL) solution. The organic layer was dried over Na_2SO_4 and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 10% acetone in toluene) to afford **S18a** (0.103 g, 37%) as white foam. TLC: (acetone: toluene = 2/8, v/v): $R_f = 0.53$; ^1H NMR (600 MHz, CDCl_3): δ 7.69-7.52 (m, 20H), 7.33-7.06 (m, 120H), 5.60 (m, 17H), 4.80-4.30 (m, 40H), 4.20-4.00 (m, 50H), 3.99-3.02 (m, 68H), 2.08 (s, 9H), 2.07 (s, 6H), 2.05 (s, 6H), 2.02 (s, 6H), 1.78-1.74 (m, 6H), 0.91-0.84 (m, 2H); ^{13}C NMR (150 MHz, CDCl_3): δ 170.9, 170.4, 170.2, 169.8, 169.4, 169.3, 132.7, 131.1, 129.0, 128.7, 128.4, 128.3, 128.2, 128.0, 127.8, 127.7, 127.6, 127.5, 68.8, 68.4, 39.0, 37.3, 33.7, 31.4, 30.6, 29.9, 29.2, 26.6, 26.0, 24.0, 23.2, 22.9, 20.1, 14.6, 14.2, 11.40; ESI-MS: m/z calcd for $\text{C}_{337}\text{H}_{361}\text{Cl}_{18}\text{N}_{11}\text{O}_{101}$; 6806.6710 found 723.5969 ($M + K$) $^{10+}$.

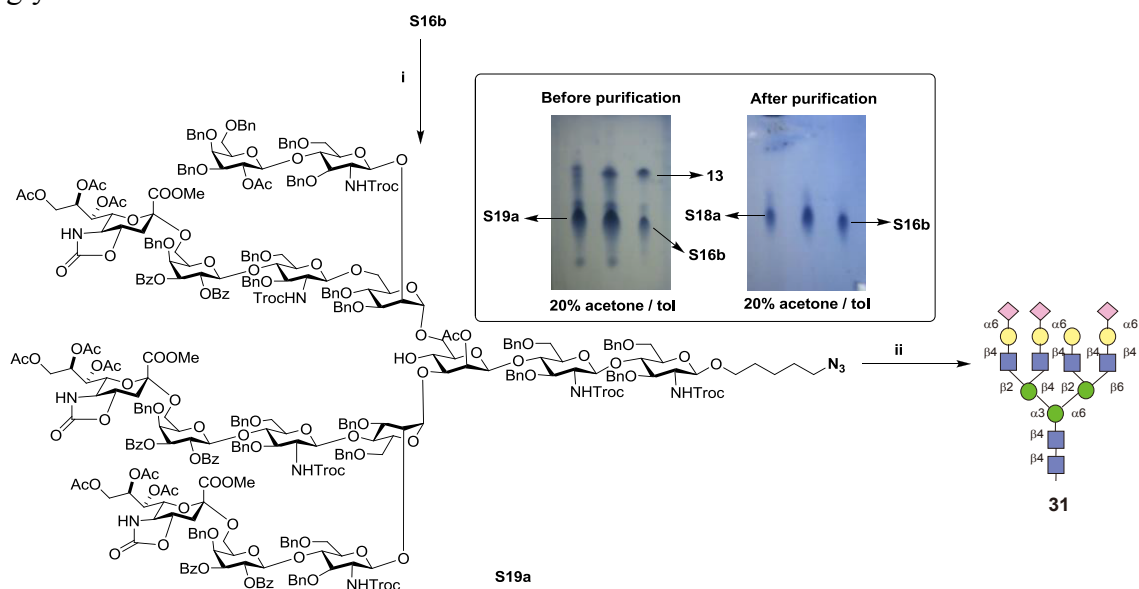


5-Aminopentyl-di-[5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate-(2 \rightarrow 6)- β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 2),(1 \rightarrow 4)- α -D-mannopyranosyl]-(1 \rightarrow 3),-di-[- β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 6),(1 \rightarrow 2)- α -D-mannopyranosyl]-(1 \rightarrow 6)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranoside **G30: Compound **S18a** (0.075 g, 0.010 mmol) was deprotected by following general procedure 2 (method 2) to afford desired glycan **G30** (0.015 mg, 45%). ^1H NMR (600 MHz, D_2O): δ 5.12 (s, 1H, H-1^d), 4.86-4.74 (m, 3H, H-1^{d',c,b}, overlapped with D_2O), 4.59-4.55 (m, 5H, H-1^{e,e',e'',e''',a}), 1.48-4.42 (m, 4H, H-1^{f,f',f'',f'''}), 4.21-4.06 (m, 5H), 3.98-3.3.20 (m, 86H), 2.98 (t, $J = 10.7$ Hz, 2H, linker CH_2), 2.66 (dd, $J = 3.7$ & 10.1 Hz, 2H, 2 x H-3_{equi.}^{g,g'}), 2.12 (s, 3H, -C(O)CH₃), 2.09 (s, 3H, -C(O)CH₃), 2.06 (s, 3H, -C(O)CH₃), 2.05 (s, 3H, -C(O)CH₃), 2.03 (s, 12H, -C(O)CH₃), 1.75-1.54 (m, 6H, 2 x H-3_{axial.}^{g,g'} and linker CH_2), 1.42-1.36 (m, 2H, linker); ^{13}C NMR (150 MHz, D_2O): δ 174.7, 174.2, 174.1, 103.4, 102.7, 101.3, 101.2, 101.0, 100.8, 100.6, 99.9, 98.9, 97.9, 78.1, 75.1, 74.3, 73.4, 72.3, 72.1, 71.7, 70.7, 70.4, 69.8, 68.3, 68.0, 59.9, 59.7, 54.7, 51.6, 39.9, 27.8, 22.7, 22.0, 21.9, 21.8; ESI-MS (negative mode): m/z calcd for $\text{C}_{117}\text{H}_{195}\text{N}_9\text{O}_{82}$; 3039.8280 found 1518.5585 ($M - \text{H}$)²⁻.**

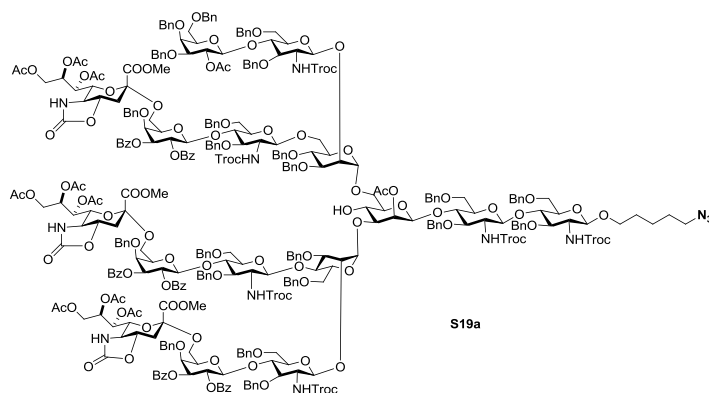
Synthesis of glycan **G31**.

Intermediate **S16b** derived from scheme S16 was used for α -selective glycosylation using donor **13** in presence of $\text{AgOTf}/\text{Cp}_2\text{HfCl}_2$ to get **S19a** in 74% yield. However, the newly formed

product was overlapped with acceptor on TLC (Scheme S19). S19a was then further deprotected to furnish glycan **G31**.

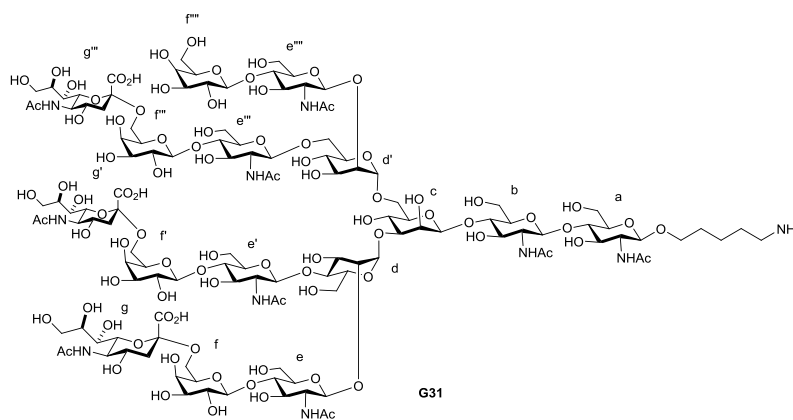


Scheme S19 | Preparation of G31. i, **13**, AgOTf, Cp₂HfCl₂, toluene, -50 °C, 74%; ii, (1) LiOH, 1,4-dioxane, 90 °C, (2) Ac₂O, pyridine, (3) NaOMe, MeOH, (4) Pd(OH)₂, MeOH: H₂O, H₂, 47%.



Compound S19a: A mixture of silver triflate (0.042 g, 0.165 mmol), bis (cyclopentadienyl) hafnium dichloride (0.043 g, 0.115 mmol) and 4 Å activated molecular sieves in dry toluene (5 mL) was stirred at rt for 1 h. The reaction mixture was then cooled to -50 °C, a solution of donor **13** (0.100 g, 0.036 mmol) and acceptor **S16b** (0.150 g, 0.033 mmol) in 5 mL toluene was added. The reaction mixture was stirred for 2 h at -20 °C, quenched with Et₃N, diluted with EtOAc and

filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), and a brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 15% acetone in toluene) to afford **S19a** (0.183 g, 74%) as white foam. TLC: (acetone: toluene = 2/8, v/v): R_f = 0.50; ¹H NMR (600 MHz, CDCl₃): δ 7.90-7.83 (m, 9H), 7.69-7.67 (m, 3H), 7.57-7.50 (m, 3H), 7.38-7.04 (m, 125H), 5.77 (t, *J* = 10.2 Hz, 4H), 5.54-5.40 (m, 20H), 5.39 (t, *J* = 8.4 Hz, 4H), 5.08 (t, *J* = 7.8 Hz, 4H), 4.98-4.60 (m, 25H), 4.53-4.10 (m, 30H), 4.00-3.80 (m, 42H), 3.21-3.18 (m, 44H), 2.93-2.78 (m, 4H), 2.08 (s, 6H), 2.07 (s, 6H), 2.05 (s, 3H), 2.02 (s, 3H), 1.98 (s, 6H), 1.94 (s, 6H), 1.74-1.18 (m, 7H), 0.91-0.84 (m, 2H); ¹³C NMR (150 MHz, CDCl₃): δ 171.8, 171.7, 170.8, 169.9, 169.8, 168.5, 168.0, 166.1, 165.4, 159.5, 132.7, 131.7, 130.1, 129.9, 129.2, 128.9, 128.8, 128.7, 128.6, 128.4, 128.3, 128.2, 128.1, 128.0, 127.9, 127.8, 127.7, 127.6, 127.5, 100.7, 100.6, 100.4, 95.8, 74.9, 74.6, 74.2, 73.9, 73.7, 73.6, 73.2, 72.9, 72.2, 71.9, 71.0, 70.5, 69.3, 68.5, 68.3, 68.0, 67.3, 66.1, 65.3, 62.3, 61.2, 53.4, 51.5, 39.0, 37.3, 33.7, 32.4, 32.2, 30.6, 29.9, 29.6, 29.4, 29.3, 28.8, 26.6, 24.0, 23.9, 23.5, 23.4, 23.1, 21.2, 20.1, 15.5, 14.6, 14.4, 14.3, 11.2; HRMS: *m/z* calcd for C₃₅₂H₃₇₆Cl₁₈N₁₂O₁₁₃; 7220.9510 found 3654.9220 (*M* + 2Na)²⁺.



5-Aminopentyl-di-[5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate-(2→6)- β -D-galactopyranosyl-(1→4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1→2),(1→4)- α -D-mannopyranosyl]-(1→3),-[5-Acetamido-3,5-dideoxy-D-glycero- α -D-

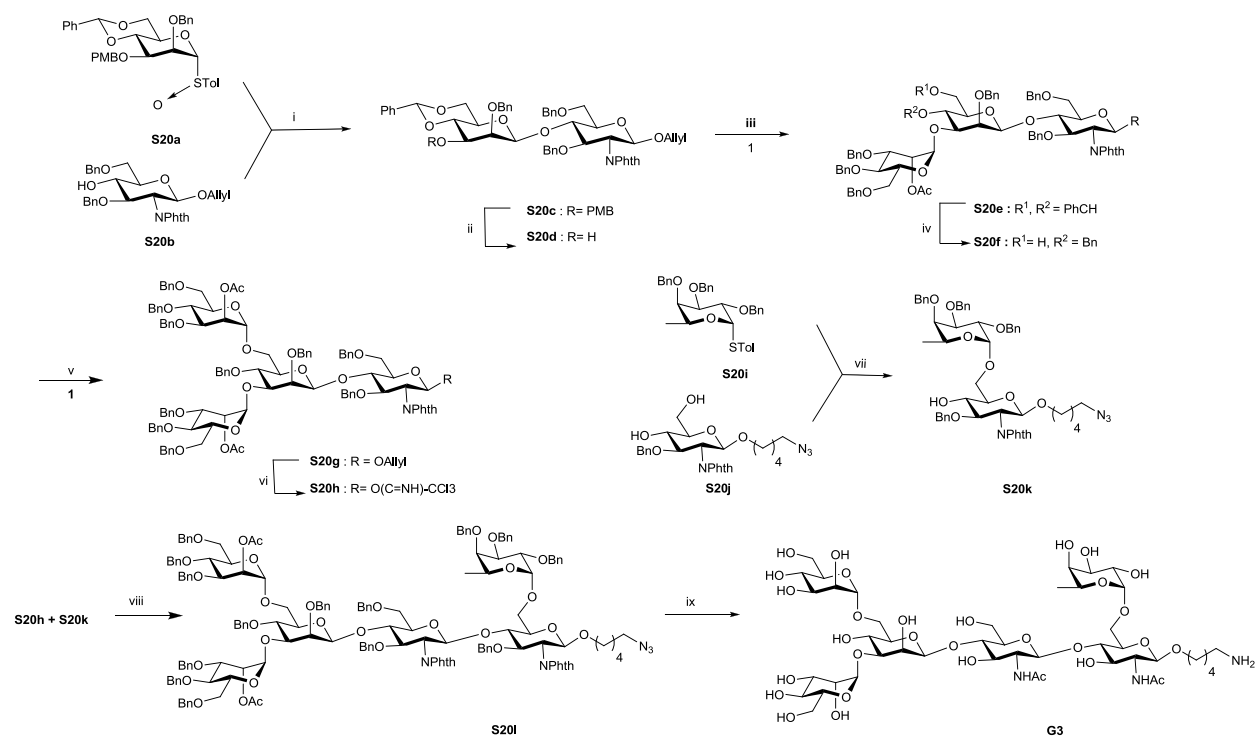
galacto-2-nonulopyrano-sylionate-(2→6)-β-D-galactopyranosyl-(1→4)-2-acetamido-2-deoxy-β-D-glucopyranosyl-(1→6)- β-D-galactopyranosyl-(1→4)-2-acetamido-2-deoxy-β-D-glucopyranosyl-(1→2)-α-D-mannopyranosyl-(1→6)-β-D-mannopyranosyl-(1→4)-2-acetamido-2-deoxy-β-D-glucopyranosyl-(1→4)-2-acetamido-2-deoxy-β-D-glucopyranoside

G31: Compound **S19a** (0.180 g, 0.024 mmol) was deprotected by following general procedure 2 (method 2) to afford desired glycan **G31** (0.039 g, 47%). ¹H NMR (600 MHz, D₂O): δ 5.12 (s, 1H, H-1^d), 4.87 (s, 2H, H-1^{d',c}), 4.75 (d, *J* = 8.4 Hz, 2H, H-1^{b, e}), 5.59-4.55 (m, 5H, overlapped H-1^{a, c', e'', e''', f}), 4.49-4.42 (m, 5H, overlapped H-1^{f', f'''}), 4.21-4.19 (m, 4H), 4.06-3.54 (m, 83H), 2.97 (t, *J* = 10.2 Hz, 2H, linker -CH₂), 2.66 (dd, *J* = 3.2 & 7.8 Hz, 3H, H-3^{g, g', g'' equi}), 2.09 (s, 3H), 2.06 (s, 9H), 2.04 (s, 3H), 2.02 (s, 12H), 1.74 (m, 2H, linker), 1.64 (t, *J* = 10.2 Hz, 3H, H-3^{g, g', g'' axial}), 1.59-1.57 (m, 2H, linker), 1.39-1.37 (m, 2H, linker); ¹³C NMR (150 MHz, D₂O): δ 174.8, 174.7, 174.5, 174.4, 174.3, 174.1, 173.3, 103.5, 102.9, 101.7, 101.4, 101.2, 100.2, 100.0, 99.8, 99.6, 99.3, 80.8, 80.6, 80.3, 79.3, 78.4, 78.3, 76.7, 75.3, 74.3, 74.2, 74.1, 74.0, 73.8, 72.9, 72.8, 72.6, 72.4, 72.3, 72.2, 72.1, 72.0, 71.9, 71.7, 71.3, 70.8, 70.4, 70.3, 70.2, 68.4, 68.4, 63.4, 62.9, 60.4, 60.3, 60.2, 59.8, 54.8, 54.3, 54.2, 51.8, 51.0, 43.7, 43.3, 39.8, 39.5, 34.4, 27.5, 26.3, 22.6, 22.4, 22.4, 22.2, 22.0, 21.9 ; ESI-MS (negative mode): *m/z* calcd for C₁₂₈H₂₁₂N₁₀O₉₀; 3331.0840 found 1664.1108 (*M* - H)²⁻.

viii. Synthesis of core fucosylated glycans G3 and G19

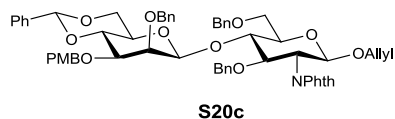
The pauci mannose type structure bearing core fucose **G3**¹² was assembled from the reducing end disaccharide **S20k** through fucosylation of diol acceptor **S20j** by donor **S20i** (Scheme S20). The α-selectivity was enhanced by in situ anomarization protocol (CuBr₂, tetrabutyl ammonium bromide), which was first developed by Lemieux and co-workers¹⁴, to get disaccharide **S20k** in 67% yield with its regio- and stereo-chemistry confirmed by NMR analysis. The formation of the key β-mannosyl linkage using sulfoxide donor **S20a**¹³ and acceptor **S20b** was performed using DTBP and Tf₂O to get disaccharide **S20c** in 67% yield with α/β ratio of 1:7. The *p*-methoxy benzyl ether protection at 3'' was removed by using DDQ to afford 3''-OH **S20d**, which was further subjected to 3''-O α-mannosylation by using imidate **1** under the promotion of BF₃.OEt₂

to obtain trisaccharide **S20e**. Selective benzylidene opening of **S20e** produced **S20f**, which further underwent α -mannosylation at 6''-O position to get tetrasaccharide **S20g**. Compound **S20g** was then converted into glycosyl imidate **S20h** by treatment with PdCl₂ and then trichloroacetimidate and DBU. Finally the condensation of imidate **S20h** and disaccharide **S20k**¹² in the presence of BF₃.OEt₂ provided desired hexasaccharide **S20i**. The global deprotection afforded the paucimannose type oligosaccharide **G3**.



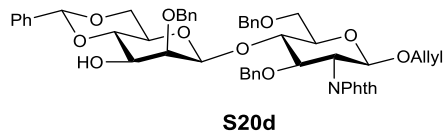
Scheme S20 | Preparation of glycan G3. i, DTBP, Tf₂O, CH₂Cl₂, -60 °C, 3 h, 67% (α/β 1:7); ii, DDQ, CH₂Cl₂ : H₂O = 10/1, 3 h, 70%; iii, **1**, BF₃.OEt₂, CH₂Cl₂, 4 Å MS, -40 °C, 1 h, 81%; iv, Triethyl silane, PhBCl₂, CH₂Cl₂, 1 h, 73%; v, BF₃.OEt₂, CH₂Cl₂, 4 Å MS, -20 °C, 2 h, 65%; vi, (i) PdCl₂, MeOH:CH₂Cl₂, RT, 6 h, (ii) DBU, trichloroacetonitrile, CH₂Cl₂, 63% over 2 steps; vii, Cu(II)Br, TBAB, DMF:CH₂Cl₂, 4 Å MS, 0 °C to RT, overnight, 76%; viii, BF₃.OEt₂, CH₂Cl₂, 4 Å MS, -70 °C, 2 h, 45%; ix, (i) NH₂CH₂CH₂NH₂, *n*BuOH, 100 °C; (ii) Ac₂O, pyridine, 0 °C to RT;

(iii) NaOMe, MeOH; (iv) Pd(OH)₂, MeOH: H₂O: HCOOH (5:3:2), H₂, 22% over 4 steps. TBAB = Tetrabutyl ammonium bromide.

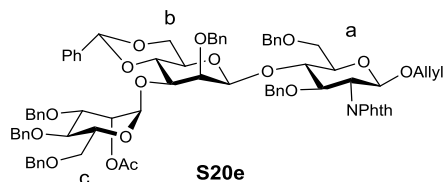


Allyl-O-2-O-benzyl-3-O-*p*-methoxy-benzyl-4,6-O-benzylidene-β-D-mannopyranosyl-(1→4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranoside (S20c): A mixture of donor **S20a** (0.272 g, 0.483 mmol) and activated 4 Å molecular sieves in CH₂Cl₂ (10 mL) was stirred at rt for 1 h. The reaction mixture was cooled to -60 °C, 2,6-di-*tert*-butyl pyridine (268 μL, 1.24 mmol) followed by Tf₂O (79 μL, 0.483 mmol) was added and stirred for 40 minutes. A solution of acceptor **S20b** (0.2 g, 0.378 mmol) in CH₂Cl₂ (5 mL) was added slowly and the resulting reaction mixture was stirred for 1 h until TLC (ethyl acetate: toluene, 1/9) indicated formation of a product with consumption of the starting material. The reaction mixture was quenched with Et₃N, diluted with CH₂Cl₂, filtered through Celite and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 10% EA in toluene) to afford **S20c** (0.230 g, 67%) as white solid. TLC: (ethyl acetate: toluene = 1/9, v/v): R_f = 0.52; ¹H NMR (600 MHz, CDCl₃): δ 7.65-7.45 (m, 4H, Ar-H), 7.35-7.21 (m, 13H, Ar-H), 6.91-6.90 (m, 2H, Ar-H), 6.90-6.81 (m, 10H, Ar-H), 5.67-5.65 (m, 1H, allyl), 5.49 (s, 1H, Ph-CH, benzylidene), 5.13 (d, *J* = 8.4 Hz, 1H, H-1^a), 5.07 (d, *J* = 8.8 Hz, 1H), 4.99 (d, *J* = 10.4 Hz, 1H), 4.87-4.82 (m, 3H), 4.65 (d, *J* = 5.4 Hz, 1H), 4.63 (d, *J* = 5.4 Hz, 1H), 4.53 (s, 1H), 4.50 (d, *J* = 12.0 Hz, 1H), 4.41 (d, *J* = 6.0 Hz, 1H), 4.39 (d, *J* = 6.0 Hz, 1H), 4.24-4.21 (m, 2H), 4.05-3.96 (m, 3H), 3.81 (s, 3H, OMe of PMB), 3.71 (d, *J* = 3.0 Hz, 1H), 3.64 (d, *J* = 10.8 Hz, 1H), 3.56-3.52 (m, 2H), 3.45-3.47 (d, *J* = 10.0 Hz, 1H), 3.40 (dd, *J* = 3.0, 6.6 Hz, 1H), 3.13-3.12 (m, 1H); ¹³C NMR (150 MHz, CDCl₃): δ 159.46, 139.08, 138.99, 138.12, 138.00, 134.05, 130.93, 129.37, 129.22, 128.84, 128.60, 128.54, 128.47, 128.45, 128.26, 128.12, 128.09, 128.07, 127.83, 127.21, 126.41, 117.60, 114.04, 102.28,

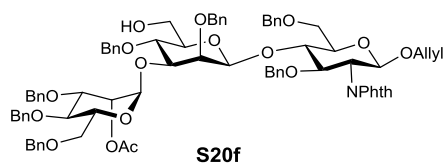
101.63, 97.70, 79.85, 78.99, 78.33, 77.44, 75.29, 75.03, 74.95, 73.86, 72.61, 69.99, 68.86, 68.84, 67.66, 55.96, 55.58; ESI-MS: m/z calcd for $C_{59}H_{59}NO_{13}$: 989.3879; found 1012.3888 ($M + Na$)⁺.



Allyl-O-2-O-benzyl-4,6-O-benzylidene-β-D-mannopyranosyl-(1→4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido-β-D-glucopyranoside (S20d): To a solution of **S20c** (0.5 g, 0.505 mmol) in 10 mL $CH_2Cl_2 : H_2O$ (10:1) was added DDQ (0.219g, 1.01 mmol) at 0 °C and the resulting reaction mixture was stirred for 3 h. The reaction mixture was then filtered, organic layer washed with H_2O (2 x 30 mL). The aqueous layer was further extracted with CH_2Cl_2 (2 x 50 mL). The combined organic layers were washed with brine solution (40 mL), dried over Na_2SO_4 and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 10% EA in toluene) to afford **S20d** (0.308 g, 70%) as colorless foam. TLC: (ethyl acetate: toluene = 2/8, v/v): R_f = 0.42; 1H NMR (600 MHz, $CDCl_3$): δ 7.65-7.44 (m, 4H, Ar-H), 7.43-7.28 (m, 13H, Ar-H), 6.92-6.90 (m, 2H, Ar-H), 6.83-6.81 (m, 5H, Ar-H), 5.68-5.64 (m, 1H, Allyl), 5.42 (s, 1H, Ph-CH, benzylidene), 5.14 (d, J = 8.4 Hz, 1H, H-1^a), 5.08 (d, J = 12.8 Hz, 1H), 5.01-4.98 (m, 2H), 4.85 (d, J = 12.6 Hz, 1H), 4.74 (d, J = 12.0 Hz, 1H), 4.65 (d, J = 12.0 Hz, 1H), 4.62 (s, 1H, H-1^b), 4.84 (d, J = 12.0 Hz, 1H), 4.40 (d, J = 12.0 Hz, 1H), 4.25-4.16 (t, J = 9.6 Hz, 1H), 3.99 (dd, J = 13.0 & 6.3 Hz, 1H), 3.73-3.65 (m, 4H), 3.54-3.47 (m, 3H), 3.13-3.09 (m, 1H); ^{13}C NMR (150 MHz, $CDCl_3$): 138.97, 138.44, 137.93, 137.56, 133.98, 129.37, 128.92, 128.79, 128.53, 128.43, 128.35, 128.26, 128.21, 128.10, 127.99, 127.27, 126.58, 117.64, 102.33, 102.25, 97.66, 79.91, 79.42, 79.15, 77.43, 76.00, 75.03, 74.89, 74.01, 71.24, 69.99, 68.78, 68.67, 67.17, 55.90; ESI-MS: m/z calcd for $C_{51}H_{51}NO_{12}$: 869.3303; found 892.3314 ($M + Na$)⁺.

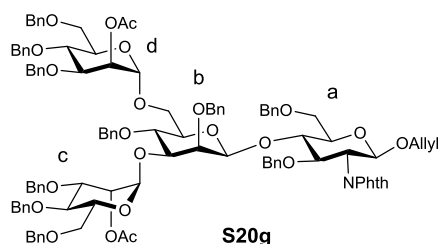


Allyl-O-(2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3)-2-O-benzyl-4,6-O-benzylidene- β -D-mannopyranosyl-(1 \rightarrow 4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranoside (S20e):- A mixture of donor **1** (0.537 g, 0.863 mmol), acceptor **S20d** (0.50 g, 0.574 mmol) and activated 4 Å molecular sieves in dry CH₂Cl₂ (10 mL) was stirred at rt for 30 minutes. The reaction was cooled to -40 °C, boron trifluoride ethyl etherate (32 μ L, 0.288 mmol) was then added slowly and the resulting reaction mixture was stirred for 1 h. The reaction was quenched by adding Et₃N, diluted with CH₂Cl₂, filtered through Celite and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 20% EA in hexane) to afford **S20e** (0.630 g, 81%) as colorless oil. TLC: (ethyl acetate: hexane = 3/7, v/v): R_f = 0.62; ¹H NMR (600 MHz, CDCl₃): δ 7.41-7.39 (m, 4H, Ar-H), 7.39-7.25 (m, 30H, Ar-H), 7.16-6.82 (m, 5H, Ar-H), 5.57-5.57 (m, 1H, Allyl), 5.59 (d, *J* = 3.0 Hz, 1H), 5.47 (s, 1H, Ph-CH₂, benzylidene), 5.29 (d, *J* = 2.1 Hz, 1H, H-1^c), 5.13 (d, *J* = 8.4 Hz, 1H, H-1^a), 5.08 (dd, *J* = 6.0 & 12.2 Hz, 1H), 5.00 (dd, *J* = 6.0, 10.2 Hz, 1H), 4.87-4.82 (m, 2H), 4.79-4.78 (m, 2H), 4.66-4.59 (m, 3H), 4.54 (s, 1H, H-1^b), 4.46-4.44 (m, 3H), 4.22-4.20 (m, 2H), 4.17-4.14 (m, 2H), 4.03-3.93 (m, 4H), 3.82-3.71 (m, 5H), 3.64-3.62 (m, 2H), 3.60-3.52 (m, 2H), 3.50 (d, *J* = 8.2 Hz, 1H), 3.19 (m, 1H), 2.34 (s, 3H, -C(O)CH₃); ¹³C NMR (150 MHz, CDCl₃): δ 170.26, 163.58, 162.83, 138.98, 138.77, 138.60, 138.40, 138.09, 138.04, 137.58, 133.97, 131.97, 129.01, 128.79, 128.71, 128.64, 128.62, 128.56, 128.52, 128.38, 128.32, 128.24, 128.12, 128.05, 127.97, 127.94, 127.84, 127.18, 126.26, 123.57, 117.58, 101.76, 101.35, 98.96, 97.63, 79.24, 78.91, 78.56, 78.22, 75.76, 75.62, 75.03, 74.77, 74.44, 73.75, 72.55, 71.83, 69.94, 69.32, 68.64, 68.54, 68.39, 67.16, 55.88, 36.75, 31.70, 21.27; ESI-MS: *m/z* calcd for C₈₀H₈₁NO₁₈: 1343.5346; found 1366.5369 (*M* + Na)⁺.



Ally-O-(2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3)-2,4-O-di-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranoside

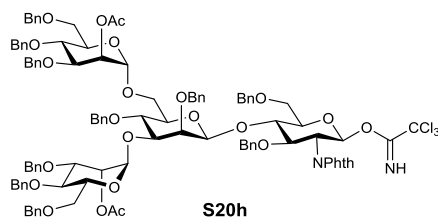
(S20f): A mixture of compound **S20e** (0.630 g, 0.468 mmol) and activated 4 Å molecular sieves in 20 mL CH₂Cl₂ was stirred for 1 h at rt. The reaction mixture was cooled to -78 °C, triethyl silane (148 μL, 0.946 mmol) followed by dichlorophenyl borane (146 μL, 1.18mmol) was added and stirred for 1 h. The reaction was quenched by Et₃N, methanol, filtered through Celite and concentrated in *vacuo*. The residue was co-distilled with methanol 2-3 times before being purified by flash column chromatography (0% → 10% EA in toluene) to give alcohol **S20f** (0.460 g, 73%). TLC: (ethyl acetate: toluene = 2/8, v/v): R_f = 0.42; ¹H NMR (600 MHz, CDCl₃): δ 7.73-7.57 (m, 4H, Ar-H), 7.50-7.08 (m, 31H, Ar-H), 6.83-6.81 (m, 4H, Ar-H), 5.76-5.69 (m, 1H, Allyl), 5.44 (t, *J* = 3.0 Hz, 1H), 5.13 (s, 1H, H-1^c), 5.12 (s, 1H), 5.08 (d, *J* = 7.8Hz, 1H, H-1^a), 5.01 (d, *J* = 11Hz, 1H), 4.93 (d, *J* = 11.0 Hz, 1H), 4.87 (d, *J* = 8.6 Hz, 1H), 4.83 (d, *J* = 8.0 Hz, 1H), 4.76 (d, *J* = 12.0 Hz, 1H), 4.69 (d, *J* = 11.0 Hz, 1H), 4.62 (t, *J* = 8.6 Hz, 1H), 4.57 (t, *J* = 7.8 Hz, 2H), 4.50 (d, *J* = 10.8 Hz, 1H), 4.44 (d, *J* = 2.5 Hz, 1H), 4.42 (s, 1H), 4.40 (d, *J* = 8.0 Hz, 1H), 4.35 (d, *J* = 12.0 Hz, 1H), 4.22-4.181 (m, 3H), 4.00-3.99 (m, 2H), 4.90-3.91 (s, 1H), 3.83-3.82 (m, 1H), 3.83 (d, *J* = 2.4 Hz, 1H), 3.77-3.78 (m, 2H), 3.64-3.54 (m, 6H), 3.41-3.39 (m, 2H), 3.19-3.18 (m, 1H), 2.08 (s, 3H); ¹³C NMR (150 MHz, CDCl₃): δ 170.34, 138.90, 138.77, 138.73, 138.36, 138.14, 138.07, 138.03, 135.91, 134.50, 134.03, 133.86, 132.97, 131.94, 131.40, 129.31, 128.86, 128.68, 128.66, 128.61, 128.57, 128.50, 128.45, 128.38, 128.26, 128.20, 128.12, 128.11, 128.03, 127.97, 127.94, 127.79, 127.76, 127.72, 127.47, 127.35, 123.50, 117.59, 101.04, 99.89, 97.61, 80.92, 78.64, 78.48, 78.41, 78.21, 75.93, 75.26, 75.20, 75.19, 75.16, 74.82, 74.59, 74.52, 73.78, 73.75, 72.61, 72.11, 69.91, 69.50, 68.99, 68.43, 62.25, 55.86, 21.29; ESI-MS: *m/z* calcd for C₈₀H₈₃NO₁₈: 1345.5502; found 1368.5527 (*M* + Na)⁺.



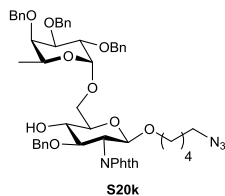
Allyl-O-di-(2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3)),(1 \rightarrow 6)-2,4-O-di-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-

glucopyranoside (S20g): A mixture of donor **1** (0.429 g, 0.690 mmol), acceptor **S20f** (0.460 g, 0.345 mmol) and activated 4 Å molecular sieves in dry CH₂Cl₂ (10 mL) was stirred at rt for 30 minutes. The reaction was cooled to -20 °C, boron trifluoride ethyl etherate (19 μ L, 0.173 mmol) was then added slowly and the resulting reaction mixture was stirred for 2 h. The reaction was quenched by adding Et₃N, diluted with CH₂Cl₂, filtered through Celite and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 20% EA in hexane) to afford **S57** (0.380 g, 65%) as colorless foam. TLC: (ethyl acetate: hexane = 3/7, v/v): R_f = 0.56; ¹H NMR (600 MHz, CDCl₃): δ 7.60-7.29 (m, 4H, Ar-H), 7.28-7.08 (m, 37H, AR-H), 7.07-7.08 (m, 8H, Ar-H), 7.07-6.93 (d, *J* = 8.0 Hz, 1H, Ar-H), 6.82-6.59 (m, 4H, Ar-H), 5.61-5.59 (m, 1H, Allyl), 5.43 (d, *J* = 6.0 Hz, 1H, H-2^c), 5.31 (d, *J* = 6.0 Hz, 1H, H-2^d), 5.10 (d, *J* = 3.0 Hz, 1H, H-1^c), 5.06 (d, *J* = 8.0 Hz, 1H, H-1^a), 4.99 (d, *J* = 10.0 Hz, 1H), 4.85 (d, *J* = 8.0 Hz, 1H), 4.83 (s, 1H, H-1^d), 4.67 (d, *J* = 8.0 Hz, 1H), 4.64-4.62 (q, 4H), 4.56 (d, *J* = 3.0 Hz, 1H, H-1^b), 4.55-4.45 (m, 8H), 4.45-4.39 (m, 9H), 4.38 (d, *J* = 10.0 Hz, 1H), 4.21 (d, *J* = 9.0 Hz, 1H), 4.19-4.10 (m, 2H), 4.08-3.70 (m, 10H), 3.69-3.49 (m, 7H), 3.39 (d, *J* = 9.1 Hz, 1H), 3.19 (d, *J* = 9.1 Hz, 1H), 2.07 (s, 3H, -C(O)CH₃), 1.82 (s, 3H, -C(O)CH₃); ¹³C NMR (150 MHz, CDCl₃): δ 170.31, 170.11, 163.43, 139.07, 138.93, 138.83, 138.80, 138.65, 138.35, 138.32, 138.21, 138.09, 134.08, 133.77, 132.00, 128.87, 128.76, 128.66, 128.56, 128.43, 128.37, 128.16, 127.99, 127.91, 127.82, 127.73, 127.13, 123.46, 117.45, 102.07, 99.92, 99.74, 98.62, 97.55, 81.40, 79.81, 78.31, 78.23, 76.84,

75.87, 75.77, 75.66, 74.87, 74.65, 74.44, 74.38, 31.21, 21.29, 21.23; ESI-MS: m/z calcd for $C_{109}H_{113}NO_{24}$: 1820.7578; found 1843.7676 ($M + Na$)⁺.

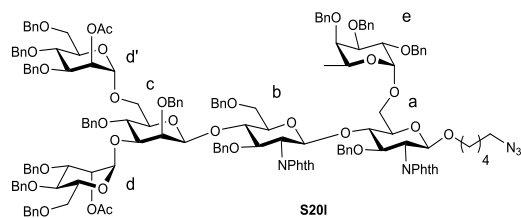


Di-(2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3)),(1 \rightarrow 6)-2,4-O-di-benzyl- β -D-mannopyranosyl-(1 \rightarrow 4)-O-(3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl trichloroacetimidate (S20h): To a solution of **S20g** (0.2 g, 0.109 mmol) in 10 mL CH_2Cl_2 :MeOH (1/1) was added $PdCl_2$ (20 mg) and stirred at room temperature for 6 h until TLC (ethyl acetate: toluene, 2/8) indicated formation of a product with consumption of the starting material. The reaction mixture was then concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 10% EA in toluene) to afford alcohol (0.140 g, 60%) as colorless foam. To a solution of alcohol (0.150 g, 0.084 mmol) in CH_2Cl_2 (10 mL) was added trichloroacetimidate (33 μ L, 0.336 mmol) followed by DBU (12 μ L, 0.033 mmol) at 0 $^\circ$ C and stirred at rt for 8 h. The reaction was quenched with Et_3N and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 10% EA in toluene) to afford **S20h** (0.121 g, 67%) as colorless oil. TLC: (ethyl acetate: toluene = 2/8, v/v): R_f = 0.47; 1H NMR (600 MHz, $CDCl_3$): δ 8.51 (s, 1H, C=NH), 7.53-7.42 (m, 4H, Ar-H), 7.40-7.00 (m, 48H, Ar-H), 6.58 (d, J = 6.6 Hz, 2H, Ar-H), 6.52-6.50 (m, 3H, Ar-H), 5.43 (s, 1H, H-1^a), 5.32 (s, 1H, H-1^c), 5.03 (s, 1H), 5.01 (t, J = 7.8 Hz, 1H), 4.90-4.78 (m, 6H), 4.67-4.45 (m, 7H), 4.43-4.30 (m, 9H), 4.28-4.20 (m, 2H), 4.10 (t, J = 7.8 Hz, 1H), 3.39-3.42 (m, 18H), 3.19 (d, J = 9.3 Hz, 1H), 2.09 (s, 3H, -C(O)CH₃), 1.83 (s, 3H, -C(O)CH₃).



5-Azidopentyl-O-2,3,4-tri-O-benzyl- α -L-fucopyranosyl-(1 \rightarrow 6)-3-O-benzyl-2-deoxy-2-

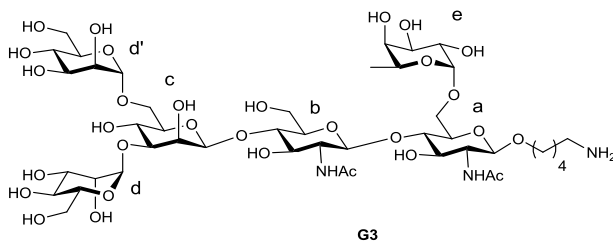
phthalimido- β -D-glucopyranoside (S20k): A mixture of CuBr₂ (0.450 g, 2.01 mmol), Bu₄NBr (0.655 g, 2.03 mmol) and activated 4 Å molecular sieves in 10 mL of CH₂Cl₂ : DMF (2/1) was stirred and cooled over an ice water bath. A solution of donor **S20i** (0.5 g, 0.925 mmol) and acceptor **S20j** (0.450 g, 0.882 mmol) in CH₂Cl₂ (10 mL) was added drop wise and the resulting reaction mixture was stirred at rt for overnight. TLC (ethyl acetate: hexane, 4/6) indicated formation of a product with consumption of the starting material. The reaction was quenched with aq. NaHCO₃, diluted with ethyl acetate (50 mL) and filtered through Celite. The filtrate was washed with aq. NaHCO₃, brine (50 mL), dried over sodium sulfate. The organic layer was concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 30% EA in hexane) to afford **S20k** (0.604 g, 76%) as colorless oil. TLC: (ethyl acetate: hexane = 4/6, v/v): R_f = 0.47; ¹H NMR (600 MHz, CDCl₃): δ 7.80-7.64 (m, 4H, Ar-H), 7.41-7.23 (m, 15H, Ar-H), 7.00-6.98 (m, 2H, Ar-H), 6.90-6.87 (m, 3H, Ar-H), 5.06 (d, *J* = 8.4 Hz, 1H, H-1^a), 4.96 (d, *J* = 11.4 Hz, 1H), 4.86 (d, *J* = 11.4 Hz, 1H), 4.82-4.72 (m, 4H), 4.67 (d, *J* = 12.0 Hz, 1H), 4.64 (d, *J* = 12.0 Hz, 1H), 4.48 (d, *J* = 12.6 Hz, 1H), 4.16-4.13 (m, 1H), 4.08-4.05 (m, 2H), 3.39-3.95 (m, 2H), 3.90-3.83 (m, 3H), 3.78 (m, 1H), 3.73-3.69 (m, 2H), 3.56-3.53 (m, 1H), 3.35-3.21 (m, 1H), 2.00-2.03 (m, 2H, -CCH₂C-, linker), 1.38-1.30 (m, 4H, -CCH₂C-, linker), 1.12 (d, *J* = 8.4 Hz, 3H, Me of Fucose), 1.00-0.96 (m, 2H, -CCH₂C-, linker); ¹³C NMR (150, MHz, CDCl₃): δ 138.83, 138.77, 138.36, 134.13, 128.90, 128.77, 128.67, 128.60, 128.52, 128.31, 128.26, 128.17, 127.93, 127.88, 127.52, 123.64, 98.81, 98.59, 79.60, 78.07, 77.77, 76.67, 75.22, 74.54, 74.51, 74.24, 73.35, 73.20, 69.37, 68.79, 67.15, 55.88, 51.39, 29.00, 28.58, 23.30, 16.89; ESI-MS: *m/z* calcd for C₅₃H₅₈N₄O₁₁: 926.3994; found 949.3998 (*M* + Na)⁺.



5-Azidopentyl-O-2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 6)-(2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranosyl-(1 \rightarrow 3))-2,4-di-O-benzyl- β -mannopyranosyl-(1 \rightarrow 4)-O-3,6-di-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl-(1 \rightarrow 4)-(2,3,4-tri-O-benzyl- α -L-fuco pyranosyl-(1 \rightarrow 6))-3-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranoside (S20l):

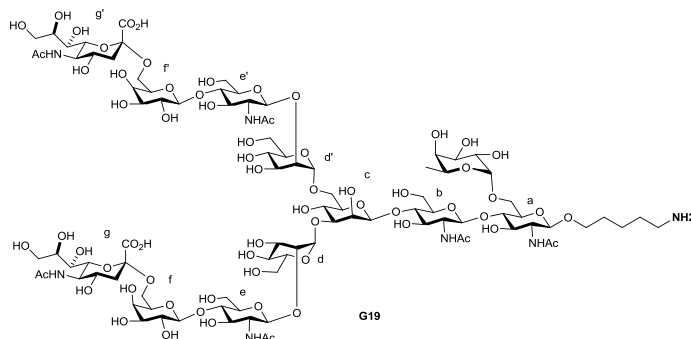
A mixture of donor **S20h** (0.050 g, 0.025 mmol), acceptor **S20k** (0.028 g, 0.031 mmol) and activated 4 Å molecular sieves in dry CH₂Cl₂ (10 mL) was stirred at rt for 30 minutes. The reaction was cooled to -70 °C, boron trifluoride ethyl etherate (0.725 μL, 0.0062 mmol) was then added slowly and the resulting reaction mixture was stirred for 2 h. The reaction was quenched by adding Et₃N, diluted with CH₂Cl₂, filtered through Celite and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 20% EA in hexane) to afford **S20l** (0.027 g, 45%) as a white foam. TLC: (ethyl acetate : hexane = 3/7, v/v): R_f = 0.50; ¹H NMR (600 MHz, CDCl₃): δ 7.83-7.62 (m, 8H, Ar-H), 7.49-7.30 (m, 8H, Ar-H), 7.30-7.10 (m, 47H, Ar-H), 7.09-7.03 (m, 10H, AR-H), 7.03-6.99 (m, 5H, Ar-H), 6.55 (t, *J* = 7.8 Hz, 2H, H-2^{d,d'}), 5.40 (s, 1H, H-1^d), 5.29-5.29 (m, 1H), 5.09 (d, *J* = 1.3 Hz, 1H, H^{1d'}), 4.88-4.76 (m, 13H), 4.67-4.65 (m, 2H), 4.58-5.48 (m, 9H), 4.42-4.40 (m, 3H), 4.36-4.30 (m, 5H), 4.20-4.10 (m, 5H), 4.05-4.00 (m, 2H), 3.92-3.80 (m, 7H), 3.79-3.60 (m, 7H), 3.62-3.41 (m, 10H), 3.25(dd, *J* = 3.2, 8.2 Hz, 1H), 3.18-3.17 (m, 2H), 2.85-2.79 (m, 2H), 2.06 (s, 3H, -C(O)CH₃), 1.76 (s, 3H, -C(O)CH₃), 1.42-1.34 (m, 4H, -CCH₂C-, linker), 1.03-0.99 (m, 5H, -CCH₂C-, linker and CH₃ of Fucose); ¹³C NMR (150 MHz, CDCl₃): δ 170.30, 169.91, 168.15, 167.76, 139.16, 138.99, 138.88, 138.82, 138.77, 138.54, 138.28, 138.21, 138.10, 138.08, 138.05, 133.86, 132.02, 131.72, 128.80, 128.62, 128.60, 128.57, 128.55, 128.33, 128.23, 128.40, 127.98, 127.92, 127.70, 127.55, 127.26, 127.06, 123.33, 101.97, 99.62, 98.41, 97.95, 97.09, 97.05, 81.42, 79.71, 78.36, 74.98, 74.75, 74.68, 73.65, 72.57,

72.51, 72.03, 71.38, 68.93, 68.86, 68.12, 66.63, 66.12, 56.76, 56.01, 51.26, 28.82, 28.56, 23.20, 21.23, 20.91; ESI-MS: m/z calcd for $C_{159}H_{165}N_5O_{34}$: 2689.1261; found 2712.1338 ($M + Na$)⁺.



5-Aminopentyl-di-(α -D-mannopyranosyl)-(1 \rightarrow 3),(1 \rightarrow 6)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-(α -L-fucopyranosyl-(1 \rightarrow 6)-2-acetamido-2-deoxy- β -D-glucopyranoside (G3**):** Compound **S20h** (0.030 g, 0.011 mmol) was deprotected by

following general procedure 2 (method 1) to obtain the title compound **G3** (0.007 g, 22%) as a white powder. ¹H NMR (600 MHz, D₂O): δ 5.09 (s, 1H, H-1^d), 4.91 (s, 1H, H-1^{d'}), 4.89 (d, $J = 3.2$ Hz, 1H, H-1^c), 4.78 (s, 1H, H-1^c), 4.65 (d, $J = 7.8$ HZ, 1H, H-1^a), 4.49 (d, $J = 7.8$ Hz, 1H, H-1^b), 4.25 (s, 1H), 4.15-4.10 (m, 1H), 4.05 (s, 1H), 3.95 (s, 1H), 3.95-3.80 (m, 8H), 3.80-3.50 (m, 23H), 3.00 (m, 2H), 2.10 (s, 3H, -C(O)CH₃), 2.09 (s, 3H, -C(O)CH₃), 1.67-1.60 (m, 2H, -CCH₂C-), 1.60-1.57 (m, 2H, -CCH₂C-), 1.41-1.37 (m, 2H, -CCH₂C-), 1.25 (d, $J = 7.8$ Hz, 3H, -CH₃ of Fucose); ¹³C NMR (150 MHz, D₂O): δ 174.67, 174.39, 170.97, 102.51, 101.10, 101.04, 100.38, 99.60, 99.39, 80.46, 79.72, 78.60, 74.33, 74.15, 73.42, 72.65, 72.30, 71.95, 71.85, 70.29, 70.12, 70.00, 69.97, 69.85, 69.45, 68.15, 66.77, 66.14, 65.85, 65.80, 61.11, 60.92, 59.97, 55.05, 54.86, 46.66, 39.28, 38.61, 28.05, 26.36, 22.27, 22.15, 21.08, 21.50, 15.37, 15.34; ESI-MS: m/z calcd for $C_{45}H_{80}N_3O_{30}$: 1141.4821; found 1142.4827 ($M + H$)⁺.



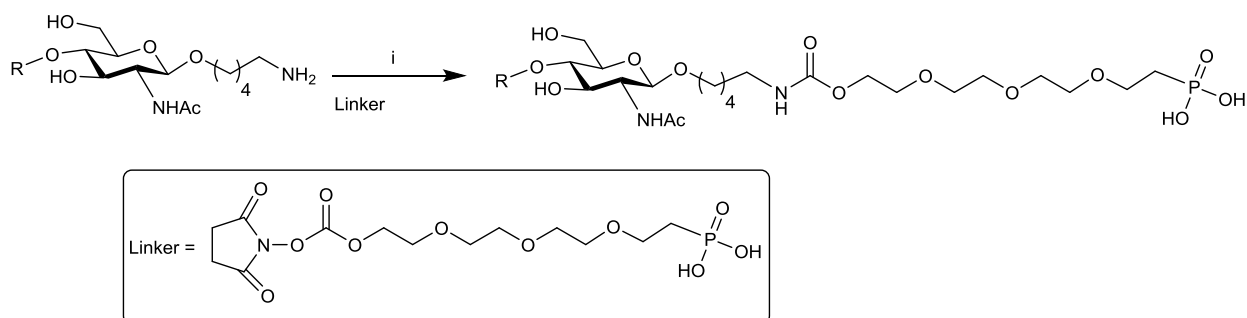
5-Aminopentyl-di-[5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate-(2 \rightarrow 6)- β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl]-(1 \rightarrow 3),(1 \rightarrow 6)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-(α -L-fucopyranosyl-(1 \rightarrow 6)-2-acetamido-2-deoxy- β -D-

glucopyranoside (G19): Sialylated biantennary with core fucose glycan **G19** was prepared by using reported procedure^{11,12}. ¹H NMR (600 MHz, D₂O): δ 4.90 (s, 1H, H-1^d), 4.72 (s, 1H, H-1^d), 4.66 (d, J = 3.6 Hz, 1H, H-1^c), 4.56 (s, 1H), 4.43 (d, J = 7.2 Hz, 1H, H-1^b), 4.37 (d, J = 7.2 Hz, 2H, H-1^{e,c}), 4.25 (d, J = 7.8 Hz, 1H, H-1^a), 4.21 (d, J = 7.8 Hz, 2H, H-1^f), 4.04 (s, 1H), 3.97 (s, 1H), 3.90 (t, J = 10.1 Hz, 2H), 3.76-3.25 (m, 65H), 2.74 (t, J = 7.8 Hz, 2H, linker), 2.43 (dd, J = 3.8 & 7.2 Hz, 1H, H-3^{equi.g,g'}), 1.87 (s, 3H), 1.84 (s, 3H, -C(O)CH₃), 1.83 (s, 3H, -C(O)CH₃), 1.80 (s, 6H, -C(O)CH₃), 1.51 (t, J = 12 H, 2H, H-3^{axial.g,g'}), 1.46-1.41 (m, 2H), 1.37-1.34 (m, 2H), 1.18-1.15 (m, 2H), 1.00 (d, J = 6.6 Hz, 3H, -CH₃ of Fucose); ¹³C NMR (150 MHz, D₂O): δ 174.5, 174.3, 174.1, 173.3, 103.3, 100.9, 100.7, 100.2, 99.7, 99.6, 99.3, 99.1, 98.9, 96.5, 80.2, 79.5, 77.8, 75.9, 75.6, 74.0, 73.3, 73.2, 73.1, 72.4, 72.2, 71.8, 71.7, 71.3, 70.3, 69.9, 69.7, 69.3, 69.1, 68.0, 67.9, 67.0, 66.9, 66.5, 65.7, 65.4, 63.0, 62.2, 61.4, 61.3, 59.8, 59.4, 54.7, 54.4, 54.2, 51.5, 39.7, 38.9, 27.7, 26.1, 22.1, 22.0, 22.0, 21.8, 21.7, 15.1 ; ESI-MS: m/z calcd for C₉₅H₁₅₉N₇O₆₆; 2451.9121; found 1226.4513 ($M - H$)²⁻.

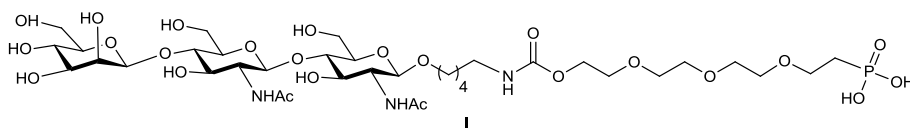
ix. Synthesis of glycans with phosphonic acid tails.

General procedure: To a solution of sugars with amine tail (3-5 μ mol) in DMF (400 μ L) was added linker [2(2(2(bis(benzyloxy)phosphoryl)ethoxyethoxy)ethyl(2,5-dioxopyrrolidin-1-yl)carbonate)] (15-25 μ mol), and the resulting reaction mixture was stirred at rt for 5 h. DMF was removed by using high vacuum and the product was purified by using Bio-Gel P-2 chromatography (eluent H₂O). The solid was dissolved in 2 mL of H₂O, added Pd(OH)₂ (50% by weight) and hydrogenated for overnight. The reaction mixture was filtered through Celite and

concentrated in *vacuo*. The residue was purified by Bio-Gel P-2 (BIO-RAD) column chromatography using water as eluent. The product was the lypholysed to obtain the desired sugar with phosphonic acid tail as white color powder.

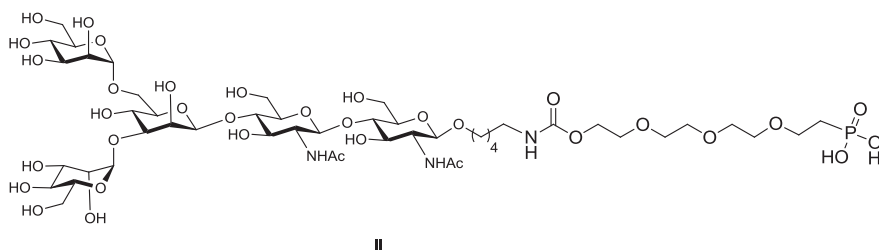


Scheme S21 | Preparation of sugars with phosphonic acid linker. i, (1) Linker, DMF, RT, 5 h;
(2) Pd(OH)₂, H₂O, H₂, RT, overnight.

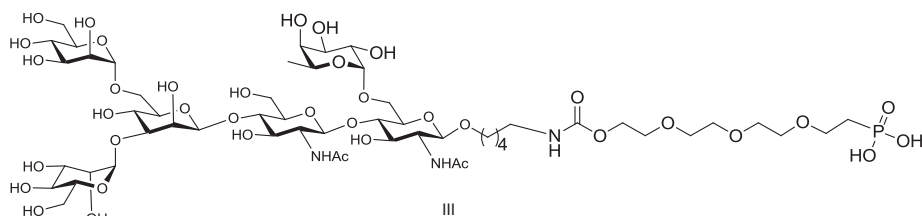


(Phosphonate-tetra-(ethyleneglycol)-carbonyl-amino)-pentyl-β-D-mannopyranosyl-(1→4)-2-acetamido-2-deoxy-β-D-glucopyranosyl-(1→4)-2-acetamido-2-deoxy-β-D-

glucopyranoside (I): Compound **G1** (3.5 mg, 5.4 μmol) was modified by above general procedure to afford compound **I** (2.2 mg, 61%), as white solid. TLC (MeOH: EA:AcOH: H₂O, 7/1/1/1, v/v). ¹H NMR (600 MHz, D₂O): δ 4.60 (d, *J* = 8.4 Hz, 1H), 4.48 (d, *J* = 7.8 Hz, 1H), 4.19 (s, 2H), 4.05 (d, *J* = 3.2 Hz, 1H), 3.93-3.49 (m, 33H), 1.75 (t, *J* = 7.8 Hz, 2H), 2.06 (s, 3H), 1.95 (s, 3H), 1.95-1.92 (m, 2H), 1.58-1.54 (m, 2H), 1.50-1.47 (m, 2H), 1.33-1.30 (m, 2H). HRMS (MALDI-TOF): *m/z* calcd for C₃₆H₆₆N₃O₂₄P; 955.8887 found 954.3099 (*M* -H).

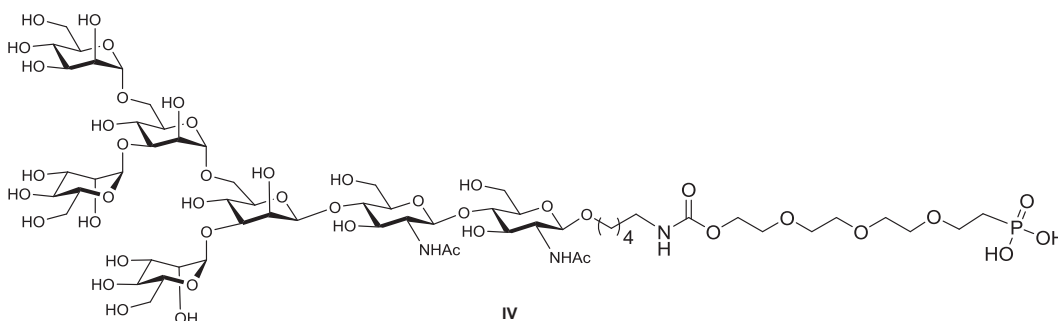


(Phosphonate-tetra-(ethyleneglycol)-carbonyl-amino)-pentyl- α -D-manno-pyranosyl(1 \rightarrow 3)- α -D-mannopyranosyl]-(1 \rightarrow 6)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranoside (II): Glycan G2 (4 mg, 2.3 μ mol) was modified by above general procedure to afford compound II (3.2 mg, 71%), as white solid. TLC (MeOH: EA: AcOH: H₂O, 7/1/1/1, v/v). ¹H NMR (600 MHz, D₂O): δ 5.01 (s, 1H), 4.83 (s, 1H), 4.51 (d, J = 6.1 Hz, 1H), 4.40 (d, J = 6.2 Hz, 1H), 4.17 (s, 1H), 4.11 (s, 2H), 3.98 (s, 1H), 3.88-3.39 (m, 42H), 3.03 (t, J = 7.8 Hz, 2H), 1.99 (s, 3H), 1.95 (s, 3H), 1.95-1.91 (m, 2H), 1.50-1.45 (m, 2H), 1.42-1.38 (m, 2H), 1.26-1.22 (m, 2H). HRMS (MALDI-TOF) Negative mode: m/z calcd for C₄₈H₈₆N₃O₃₄P; 1279.4830 found 1278.5001 ($M - H$).

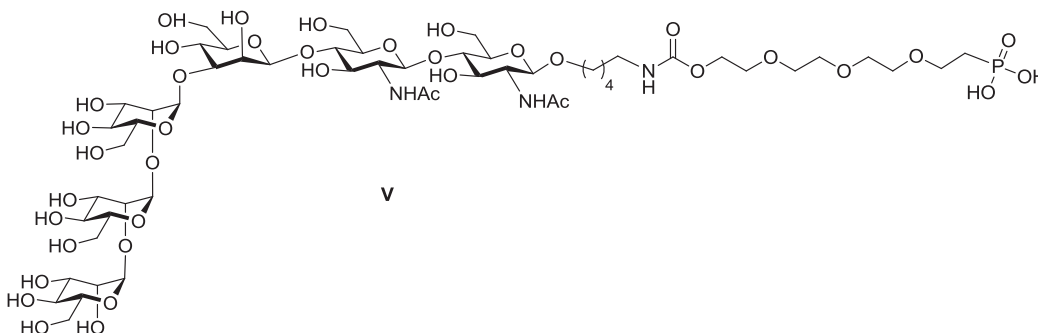


(Phosphonate-tri-(ethyleneglycol)-carbonyl-amino)-pentyl- α -D-manno-pyranosyl(1 \rightarrow 3)- α -D-mannopyranosyl]-(1 \rightarrow 6)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-(α -L-fucopyranosyl-(1 \rightarrow 6)-2-acetamido-2-deoxy- β -D-glucopyranoside) (III): Glycan G3 (1.2 mg, 1.0 μ mol) was modified by above general procedure to afford compound III (1.0 mg, 91%), as white solid. TLC (MeOH: EA: AcOH: H₂O, 7/1/1/1, v/v). ¹H NMR (600 MHz, D₂O): δ 5.09 (s, 1H), 4.90 (s, 1H), 4.88 (d, J = 7.4 Hz, 1H), 4.66 (d, J = 7.5 Hz, 1H), 4.48 (d, J = 7.2 Hz, 1H), 4.25 (s, 1H), 4.19 (s, 2H), 4.12-4.10 (m, 1H), 4.06 (s, 1H), 3.95 (s, 1H), 3.92-3.84 (m, 14H), 3.79-3.69 (m, 31H), 3.11-3.09 (m, 2H), 2.08 (s,

3H), 2.02 (s, 3H), 1.99-1.93 (m, 2H), 1.58-1.53 (m, 2H), 1.49-1.47 (m, 2H), 1.34-1.30 (m, 2H), 1.22 (d, 3H). HRMS (MALDI-TOF): m/z calcd for $C_{54}H_{96}N_3O_{38}P$; 1427.6754 found 1450.5709 ($M + Na$).

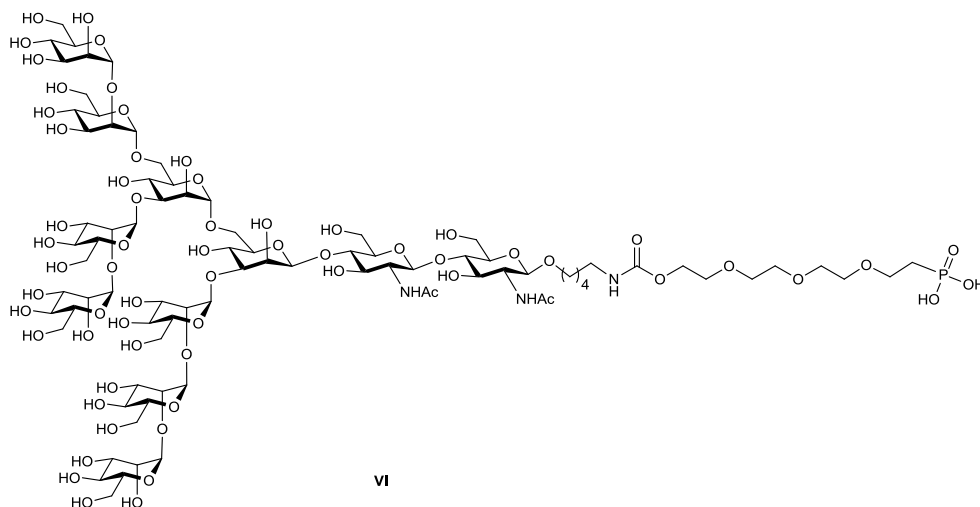


(Phosphonate-tetra(ethyleneglycol)-carbonyl-amino)-pentyl- α -D-mannopyranosyl(1 \rightarrow 3),[di-(α -D-mannopyranosyl)-(1 \rightarrow 3),(1 \rightarrow 6)- α -D-mannopyranosyl)-(1 \rightarrow 6)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranoside (IV): Glycan **G4** (4 mg, 3.1 μ mol) was modified by above general procedure to afford compound **IV** (3.7 mg, 78%), as white solid. TLC (MeOH: EA: AcOH: H₂O, 7/1/1/1, v/v). ¹H NMR (600 MHz, D₂O): δ 5.10 (d, $J = 9.6$ Hz, 2H), 4.93 (s, 1H), 4.70 (s, 1H), 4.57 (d, $J = 7.2$ Hz, 1H), 4.48 (d, $J = 7.8$ Hz, 1H), 4.24 (d, $J = 3.2$ Hz, 2H), 4.19-4.13 (m, 4H), 4.06-4.04 (m, 3H), 3.95-3.40 (m, 49H), 3.38-3.37 (m, 1H), 2.10 (t, $J = 7.8$ Hz, 2H), 2.07 (s, 6H), 2.02 (s, 6H), 1.98-1.93 (m, 2H), 1.58-1.47 (m, 2H), 1.34-1.29 (m, 2H), 1.27-1.25 (m, 2H); HRMS (MALDI-TOF): m/z calcd for $C_{60}H_{106}N_3O_{44}P$; 1604.5411 found 1607.0452 ($M + 3H$).



(Phosphonate-tetra-(ethylene glycol)-carbonyl-amino)-pentyl- α -D-mannopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl-(1 \rightarrow 3)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-

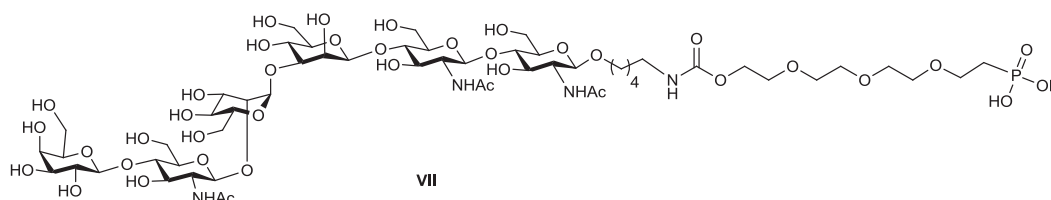
glucopyranoside (V): Glycan **G5** (5 mg, 4.4 μ mol) was modified by above general procedure to afford compound **V** (3.5 mg, 58%), as white solid. TLC (MeOH: EA: AcOH: H₂O, 7/1/1/1, v/v). ¹H NMR (600 MHz, D₂O): δ 5.34 (s, 1H), 5.29 (s, 1H), 5.03 (s, 1H), 4.58 (d, $J = 7.8$ Hz, 1H), 4.47 (d, $J = 7.8$ Hz, 1H), 4.20 (bs, 3H), 4.09-4.05 (m, 3H), 3.90-3.20 (m, 47H), 3.10 (t, $J = 7.8$ Hz, 2H), 2.05 (s, 3H), 2.01 (s, 3H), 2.00-1.92 (m, 2H), 1.56-1.53 (m, 2H), 1.49-1.46 (m, 2H), 1.33-1.30 (m, 2H); HRMS (MALDI-TOF): m/z calcd for C₅₄H₉₆N₃O₃₉P; 1442.3105 found 1444.5470 ($M + 2H$).



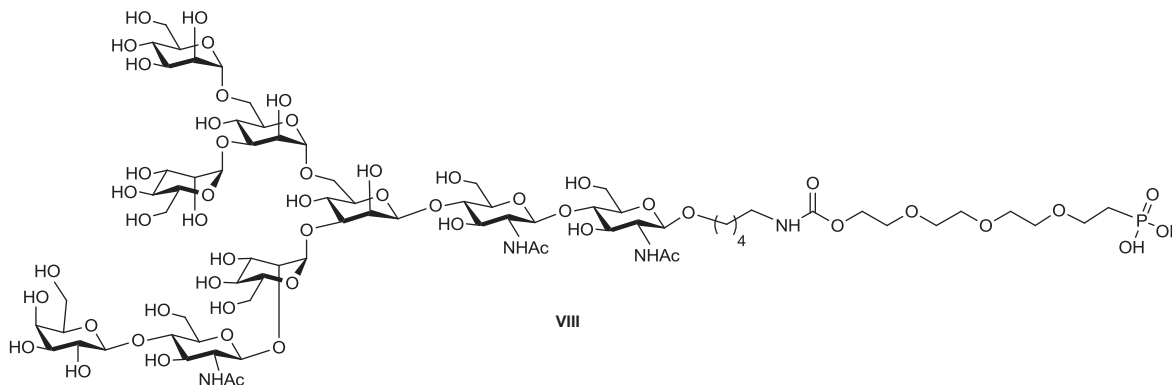
(Phosphonate-tetra-(ethyleneglycol)-carbonyl-amino)-pentyl- α -D-mannopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl-(1 \rightarrow 3)-{ α -D-mannopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl-(1 \rightarrow 3)-[α -D-mannopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl-(1 \rightarrow 6)]- α -D-mannopyranosyl-(1 \rightarrow 6)}- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-

glucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranoside (VI): Glycan **G6** (4 mg, 2.0 μ mol) was modified by above general procedure to afford compound **VI** (3.7 mg, 66%), as white solid. TLC (MeOH: EA: AcOH: H₂O, 7/1/1/1, v/v). ¹H NMR (600 MHz, D₂O): δ 5.53 (s,

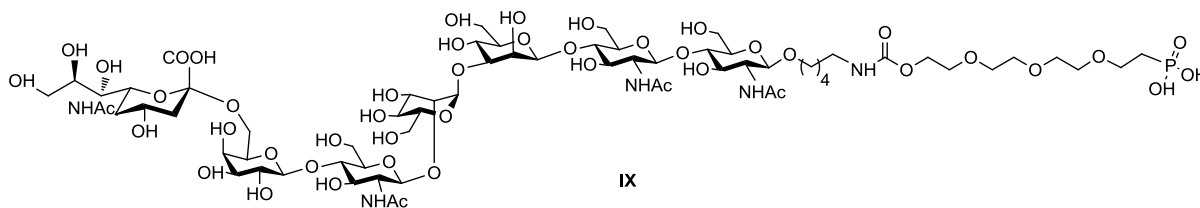
1H), 5.39 (d, $J = 10.2$ Hz, 1H), 5.26 (d, $J = 9.8$ Hz, 1H), 5.23 (s, 1H), 5.16 (d, $J = 4.8$ Hz, 1H), 5.08-5.05 (m, 5H), 4.60-3.20 (m, 83H), 2.08 (s, 3H), 2.02 (s, 3H), 1.38-1.03 (m, 2H), 1.58-1.56 (m, 2H), 1.52-1.48 (m, 2H), 1.35-1.32 (m, 2H); HRMS (MALDI-TOF): m/z calcd for $C_{84}H_{146}N_3O_{64}P$; 2253.0310 found 2257.0812 ($M + 4H$).



(Phosphonate-tetr-(ethyleneglycol)-carbonyl-amino)-pentyl- β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl -(1 \rightarrow 2)- α -D-mannopyranosyl]-(1 \rightarrow 3)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranoside (VII): Glycan **G9** (6.0 mg, 5.1 μ mol) was modified by above general procedure to afford compound **VII** (3.5 mg, 50%), as white solid. TLC (MeOH: EA: AcOH: H_2O , 7/1/1/1, v/v). 1H NMR (600 MHz, D_2O): δ 5.03 (d, $J = 8.4$ Hz, 2H), 4.58 (d, $J = 7.2$ Hz, 1H), 4.48 (d, $J = 7.2$ Hz, 1H), 4.45 (d, $J = 7.8$ Hz, 1H), 4.26 (dd, $J = 3.0$ & 9.2 Hz, 2H), 4.19 (bs, 2H), 4.00-3.38 (m, 49H), 3.11 (t, $J = 7.8$ Hz, 2H), 2.06 (s, 3H), 2.05 (s, 3H), 2.02 (s, 3H), 1.99-1.93 (m, 2H), 1.56-1.54 (m, 2H), 1.50-1.48 (m, 2H), 1.31-1.27 (m, 2H); HRMS (MALDI-TOF): m/z calcd for $C_{56}H_{99}N_4O_{39}P$; 1482.5624 found 1481.5069 ($M - H$).



(Phosphonate-tetra-(ethyleneglycol)-carbonyl-amino)-pentyl- β -D-galactopyranosyl-(1 \rightarrow 4)-[2-acetamido-2-deoxy- β -D-gluco-pyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl-(1 \rightarrow 3),[di-(α -D-mannopyranosyl)-(1 \rightarrow 3),(1 \rightarrow 6)- α -D-manno-pyranosyl](1 \rightarrow 6)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-gluco-pyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-gluco-pyranoside (VIII): Glycan **G12** (5.0 mg, 3.0 μ mol) was modified by above general procedure to afford compound **VIII** (3.1 mg, 54%), as white solid. TLC (MeOH: EA: AcOH: H₂O, 7/1/1/1, v/v). ¹H NMR (600 MHz, D₂O): δ 5.11 (s, 2H), 5.02 (d, $J = 7.8$ Hz, 2H), 4.68 (d, $J = 3.2$ Hz, 2H), 4.48 (d, $J = 7.2$ Hz, 1H), 4.45 (d, $J = 7.2$ Hz, 1H), 4.27 (s, 2H), 4.22 (s, 2H), 4.15 (s, 1H), 4.08 (s, 2H), 4.00-3.30 (m, 64H), 3.10 (t, $J = 7.8$ Hz, 2H), 2.08 (s, 3H), 2.05 (s, 3H), 2.02 (s, 3H), 1.93-1.87 (m, 2H), 1.56-1.54 (m, 2H), 1.51-1.47 (m, 2H), 1.33-1.30 (m, 2H); HRMS (MALDI-TOF): m/z calcd for C₇₄H₁₂₉N₄O₅₄P; 1968.6502 found 1967.6622 ($M - H$).

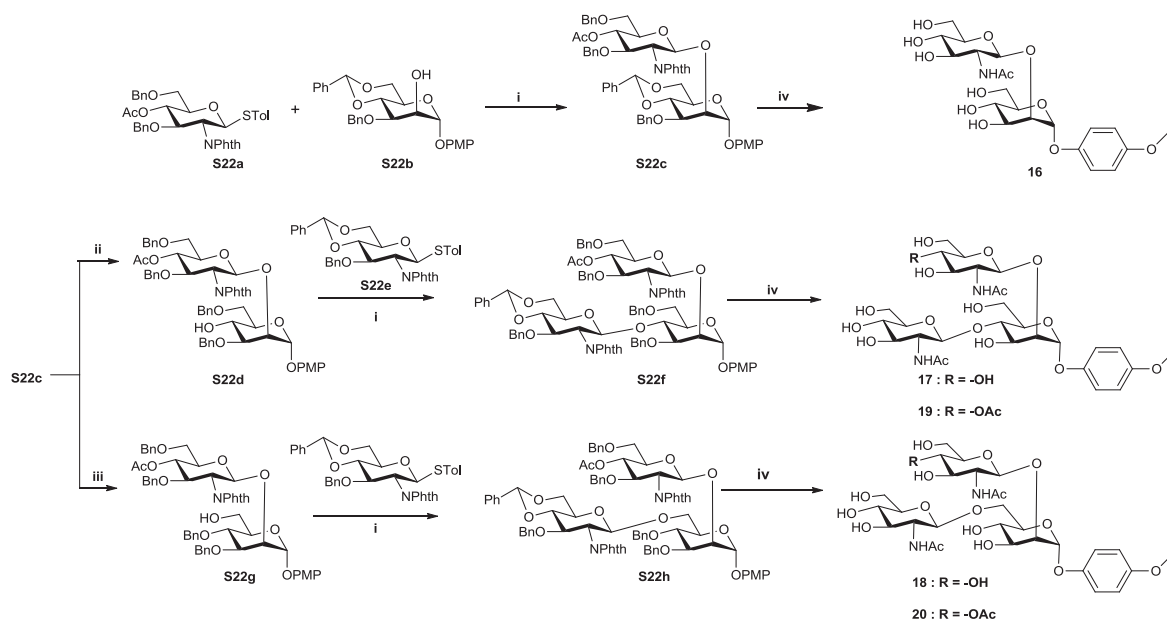


Phosphonate-tetra-(ethyleneglycol)-carbonyl-amino)-pentyl-[5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosyl-(2 \rightarrow 6)- β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-gluco-pyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl](1 \rightarrow 3)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-gluco-pyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-gluco-pyranoside (IX): Glycan **G10** (5.0 mg, 2.5 μ mol) was modified by above general procedure to afford compound **IX** (3.8 mg, 63%), as white solid. TLC (MeOH: EA: AcOH: H₂O, 7/1/1/1, v/v). ¹H NMR (600 MHz, D₂O): δ 5.08 (d, $J = 8.4$ Hz, 2H), 4.93 (s, 1H), 4.62 (d, $J = 7.2$ Hz, 1H), 4.51 (d, $J = 7.8$ Hz, 1H), 4.45 (d, $J = 7.8$ Hz, 1H), 4.30 (d, $J = 3.2$ Hz, 1H), 4.28 (s, 1H), 4.22 (s, 2H), 4.02-3.30 (m, 54H), 3.13 (t, $J = 7.8$ Hz, 2H), 2.68 (dd, $J = 3.2, 7.8$ Hz, 1H), 2.09 (s, 6H), 2.05 (s, 6H), 2.01-1.93 (m, 2H), 1.74 (t, $J = 12.1$ Hz, 1H), 1.60-1.57

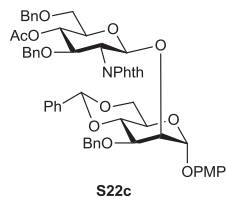
Phosphonate-tetra-(ethyleneglycol)-carbonyl-amino)-pentyl-di-[5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyrano-sylonate-(2 \rightarrow 6)- β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl]-(1 \rightarrow 3),(1 \rightarrow 6)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranoside (XI): Glycan **G16** (3.5 mg, 1.5 μ mol) was modified by above general procedure to afford compound **XI** (2.6 mg, 69%) as a white solid. TLC (MeOH: AcOH: H₂O, 8/1/1, v/v). ¹H NMR (600 MHz, D₂O): δ 5.05 (d, J = 8.4 Hz, 1H), 4.63 (d, J = 6.6 Hz, 1H), 4.51 (s, 1H), 4.46 (t, J = 7.8 Hz, 2H), 4.32 (s, 1H), 4.29 (s, 2H), 4.22 (s, 2H), 4.09-3.48 (m, 83H), 3.13 (t, J = 7.8 Hz, 2H), 2.74-2.68 (m, 2H), 2.10 (s, 9H), 2.05 (s, 9H), 2.12-2.00 (m, 2H), 1.76-1.73 (m, 2H), 1.58-1.55 (m, 2H), 1.51-1.50 (m, 2H), 1.35-1.29 (m, 2H); HRMS (MALDI-TOF negative mode): m/z calcd for C₉₈H₁₆₆N₇O₇₀P; 2592.9383 found 1294.9685 ($M - 2H$)²⁻.

x. Chemo-enzymatic synthesis of D1 and D2/D3 arm modules.

Our chemo-enzymatic strategy commenced with preparation of acceptor substrates **16-20**. As depicted in scheme S22, mannosyl acceptor **S22b** was glycosylated with donor **S22a** in the presence of NIS/TfOH to afford the disaccharide **S22c** in 60% yield. Benzylidene ring of **S22c** was opened both at 4-OH, **S22d** or 6-OH, **S22g** under different reaction conditions. Donor **S22e** was installed separately at 4-O and 6-O positions of **S22d** and **S22g** respectively. Finally, global deprotection of intermediates **S22c**, **S22f** and **S22h** was performed. In case of compounds **S22j** and **S22i**, the GlcNAc residues at Man 4 and 6-O positions were differentiated from 2-O GlcNAc through acetylation of 4-OH, for preparation of asymmetric modules.

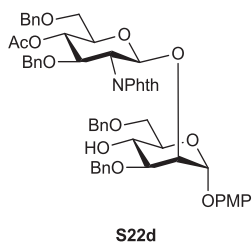


Scheme S22 | Preparation of acceptor substrates. **i**, NIS, TfOH, CH₂Cl₂, -50 °C; **ii**, Triethyl silane, TFA, 4 Å MS, CH₂Cl₂, 0 °C, 2 h, 58%; **iii**, BH₃.THF, Bu₂BOTf, CH₂Cl₂, 4 Å MS, 0 °C, 3 h, 71%; **iv**, (1) NH₂CH₂CH₂NH₂, *n*-BuOH, 100 °C; (2) Ac₂O, pyridine, 0 °C to RT; (3) Pd(OH)₂, MeOH: H₂O: HCOOH (5:3:2), H₂;

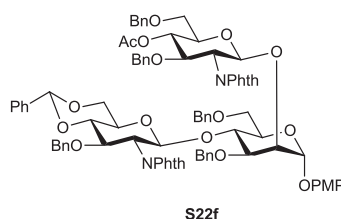


p-methoxyphenyl-O-[4-O-acetyl-3,6-O-di-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl-(1 \rightarrow 2)-O-4,6-O-benzylidene-3-O-benzyl- α -D-mannopyranoside (S22c):

Activated 4Å molecular sieves were added to a solution of acceptor **S22b** (0.250 g, 0.536 mmol) and donor **S22a** (0.410 g, 0.643 mmol) in anhydrous CH₂Cl₂ (10 mL). The reaction mixture was stirred for 1 h at rt then cooled to -50 °C. NIS (0.241 g, 1.07 mmol) and TfOH (11.8 μ L, 0.134 mmol) were added slowly, and the resulting reaction mixture was stirred for 1 h. When TLC (ethyl acetate: toluene, 2/8) indicated formation of product with consumption of starting material, the reaction was quenched by adding Et₃N then filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), aqueous Na₂S₂O₃ (2 x 50 mL), and brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 10% EA in toluene) to afford **S22c** (0.403 g, 76%) as a white foam. TLC (ethyl acetate: toluene = 2/8, v/v); R_f = 0.49; ¹H NMR (400 MHz, CDCl₃): δ 7.69-3.56 (m, 4H), 7.36-7.22 (m, 15H), 7.20-6.96 (m, 5H), 6.53 (d, *J* = 8.4 Hz, 2H), 6.50 (d, *J* = 8.2 Hz, 2H), 5.41 (s, 1H), 5.24 (d, *J* = 7.8 Hz, 1H), 5.10 (t, 1H), 4.93 (s, 1H), 4.73-4.43 (m, 10H), 4.00-3.20 (m, 5H), 1.98 (s, 3H); ¹³C NMR (150 MHz, CHCl₃): δ 169.2, 152.3, 135.1, 134.9, 133.9, 128.7, 128.6, 128.4, 128.1, 128.1, 128.0, 127.7, 127.6, 127.5, 117.8, 114.7, 97.1, 96.3, 74.0, 73.9, 73.1, 72.9, 72.8, 72.8, 72.5, 72.2, 71.4, 65.7, 55.4, 54.2, 22.5; ESI-MS: *m/z* calcd for C₅₇H₅₅NO₁₄; 977.3623 found 1000.3490 (*M* + Na)⁺.

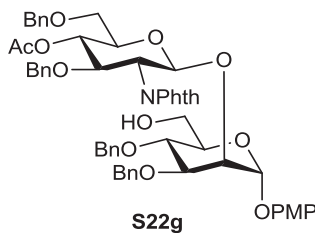


p-methoxyphenyl-O-[4-O-acetyl-3,6-O-di-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl-(1 \rightarrow 2)-O-3,6-O-dibenzyl- α -D-mannopyranoside (S22d): To a solution of **S22c** (1.01 g, 1.02 mmol) in anhydrous CH₂Cl₂ (10 mL) was added triethyl silane (1.63 mL, 10.2 mmol) followed by trifluoroacetic acid (0.758 mL, 10.2 mmol) at 0 °C. The resulting reaction mixture was stirred for 2 h. After 2 h, TLC (ethyl acetate: toluene, 1.5/8.5 v/v) indicated product formation with consumption of starting material. The reaction mixture was washed with sat. NaHCO₃ (2 x 50 mL). The aqueous layer was further extracted with CH₂Cl₂ (3 x 30 mL), and the combined organic layer were washed with brine solution (100 mL), dried over MgSO₄, filtered and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% – 10% EA in toluene) to afford **S22d** (0.580 g, 58%) as clear oil. TLC (ethyl acetate: toluene = 1.5/8.5, v/v): R_f = 0.35; ¹H NMR (400 MHz, CDCl₃): δ 7.63-7.35 (m, 4H), 7.30-7.10 (m, 14H), 7.02-6.97 (m, 3H), 6.93-6.86 (m, 3H), 6.72-6.66 (m, 4H), 5.27 (d, *J* = 8.4 Hz, 1H), 5.11 (t, *J* = 10.2 Hz, 1H), 5.02 (d, *J* = 2.8 Hz, 1H), 4.88 (s, 2H), 4.80 (s, 1H), 4.60 (d, *J* = 10.2 Hz, 2H), 4.58-4.24 (m, 6H), 4.01 (s, 2H), 3.79-3.74 (m, 2H), 3.72 (s, 3H), 3.64-3.56 (m, 3H), 3.35 (dd, *J* = 2.8 & 8 Hz, 1H), 2.93 (dd, *J* = 2.3 & 7.8 Hz, 1H), 1.94 (s, 3H); ¹³C NMR (150 MHz, CHCl₃): δ 150.33, 138.18, 137.98, 133.96, 128.70, 128.65, 128.42, 128.18, 128.12, 128.02, 127.76, 127.62, 127.52, 117.89, 114.71, 97.19, 96.33, 74.08, 73.96, 73.10, 72.95, 72.80, 71.86, 70.96, 70.75, 70.48, 67.72, 55.89, 55.69, 21.21; ESI-MS: *m/z* calcd for C₅₇H₅₇NO₁₄; 979.3779 found 1002.3660 (*M* + Na)⁺.



p-methoxyphenyl-O-4-O-acetyl-3,6-O-di-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl-(1 \rightarrow 2)-[O-4,6-O-benzylidene-3-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl-(1 \rightarrow 4)]-3,6-O-di-benzyl- α -D-mannopyranoside (S33f): Activated 4 Å molecular sieves were added to a solution of acceptor **S22d** (0.580 g, 0.590 mmol) and donor **S22e** (0.525 g,

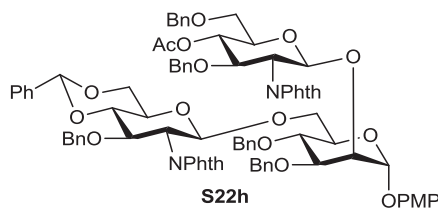
0.880 mmol) in anhydrous CH_2Cl_2 (10 mL). The reaction mixture was stirred for 1 h at rt then cooled to $-50\text{ }^\circ\text{C}$. NIS (0.265 g, 1.14 mmol) and TfOH (13 μL , 0.147 mmol) were added slowly, and the resulting reaction mixture was stirred for 2 h. When TLC (ethyl acetate: toluene, 1.5/8.5) indicated formation of product with consumption of starting material, the reaction was quenched by adding Et_3N then filtered through Celite. The filtrate was washed with aqueous NaHCO_3 (2 x 50 mL), aqueous $\text{Na}_2\text{S}_2\text{O}_3$ (2 x 50 mL), and brine (50 mL) solution. The organic layer was dried over Na_2SO_4 and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 10% EA in toluene) to afford **S22f** (0.730 g, 85%) as a pale yellow solid. TLC (ethyl acetate: toluene = 1.5/8.5, v/v): $R_f=0.60$; ^1H NMR (400 MHz, CDCl_3): δ 7.69- 6.67 (m, 8H), 7.59-7.18 (m, 20H), 6.96-6.75 (m, 10H), 6.57-6.52 (m, 4H), 5.44 (s, 1H), 5.27 (d, $J = 8.4$ Hz, 1H), 5.21 (d, $J = 8.8$ Hz, 1H), 5.11 (t, $J = 10.7$ Hz, 1H), 3.29-3.24 (m, 1H), 4.94 (d, $J = 8.4$ Hz, 1H), 4.76-4.69 (m, 2H), 4.57 (d, $J = 7.8$ Hz, 1H), 4.53 (d, $J = 7.2$ Hz, 1H), 4.45-4.19 (m, 9H), 4.10-4.06 (m, 2H), 3.99-3.94 (m, 2H), 3.63 (s, 3H), 3.56 -3.48 (m, 8H), 3.00 (dd, $J = 2.3$ and 7.8 Hz, 2H), 1.99 (s, 3H); ^{13}C NMR (150 MHz, CHCl_3): δ 169.98, 168.04, 138.94, 138.46, 138.14, 137.96, 133.90, 131.78, 129.24, 128.69, 128.52, 128.34, 128.25, 128.18, 128.08, 127.72, 127.33, 126.38, 123.56, 117.90, 114.47, 101.43, 99.38, 83.11, 78.85, 74.79, 74.27, 73.94, 73.76, 72.80, 72.34, 71.43, 70.26, 69.23, 68.90, 66.00, 56.49, 55.85, 55.70, 21.70; ESI-MS: m/z calcd for $\text{C}_{85}\text{H}_{80}\text{N}_2\text{O}_{20}$; 1448.5153 found 1471.5156 ($M + \text{Na}$) $^+$.



p-methoxyphenyl-O-[4-O-acetyl-3,6-O-di-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranosyl-(1 \rightarrow 2)-O-3,4-O-dibenzyl- α -D-mannopyranoside (S22g):

To a mixture of compound **S22c** (0.800 g, 0.816 mmol) and activated molecular sieves in anhydrous CH_2Cl_2 (10 mL) was added borane.THF complex (0.781 mL of a 1M solution in THF, 8.15 mmol) and

Bu₂BOTf (0.351 mL of a 1M solution in CH₂Cl₂, 1.63 mmol) were added at 0°C. The reaction mixture was allowed to stirred at room temperature for 3 h. TLC (acetone: toluene, 1/9) indicated formation of a product with consumption of the starting material. Triethyl amine was added to the reaction mixture followed by slow addition of methanol at 0 °C. When no more hydrogen was produced, the reaction mixture was filtered through Celite, The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), and brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% →10% EA in toluene) to afford **S22g** (0.500 g, 71%) as clear foam. TLC (acetone: toluene = 2/8, v/v): R_f = 0.41; ¹H NMR (400 MHz, CDCl₃): δ 7.84-7.55 (m, 4H), 7.40-6.90 (m, 10H), 6.87-6.80 (m, 7H), 6.93-6.86 (m, 3H), 6.72-6.66 (m, 4H), 5.27 (d, *J* = 8.4 Hz, 1H), 5.11 (t, *J* = 10.2 Hz, 1H), 5.02 (d, *J* = 2.8 Hz, 1H), 4.88 (s, 2H), 4.80 (s, 1H), 4.60 (d, *J* = 10.2 Hz, 2H), 4.58-4.24 (m, 6H), 4.01 (s, 2H), 3.79-3.74 (m, 2H), 3.72 (s, 3H), 3.64-3.56 (m, 3H), 3.35 (dd, *J* = 2.8 & 8 Hz, 1H), 2.93 (dd, *J* = 2.3 & 7.8 Hz, 1H), 1.94 (s, 3H); ¹³C NMR (150 MHz, CHCl₃): δ 154.2, 138.6, 136.5, 133.5, 128.0, 127.5, 127.4, 128.18, 127.1, 127.0, 126.7, 126.6, 126.5, 117.89, 114.71, 97.19, 96.33, 74.08, 73.96, 73.10, 72.9, 72.8, 72.5, 72.6, 71.9, 71.4, 65.7, 56.8, 56.6, 21.9; ESI-MS: *m/z* calcd for C₅₇H₅₇NO₁₄; 979.2640; found 979.2012.



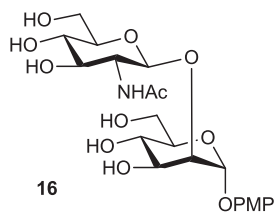
p-methoxyphenyl-O-4-O-acetyl-3,6-O-di-benzyl-2-deoxy-2-phthalimido-β-D-gluco

pyranosyl-(1→2)-[-O-4,6-O-benzylidene-3-O-benzyl-2-deoxy-2-phthalimido-β-D-gluco

pyranosyl-(1→6)]-3,6-O-di-benzyl-α-D-mannopyranoside (S22h**):** Activated 4 Å molecular

sieves were added to a solution of acceptor **S22g** (0.500 g, 0.509 mmol) and donor **S22e** (0.452 g, 0.763 mmol) in anhydrous CH₂Cl₂ (10 mL). The reaction mixture was stirred for 1 h at room temperature then cooled to -50 °C. NIS (0.229 g, 1.01 mmol) and TfOH (22 μL, 0.250 mmol)

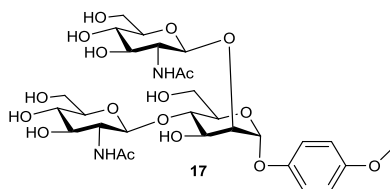
were added slowly, and the resulting reaction mixture was stirred for 1 h. When TLC (ethyl acetate: toluene, 1/9) indicated formation of product with consumption of starting material, the reaction was quenched by adding Et₃N then filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 50 mL), aqueous Na₂S₂O₃ (2 x 50 mL), and brine (50 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 10% EA in toluene) to afford **S22h** (0.610 g, 82%) as a pale yellow solid. TLC (ethyl acetate: toluene = 1/9, v/v): ¹H NMR (400 MHz, CDCl₃): δ 7.75-7.50 (m, 8H), 7.47-7.05 (m, 20H), 7.02-6.82 (m, 10H), 6.66 (d, *J* = 8.5 Hz, 2H), 6.48 (d, *J* = 8.6 Hz, 2H), 5.38 (s, 1H), 5.17 (d, *J* = 8.4 Hz, 1H), 5.21 (t, *J* = 10.3 Hz, 1H), 4.85 (d, *J* = 8.6 Hz, 1H), 3.29-3.24 (m, 1H), 4.94 (d, *J* = 8.4 Hz, 1H), 4.76-4.69 (m, 2H), 4.57 (d, *J* = 7.8 Hz, 1H), 4.53 (d, *J* = 7.2 Hz, 1H), 4.45-4.19 (m, 9H), 4.10-4.06 (m, 2H), 3.99-3.94 (m, 2H), 3.63 (s, 3H), 3.56 - 3.55 (m, 6H), 3.44-3.25 (m, 2H), 3.00 (t, *J* = 10.3 Hz, 2H), 1.99 (s, 3H); ¹³C NMR (150 MHz, CHCl₃): δ 170.0, 154.9, 150.7, 146.8, 138.6, 137.9, 137.4, 137.1, 136.9, 133.9, 132.7, 129.24, 128.69, 128.52, 128.34, 128.25, 127.9, 127.72, 127.33, 126.38, 123.56, 117.90, 114.47, 101.43, 99.38, 83.11, 78.85, 74.79, 74.27, 73.94, 73.76, 72.80, 72.34, 71.43, 70.26, 69.23, 68.90, 66.00, 58.49, 56.8, 56.7, 55.90, 55.8, 21.8; ESI-MS: *m/z* calcd for C₈₅H₈₀N₂O₂₀; 1448.5153 found 1449.5678 (*M* + H)⁺.



p-methoxyphenyl-O-2-acetamido-2-deoxy-β-D-glucopyranosyl-(1→2)-α-D-

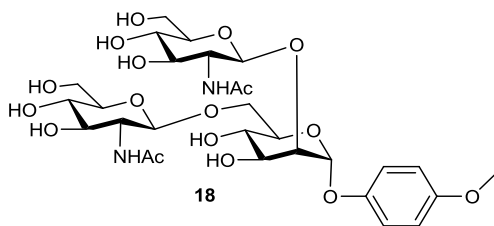
mannopyranoside (16): Compound **S22c** (0.105 g, 0.107 mmole) was deprotected by following general procedure 2 (method 1) to get the title compound **16** (0.035 g, 66%) as a white solid. ¹H NMR (400 MHz, D₂O): δ 7.08 (d, *J* = 9.6 Hz, 2H), 6.92 (d, *J* = 9.1 Hz, 2H), 5.43 (s, 1H), 4.60 (d, *J* = 8.4 Hz, 1H), 4.25 (t, *J* = 2.3 Hz, 1H), 4.01 (dd, *J* = 3.2 & 10.1 Hz, 1H), 3.87 (dd, *J* = 3.1 & 12.2 Hz, 1H), 3.80 (dd, *J* = 3.0 & 12.8 Hz, 1H), 3.78 (s, 3H), 3.72- 3.67 (m, 3H), 3.61-3.52 (m,

3H), 3.46- 3.40 (m, 2H), 1.99 (s, 3H), ^{13}C NMR (150 MHz, D_2O): δ 174.64, 170.85, 154.46, 149.57, 118.60, 114.83, 99.57, 96.50, 76.24, 75.67, 73.35, 73.10, 69.67, 69.28, 66.90, 61.14, 60.40, 55.55, 55.20; ESI-MS: m/z calcd for $\text{C}_{21}\text{H}_{31}\text{NO}_7$; 489.1738 found 512.1715 ($M + \text{Na}$) $^+$.



***p*-methoxyphenyl-O-di-(2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 2),(1 \rightarrow 4)- α -D-**

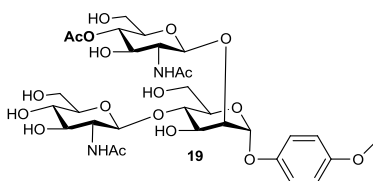
mannopyranoside (17): Compound **S22f** (0.125 g, 0.086 mmol) was deprotected by following general procedure 2 (method 1) to get the title compound **17** (0.42 g, 66%) as a white solid. ^1H NMR (400 MHz, D_2O): δ 7.10 (d, $J = 8.8$ Hz, 2H), 7.08 (d, $J = 8.2$ Hz, 2H), 5.39 (s, 1H), 4.53 (d, $J = 8.2$ Hz, 1H), 4.50 (d, $J = 7.8$ Hz, 1H), 4.25 (s, 1H), 4.20 (d, $J = 7.8$ Hz, 1H), 4.00 (dd, $J = 3.2$ and 7.2 Hz, 1H), 3.71 (s, 3H), 3.67-3.74 (m, 15H), 2.02 (s, 3H), 1.91 (s, 3H); ^{13}C NMR (150 MHz, D_2O): δ 174.48, 147.04, 154.49, 149.86, 118.53, 114.97, 100.80, 99.64, 96.55, 76.40, 75.71, 75.66, 73.74, 73.01, 72.63, 69.74, 69.27, 69.19, 67.32, 60.72, 60.60, 60.45, 59.05, 55.62, 55.34, 55.22, 22.24, 22.01; ESI-MS: m/z calcd for $\text{C}_{29}\text{H}_{44}\text{N}_2\text{O}_{17}$; 693.2713 found 693.2688 ($M + \text{H}$) $^+$.



***p*-methoxyphenyl-O-di-(2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 2),(1 \rightarrow 6)- α -D-**

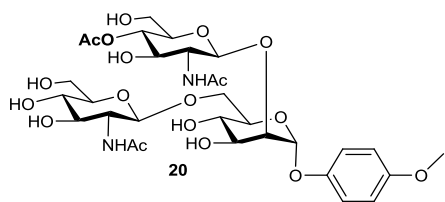
mannopyranoside (18): Compound **S22h** (0.200 g, 0.138 mmol) was deprotected by following general procedure 2 (method 1) to get the title compound **18** (0.60 g, 63%) as a white solid. ^1H NMR (400 MHz, D_2O): δ 7.10 (d, $J = 8.8\text{Hz}$, 2H), 6.96 (d, $J = 8.2$ Hz, 2H), 5.44 (s, 1H), 4.60 (d,

$J = 7.5\text{Hz}$, 1H), 4.52 (d, $J = 7.3\text{Hz}$, 1H), 4.31 (s, 1H), 4.18 (d, $J = 2.8\text{ Hz}$, 1H), 3.90 (t, $J = 10.7\text{ Hz}$, 2H), 3.80 (s, 3H), 3.77-3.70 (m, 6H), 3.56-3.44 (m, 8H), 2.02 (s, 6H); ^{13}C NMR (150 MHz, CHCl_3): δ 172.7, 171.9, 157.9, 150.3, 118.5, 114.9, 100.2, 99.4, 99.0, 75.74, 74.2, 73.9, 72.1, 70.3, 68.7, 68.3, 68.2, 55.9, 55.6, 55.1, 20.1; ESI-MS: m/z calcd for $\text{C}_{29}\text{H}_{44}\text{N}_2\text{O}_{17}$; 692.2532 found 715.2518 ($M + \text{Na}$) $^+$.



***p*-methoxyphenyl-O-(4-O-acetyl-2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 2)-O-(2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 4)- α -D-mannopyranoside (19):**

A mixture of **S22f** (0.610 g, 0.421 mmol) and 10 mL of ethylene diamine: *n*-BuOH (1:4) was stirred at 90 °C for overnight. Volatiles were then evaporated and the crude product was reacted with 10 mL Ac_2O /pyridine (1:2) for overnight. The solvents were removed using high vacuum and product was purified by flash column chromatography (acetone: toluene, 2/8, v/v). The product was dissolved in 10 mL MeOH: H_2O : HCOOH (6:3:1), $\text{Pd}(\text{OH})_2$ (50% by weight) was added and the reaction mixture was hydrogenated for overnight. The reaction mixture was filtered through Celite and concentrated in *vacuo*. The residue was purified by Bio-Gel P-2 (BIO-RAD) column chromatography using water as eluent, and the product was the lyophilized to get **19** (0.210 g, 67%) as a white color powder. ^1H NMR (400 MHz, D_2O): δ 7.01 (d, $J = 9.2\text{ Hz}$, 2H), 6.90 (d, $J = 9.8\text{ Hz}$, 2H), 5.40 (s, 1H), 4.61 (d, $J = 8.0\text{ Hz}$, 1H), 4.52 (d, $J = 8.4\text{ Hz}$, 1H), 4.25 (d, $J = 2.1\text{ Hz}$, 1H), 4.73 (dd, $J = 1.2$ & 7.2 Hz , 1H), 3.92 (d, $J = 12.3\text{ Hz}$, 1H), 3.74 (s, 3H), 3.72-3.67 (m, 8H), 3.60-3.41 (m, 7H), 2.12 (s, 3H), 2.03 (s, 6H); ^{13}C NMR (150 MHz, D_2O): δ 174.67, 174.17, 172.91, 154.43, 149.58, 118.53, 118.37, 114.81, 101.48, 99.70, 96.04, 77.51, 76.00, 75.75, 73.50, 73.39, 71.74, 71.06, 70.93, 69.57, 69.34, 68.04, 60.58, 60.47, 60.00, 55.55, 55.39, 55.13, 22.24, 21.95, 20.17, 20.03; ESI-MS: m/z calcd for $\text{C}_{31}\text{H}_{46}\text{N}_2\text{O}_{18}$; 735.2818 found 735.2780 ($M + \text{H}$) $^+$.

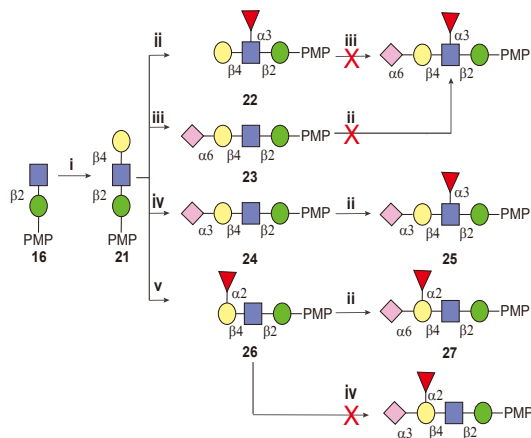


***p*-methoxyphenyl-O-(4-O-acetyl-2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 2)-O-(2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 6)- α -D-mannopyranoside (**20**):**

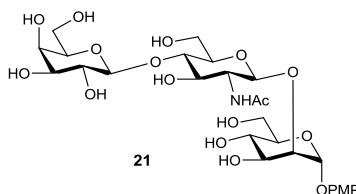
A mixture of **S22h** (0.350 g, 0.241 mmol) and 10 mL of ethylene diamine: *n*BuOH (1:4) was stirred at 90 °C for overnight. Volatiles were then evaporated and the crude product was reacted with 10 mL Ac₂O/pyridine (1:2) for overnight. The solvents were removed using high vacuum and product was purified by flash column chromatography (acetone: toluene, 2/8, v/v). The product was dissolved in 10 mL MeOH: H₂O: HCOOH (6:3:1), Pd (OH)₂ (50% by weight) was added and the reaction mixture was hydrogenated for overnight. The reaction mixture was filtered through Celite and concentrated in *vacuo*. The residue was purified by Bio-Gel P-2 (BIO-RAD) column chromatography using water as eluent, and the product was lyophilized to get **20** (0.120 g, 70%) as a white color powder. ¹H NMR (400 MHz, D₂O): δ 7.08 (d, *J* = 8.4 Hz, 2H), 6.91 (d, *J* = 8.2 Hz, 2H), 5.38 (s, 1H), 4.66 (d, *J* = 8.0 Hz, 1H), 4.48 (d, *J* = 8.2 Hz, 1H), 4.38-4.27 (m, 2H), 4.19 (t, *J* = 2.8 Hz, 1H), 4.09 (d, *J* = 12.2 Hz, 1H), 3.98 (dd, *J* = 2.8 and 7.8 Hz, 1H), 3.84 (d, *J* = 12.3 Hz, 2H), 3.75 (s, 3H), 3.67-3.33 (m, 11H), 1.93 (s, 3H), 1.91 (s, 3H), 1.78 (s, 3H); ¹³C NMR (150 MHz, D₂O): δ 174.41, 173.93, 173.75, 154.47, 149.86, 118.53, 118.48, 114.93, 100.84, 99.71, 96.50, 76.76, 75.66, 73.75, 73.27, 72.74, 72.60, 69.76, 69.47, 69.24, 69.12, 67.28, 62.94, 60.62, 55.60, 55.32, 55.22, 22.30, 22.06, 20.03; ESI-MS: *m/z* calcd for C₃₁H₄₆N₂O₁₈; 735.2818 found 735.2769 (*M* + H)⁺.

Preparation of linear module. As depicted in scheme S23, the preparation of linear modules was commenced with enzymatic β -1,4-galctosylation of GlcNAc residue of acceptor **16** to form LacNAc moiety **21**. Modules **21** was then underwent action of α -1,3- fucosyltransferase, α -2,6-sialyltransferase, α -2,3- sialyltransferase, and α -1,3- fucosyltransferase to obtain modules **22**, **23**, **24**, and **25** respectively. Presence of α -1,3 fucose residue on GlcNAc restrict addition of sialic

acid on adjacent galactose at both 3 and 6 positions. Interestingly, α -2,3-sialylated LacNAc **24** was found to be the substrate of α -1,3- fucosyltransferase to get **25**, but not the α -2,6-sialylated LacNAc **23**. More striking to us was the α -1,2- fucosylated LacNAc **26** was the substrate of α -2,6- sialyltransferase, but not of α -2,3- sialyltransferase. Unique substrate specificities of FucTs and SiaTs allowed us to establish a rapid access to more diverse type of modules for *N*-glycan synthesis.



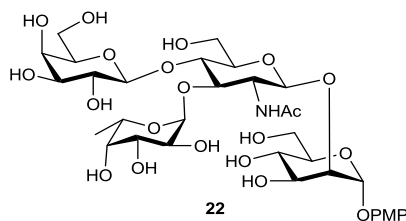
Scheme S23 | Preparation of linear modules. i, UDP-galactose, β 1, 4-GalT; ii, GDP-fucose, α 1, 3-FucT; iii, CMP-Neu5Ac, α 2, 6-SiaT; iv, CMP-Neu5Ac, α 2, 3-SiaT; v, GDP-fucose, α 1, 2-FucT.



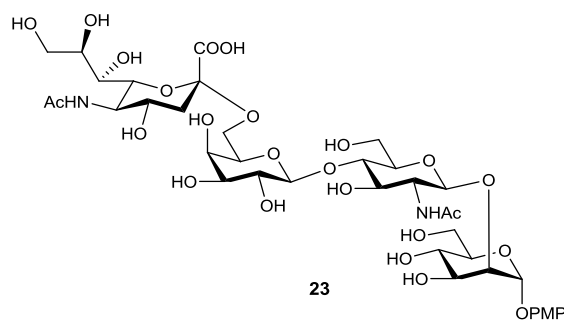
***p*-methoxyphenyl-O- β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-**

glucopyranosyl-(1 \rightarrow 2)- α -D-mannopyranoside (21): Compound **16** (100 mg, 0.204 mmol) was galactosylated by using general procedure **4** to get **21** (115 mg, 86 %) as amorphous white solids.

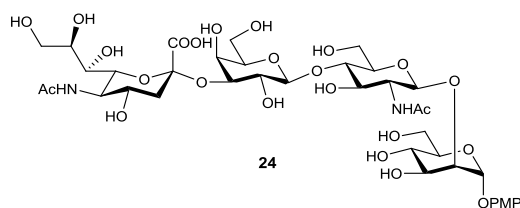
^1H NMR (400 MHz, D_2O): δ 7.10 (d, J = 8.8 Hz, 2H), 6.96 (d, J = 8.1 Hz, 2H), 5.44 (s, 1H), 4.59 (d, J = 8.4 Hz, 1H), 4.49 (d, J = 8.2 Hz, 1H), 4.45 (s, 1H), 4.00-3.75 (m, 17H), 3.60 (s, 3H), 2.01 (s, 3H); ESI-MS: m/z calcd for $\text{C}_{27}\text{H}_{41}\text{NO}_{17}$; 652.2447 found 674.2262 ($M + \text{H}$) $^+$.



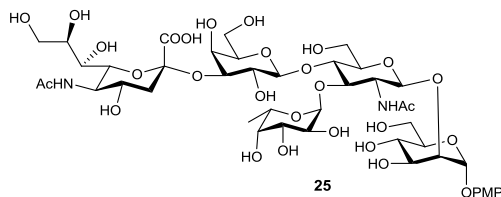
***p*-methoxyphenyl-O- β -D-galactopyranosyl-(1 \rightarrow 4)-[α -L-fucopyranosyl-(1 \rightarrow 3)-2-acetamido-2-deoxy- β -D-glucopyranosyl]-(1 \rightarrow 2)- α -D-mannopyranoside (22):** Compound **21** (80 mg, 0.122 mmol) was fucosylated by using general procedure **5** to afford **22** (82 mg, 84%) as white solid after lyophilization. ^1H NMR (400 MHz, D_2O): δ 7.15 (d, $J = 9.2$ Hz, 2H), 7.00 (d, $J = 9.1$ Hz, 2H), 5.45 (s, 1H), 5.12 (d, $J = 4.1$ Hz, 1H), 4.67 (d, $J = 7.8$ Hz, 2H), 4.44 (d, $J = 8.4$ Hz, 1H), 4.05 (s, 1H), 4.01 (dd, $J = 3.2$ and 7.8 Hz, 1H), 3.98 (t, $J = 10.2$ Hz, 2H), 3.80-3.70 (m, 6H), 3.79 (s, 3H), 3.70-3.40 (m, 11H), 2.06 (s, 3H), 1.18 (d, $J = 6.4$ Hz, 3H); ESI-MS: m/z calcd for $\text{C}_{33}\text{H}_{51}\text{NO}_{21}$; 797.7570 found 820.2837 ($M + \text{Na}$) $^+$.



***p*-methoxyphenyl-O-[5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylate-(2 \rightarrow 6)- β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl]-(1 \rightarrow 2)- α -D-mannopyranoside (23):** Compound **21** (150 mg, 0.184 mmol) was α 2,6-sialylated by using general procedure **3** to afford **23** (169 mg, 90%) as white solid after lyophilization. ^1H NMR (400 MHz, D_2O): δ 7.10 (d, $J = 9.2$ Hz, 2H), 7.08 (d, $J = 9.1$ Hz, 2H), 5.49 (s, 1H), 4.64 (d, $J = 8.1$ Hz, 1H), 4.40 (d, $J = 8.2$ Hz, 1H), 4.26 (t, $J = 2.1$ Hz, 1H), 4.01-3.80 (m, 7H), 3.74 (s, 3H), 3.66-3.58 (m, 17H), 2.63 (dd, $J = 4.4$ and 12.8 Hz, 1H), 2.07 (s, 3H), 1.97 (s, 3H), 1.68 (t, 1H); ESI-MS: m/z calcd for $\text{C}_{38}\text{H}_{58}\text{N}_2\text{O}_{25}$; 942.3245 found 941.3269 ($M - \text{H}$) $^-$.

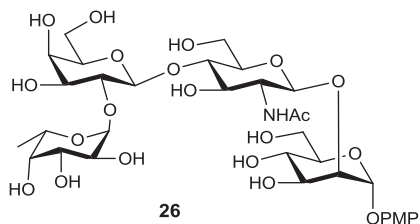


***p*-methoxyphenyl-O-[5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate-(2 \rightarrow 3)- β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 2)- α -D-mannopyranoside (24):** Compound **21** (10 mg, 15.3 μ mol) was α 2,3-sialylated by using general procedure **3** to afford **24** (11.5 mg, 80%) as white solid after lyophilization. ^1H NMR (400 MHz, D_2O): δ 7.16 (d, $J = 9.2$ Hz, 2H), 7.03 (d, $J = 9.5$ Hz, 2H), 5.49 (s, 1H), 4.67 (d, $J = 8.4$ Hz, 1H), 4.56 (d, $J = 8.4$ Hz, 1H), 4.32 (t, $J = 2.1$ Hz, 1H), 4.15 (dd, $J = 3.2$ & 7.5 Hz, 1H), 4.10-3.98 (m, 4H), 3.95-3.85 (m, 6H), 3.84 (s, 3H), 3.80-3.50 (m, 13H), 2.79 (dd, $J = 4.8$ and 12.1 Hz, 1H), 2.09 (s, 3H), 2.07 (s, 3H), 1.85 (t, 1H) ; ESI-MS: m/z calcd for $\text{C}_{38}\text{H}_{58}\text{N}_2\text{O}_{25}$; 942.3245 found 941.3312 ($M-H$) $^-$.

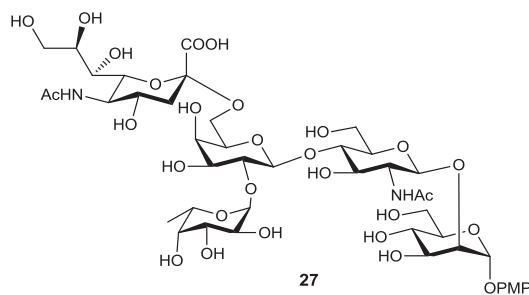


***p*-methoxyphenyl-O-[5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate-(2 \rightarrow 3)- β -D-galactopyranosyl-(1 \rightarrow 4)-[α -L-fucopyranosyl-(1 \rightarrow 3)-2-acetamido-2-deoxy- β -D-glucopyranosyl]-(1 \rightarrow 2)- α -D-mannopyranoside (25):** Compound **24** (8 mg, 8.5 μ mol) was fucosylated by using general procedure **5** to afford **25** (6.5 mg, 70%) as white solid after lyophilization. ^1H NMR (400 MHz, D_2O): δ 7.14 (d, $J = 6.8$ Hz, 2H), 6.98 (d, $J = 7.2$ Hz, 2H), 5.46 (s, 1H), 5.12 (d, $J = 4.2$ Hz, 1H), 4.67 (d, $J = 8.4$ Hz, 1H), 4.30 (t, $J = 2.3$ Hz, 1H), 4.10 (dd, $J = 3.0$ & 7.8 Hz, 1H), 4.00-3.80 (m, 14H), 3.79 (s, 3H), 3.75-3.50 (m, 13H), 2.75 (dd,

$J = 4.0$ and 12.0 Hz, 1H), 2.03 (s, 3H), 2.01(s, 3H), 1.76 (t, 1H), 1.17 (d, 3H, Fuc-Me); ESI-MS: m/z calcd for $C_{44}H_{68}N_2O_9$; 1088.3857 found 1087.3850 ($M - H$)⁻.



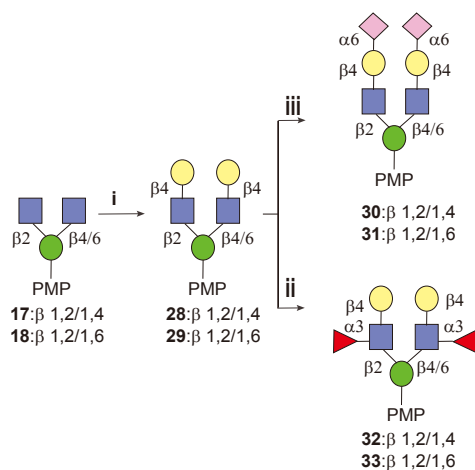
p-methoxyphenyl-O-[α -L-fucopyranosyl-(1 \rightarrow 2)- β -D-galactopyranosyl]-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl]-(1 \rightarrow 2)- α -D-mannopyranoside (26): Compound **21** (3 mg, 4.6 μ mol) was fucosylated by using general procedure **5** to afford **26** (2.1 mg, 53%) as white solid after lyophilization. 1H NMR (400 MHz, D_2O): δ 7.17 (d, $J = 7.2$ Hz, 2H), 7.01 (d, $J = 10.1$ Hz, 2H), 5.49 (s, 1H), 5.33 (s, 1H), 4.67 (d, $J = 8.2$ Hz, 1H), 4.56 (d, $J = 8.2$ Hz, 1H), 4.33 (m, 1H), 4.25 (q, 1H), 3.98 (dd, $J = 3.1$ and 7.2 Hz, 1H), 3.96 (dd, $J = 1.8$ and 7.2 Hz, 1H), 3.85-3.50 (m, 20H), 3.40 (m, 1H), 1.97 (s, 3H), 1.14 (d, $J = 6.4$ Hz, 3H) ; ESI-MS: m/z calcd for $C_{33}H_{51}NO_{21}$; 797.2954 found 820.2922 ($M + Na$)⁺.



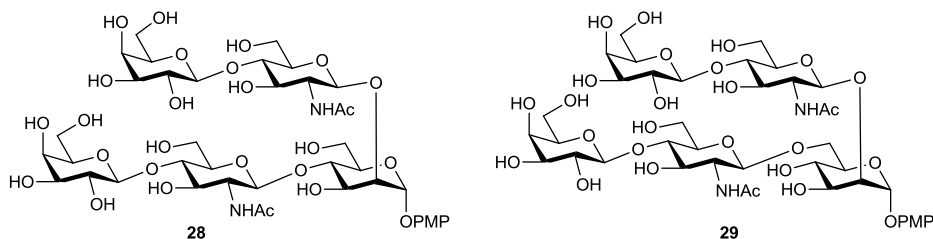
p-methoxyphenyl-O-[5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylate-(2 \rightarrow 6)-[α -L-fucopyranosyl-(1 \rightarrow 2)- β -D-galactopyranosyl]-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl]-(1 \rightarrow 2)- α -D-mannopyranoside (27): Compound **26** (2.0 mg, 2.7 μ mol) was α 2,6-sialylated by using general procedure **3** to afford **27** (1.8 mg, 66%) as white solid after lyophilization. 1H NMR (400 MHz, D_2O): δ 7.08 (d, $J = 9.5$ Hz, 2H), 6.91 (d, $J = 9.2$ Hz, 2H), 5.49 (s, 1H), 5.27 (s, 1H), 4.59 (d, $J = 8.2$ Hz, 1H), 4.47 (d, $J = 8.1$ Hz, 1H), 4.22 (bs, 1H), 4.12 (d, $J = 6.8$ Hz, 1H), 4.00-3.49 (m, 30H), 2.63 (dd, $J = 3.2$ and 12.0 Hz, 1H), 2.01(s,

3H), 1.97 (s, 3H), 1.67 (t, 1H), 1.19 (d, 3H, Fuc-Me) ; ESI-MS: m/z calcd for $C_{44}H_{68}N_2O_{29}$; 1088.3857 found 1087.3814 ($M - H$)⁻.

Preparation of symmetrically branched modules.



Scheme S24 | Preparation of symmetrically branched modules. i, β -1, 4 GalT, UDP-Gal; ii, α 1,3- fucosyltransferase, GDP-fucose; iii, α 2,6- sialyltransferase, CMP- β -D-Sialic acid; .



Compound 28 and 29: Compound 17 (10 mg, 14.3 μ mol) and 18 (10 mg, 14.3 μ mol) were galactosylated by using general procedure 4 to get 28 (10 mg, 71%) and 29 (9 mg, 61%) as amorphous white solids.

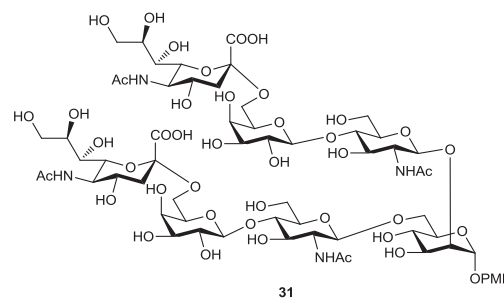
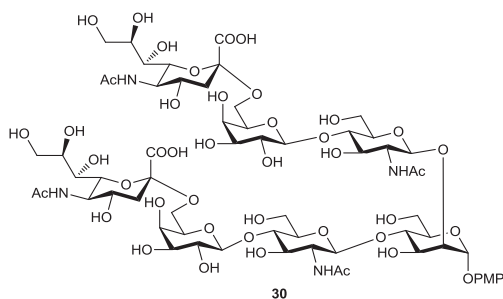
p-methoxyphenyl-O-di-[β -D-galactopyranosyl-(1 \rightarrow 4) -2-acetamido-2-deoxy- β -D-

glucopyranosyl]-(1 \rightarrow 2), (1 \rightarrow 4) - α -D-mannopyranoside (28): ¹H NMR (400 MHz, D₂O): δ 7.11 (d, J = 9.2 Hz, 2H), 6.95 (d, J = 8.5 Hz, 2H), 5.44 (s, 1H), 4.65 (d, J = 8.9 Hz, 1H), 4.58 (d, J = 9.2 Hz, 1H), 4.45 (dd, J = 3.8 & 9.2 Hz, 2H), 4.30 (t, J = 2.1 & 6.5 Hz, 1H), 4.18 (dd, J = 3.2

& 7.8 Hz, 1H), 4.00 (t, $J = 10.8$ Hz, 2H), 3.90 (d, $J = 8.2$ Hz, 2H), 3.78 (s, 3H), 3.75-3.64 (m, 20H), 3.58-3.49 (m, 4H), 2.02 (s, 3H), 2.01 (s, 3H); ESI-MS: m/z calcd for $C_{41}H_{64}N_2O_{27}$; 1016.9564 found 1039.3607 ($M + Na$)⁺.

p-methoxyphenyl-O-di-[β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-

glucopyranosyl]-(1 \rightarrow 2), (1 \rightarrow 6)- α -D-mannopyranoside (29): ¹H NMR (400 MHz, D₂O): δ 7.02 (d, $J = 9.0$ Hz, 2H), 6.92 (d, $J = 9.0$ Hz, 2H), 5.32 (s, 1H), 4.55 (d, $J = 4.3$ Hz, 1H), 4.41 (d, $J = 8.2$ Hz, 1H), 4.34 (dd, $J = 13.1, 7.9$ Hz, 2H), 4.16 (s, 1H), 4.00 (d, $J = 11.0$ Hz, 1H), 3.84-3.95 (m, 2H), 3.70-3.81 (m, 9H), 3.49-3.68 (m, 16H), 3.37-3.44 (m, 4H), 1.94 (s, 3H), 1.82 (s, 3H); ESI-MS: m/z calcd for $C_{41}H_{64}N_2NaO_{27}$; 1039.3594 found 1039.3551.

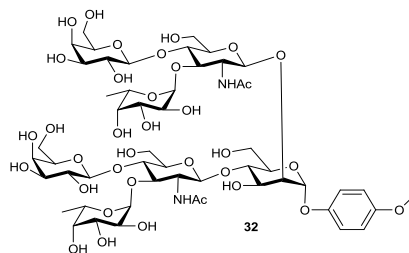


Compound 30 and 31: Compound 28 (8 mg, 7.3 μ mol) and 29 (8 mg, 7.3 μ mol) were sialylated by using general procedure 3 to afford 30 (9 mg, 78%) and 31 (7.5 mg, 65%) as a white solid after lyophilization.

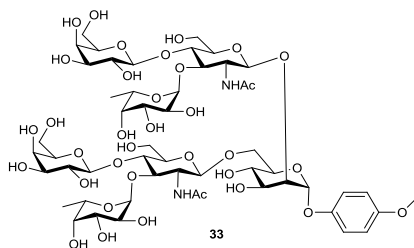
p-methoxyphenyl-O-di-[5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate-(2 \rightarrow 6)- β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl]-

(1 \rightarrow 2), (1 \rightarrow 4)- α -D-mannopyranoside (30): ¹H NMR (400 MHz, D₂O): δ 7.12 (d, $J = 9.8$ Hz, 2H), 6.98 (d, $J = 9.3$ Hz, 2H), 5.46 (s, 1H), 4.63 (d, $J = 8.4$ Hz, 1H), 4.58 (d, $J = 8.2$ Hz, 1H), 4.42 (d, $J = 7.8$ Hz, 2H), 4.32 (t, $J = 3.2$ Hz, 1H), 4.20 (dd, $J = 3.2$ & 7.2 Hz, 1H), 4.03-3.85 (m, 4H), 3.82-3.40 (m, 41H), 2.58 (dd, $J = 3.2$ & 10.3 Hz, 2H), 2.05 (s, 3H), 2.04 (s, 3H), 2.02 (s, 3H), 2.00 (s, 3H), 1.53 (m, 2H); ESI-MS: m/z calcd for $C_{63}H_{98}N_4O_{43}$; 1599.4620 found 798.2775 ($M - H$)²⁻.

***p*-methoxyphenyl-O-di-[5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate-(2 \rightarrow 6)- β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl]-(1 \rightarrow 2), (1 \rightarrow 6)- α -D-mannopyranoside (31):** ^1H NMR (600 MHz, D_2O): δ 7.05 (d, $J = 9.2$ Hz, 2H), 6.95 (d, $J = 9.2$ Hz, 2H), 5.36 (d, $J = 1.6$ Hz, 1H), 4.59 (d, $J = 8.2$ Hz, 1H), 4.46 (d, $J = 8.1$ Hz, 1H), 4.36 (d, $J = 7.9$ Hz, 1H), 4.32 (d, $J = 7.9$ Hz, 1H), 4.20 (dd, $J = 3.4, 1.8$ Hz, 1H), 4.06 (d, $J = 10.3$ Hz, 1H), 3.86-3.98 (m, 4H), 3.64-3.86 (m, 19H), 3.53-3.62 (m, 13H), 3.37-3.50 (m, 9H), 2.58 (ddd, $J = 12.0, 4.4, 2.1$ Hz, 2H), 1.98 (s, 3H), 1.94 (s, 3H), 1.94 (s, 3H), 1.87 (s, 3H), 1.63 (td, $J = 12.2, 8.6$ Hz, 2H); ESI-MS : m/z calcd for $\text{C}_{63}\text{H}_{98}\text{N}_4\text{NaO}_{43}$; 1621.5479 found 1621.5473.



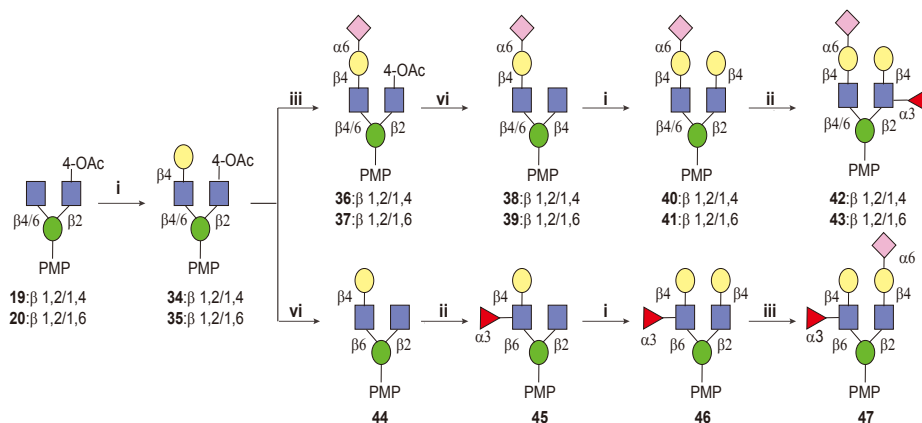
***p*-methoxyphenyl-O-di- $\{\beta$ -D-galactopyranosyl-(1 \rightarrow 4)-[α -L-fucopyranosyl-(1 \rightarrow 3)-2-acetamido-2-deoxy- β -D-glucopyranosyl]-(1 \rightarrow 2), (1 \rightarrow 4)- α -D-mannopyranoside (32):**
 Compound **28** (15 mg, 14.7 μmol) was fucosylated by using general procedure 5 to get **32** (14 mg, 73%) as amorphous white solids. ^1H NMR (400 MHz, D_2O): δ 7.13 (d, $J = 9.2$ Hz, 2H), 6.99 (d, $J = 9.0$ Hz, 2H), 5.44 (d, $J = 3.2$ Hz, 1H), 5.12 (d, $J = 4.1$ Hz, 1H), 5.10 (d, $J = 4.0$ Hz, 1H), 4.81 (d, $J = 8.4$ Hz, 2H), 4.68 (d, $J = 7.4$ Hz, 1H), 4.58 (d, $J = 7.2$ Hz, 1H), 4.45 (dd, $J = 3.2$ & 8.4 Hz, 2H), 4.31 (t, $J = 3.2$ Hz, 1H), 4.20 (dd, $J = 3.2$ & 8.4 Hz, 1H), 4.09-3.84 (m, 20H), 3.82 (s, 3H), 3.80-3.40 (m, 14H), 2.06 (s, 6H), 1.17 (d, $J = 6.4$ Hz, 6H); ESI-MS: m/z calcd for $\text{C}_{53}\text{H}_{84}\text{N}_2\text{O}_{35}$; 1308.4855 found 1309.4911 ($M + \text{H}$) $^+$.



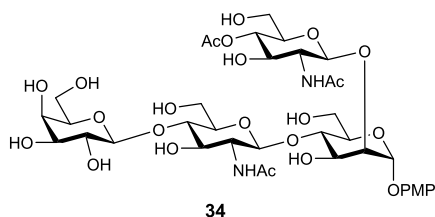
***p*-methoxyphenyl-O-di- $\{\beta$ -D-galactopyranosyl-(1 \rightarrow 4)- $\{\alpha$ -L-fucopyranosyl-(1 \rightarrow 3)-2-acetamido-2-deoxy- β -D-glucopyranosyl $\}$ -(1 \rightarrow 2), (1 \rightarrow 6)- α -D-mannopyranoside (33):**

Compound **29** (15 mg, 14.7 μ mol) was fucosylated by using general procedure 5 to get **33** (15.5 mg, 81%) as amorphous white solids. ^1H NMR (400 MHz, D_2O): δ 7.17 (d, $J = 8.2$ Hz, 2H), 7.06 (d, $J = 8.0$ Hz, 2H), 5.42 (s, 1H), 5.14 (d, $J = 4.0$ Hz, 1H), 5.09 (d, $J = 4.0$ Hz, 1H), 4.87-4.85 (m, 2H), 4.60 (d, $J = 7.2$ Hz, 1H), 4.56 (q, 2H), 4.30 (m, 1H), 4.15 (d, $J = 12.2$ Hz, 1H), 4.06 (dd, $J = 3.2$ & 7.8 Hz, 1H), 4.00-3.45 (m, 37), 2.06 (s, 3H), 1.95 (s, 3H), 1.19 (d, $J = 6.5$ Hz, 6H); ESI-MS : m/z calcd for $\text{C}_{53}\text{H}_{84}\text{N}_2\text{O}_{35}$; 1308.4855 found 1331.3561 ($M + \text{Na}$) $^+$.

Preparation of asymmetrically branched module.

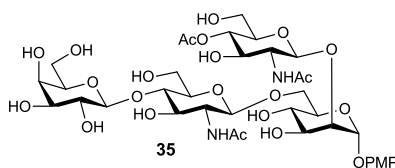


Scheme S25 | Preparation of assymetric module. UDP-galactose, β 1, 4-GalT; ii, GDP-fucose α 1, 3-FucT; iii, CMP-Neu5Ac, α 2, 6-SiaT; vi, NaOH.



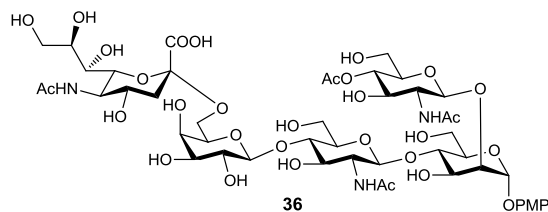
p-methoxyphenyl-O-(4-O-acetyl-2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 2)-O-(β -D-galactopyranosyl-(1 \rightarrow 4)-O-2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 4)- α -D-

mannopyranoside (34): Compound **19** (15 mg, 20.4 μ mol) was galactosylated using general procedure 4 to afford the **34** (12.5 mg, 68%) as amorphous white solids. ^1H NMR (600 MHz, D_2O): δ 7.01 (d, $J = 9.2$ Hz, 2H), 6.88 (d, $J = 9.2$ Hz, 2H), 5.34 (d, $J = 1.8$ Hz, 1H), 4.56 (d, $J = 8.5$ Hz, 1H), 4.45 (d, $J = 8.1$ Hz, 1H), 4.37 (d, $J = 7.7$ Hz, 1H), 4.20-4.33 (m, 2H), 4.16-4.20 (m, 1H), 4.07 (dd, $J = 8.7, 3.0$ Hz, 1H), 3.86-3.94 (m, 1H), 3.81 (d, $J = 3.4$ Hz, 1H), 3.31-3.78 (m, 21H), 2.03 (s, 3H), 1.93 (s, 3H), 1.92 (s, 3H); ESI-MS: m/z calcd for $\text{C}_{37}\text{H}_{56}\text{N}_2\text{NaO}_{23}$; 919.9166 found 919.3156.

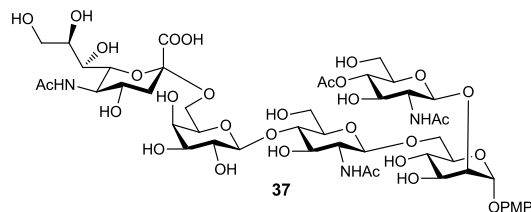


p-methoxyphenyl-O-(4-O-acetyl-2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 2)-O-(β -D-galactopyranosyl-(1 \rightarrow 4)-O-2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 6)- α -D-

mannopyranoside (S27a): Compound **20** (8 mg, 10.9 μ mol) was galactosylated by using general procedure 4 to give the **35** (5 mg, 75 %) as amorphous white solids. ^1H NMR (600 MHz, D_2O): δ 7.12 (d, $J = 9.1$ Hz, 2H), 7.01 (d, $J = 9.8$ Hz, 2H), 5.40 (s, 1H), 4.67 (d, $J = 8.4$ Hz, 1H), 4.49 (d, $J = 8.4$ Hz, 1H), 4.40 (d, $J = 8.3$ Hz, 2H), 4.31 (dd, $J = 3.1$ & 7.8 Hz, 1H), 4.21 (t, $J = 3.1$ Hz, 1H), 4.08 (d, $J = 12.3$ Hz, 2H), 4.00 (dd, $J = 3.2$ & 8.1 Hz, 1H), 3.90-3.48 (m, 3H), 3.80 (s, 3H), 3.75-3.47 (m, 15H), 2.15 (s, 3H), 2.0 (s, 3H), 1.98 (s, 3H); ESI-MS: m/z calcd for $\text{C}_{37}\text{H}_{56}\text{N}_2\text{O}_{23}$; 896.3162 found 919.3164 ($M + \text{Na}$) $^+$.

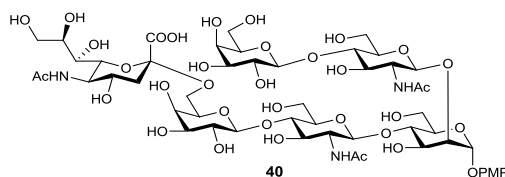


***p*-methoxyphenyl-O-(4-O-acetyl-2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 2)-O-(5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyrano-sylonate-(2 \rightarrow 6)- β -D-galactopyranosyl-(1 \rightarrow 4)-O-2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 4)- α -D-mannopyranoside (36):** Compound **34** (10 mg, 11.1 μ mol) was α 2,6-sialylated by using general procedure 3 to afford **36** (11.2 mg, 84%) as white solid after lyophilization. ^1H NMR (600 MHz, D_2O): δ 7.02 (d, J = 9.2 Hz, 2H), 6.89 (d, J = 9.2 Hz, 2H), 5.35 (d, J = 2.0 Hz, 1H), 4.55 (dd, J = 18.4, 8.4 Hz, 1H), 4.48 (d, J = 7.9 Hz, 1H), 4.17-4.37 (m, 4H), 4.05-4.12 (m, 1H), 3.29-3.97 (m, 30H), 2.56 (dd, J = 12.4, 4.7 Hz, 1H), 2.04 (s, 3H), 1.96 (s, 3H), 1.92 (s, 3H), 1.92 (s, 3H), 1.60 (t, J = 12.2 Hz, 1H); ESI-MS: m/z calcd for $\text{C}_{48}\text{H}_{72}\text{N}_3\text{O}_{31}$; 1186.4155 found 1186.4175.

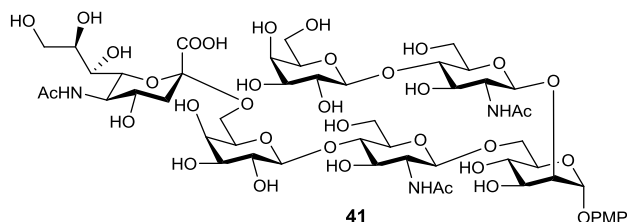


***p*-methoxyphenyl-O-(4-O-acetyl-2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 2)-O-(5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyrano-sylonate-(2 \rightarrow 6)- β -D-galactopyranosyl-(1 \rightarrow 4)-O-2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 6)- α -D-mannopyranoside (37):** Compound **35** was α 2,6-sialylated by using general procedure 3 to afford **37** (5.1 mg, 77%) as white solid after lyophilization. ^1H NMR (600 MHz, D_2O): δ 7.11 (d, J = 9.1 Hz, 2H), 7.00 (d, J = 9.2 Hz, 2H), 5.38 (s, 1H), 4.50 (d, J = 8.4 Hz, 1H), 4.36 (d, J = 8.2 Hz, 1H), 4.22 (t, J = 2.1 Hz, 1H), 4.10 (dd, J = 3.1 & 7.8 Hz, 1H), 3.97-3.85 (m, 4H), 3.77 (s, 3H), 3.73-3.46 (m, 26H), 2.63 (dd, J = 3.2 and 7.8 Hz, 1H), 2.09 (s, 3H), 2.01 (s, 3H), 1.99 (s,

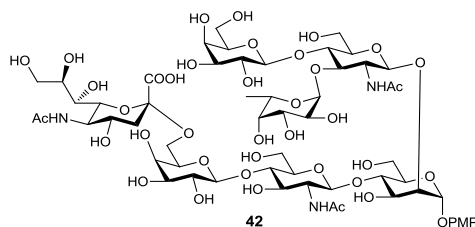
3H), 1.94 (s, 3H), 1.65 (t, 1H); ESI-MS: m/z calcd for $C_{48}H_{73}N_3O_{31}$; 1187.4144 found 1186.4133 ($M-H$)⁻.



***p*-methoxyphenyl-O-(β -D-galactopyranosyl-(1 \rightarrow 4)-O-2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 2)-O-(5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyrano-sylonate-(2 \rightarrow 6)- β -D-galactopyranosyl-(1 \rightarrow 4)-O-2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 4)- α -D-mannopyranoside (40):** To a solution of compound **36** (7 mg, 5.8 μ mol) in 0.5 mL H₂O was added NaOH (9.4 mg, 23.6 μ mol) and stirred for 4 h. Reaction was neutralized and product was purified by Bio-Gel P-2 chromatography (eluent H₂O) to afford **38** (6.1 mg, 89%) as white solid after lyophilization. Compound **38** (6 mg, 5.3 μ mol) and UDP galactose (6.4 mg, 10.6 μ mol) were dissolved in Tris buffer (25 mM, pH 7.5) and MnCl₂ (20 mM). GalT-1 (150 units) were added to achieve a final concentration of glycan to 5 mM. The resulting reaction mixture was incubated at 37 °C for 48 h. The reaction mixture was centrifuged and the supernatant subjected to gel filtration over P2-Biogel (eluent water). Fractions containing the product were combined and lyophilized to give the **40** (5 mg, 73%) as amorphous white solids. ¹H NMR (400 MHz, D₂O) δ 7.02 (d, J = 9.2 Hz, 2H), 6.88 (d, J = 9.2 Hz, 2H), 5.35 (d, J = 1.8 Hz, 1H), 4.56 (d, J = 7.9 Hz, 1H), 4.48 (d, J = 7.9 Hz, 1H), 4.35 (dd, J = 9.6, 7.7 Hz, 2H), 4.27-4.30 (m, 2H), 4.21-4.24 (m, 1H), 4.02-4.20 (m, 6H), 3.87-3.92 (m, 3H), 3.39-3.85 (m, 28H), 2.56 (dd, J = 12.4, 4.6 Hz, 1H), 1.96 (s, 3H), 1.93 (s, 3H), 1.92 (s, 3H), 1.60 (t, J = 12.2 Hz, 1H); ESI-MS: m/z calcd for $C_{52}H_{80}N_3O_{35}$; 1306.4578 found 1306.4617.

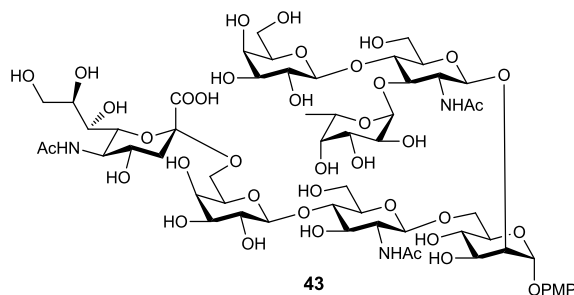


***p*-methoxyphenyl-O-(β -D-galactopyranosyl-(1 \rightarrow 4)-O-2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 2)-O-(5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyrano-sylonate-(2 \rightarrow 6)- β -D-galactopyranosyl-(1 \rightarrow 6)-O-2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 4)- α -D-mannopyranoside (41):** To a solution of compound **37** (5 mg, 4.2 μ mol) in 0.5 mL H₂O was added NaOH (6.7 mg, 16.8 μ mol) and stirred for 4 h. Reaction was neutralized and product was purified by Bio-Gel P-2 chromatography (eluent H₂O) to afford **39** (3.8 mg, 79%) as white solid after lyophilization. Compound **39** (3.5 mg, 3.0 μ mol) and UDP galactose (3.9 mg, 6.1 μ mol) were dissolved in Tris buffer (25 mM, pH 7.5) and MnCl₂ (20 mM). GalT-1 (150 units) were added to achieve a final concentration of glycan to 5 mM. The resulting reaction mixture was incubated at 37 °C for 48 h. The reaction mixture was centrifuged and the supernatant subjected to gel filtration over P2-Biogel (eluent water). Fractions containing the product were combined and lyophilized to give the **41** (3.3 mg, 75%) as amorphous white solids. ¹H NMR (400 MHz, D₂O): δ 7.11 (d, *J* = 9.1 Hz, 2H), 7.00 (d, *J* = 9.2 Hz, 2H), 5.38 (s, 1H), 4.64 (d, *J* = 8.1 Hz, 1H), 4.50 (d, *J* = 8.2 Hz, 1H), 4.23 (d, *J* = 8.3 Hz, 1H), 4.38 (d, *J* = 8.0 Hz, 1H), 4.22 (t, *J* = 2.1 Hz, 1H), 4.10 (d, *J* = 12.2 Hz, 1H), 4.01-3.95 (m, 4H), 3.87-3.63 (m, 26H), 3.52-3.45 (m, 7H), 2.63 (dd, *J* = 2.1 and 7.8 Hz, 1H), 2.63 (dd, *J* = 3.2 and 7.8 Hz, 1H), 2.01 (s, 3H), 1.99 (s, 3H), 1.93 (s, 3H), 1.67 (t, 1H); ESI-MS: *m/z* calcd for C₅₂H₈₁N₃O₃₅; 1307.4567 found 1306.4609 (*M - H*)⁻.

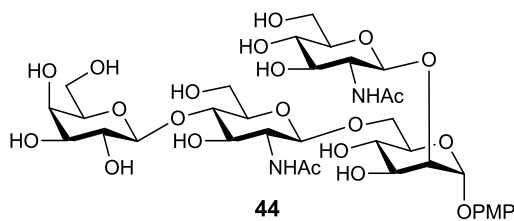


***p*-methoxyphenyl-O-(β -D-galactopyranosyl-(1 \rightarrow 4)-O-[α -L-fucopyranosyl-(1 \rightarrow 3)-2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 2)-O-(5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyrano-sylonate-(2 \rightarrow 6)- β -D-galactopyranosyl-(1 \rightarrow 4)-O-2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 4)- α -D-mannopyranoside (42):** Compound **40** (2 mg, 1.52

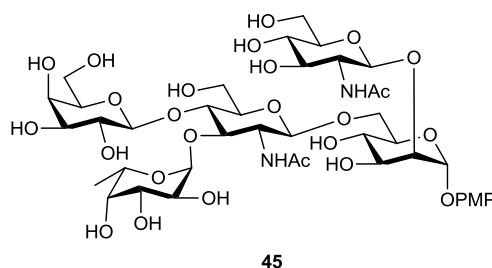
μmol) was α 1,3-fucosylated by using general procedure 5 to get **42** (2 mg, 85%) as amorphous white solids. ^1H NMR (400 MHz, D_2O): δ 7.16 (d, $J = 8.8$ Hz, 2H), 7.06 (d, $J = 8.1$ Hz, 2H), 5.45 (s, 1H), 5.08 (d, $J = 4.1$ Hz, 1H), 4.60 (d, $J = 4.2$ Hz, 1H), 4.44 (dd, $J = 3.5$ & 7.9 Hz, 2H), 4.30 (bs, 1H), 4.15 (d, $J = 8.4$ Hz, 1H), 4.09-3.40 (m, 43H), 2.70 (d, $J = 11.2$ Hz, 1H), 2.08 (s, 3H), 2.05 (s, 3H), 1.93 (s, 3H), 1.74 (t, 1H), 1.20 (d, $J = 6.4$ Hz, 3H); ESI-MS: m/z calcd for $\text{C}_{58}\text{H}_{91}\text{N}_3\text{O}_{39}$; 1453.5245 found 1452.5157 ($M - \text{H}$) $^-$.



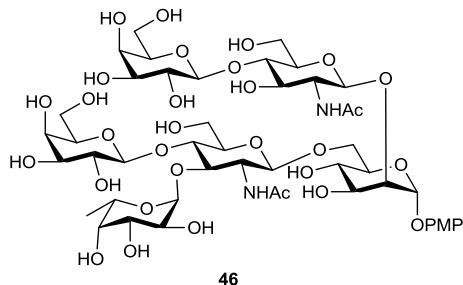
***p*-methoxyphenyl-O-(β -D-galactopyranosyl-(1 \rightarrow 4)-O-[α -L-fucopyranosyl-(1 \rightarrow 3)-2-acetamido-2-deoxy- β -D-glucopyranosyl]-(1 \rightarrow 2)-O-(5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate-(2 \rightarrow 6)- β -D-galactopyranosyl-(1 \rightarrow 4)-O-2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 6)- α -D-mannopyranoside (**43**):** Compound **41** (4 mg, 3.05 μmol) was α 1,3-fucosylated by using general procedure 5 to get **43** (3.2 mg, 72%) as amorphous white solids. ^1H NMR (400 MHz, D_2O): δ 7.21 (d, $J = 8.2$ Hz, 2H), 7.06 (d, $J = 8.0$ Hz, 2H), 5.74 (s, 1H), 5.72 (d, $J = 8.1$ Hz, 2H), 5.52 (dd, $J = 3.1$ & 7.2 Hz, 1H), 5.40 (s, 1H), 5.01 (d, $J = 4.2$ Hz, 1H), 4.50-3.25 (m, 44H), 2.50 (dd, $J = 3.1$ & 7.2 Hz, 1H), 2.01 (s, 3H), 1.99 (s, 6H), 1.56 (t, 1H), 1.05 (d, 3H, Fuc-Me); ESI-MS: m/z calcd for $\text{C}_{58}\text{H}_{91}\text{N}_3\text{O}_{39}$; 1453.5245 found 1452.5445 ($M - \text{H}$) $^-$.



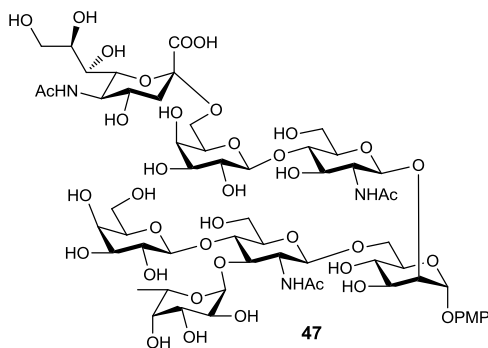
***p*-methoxyphenyl-O-(2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 2)-O-(β -D-galactopyranosyl-(1 \rightarrow 4)-O-2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 6)- α -D-mannopyranoside (44):** To a solution of compound **35** (20 mg, 22.3 μ mol) in 0.5 mL H₂O was added NaOH (3.57 mg, 89.2 μ mol) and stirred for 4 h. Reaction was neutralized and product was purified by Bio-Gel P-2 chromatography (eluent H₂O) to afford **44** (15.3 mg, 80%) as white solid after lyophilization. ¹H NMR (400 MHz, D₂O): δ 7.15 (d, *J* = 9.2 Hz, 2H), 7.06 (d, *J* = 9.8 Hz, 2H), 5.42 (s, 1H), 4.67 (d, *J* = 8.2 Hz, 1H), 4.54 (d, *J* = 8.0 Hz, 1H), 4.47 (d, *J* = 8.2 Hz, 1H), 4.32 (t, *J* = 2.1 Hz, 1H), 4.10 (d, *J* = 12.2 Hz, 1H), 4.09 (dd, *J* = 3.2 & 8.2 Hz, 1H), 4.01-3.91 (m, 4H), 3.85 (s, 3H), 3.80-3.40 (m, 17H), 2.03 (s, 3H), 1.98 (s, 3H); ESI-MS: *m/z* calcd for C₃₅H₅₄N₂O₂₂; 854.3125 found 877.3062 (*M* + Na)⁺.



***p*-methoxyphenyl-O-(2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 2)-O-(β -D-galactopyranosyl-(1 \rightarrow 4)-O-[α -L-fucopyranosyl-(1 \rightarrow 3)-2-acetamido-2-deoxy- β -D-glucopyranosyl]-1 \rightarrow 6)- α -D-mannopyranoside (45):** Compound **44** (8 mg, 9.3 μ mol) was α 1,3-fucosylated by using general procedure **5** to afford **45** (6 mg, 66%) as white solid after lyophilization. ¹H NMR (400 MHz, D₂O): δ 7.14 (d, *J* = 8.2 Hz, 2H), 7.02 (d, *J* = 8.1 Hz, 2H), 5.41 (s, 1H), 5.07 (d, *J* = 4.0 Hz, 1H), 4.64 (d, *J* = 8.2 Hz, 2H), 4.58 (d, *J* = 7.4 Hz, 1H), 4.44 (d, *J* = 8.5 Hz, 1H), 4.28 (t, *J* = 2.1 Hz, 1H), 4.10 (d, *J* = 12.2 Hz, 1H), 4.05 (dd, *J* = 3.1 & 7.2 Hz, 1H), 3.95-3.85 (m, 8H), 3.82 (s, 3H), 3.80-3.33 (m, 16H), 2.04 (s, 3H), 1.93 (s, 3H), 1.17 (d, *J* = 6.8 Hz, 3H); ESI-MS: *m/z* calcd for C₄₁H₆₄N₂O₂₆; 1000.2356 found 1023.3599 (*M* + Na)⁺.



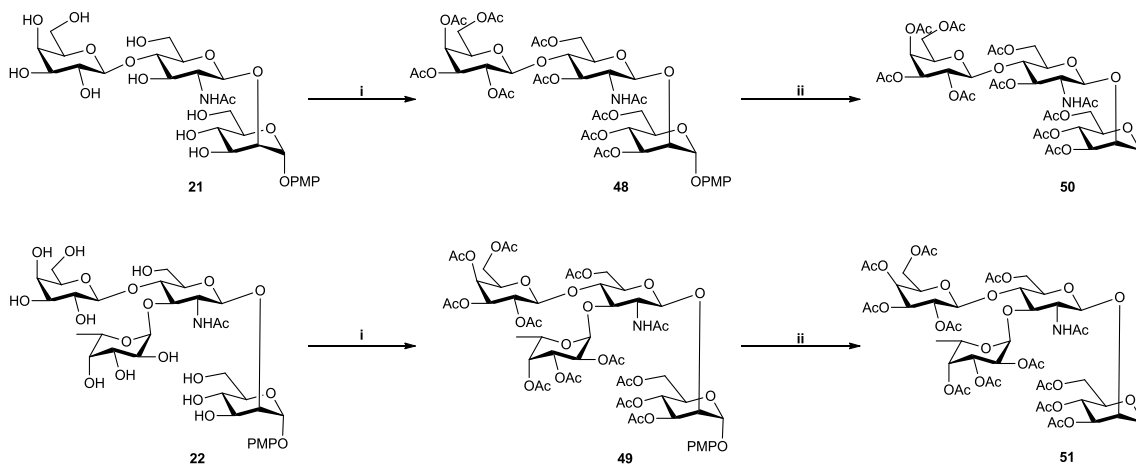
***p*-methoxyphenyl-O-(β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 2)-O-(β -D-galactopyranosyl-(1 \rightarrow 4)-O-[α -L-fucopyranosyl-(1 \rightarrow 3)-2-acetamido-2-deoxy- β -D-glucopyranosyl]-(1 \rightarrow 6)- α -D-mannopyranoside (46):** Compound **45** (7 mg, 7.2 μ mol) was β 1,4-galactosylated by using general procedure 4 to give the **46** (6.5 mg, 80 %) as amorphous white solids. ^1H NMR (400 MHz, D_2O): δ 7.16 (d, J = 9.4 Hz, 2H), 7.05 (d, J = 9.3 Hz, 2H), 5.43 (s, 1H), 5.09 (d, J = 4.2 Hz, 1H), 4.83 (d, J = 7.5 Hz, 1H), 4.78 (d, J = 7.2 Hz, 1H), 4.58 (d, J = 8.2 Hz, 1H), 4.48 (d, J = 9.5 Hz, 1H), 4.42 (d, J = 8.5 Hz, 1H), 4.30 (dd, J = 2.1 & 3.4 Hz, 1H), 4.14 (s, 1H), 4.09-3.92 (m, 14H), 3.85 (s, 3H), 3.84-3.50 (m, 17H), 2.06 (s, 3H), 1.95 (s, 3H), 1.20 (d, J = 6.5 Hz, 3H); ESI-MS: m/z calcd for $\text{C}_{47}\text{H}_{74}\text{N}_2\text{O}_{31}$; 1162.4309 found 1185.4076 ($M + \text{Na}$) $^+$.



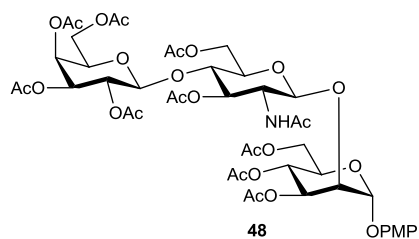
***p*-methoxyphenyl-O-(β -D-galactopyranosyl-(1 \rightarrow 4)-O-[α -L-fucopyranosyl-(1 \rightarrow 3)-2-acetamido-2-deoxy- β -D-glucopyranosyl]-(1 \rightarrow 6)-O-(5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyranosylonate-(2 \rightarrow 6)- β -D-galactopyranosyl-(1 \rightarrow 4)-O-2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 2)- α -D-mannopyranoside (47):** Compound **46** (5 mg, 4.31 μ mol) was α 2,6-sialylated by using general procedure 3 to get **47** (5.2 mg, 83%) as amorphous

white solids. ^1H NMR (400 MHz, D_2O): δ 7.08 (d, $J = 8.1$ Hz, 2H), 6.97 (d, $J = 8.0$ Hz, 2H), 5.48 (s, 1H), 5.10 (d, $J = 5.1$ Hz, 1H), 4.50 (d, $J = 7.2$ Hz, 1H), 4.43 (dd, $J = 3.1$ & 8.2 Hz, 2H), 4.32 (bs, 1H), 4.08 (d, $J = 8.1$ Hz, 1H), 4.09-3.40 (m, 43H), 2.60 (dd, $J = 3.2$ & 9.2 Hz, 1H), 1.97 (s, 3H), 1.95 (s, 3H), 1.90 (s, 3H), 1.54 (t, 1H), 1.18 (d, $J = 6.4$ Hz, 3H); ESI-MS: m/z calcd for $\text{C}_{58}\text{H}_{91}\text{N}_3\text{O}_{39}$; 1453.5245 found 1452.5101 ($M - \text{H}$) $^-$.

Chemical derivatization of chemo-enzymatically prepared modules. The modules **21** and **22** generated by chemo-enzymatic way were then peracetylated in presence of Ac_2O /pyridine. The reducing end *p*-methoxy phenyl ether protection was then cleaved using cerium ammonium nitrate and free hydroxyl was changed to fluoride to obtained donors **50** and **51** (Scheme S26).

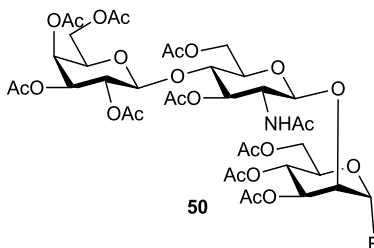


Scheme S26 | Reagents and conditions. i, Ac_2O , pyridine, RT, overnight; ii, (1) CAN, ACN: toluene: H_2O , (2) DAST, CH_2Cl_2 , -30 °C. CAN: Cerium ammonium nitrate; DAST: Diethylaminosulfur trifluoride.



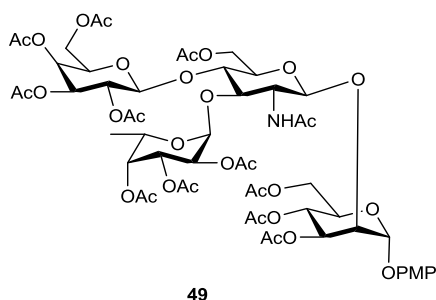
***p*-methoxyphenyl-O-[2,3,4,6-O-tetraacetyl- β -D-galactopyranosyl]-(1 \rightarrow 4)-[3,6-O-diacetyl-2-acetamido-2-deoxy- β -D-glucopyranosyl]-(1 \rightarrow 2)-3,4,6-O-triacetyl- α -D-mannopyranoside**

(48): To a solution of **21** (0.230 g, 0.360 mmol) in 10 mL pyridine at 0 °C was added acetic anhydride (6 mL) and stirred at rt for overnight. Reaction mixture was then concentrated, diluted with 50 mL of CH₂Cl₂ and extracted with sat. NaHCO₃. Combined organic layers were evaporated and product was purified by silica gel column chromatography (0% \rightarrow 20% acetone in toluene) to afford desired **48** (0.260 g, 72 %). TLC (acetone : toluene =3/7, v/v), R_f= 0.34; ¹H NMR (400 MHz, CHCl₃): δ 7.00 (d, *J* = 9.1 Hz, 2H), 6.82 (d, *J* = 9.1 Hz, 2H), 5.74 (d, *J* = 8.7 Hz, 1H), 5.33-5.35 (m, 2H), 5.27-5.30 (m, 2H), 5.22 (dd, *J* = 9.6, 8.2 Hz, 1H), 5.10 (dd, *J* = 10.5, 7.9 Hz, 1H), 4.96 (dd, *J* = 10.5, 3.4 Hz, 1H), 4.72 (d, *J* = 7.5 Hz, 1H), 4.47 (d, *J* = 7.9 Hz, 1H), 4.41 (dd, *J* = 11.8, 2.7 Hz, 1H), 4.31 (t, *J* = 2.2 Hz, 1H), 3.98-4.21 (m, 6H), 3.81-3.91 (m, 2H), 3.72-3.79 (m, 4H), 3.63 (ddd, *J* = 8.6, 5.6, 2.7 Hz, 1H), 2.14 (s, 3H), 2.11 (s, 3H), 2.08 (s, 3H), 2.06 (s, 2H), 2.03 (s, 3H), 2.03 (s, 6H), 2.02 (s, 3H), 1.96 (s, 6H); ESI-MS: *m/z* calcd for C₄₅H₅₉NO₂₆; 1029.9480 found 1030.4587 (*M* + H)⁺.



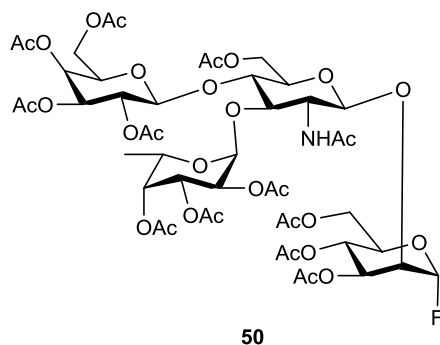
[2,3,4,6-O-tetraacetyl- β -D-galactopyranosyl]-(1 \rightarrow 4)-[3,6-O-diacetyl-2-acetamido-2-deoxy- β -D-glucopyranosyl]-(1 \rightarrow 2)-3,4,6-O-triacetyl- α -D-mannopyranosyl fluoride (50**):** Cerium ammonium nitrate (0.2 g, 0.554 mmol) was added to a solution of compound **48** (0.200 g, 0.203 mmol) in 10 mL of acetonitrile: toluene: H₂O (4:2:1). The resulting reaction mixture was stirred at RT for 3 h. The reaction was diluted with EtOAc (50 mL) and washed with H₂O (30 x 2 mL) and brine (30 mL). The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The product was purified by flash column chromatography (0% \rightarrow 25% acetone in toluene) to afford

-OH compound (0.120 g) as a foam. The residue (0.120 g, 0.130 mmol) was dissolved in CH₂Cl₂ (10 mL) at -30 °C. Then, DAST (34 μL, 0.260 mmol) was added slowly, and the resulting reaction mixture was stirred for 1 h. When TLC (acetone: toluene, 3/7) indicated formation of product with consumption of starting material, the reaction was quenched with aq. NaHCO₃. The filtrate was washed with aqueous NaHCO₃ (2 x 30 mL) and brine (20 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by silica gel column chromatography (0% → 20% acetone in toluene) to afford **50** (0.080 g, 51% over 2 steps) as white solid. TLC (acetone: toluene =3/7, v/v), R_f= 0.34; ¹H NMR (600 MHz, CHCl₃): δ 5.50 (d, *J* = 49.6 Hz, 1H), 5.30 (d, *J* = 3.8 Hz, 1H), 2.26 (t, *J* = 7.8 Hz, 1H), 5.05-4.89 (m, 4H), 4.55 (d, *J* = 8.4 Hz, 1H), 4.40 (d, *J* = 8.2 Hz, 1H), 4.32 (s, 2H), 4.18 (dd, *J* = 3.2 and 7.8 Hz, 1H), 4.05-4.01 (m, 6H), 3.81 (t, *J* = 7.8 Hz, 1H), 3.71 (t, *J* = 7.1 Hz, 1H), 3.50 (t, *J* = 3.1 Hz, 1H), 2.29 (s, 3H), 2.08 (s, 3H), 2.07 (s, 3H), 2.06 (s, 3H), 2.02 (s, 3H), 1.99 (s, 3H), 1.98 (s, 3H), 1.96 (s, 3H), 1.91 (s, 3H), 1.82 (s, 3H), ¹³C NMR (150 MHz, CHCl₃): 171.29, 171.10, 170.66, 170.56, 170.35, 170.30, 169.50, 129.21, 128.40, 125.47, 106.54, 104.31, 101.05, 100.31, 76.07, 72.82, 72.64, 72.36, 71.16, 70.95, 70.71, 69.41, 69.28, 66.86, 64.92, 62.38, 62.13, 61.04, 53.12, 23.13, 21.64, 21.22, 20.95, 20.87, 20.77, 20.68, 20.62; ESI-MS: *m/z* calcd for C₃₈H₅₂FNO₂₄; 925.8145 found 925.8354 (*M* +H)⁺.



***p*-methoxyphenyl-O-[2,3,4,6-O-tetraacetyl-β-D-galactopyranosyl]-(1→4)-[2,3,4-O-triacetyl-α-L-fucopyranosyl-(1→3)-3,6-O-diacetyl-2-acetamido-2-deoxy-β-D-glucopyranosyl]-(1→2)-3,4,6-O-triacetyl-α-D-mannopyranoside (49):** To a solution of **22** (0.100 g, 0.125 mmol) in 6 mL pyridine at 0 °C was added acetic anhydride (4 mL) and stirred at rt for overnight. Reaction

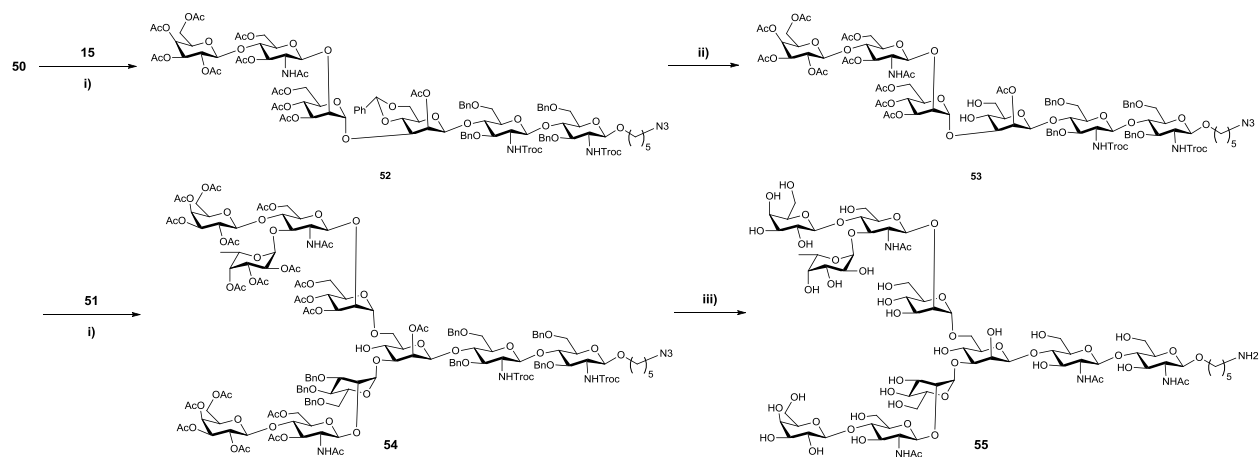
mixture was then concentrated, diluted with 50 mL of CH₂Cl₂ and extracted with sat. NaHCO₃. Combined organic layers were evaporated and product was purified by silica gel column chromatography (0% → 10% acetone in CH₂Cl₂) to afford desired **49** (0.135 g, 85 %). TLC (acetone: CH₂Cl₂ = 1/9, v/v), R_f = 0.31; ¹H NMR (400 MHz, CHCl₃): δ 7.00 (d, *J* = 6.8 Hz, 2H), 6.83 (d, *J* = 6.8 Hz, 2H), 5.43 (d, *J* = 3.2 Hz, 1H), 4.42 (d, *J* = 3.2 Hz, 1H), 3.98 (d, *J* = 2.1 Hz, 2H), 5.25 (dd, *J* = 3.2 & 8.2 Hz, 2H), 5.19 (dd, *J* = 3.8 & 8.5 Hz, 2H), 5.10 (t, *J* = 9.8 Hz, 2H), 4.89 (d, *J* = 6.3 Hz, 1H), 4.85 (d, *J* = 2.2 Hz, 1H), 4.70 (d, *J* = 9.8 Hz, 2H), 4.61 (d, *J* = 1.2 Hz, 1H), 4.40 (dd, *J* = 3.2 & 7.2 Hz, 1H), 4.31 (m, 3H), 4.22-3.89 (m, 6H), 3.80 (m, 2H), 3.78 (s, 3H), 3.60 (m, 1H), 2.10 (s, 3H), 2.08 (s, 3H), 2.08 (s, 3H), 2.02 (s, 3H), 2.00 (s, 3H), 1.99 (s, 3H), 1.98 (s, 3H), 1.96 (s, 3H), 1.93 (s, 3H), 1.90 (s, 3H), 1.89 (s, 3H), 1.88 (s, 3H), 1.11 (d, 3H); ESI-MS: *m/z* calcd for C₅₅H₇₃NO₃₂; 1259.4116 found 1282.3974 (*M* + Na)⁺.



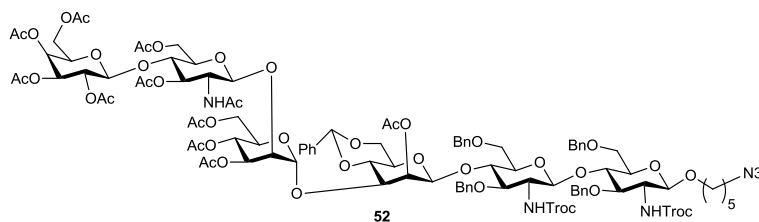
[2,3,4,6-O-tetraacetyl-β-D-galactopyranosyl]-(1→4)-[2,3,4-O-triacetyl-α-L-fucopyranosyl]-(1→3)-3,6-O-diacetyl-2-acetamido-2-deoxy-β-D-glucopyranosyl]-(1→2)-3,4,6-O-triacetyl-α-D-mannopyranosyl fluoride (50): Cerium ammonium nitrate (0.121 g, 0.142 mmol) was added to a solution of compound **49** (0.090 g, 0.071 mmol) in 7 mL of acetonitrile: toluene: H₂O (4:2:1). The resulting reaction mixture was stirred at rt for 3 h. The reaction was diluted with EtOAc (40 mL) and washed with H₂O (10 x 2 mL) and brine (10 mL). The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The product was purified by flash column chromatography (0% → 15% acetone in CH₂Cl₂) to afford **-OH** compound (0.065 g) as a foam. The residue (0.060 g,

0.052 mmol) was dissolved in CH₂Cl₂ (5 mL) at -30 °C. Then, DAST (11.6 μL, 0.104 mmol) was added slowly, and the resulting reaction mixture was stirred for 1 h. When TLC (acetone: CH₂Cl₂, 1/9) indicated formation of product with consumption of starting material, the reaction was quenched with aq. NaHCO₃. The filtrate was washed with aqueous NaHCO₃ (2 x 10 mL) and brine (10 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by silica gel column chromatography (0% → 10% acetone in CH₂Cl₂) to afford **51** (0.055 g, 67% over 2 steps) as white solid. TLC (acetone: CH₂Cl₂ = 1/9, v/v), R_f = 0.44; ¹H NMR (400 MHz, CHCl₃): δ 5.55 (d, *J* = 52.1 Hz, 1H), 5.40 (dt, *J* = 3.2 & 7.8 Hz, 3H), 5.21 (dd, *J* = 2.1 & 8.2 Hz, 2H), 5.18-5.04 (m, 3H), 5.0 (bs, 1H), 4.60 (s, 2H), 4.40 (dd, *J* = 5.6 & 8.5 Hz, 1H), 4.38-4.00 (m, 7H), 3.80 (m, 2H), 3.60 (m, 1H), 2.16 (s, 3H), 2.14 (s, 3H), 2.13 (s, 3H), 2.10 (s, 3H), 2.09 (s, 3H), 2.07 (s, 3H), 2.06 (s, 3H), 2.06 (s, 3H), 2.05 (s, 3H), 2.04 (s, 3H), 2.00 (s, 3H), 1.95 (s, 3H), 1.94 (s, 3H), 1.17 (d, *J* = 6.4 Hz, 3H) ; ESI-MS: *m/z* calcd for C₄₈H₆₆FNO₃₀; 1155.0314 found 1178.3491 (*M* + Na)⁺.

Chemical glycosylation to core trisaccharide: At this stage we are in position to investigate the coupling efficiency of fluoride donors **50** and **51** to core trisaccharide **15**. Depicted in Scheme S27, stereoselective conjugation of **50** to 3-O position of core in presence of AgOTf/Cp₂HfCl₂ provided expected hexasaccharide **52** in 56% yield. The cleavage of benzylidene ring using catalytic *p*-toluene sulfonic acid provided 4, 6-diol **53**. Taking advantage of its reactivity, primary hydroxyl group of **53** was then selectively glycosylated with **51** to afford fully protected decasaccharide **54** in moderate yield. At last, the global deprotection afforded the desired selectively fucosylated bi-antennary complex type glycan **55**.

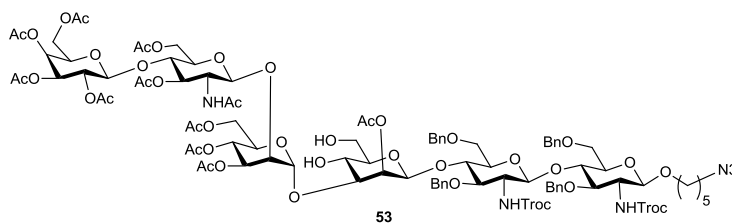


Scheme S27 | Reagents and conditions. i, AgOTf, Cp₂HfCl₂, toluene, 4 Å MS, 0 °C to rt; ii, *p*-TSA, acetonitrile, rt; iii, (1) LiOH, 1,4-dioxane: H₂O; 90 °C, overnight; (2) Ac₂O, pyridine, overnight; (3) NaOMe, MeOH, overnight; (4) Pd(OH)₂, MeOH: H₂O: HCOOH (5:3:2), H₂. AgOTf: Silver trifluoromethanesulfonate; Cp₂HfCl₂: Bis(cyclopentadienyl)hafnium dichloride, MS: molecular sieves.

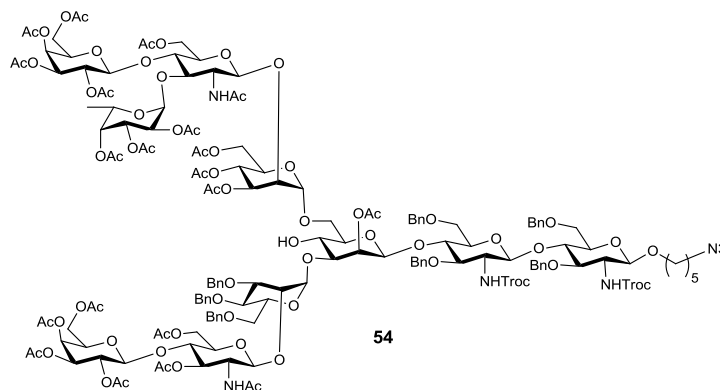


5-Azidopentyl-O-[[2,3,4,6-O-tetraacetyl-β-D-galactopyranosyl]-(1→4)-[3,6-O-diacetyl-2-acetamido-2-deoxy-β-D-glucopyranosyl]-(1→2)-[3,4,6-O-triacetyl-α-D-mannopyranosyl]]-(1→3)-[2-O-acetyl-4,6-O-benzylidene-β-D-mannopyranosyl-(1→4)-O-(3,6-di-O-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino-β-D-glucopyranosyl)-(1→4)-O-3,6-di-O-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino-β-D-glucopyranoside (52). A mixture of Silver triflate (0.039 g, 0.155 mmol), Bis (cyclopentadienyl) hafnium dichloride (0.041 g, 0.108 mmol) and 4 Å activated molecular sieves in dry toluene (3 mL) was stirred at rt

for 1 h. The reaction mixture was then cooled to 0 °C, a solution of donor **50** (0.043 g, 0.046 mmol) and acceptor **15** (0.045 g, 0.031 mmol) in 3 mL toluene was added. The mixture was stirred at RT for 3 h, quenched with Et₃N, diluted with CH₂Cl₂ and filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 20 mL), and a brine (20 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 10% acetone in CH₂Cl₂) to afford **52** (0.051 g, 70%) as colorless foam. TLC: (acetone: CH₂Cl₂ = 1.5/8.5, v/v): R_f = 0.46; ¹H NMR (400 MHz, CHCl₃): δ 7.46-7.16 (m, 25H), 5.41 (s, 1H), 5.32 (d, *J* = 9.8 Hz, 1H), 5.29 (d, *J* = 7.2 Hz, 1H), 5.20 (t, *J* = 10.2 Hz, 1H), 5.09 (t, *J* = 10.3 Hz, 1H), 4.93-4.87 (m, 5H), 4.82-4.57 (m, 5H), 4.50 (d, *J* = 12.2 Hz, 2H), 4.41-4.22 (m, 4H), 4.13-3.56 (m, 8H), 3.43-3.36 (m, 4H), 3.20 (t, *J* = 10.3 Hz, 4H), 3.19 (t, *J* = 10.2 Hz, 1H), 2.20 (s, 3H), 2.18 (s, 3H), 2.13 (s, 3H), 2.06 (s, 3H), 2.05 (s, 3H), 2.04 (s, 3H), 2.03 (s, 3H), 1.98 (s, 3H), 1.96 (s, 3H), 1.90 (s, 3H), 1.81 (s, 3H), 1.58-1.51 (m, 4H), 1.40-1.34 (m, 2H); ¹³C NMR (150 MHz, CHCl₃): δ 171.28, 170.80, 170.69, 170.49, 170.43, 170.36, 169.97, 169.68, 169.41, 154.30, 154.13, 138.83, 138.18, 137.44, 131.28, 129.25, 129.11, 128.91, 128.63, 128.54, 128.37, 128.18, 128.10, 128.00, 127.79, 127.71, 126.47, 102.55, 101.37, 100.84, 100.44, 98.60, 95.82, 76.07, 74.7474.61, 74.51, 74.19, 73.83, 73.59, 72.35, 71.24, 70.95, 70.79, 70.03, 69.62, 69.52, 69.37, 68.66, 68.42, 66.92, 66.41, 65.51, 62.42, 62.02, 61.15, 57.88, 57.23, 53.60, 51.58, 29.27, 28.82, 23.46, 23.29, 21.10, 20.96, 20.82; ESI-MS: *m/z* calcd for C₁₀₄H₁₂₆Cl₆N₆O₄₁; 2360.8510 found 2361.6131 (*M* + H)⁺.



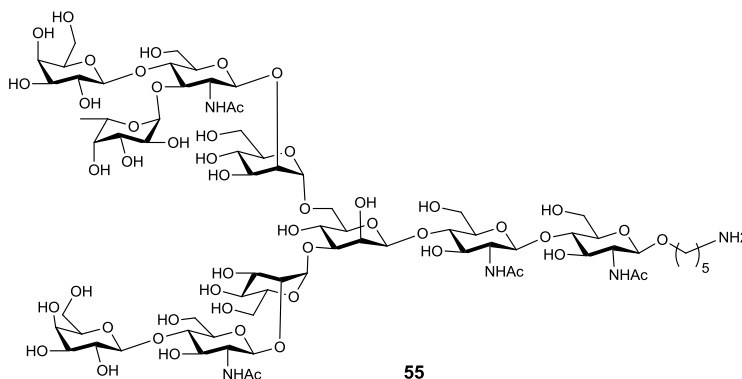
5-Azidopentyl-O-[[2,3,4,6-O-tetraacetyl- β -D-galactopyranosyl]-(1 \rightarrow 4)-[3,6-O-diacetyl-2-acetamido-2-deoxy- β -D-glucopyranosyl]-(1 \rightarrow 2)-[3,4,6-O-triacetyl- α -D-mannopyranosyl]]-(1 \rightarrow 3)-[2-O-acetyl- β -D-mannopyranosyl]-(1 \rightarrow 4)-O-(3,6-di-O-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino- β -D-glucopyranosyl]-(1 \rightarrow 4)-O-3,6-di-O-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino- β -D-glucopyranoside (53**):** *p*-Toluene sulfonic acid monohydrate (0.001 g, 0.008 mmol) was added to a solution of **52** (0.040 g, 0.016 mmol) in acetonitrile: MeOH 2/1 (3 mL) and the resulting reaction mixture was stirred at RT for 5 h. The reaction was quenched by adding Et₃N and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% \rightarrow 15% acetone in CH₂Cl₂) to give diol **53** (0.022 g, 57%). TLC: (acetone: CH₂Cl₂ = 1.5/8.5, v/v): R_f = 0.32; ¹H NMR (400 MHz, CDCl₃): δ 7.43-7.17 (m, 20H), 5.33 (d, *J* = 3.2 Hz, 1H), 5.25 (d, *J* = 3.2 Hz, 1H), 5.19-5.04 (m, 4H), 4.98 (d, *J* = 3.2 Hz, 1H), 4.95 (d, *J* = 3.1 Hz, 1H), 4.94-4.89 (m, 2H), 4.72-4.55 (m, 8H), 4.50-4.28 (m, 9H), 4.20-3.30 (m, 30H), 3.20 (t, 5H), 2.15 (s, 3H), 2.14 (s, 3H), 2.12 (s, 3H), 2.09 (s, 3H), 2.07 (s, 3H), 2.06 (s, 3H), 2.04 (s, 3H), 2.03 (s, 3H), 2.02 (s, 3H), 2.01 (s, 3H), 2.00 (s, 3H), 1.95 (s, 3H), 1.94 (s, 3H), 1.56-1.51 (m, 4H), 1.40-1.34 (m, 2H); ESI-MS: *m/z* calcd for C₉₇H₁₂₂Cl₆N₆O₄₃; 2272.7420 found 2295.5522 (*M* + Na)⁺.



5-Azidopentyl-O-[[2,3,4,6-O-tetraacetyl- β -D-galactopyranosyl]-(1 \rightarrow 4)-[3,6-O-diacetyl-2-acetamido-2-deoxy- β -D-glucopyranosyl]-(1 \rightarrow 2)-[3,4,6-O-triacetyl- α -D-mannopyranosyl]]-(1 \rightarrow 3)-[2,3,4,6-O-tetraacetyl- β -D-galactopyranosyl]-(1 \rightarrow 4)-[2,3,4-O-triacetyl- α -L-

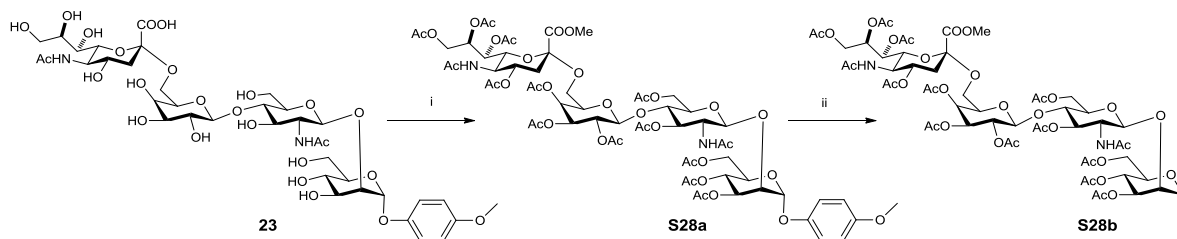
fucopyranosyl-(1→3)-3,6-O-diacetyl-2-acetamido-2-deoxy-β-D-glucopyranosyl]-(1→2)-3,4,6-O-triacetyl-α-D-mannopyranosyl}-(1→6)-[2-O-acetyl-β-D-mannopyranosyl-(1→4)-O-(3,6-di-O-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino-β-D-glucopyranosyl)-(1→4)-O-3,6-di-O-benzyl-2-deoxy-2-(2,2,2-trichloroethoxy)carbonylamino-β-D-

glucopyranoside (53): A mixture of Silver triflate (0.011 g, 44.1 μmol), Bis(cyclopentadienyl) hafnium dichloride (0.012 g, 30.8 μmol) and 4 Å activated molecular sieves in dry toluene (3 mL) was stirred at rt for 1 h. The reaction mixture was then cooled to 0 °C, a solution of donor **50** (0.015 g, 13.2 μmol) and acceptor **15** (0.020 g, 8.80 μmol) in 2 mL toluene was added. The mixture was stirred at RT for 5 h, quenched with Et₃N, diluted with CH₂Cl₂ and filtered through Celite. The filtrate was washed with aqueous NaHCO₃ (2 x 10 mL), and a brine (20 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by flash column chromatography (0% → 15% acetone in CH₂Cl₂) to afford **52** (0.010 g, 34%) as colorless foam. TLC: (acetone: CH₂Cl₂ = 1.5/8.5, v/v): R_f = 0.41; ¹H NMR (400 MHz, CHCl₃): δ 7.48-7.20 (m, 20H), 5.40-4.51 (m, 24H), 4.50-4.00 (m, 10H), 3.98-3.50 (m, 30H), 3.49-3.10 (m, 20H), 2.18 (s, 3H), 2.16 (s, 3H), 2.15 (s, 3H), 2.14 (s, 3H), 2.13 (s, 3H), 2.12 (s, 3H), 2.11 (s, 3H), 2.10 (s, 3H), 2.09 (s, 3H), 2.07 (s, 3H), 2.06 (s, 3H), 2.05 (s, 3H), 2.04 (s, 3H), 2.03 (s, 3H), 2.02 (s, 3H), 2.01 (s, 3H), 2.00 (s, 3H), 1.99 (s, 3H), 1.97 (s, 3H), 1.95 (s, 3H), 1.94 (s, 3H), 1.93 (s, 3H), 1.33-1.23 (m, 4H), 1.20-1.15 (m, 2H); ESI-MS: *m/z* calcd for C₁₄₅H₁₈₇Cl₆N₇O₇₃; 3408.7670 found 1176.3855 (*M* + K)³⁺.

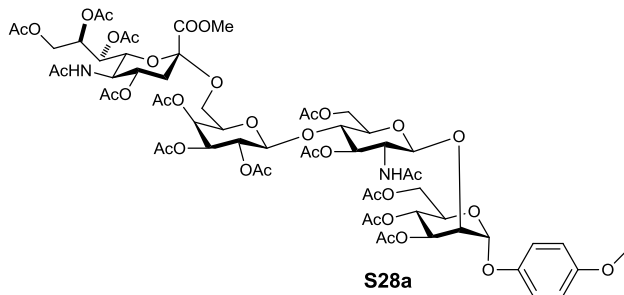


5-Aminopentyl- β -D-galactopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 2)- α -D-mannopyranosyl-(1 \rightarrow 3),-[β -D-galactopyranosyl-(1 \rightarrow 4)-(α -L-fucopyranosyl-(1 \rightarrow 3)-2-acetamido-2-deoxy- β -D-glucopyranosyl)-(1 \rightarrow 2)- α -D-mannopyranosyl-(1 \rightarrow 6)- β -D-mannopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranosyl-(1 \rightarrow 4)-2-acetamido-2-deoxy- β -D-glucopyranoside (55): A mixture of **54** (0.010 g, 2.9 μ mol) and lithium hydroxide (0.005 g, 50% by wt) in 2 mL of 1, 4 dioxane: H₂O (4:1) was stirred at 90 °C for overnight. Volatiles were then evaporated and the crude product was reacted with 3 mL Ac₂O: pyridine (1:2) for overnight. The solvents were removed using high vacuum and product was purified by C18 gel column chromatography (MeOH: H₂O as an eluent). The product was de-acetylated using sodium methoxide in MeOH (3 mL) for overnight. The reaction mixture was neutralized by using IR-120, filtered and concentrated in *vacuo*. The residue was purified by C18 gel column chromatography (MeOH: H₂O as an eluent). The product was dissolved in 3 mL MeOH: H₂O: HCOOH (6:3:1), Pd(OH)₂ (50% by weight) was added and the mixture was hydrogenated for overnight. The reaction mixture was filtered through Celite and concentrated in *vacuo*. The residue was purified by Bio-Gel P-2 (BIO-RAD) column chromatography using water as eluent. The product was the lyophilized to get desired oligosaccharides **55** (0.002 g, 36%) as a white color powder. TLC: (n-BuOH: AcOH: H₂O = 1/1/1, v/v): R_f = 0.32; ¹H NMR (400 MHz, D₂O): δ 5.15 (d, *J* = 4.2 Hz, 1H), 4.94 (s, 1H), 4.60 (m, 3H), 4.45 (q, 3H), 4.30 (s, 1H), 4.21 (d, *J* = 2.1 Hz, 1H), 4.13 (d, *J* = 2.0 Hz, 1H), 4.05-3.40 (m, 58H), 3.01 (t, 2H, linker), 2.10 (s, 3H, -Ac), 2.07 (s, 3H, -Ac), 2.06 (s, 3H, -Ac), 2.05 (s, 3H, -Ac), 1.76-1.58 (m, 4H, linker), 1.45-1.36 (m, 2H, linker), 1.21 (d, *J* = 6.5 Hz, 3H, Fuc-Me); ¹³C NMR (100 MHz, CHCl₃): δ 174.6, 174.4, 102.9, 101.8, 101.0, 99.4, 99.1, 98.5, 96.8, 80.3, 79.6, 79.3, 78.4, 76.4, 75.3, 75.2, 74.7, 74.6, 74.5, 74.3, 73.5, 72.8, 72.4, 71.9, 71.0, 70.9, 70.0, 69.5, 69.4, 69.1, 68.5, 68.3, 67.7, 67.2, 66.7, 65.8, 61.4, 60.9, 59.6, 59.5, 55.6, 54.9, 54.8, 39.3, 28.0, 26.3, 22.3, 22.2, 22.0, 15.3; ESI-MS: *m/z* calcd for C₇₃H₁₂₅N₅O₅₀; 1871.7545 found 1872.7477 (*M* + H)⁺.

Chemical derivatization of sialylated module. Preparation of sialylated antennae is of particular importance due to the complexity associated with sialic acid chemistry. Having established the synthetic protocol for non-sialylated modules (Scheme S28), we turned our attention to sialylated module **22**. The carboxylic acid functionality was esterified in presence of trimethylsilyl diazomethane without affecting free hydroxyl groups. Next, the peracetylation was performed using acetic anhydride and pyridine to afford **S30a**. The peracetylation step allows for purification of the oligosaccharide material before the removal of anomeric groups. At last, reducing end transformation was done to form anomeric fluoride donor **S30b**.



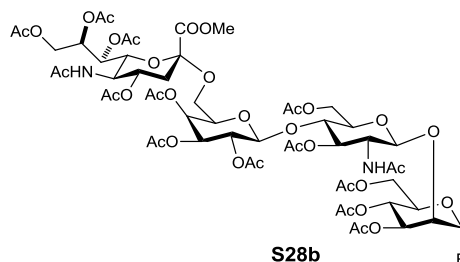
Scheme S28 | Reagents and Conditions. i, (1) Trimethylsilyl diazomethane, MeOH; (2) Ac₂O, pyridine, RT, overnight; ii, (1) CAN, ACN: toluene: H₂O, (2) DAST, CH₂Cl₂, -30 °C, 34% over 2 steps.



***p*-methoxyphenyl-O-[3,7,8,9-O-tetraacetyl-5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyrano-sylonate]-(2→6)-O-[2,3,4,6-O-tetraacetyl- β -D-galactopyranosyl]-(1→4)-[3,6-O-diacetyl-2-acetamido-2-deoxy- β -D-glucopyranosyl]-(1→2)-3,4,6-O-triacetyl- α -D-mannopyranoside (**S28a**):** To a solution of compound **23** (30 mg, 0.031 mmol) in MeOH (3 mL) was added trimethylsilyl diazomethane (1M solution in hexane, 0.046 mL, 0.31 mmol).

The resulting reaction mixture was stirred at RT for overnight. Complete consumption of starting material was confirmed by NMR and mass. The reaction mixture was concentrated to dryness and purified by C18 (eluent MeOH: H₂O). TLC: (n-butanol: H₂O: acetic acid = 2/1/1, v/v): R_f = 0.45; ¹H NMR (400 MHz, MeOD): δ 7.14 (d, *J* = 9.5 Hz, 2H), 6.97 (d, *J* = 9.8 Hz, 2H), 5.50 (s, 1H), 4.68 (d, *J* = 8.5 Hz, 1H), 4.45 (d, *J* = 8.0 Hz, 1H), 4.36 (s, 1H), 4.20-3.90 (m, 10H), 3.84 (s, 3H), 3.76-3.58 (m, 17H), 3.51 (s, 3H), 2.66 (dd, *J* = 3.8 and 11.5 Hz, 1H), 2.00 (s, 3H), 1.96 (s, 3H), 1.60 (t, 1H); ESI-MS: *m/z* calcd for C₃₉H₆₀N₂O₂₅; 942.3245 found 987.3033 (*M* + Na)⁺.

A solution of methyl ester (0.025 g, 0.025 mmol) in pyridine (2 mL) and acetic anhydride (1.5 mL) was stirred at rt for overnight. Reaction was then concentrated to dryness, dissolved in dichloromethane, extracted with sat. NaHCO₃ solution, dried and concentrated. Product was purified by C18 column (MeOH: water eluent) to afford **S28a** (0.030 g, 74%) as a white solid. TLC: (acetone: CH₂Cl₂ 4/6, v/v): R_f = 0.42; ¹H NMR (400 MHz, MeOD): δ 7.07 (d, *J* = 8.5 Hz, 2H), 6.88 (d, *J* = 8.9 Hz, 2H), 5.45 (s, 1H), 5.40 (d, *J* = 7.2 Hz, 1H), 5.36 (t, *J* = 10.3 Hz, 1H), 5.30 (d, *J* = 7.5 Hz, 1H), 5.28 (d, *J* = 7.0 Hz, 1H), 5.26 (s, 1H), 5.23 (s, 1H), 5.19 (d, *J* = 8.6 Hz, 1H), 5.17 (t, *J* = 10.3 Hz, 1H), 5.11-5.07 (m, 5H), 5.00-4.95 (m, 3H), 4.70 (dd, *J* = 4.2 & 7.8 Hz, 2H), 4.46-4.35 (m, 4H), 4.30-4.13 (m, 3H), 4.08-3.97 (m, 8H), 3.76 (s, 3H), 3.75 (s, 3H), 3.60-3.56 (m, 3H), 2.58 (dd, *J* = 4.2 & 8.5 Hz, 1H), 2.13 (s, 3H), 2.12 (s, 3H), 2.11 (s, 3H), 2.10 (s, 6H), 2.07 (s, 3H), 2.05 (s, 6H), 2.02 (s, 6H), 1.99 (s, 3H), 1.98 (s, 3H), 1.91 (s, 6H), 1.84 (t, *J* = 10.2 Hz, 1H); ESI-MS: *m/z* calcd for C₆₃H₈₄N₂O₃₇; 1461.3420 found 1486.4864 (*M* + 2H + Na)⁺.



***p*-methoxyphenyl-O-[3,7,8,9-O-tetraacetyl-5-Acetamido-3,5-dideoxy-D-glycero- α -D-galacto-2-nonulopyrano-sylonate]-(2→6)-O-[2,3,4,6-O-tetraacetyl- β -D-galactopyranosyl]-**

(1→4)-[3,6-O-diacetyl-2-acetamido-2-deoxy-β-D-glucopyranosyl]-(1→2)-3,4,6-O-triacetyl-α-D-mannopyranosyl fluoride (S28b): Cerium ammonium nitrate (0.037 g, 0.065 mmol) was added to a solution of compound **S28a** (0.020 g, 0.013 mmol) in 2 mL of acetonitrile: toluene: H₂O (4:2:1). The resulting reaction mixture was stirred at RT for 3 h. The reaction was diluted with EtOAc (10 mL) and washed with H₂O (10 x 2 mL) and brine (10 mL). The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The product was purified by C18 column chromatography (MeOH: H₂O eluent) to afford **1-OH** compound (11 mg). The residue (10 mg, 0.007 mmol) was dissolved in CH₂Cl₂ (5 mL) at -30 °C. Then, DAST (3 μL, 0.021 mmol) was added slowly, and the resulting reaction mixture was stirred for 1 h. When TLC (acetone: CH₂Cl₂, 4/6) indicated formation of product with consumption of starting material, the reaction was quenched with aq. NaHCO₃. The filtrate was washed with aqueous NaHCO₃ (2 x 5 mL) and brine (5 mL) solution. The organic layer was dried over Na₂SO₄ and concentrated in *vacuo*. The residue was purified by C18 column chromatography (MeOH: H₂O eluent) to afford **S30b** (0.007 g, 38% over 2 steps). TLC (acetone: CH₂Cl₂ = 6/6, v/v), R_f = 0.24; ¹H NMR (400 MHz, CHCl₃): δ 5.99 (d, J = 10.2 Hz, 1H), 5.50 (d, J = 52.2 Hz, 1H), 5.39 (d, J = 10.3 Hz, 1H), 5.33-5.17 (m, 7H), 4.87-4.81 (m, 2H), 4.61 (d, J = 10.2 Hz, 1H), 4.52 (d, J = 10.3 Hz, 1H), 4.40-4.026 (m, 5H), 4.21-3.83 (m, 4H), 3.70 (s, 3H), 3.49-3.47 (m, 2H), 2.54 (dd, J = 3.8 & 8.1 Hz, 1H), 2.19 (s, 6H), 2.17 (s, 6H), 2.14 (s, 6H), 2.10 (s, 3H), 2.05 (s, 6H), 2.02 (s, 6H), 1.98 (s, 3H), 1.97 (s, 6H), 1.23 (t, J = 10.1 Hz, 1H); ESI-MS: *m/z* calcd for C₅₆H₇₇FN₂O₃₅; 1357.2094 found 1339.3978 (*M* + Na)⁺.

2. Glycan Array Analysis

i. Microarray Fabrication

a) Fabrication of *N*-hydroxy succinimide-coated glass slide: All monovalent glycans (G1-33) were prepared in 10 mM concentration individually and served as mother solutions which are to be diluted with printing buffer to prepare a working solution. Microarrays were printed (BioDot Cartesian Technologies) by robotic pin (SMP3: TeleChem International) deposition of ~ 0.6 nL of 100 μ M concentration of amine-containing glycans in printing buffer from a 384 well plate onto NHS-coated glass slides. The printed slides were allowed to react in an atmosphere of 80% humidity for an hour followed by desiccation overnight. These slides were designed for 16 grids in one slide, and stored at room temperature in a desiccator prior to use. Before the binding assay, these slides were blocked with blocking solution for 3 h. The slides were then washed with PBST-BSA buffer prior to use. Unless otherwise stated, reagents were obtained from commercial suppliers and used without purification. All aqueous solutions were prepared from distilled de-ionized water filtered with a Milli-Q purification system and sterile filtered through a 0.2 μ m filter. Buffers used in the experiment include the printing buffer (pH 8.5, 300 mM phosphate buffer containing 0.05% (v/v) Tween-20), the blocking buffer (superblock blocking buffer in PBS, Pierce), the washing buffer (PBST buffer; PBS and 0.05% Tween-20) and the binding buffer (PBST-BSA Buffer; PBST buffer and 3% BSA). Printing buffer, blocking buffer, and binding buffer were prepared freshly before use.

b) Fabrication of aluminium oxide coated glass slide. The aluminum oxide glass substrate provides advantages for being able to be assessed by both mass spectrometry and fluorescence scanning. In addition, the fluorescence intensity of sugar-protein binding on the substrates was found to be more sensitive than that on transparent glass slide¹⁵. There are many reports describing the surface of micron to nano pore structures of anodized aluminum oxide. However, only a few articles have mentioned the initial planar layer of aluminum oxide¹⁶ during the treatment of surface anodization. In the development of AAO glass substrate, a computer

experimental design program (Design Expert 8.0) has been used to optimize the planar layer of surface anodized aluminum oxide to be best used in glycan microarray¹⁷.

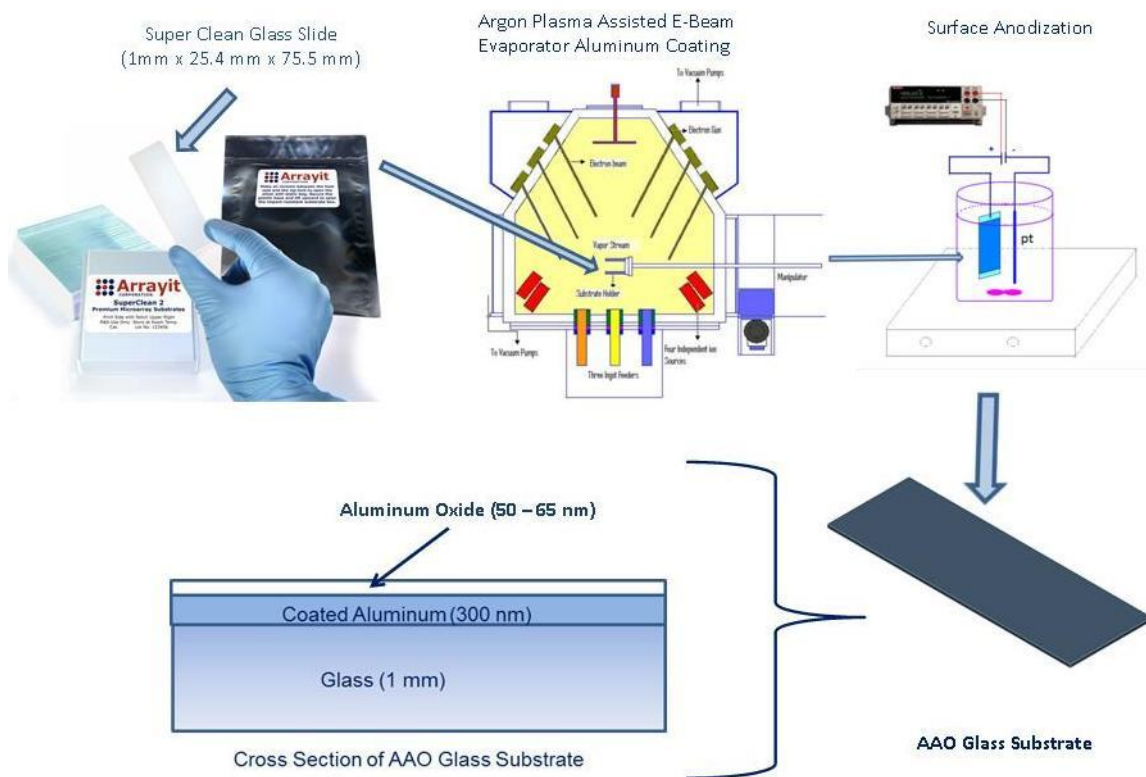


Figure S4 | Schematic drawing of the fabrication of anodized aluminum oxide (AAO) glass substrate.

As shown in Figure S4, approximately a larger of 300 nm aluminum coated on glass slide (1 x 75.5 x 24.5 mm) was fabricated using E-Beam VDP coater by The Thin Film Technology Division, Instrument Technology Research Center at National Applied Research Laboratories, (Hsinchu Science Park, Hsinchu, Taiwan, R.O.C.). The slide after Al-coating was immediately packed (one slide per container) and vacuum sealed in air-tight laminated foil pouches until the moment for surface anodization. Surface anodization of the aluminum coated glass slide has been conducted in-house via electrochemical reaction in a 0.3 M oxalic acid aqueous solution in a 4 °C chamber. The anodization reaction was controlled by voltage and reaction time. Under proper reaction conditions, a layer of approximately 50 to 65 nm (in thickness) of smooth

anodized aluminum oxide (AAO) can be grown consistently on top of aluminum coated glass slide. Unlike the conventional anodic aluminum oxide surface with pores, as the AFM picture shown in Figure S5A, we have developed a smooth AAO glass substrate with Rsm surface roughness (2.3 nm) similar to the surface of glass slide (< 3 nm). The cross section picture of a typical AAO glass substrate is shown in Figure S5B. To obtain the optimized AAO glass substrate with its physical properties shown in Figure S5, the detail experiment using computer program generated factorial design and response surface methodology has been conducted and will be reported in a separate article.

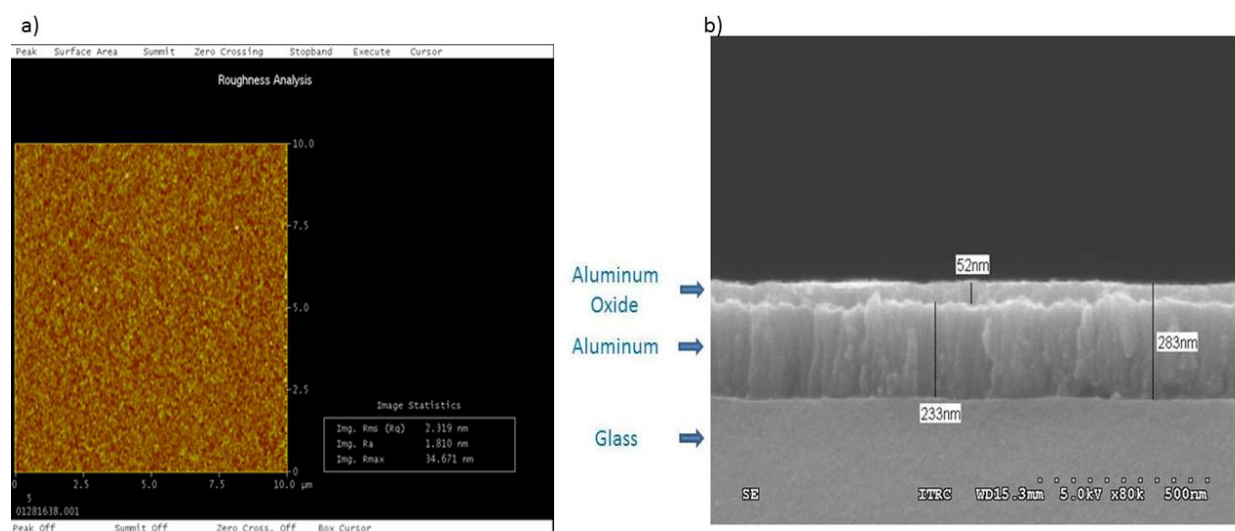


Figure S5 | (A) AFM picture – Roughness Analysis of the surface. Img. Rms (Rq) 2.319 nm and
(B) Cross Section SEM picture of the AAO Glass Substrate.

ii. Surface Comparison of AAO Glass Substrate vs. NHS-Activated Glass Slide

NHS-glass slide has been widely used for glycan microarray in our and other groups. The amide formation of the NHS functional group with the amine of sugar derivative was taken place on the surface. Its first chemical reaction was to covalently bond to the carboxyl group of the functionalized glass surface. The AAO glass substrate contains a layer of stable polymer network of amorphous aluminum oxide on the surface which is ready for the chemical reaction with the phosphonic acid tail of glycan derivative to form a phosphonate. Interface reaction of phosphonic

acid on metal oxides, either aluminum oxide or titanium oxide, is spontaneous and the resulting array is more homogeneous, and density and distribution can be easily controlled compared to the reactions on NHS slide. To identify the reactive site available for interface covalent bonding, Cy5-phosphonic acid and Cy5-amine have been used for microarray on the surfaces directly. These dyes (Figure S6) have been synthesized in-house starting from Cy5-NHS ester (Lumiprobe Corp. Item No. 63020).

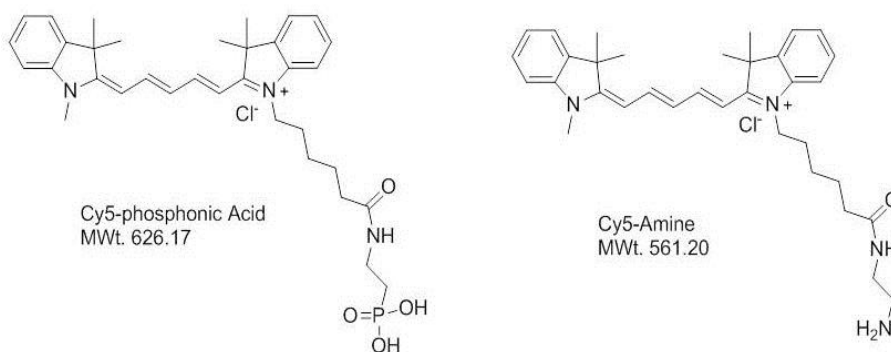


Figure S6 | Structures of Cy5-phosphonic acid linker and Cy5-amine linker

The dyes, Cy5-phosphonic acid and Cy5-amine, were dissolved in a 7:3 ratio of ethylene glycol/water mixtures (1 mM) with pH adjustment to 6 and 8.5 respectively. The arrayed slides were washed thoroughly with a mixture of 5% methanol in water, spin-dried before subjecting for GenePix (4300A) Scanning. Figure S7 shows the microarray pictures of Cy5-phosphonic acid on AAO glass substrate (Figure S7A), Cy5-amine on NHS-glass slide (Figure S7B) and their 20 spots average fluorescence intensity (Figure S7C). The representative confocal microscope pictures of these arrayed spots are given in Figure S8. Higher loading capacity and more uniformly distributed Cy5-molecules have been observed similarly to those which have been seen usually in the glycan microarray and sugar/protein binding.

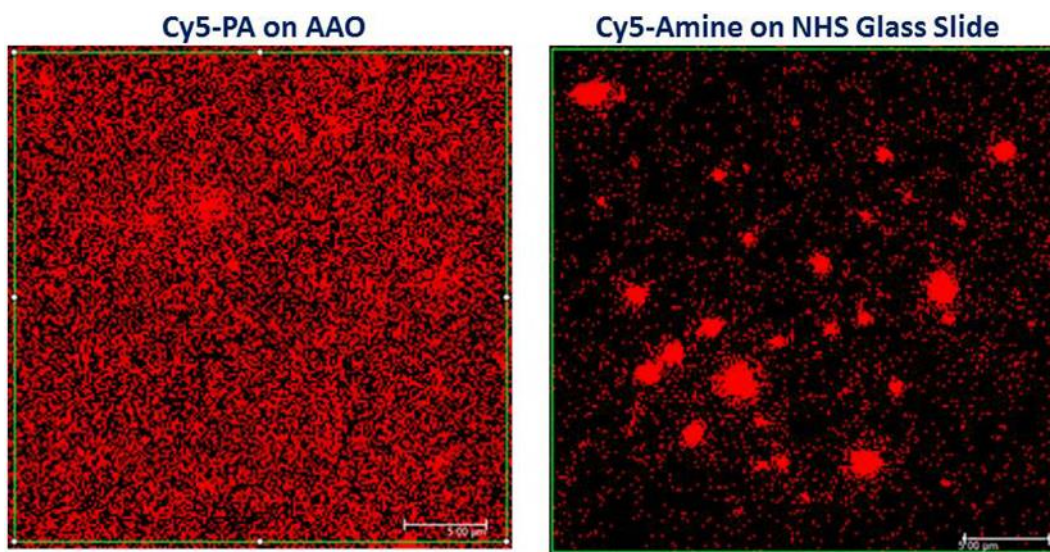


Figure S7 | GenePix Scanning (at PMT 450) of 1 mM of (A) Cy5-phosphonic Acid on AAO glass substrate (B) Cy5-Amine on NHS glass slide, and (C) their averaged 20 spots fluorescence intensity.

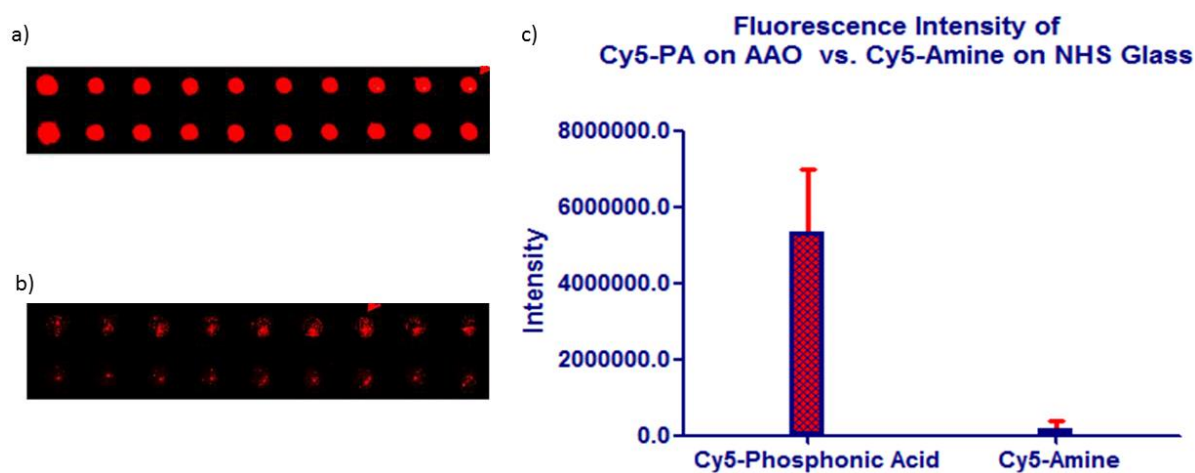


Figure S8 | Representative Confocal Microscopes pictures Cy5-phosphonic Acid, and Cy5-Amine on AAO glass substrate and NHS glass slide. Selective $900 \mu\text{m}^2$ area within the spots.

iii. Surface uniformity comparison of ACG and NHS slide using confocal microscope.

To evaluate the density and uniformity of glycan array on ACG and NHS activated glass slide, we used the mannose monosaccharide formed glycan array as a model. A mannose solution of 100 μM was used in arraying, and a solution of ConA488 of 50 $\mu\text{g}/\text{mL}$ was used in sugar/protein binding. The images of ConA488/mannose binding observed from confocal microscope further confirmed that the AAO glass substrate has denser and more uniformly distributed covalently bonded sugars. Figure S9 shows the images observed from Confocal Microscope (Leica SP5) of ConA488/mannose binding on AAO Glass Substrate vs. NHS Glass Slide.

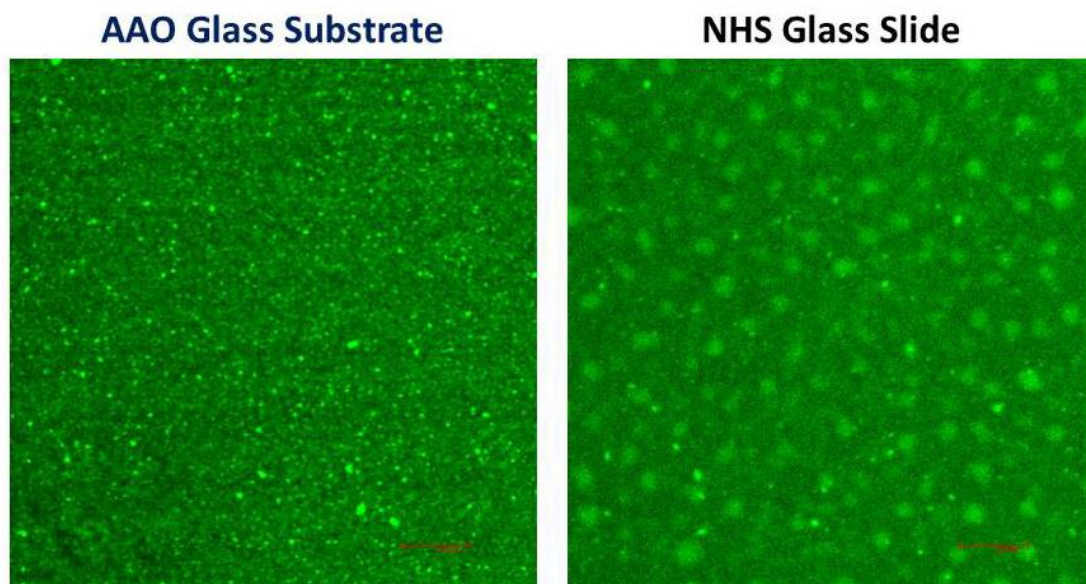


Figure S9 | Confocal Microscope of ConA488/Mannose binding on AAO glass Substrate vs. NHS-Glass Slide.

Atomic force microscopy image of sugar distribution on aluminium-oxide coated glass slide, and NHS coated glass slide suggest that the AAO glass substrate (Figure S10a) provides more uniform distribution of covalently bonded sugars than that of NHS glass slide (Figure S10a). Particle counts of these images were obtained by counting the particles over the height of one half width distribution of the maximum numbers of particle height. Mannose derivatives can

only covalently bond on the slides where the activated functional groups were available on the surface. Regardless of differences in Rms of the glass base materials, the AAO glass substrate provides more homogeneous distribution than that of NHS glass slide.

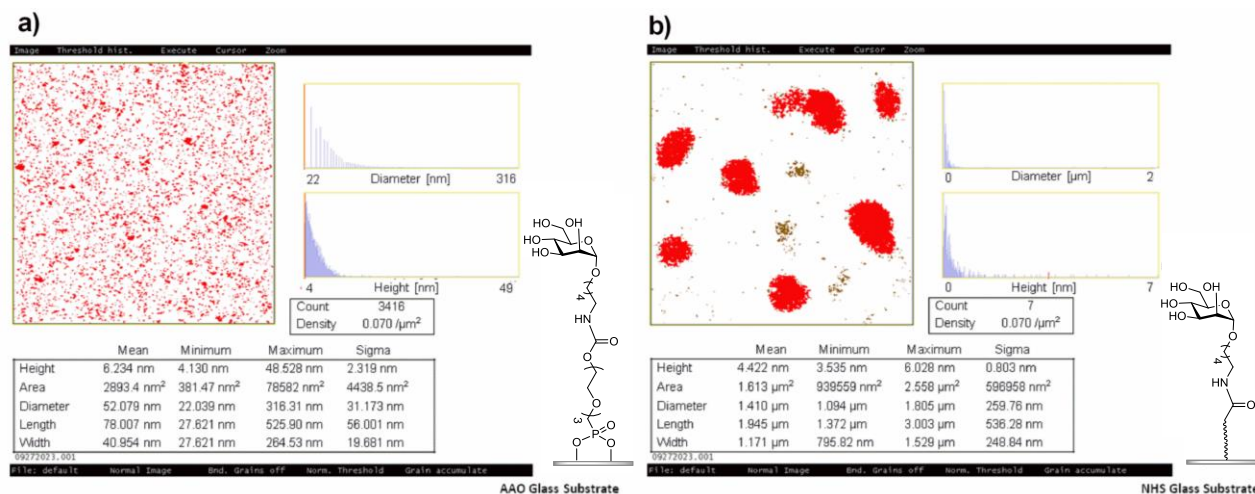


Figure S10 | Atomic force microscopy image showing sugar distribution on **a)** aluminium-oxide coated glass slide, and **b)** NHS coated glass slide.

iv. Antibody binding assay: Antibodies PG9, PG16 and PGTs 141-145 were diluted by binding buffer to 100 μg/mL prior to use. DyLight649-conjugated Donkey Anti-Human IgG antibody was then pre-complexed with primary antibodies. The final concentration of the precomplexed solution was adjusted to 50 μg/mL with binding buffer. The printed glass slide was assembled into FAST[®] frame slide holder (Whatman[®]), 80 μL of precomplexed antibody solution were then applied to each well accordingly. The antibodies binding process was maintained in 4 °C with gentle shaking and then antibody solutions were carefully pipetted out after 6 hours incubation. The glass slide was gently washed by 100 μL PBST washing buffer and then spin-dried for 3 minutes.

v. Image processing and data analysis: The slide was scanned at 635 nm with a microarray fluorescence chip reader (GenePix[®] 4300A, Molecular Devices Corporation). Scanned images were analyzed with GenePix Pro 6.0 analysis software (Molecular Devices Corporation,). The image resolution was set to 5 μm per pixel. Spots were defined as circular features with defined

diameter of 100 μm . The values of total intensity were chosen for data processing, performed with Graphpad Prism[®] 6.0. The intensities were calculated and averaged. Error bars represent the standard deviation for all data points reported.

vi. Analyzing glycan binding specificity of HIV-1 bNAbs on NHS-coated glass slide. The slide for the study of PG9, PG16 and PGTs141-145 was prepared by printing glycans **G1-33** (Figure S11) on the N-hydroxysuccinimide coated surface through covalent bond formation. Slide images obtained by fluorescent scan after secondary antibody incubation.

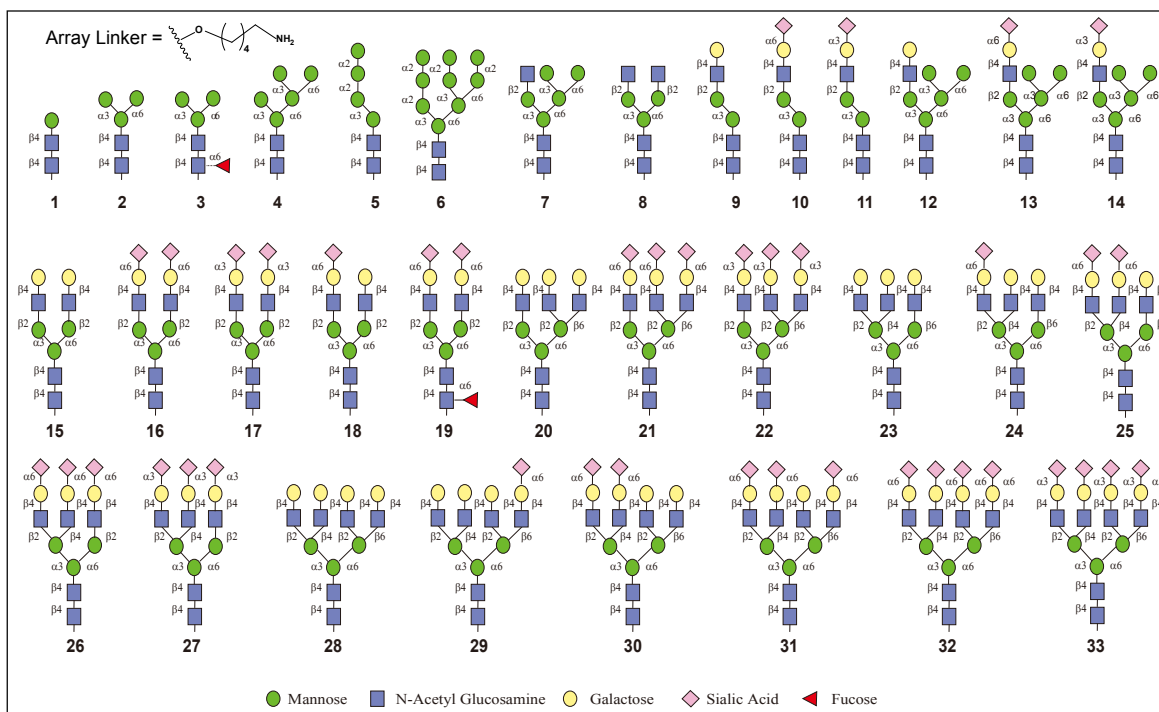


Figure S11 | Schematic representation of *N*-glycans printed on NHS coated glass slide.

Having incorporated diverse HIV-1 gp120 related *N*-glycans in our array, we next proceed to investigate binding behavior of PG16 to those glycans. Our results are consistent with our previously reported data, where, the PG16 binding is proportional to α 2,6-Neu5Ac count at the termini (Figure S12). A short conclusion from our study is presence of di-sialylated antennae in defined orientation is critical for binding in PG16 binding pocket. However, it demand deep investigation to prove the phenomenon.

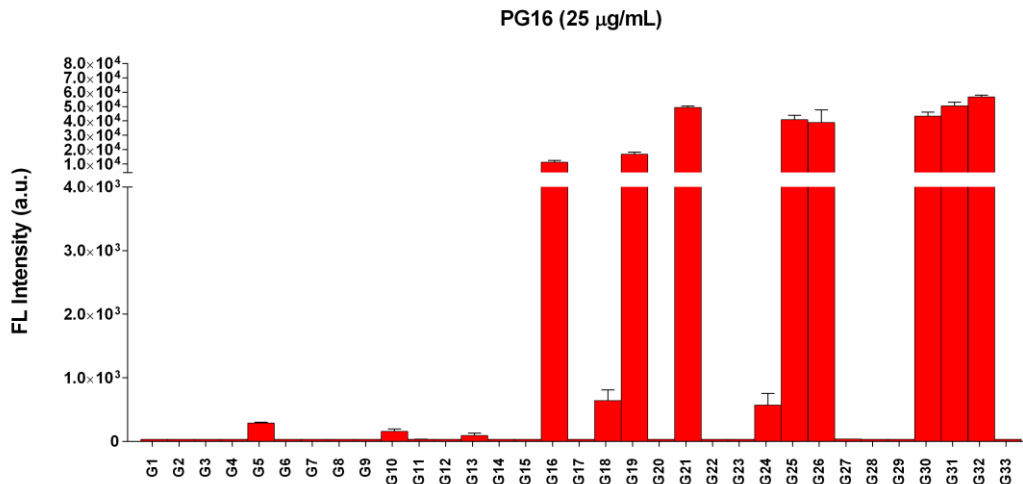


Figure S12. Binding behavior of PG16 using NHS-coated glass slide. Bindings of PG16 to panel of *N*-glycans represented in bar chart.

To our knowledge, there is no any array based study reported before, that showed defined set of glycans binds to antibodies PG9 and PGTs141-45. To get into the details, we scened PG9 and PGTs141-145 on our NHS-coated glass slide array.

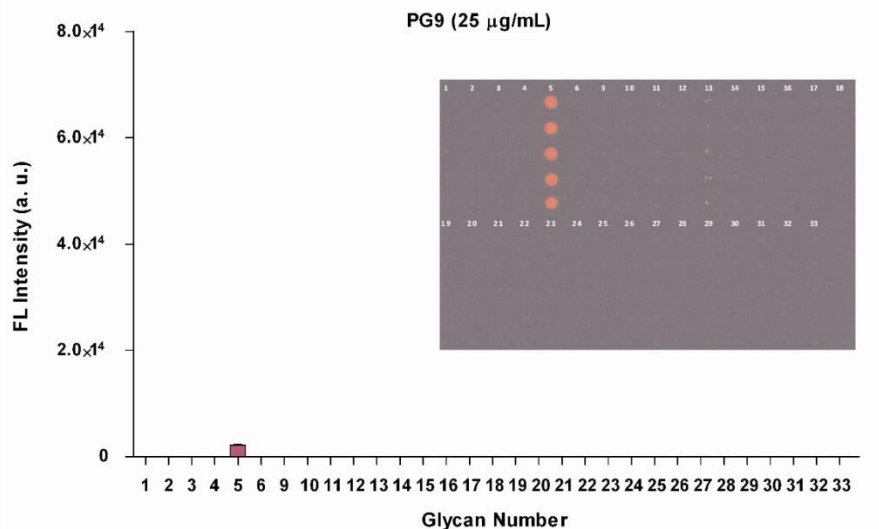


Figure S13. Binding behavior of PG9 using NHS-coated glass slide.

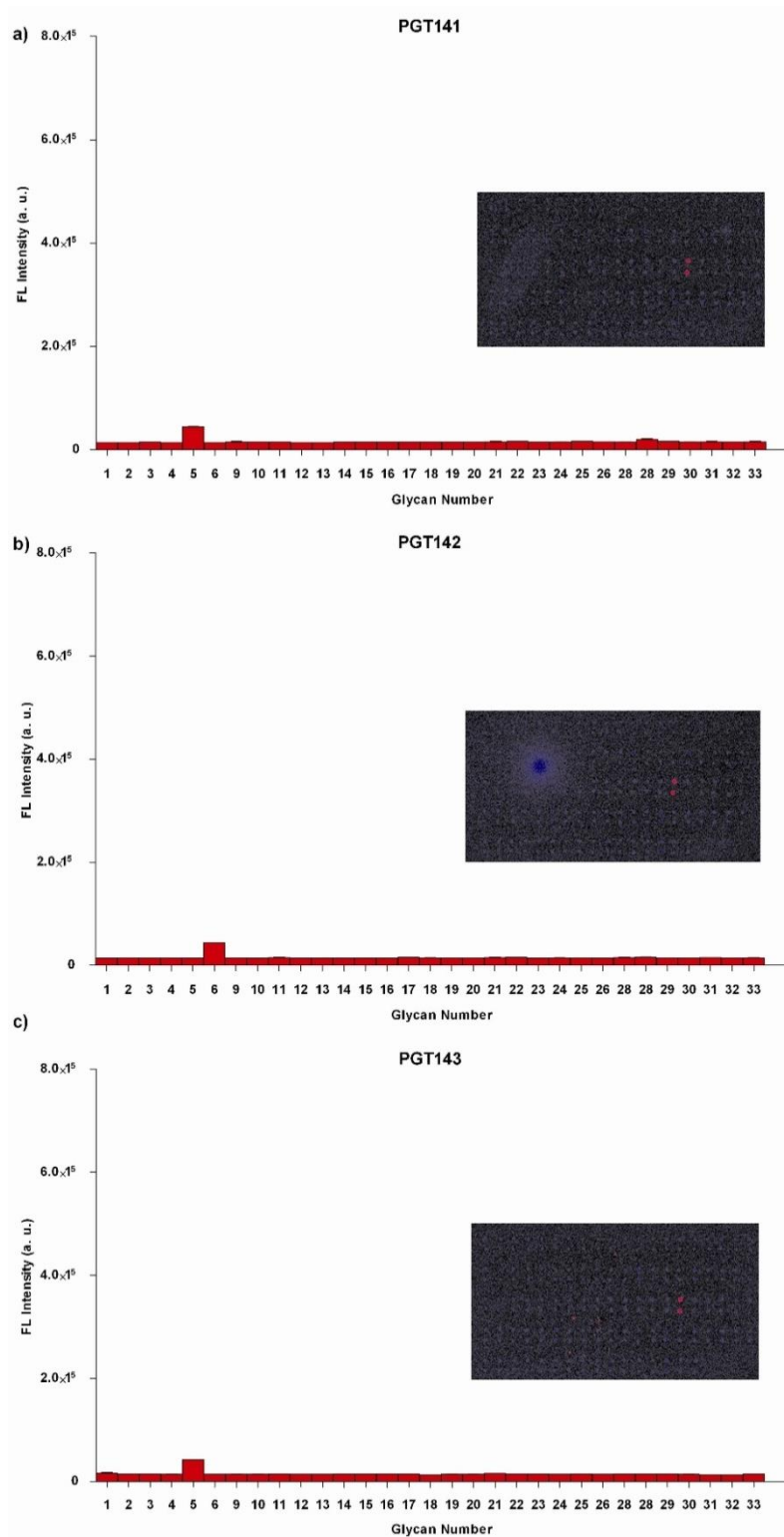


Figure S14. Binding behavior of PGTs 141-143 using NHS-coated glass slide. Antibody concentrations used here are 25 $\mu\text{g}/\text{mL}$

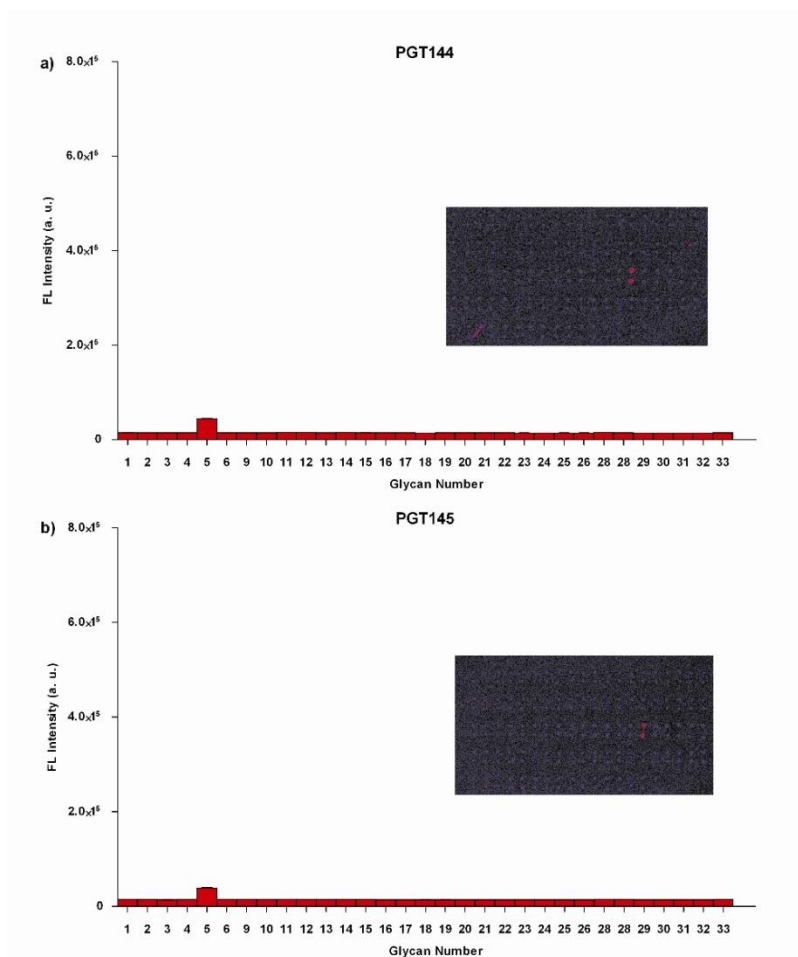


Figure S15. Binding behavior of PGTs14-145 using NHS-coated glass slide. Antibody concentrations used here are 25 $\mu\text{g/mL}$

In our binding experiment, we could not observe a detectable binding for antibodies PG9 (Figure. S13) and PGTs 141-145 (Figure. S14-15) towards any of the glycans printed on array. The possible explanations are 1) the extremely weak binding abilities of these antibodies to peptide/protein free glycans, 2) the glycan density on NHS-coated array may not be sufficient enough for detectable binding, and 3) the hydrolyzing tendency of N-hydroxy succinimide activation of carboxylic acid groups on array might affect the efficient immobilization of glycans on array. However, throughout our experiments (Figure S13-15), we observed a strong signal for glycan **5** ($\text{Man}_4\text{GlcNAc}_2$). In order to confirm whether it's come from non-specific binding between secondary antibody and glycan **5**, we performed a control experiment where the

secondary antibody was applied directly on array without pre-complexed with primary antibody. Indeed, in absence of primary antibody we observed the fluorescent signal for glycan **5** (Figure S16). Although the reason is unknown but it confirmed that secondary antibody showed non-specific binding to glycan **5**.

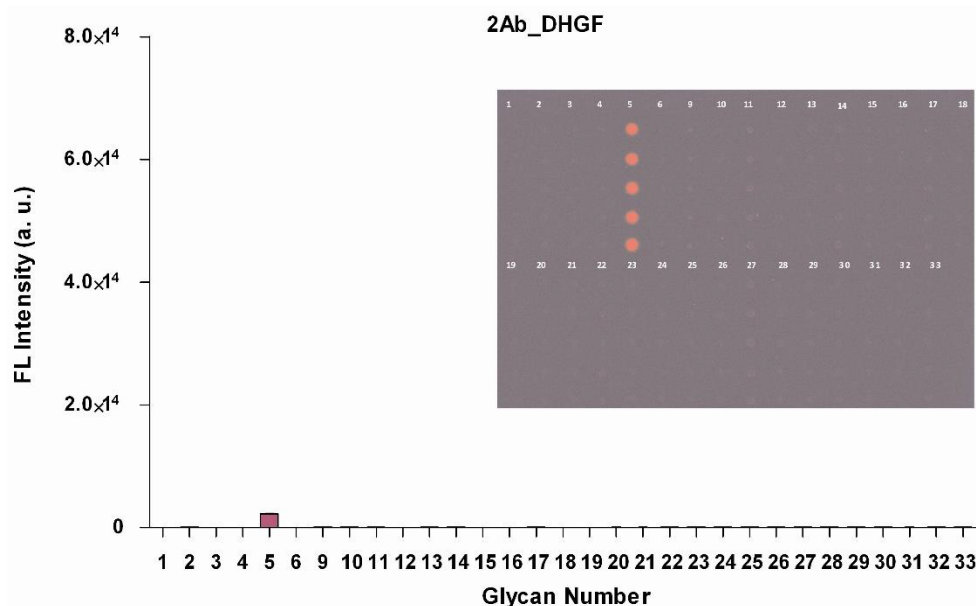


Figure S16. Experimental proof of secondary antibody binding to glycan 5.

vii. Analyzing glycan binding specificity of HIV-1 bNAbs on ACG slide. Having demonstrated the applicability of ACG slide over NHS-coated slide in terms of glycan density and enhancement in signal intensity, we printed glycan glycans **I-XI** on ACG array using phosphonate chemistry (Figure S17). The binding specificities of PGTs141-144 are shown below.

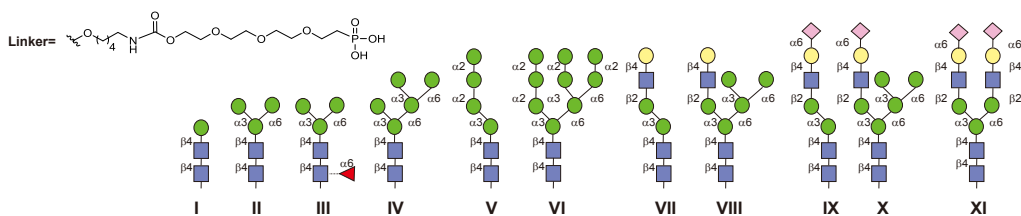


Figure S17. Cartoon represents glycans printed on ACG array. The structure of linker is shown on the upper left corner.

The binding specificities of PGTs141-144 are shown in Figure S18. However, we could not find the binding for PGT145 on ACG array, most probably because of its weak glycan binding affinity.

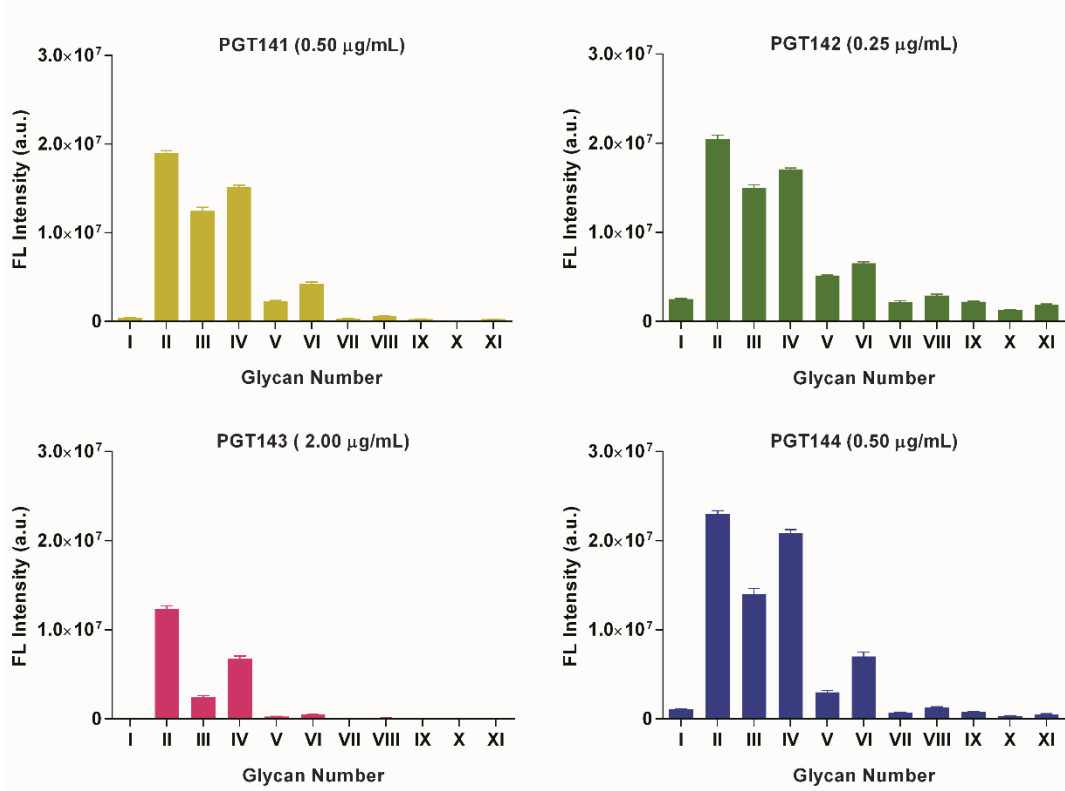


Figure S18. Bindings of PGTs 141-144 towards panel of N-glycans on ACG array is shown in bar chart.

viii. Determination of surface dissociation constants ($K_{D,surf}$) on ACG slide: The aluminium oxide coated glass slides for the determination of dissociation constants were spotted with 100 and 50 µM concentrations of glycans $\text{Man}_5\text{GlcNAc}_2$ (V), hybrid type (XI), bi-antennary complex type (XII), V+XI (1:1 mole ratio), V+XII, and XI+XII. Antibody PG16 was serially diluted to 3.32, 1.66, 0.833, 0.415, 0.207, 0.103, 0.051 and 0.025 µM with 3% BSA-PBST buffer. DyLight649-conjugated Donkey Anti-Human IgG antibody was pre-complexed with primary antibody PG16 (1:1 by weight). The pre-complexed solution (100 µL) was applied to each well

and incubated at 4 °C for 6 h in the dark. Finally, slides were washed with PBST, spin dried and scanned at 450 nm with a microarray fluorescence chip reader. Scanned images were analyzed with GenePix Pro 6.0 analysis software. The signal intensities for binding of PG16 to Man₅GlcNAc₂ (V) and mixture of V+XI (1:1 mole ratio) were too weak to determine the K_D . The binding curves for the rest of samples printed on array are shown in Figure S19 and K_D values are summarized in Table S2.

In the case of PG9, because of its very weak glycan binding affinity, we were not able to achieve the signal saturation to measure the binding constants, however, increasing PG9 concentration to 700 µg/mL resulted in a precipitation on array surface.

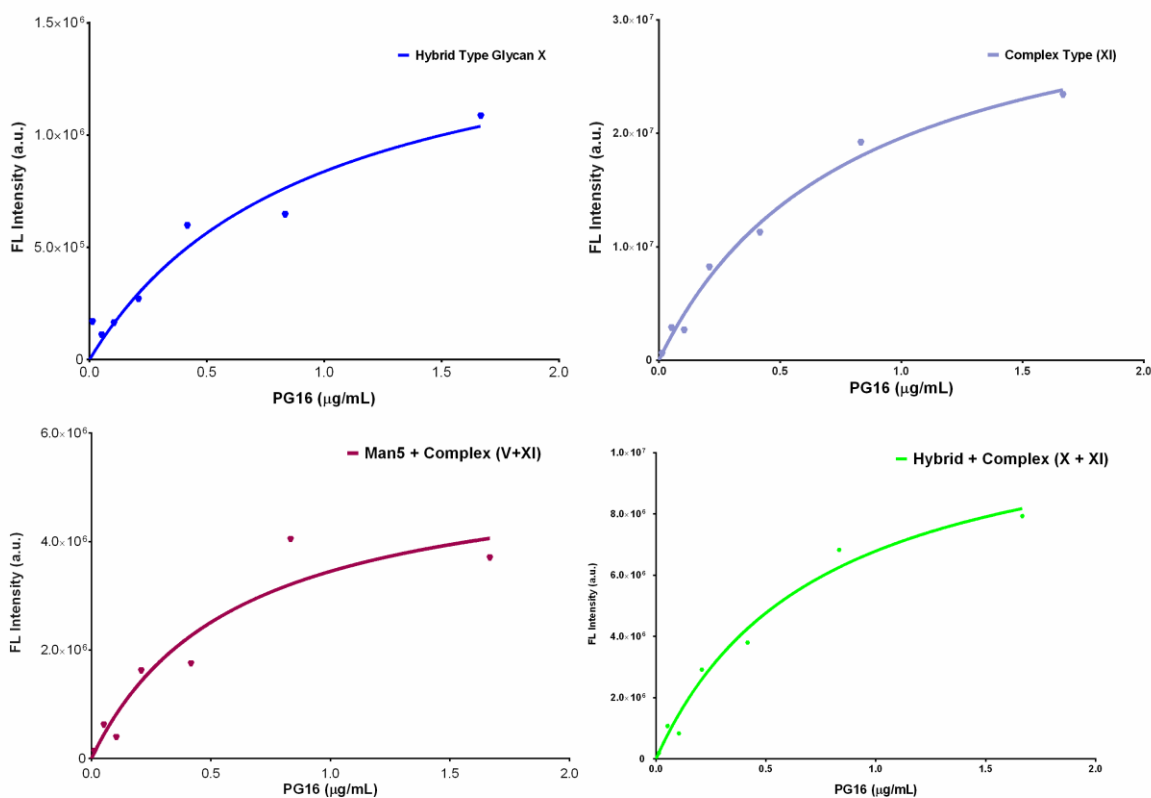


Figure S19. Antibody PG16 binding curves observed for glycans **X**, **XI** and mixtures **V+XI** and **X+XI** at 100 µM concentration. The curves were obtained by using DyLight649-conjugated donkey anti-Human IgG secondary antibody.

Table S1. $K_{D,surf}$ (μM) values of antibody PG16 and glycans X, XI and mixtures V+XI and X+XI.

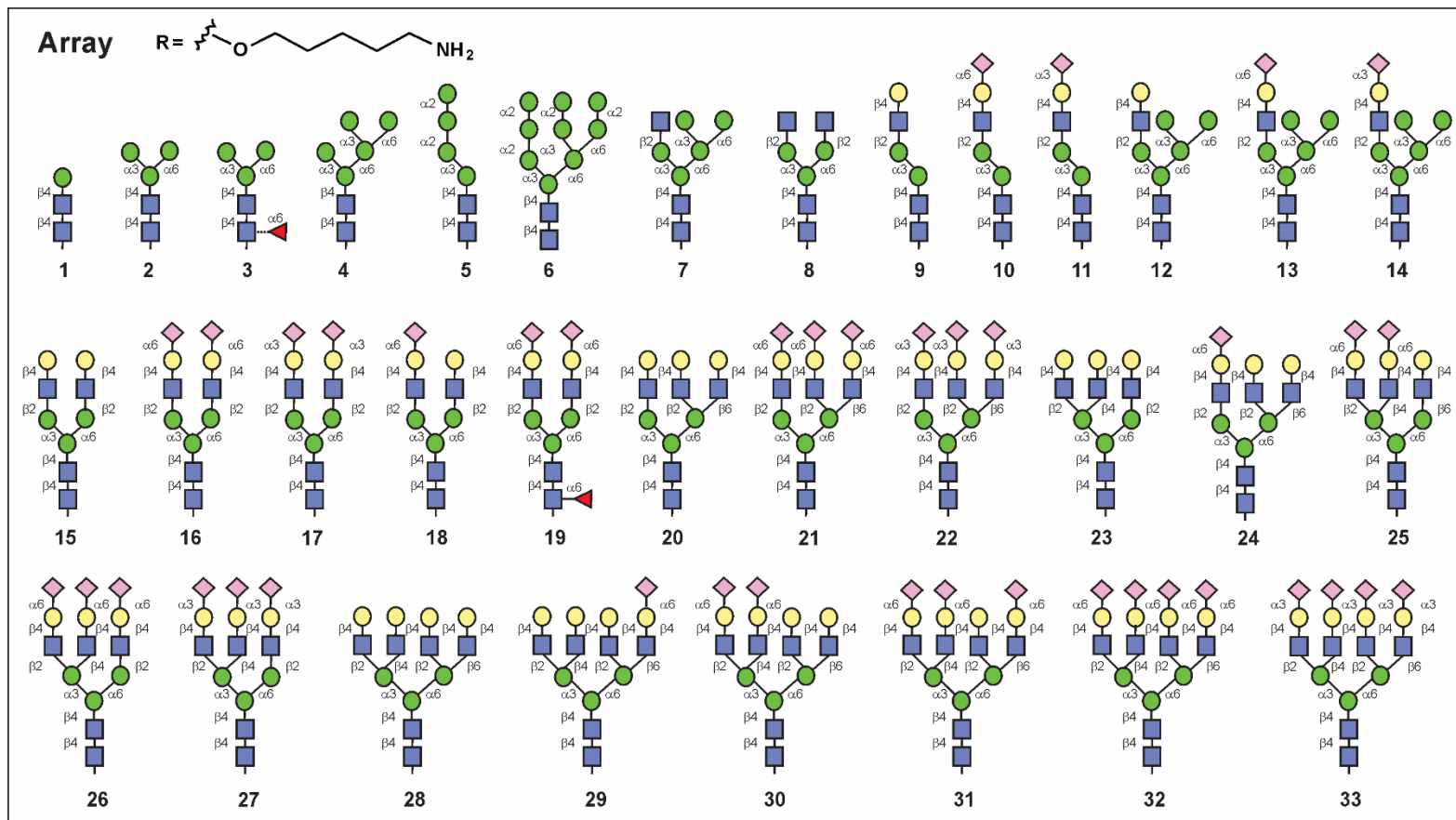
Glycan no.	$K_{D,surf}$ (μM) \pm SD (μM)
X	$0.935 \pm 0.026 \mu\text{M}$
XI	$0.320 \pm 0.125 \mu\text{M}$
V + XI	$0.827 \pm 0.200 \mu\text{M}$
X + XI	$0.988 \pm 0.223 \mu\text{M}$

References:

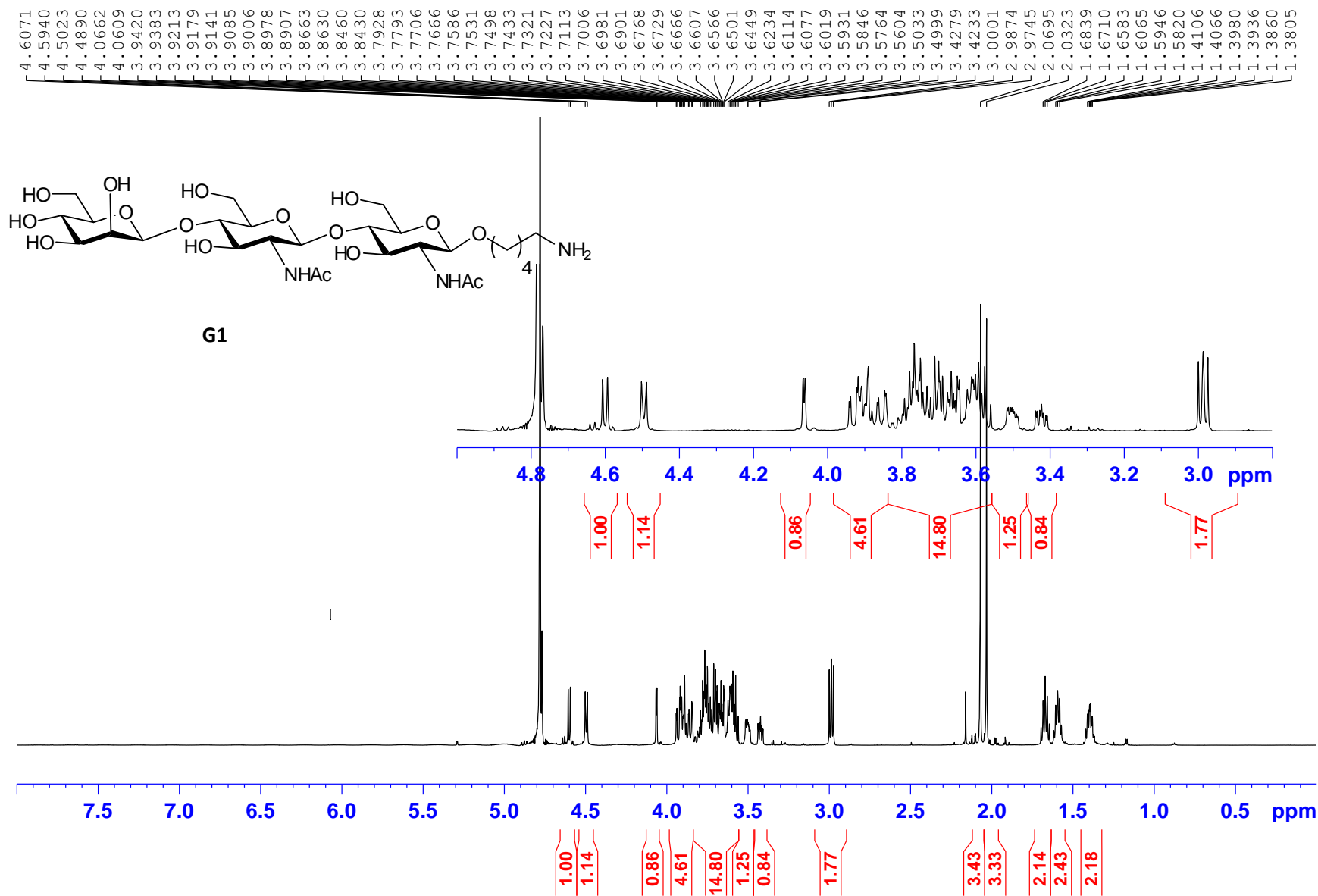
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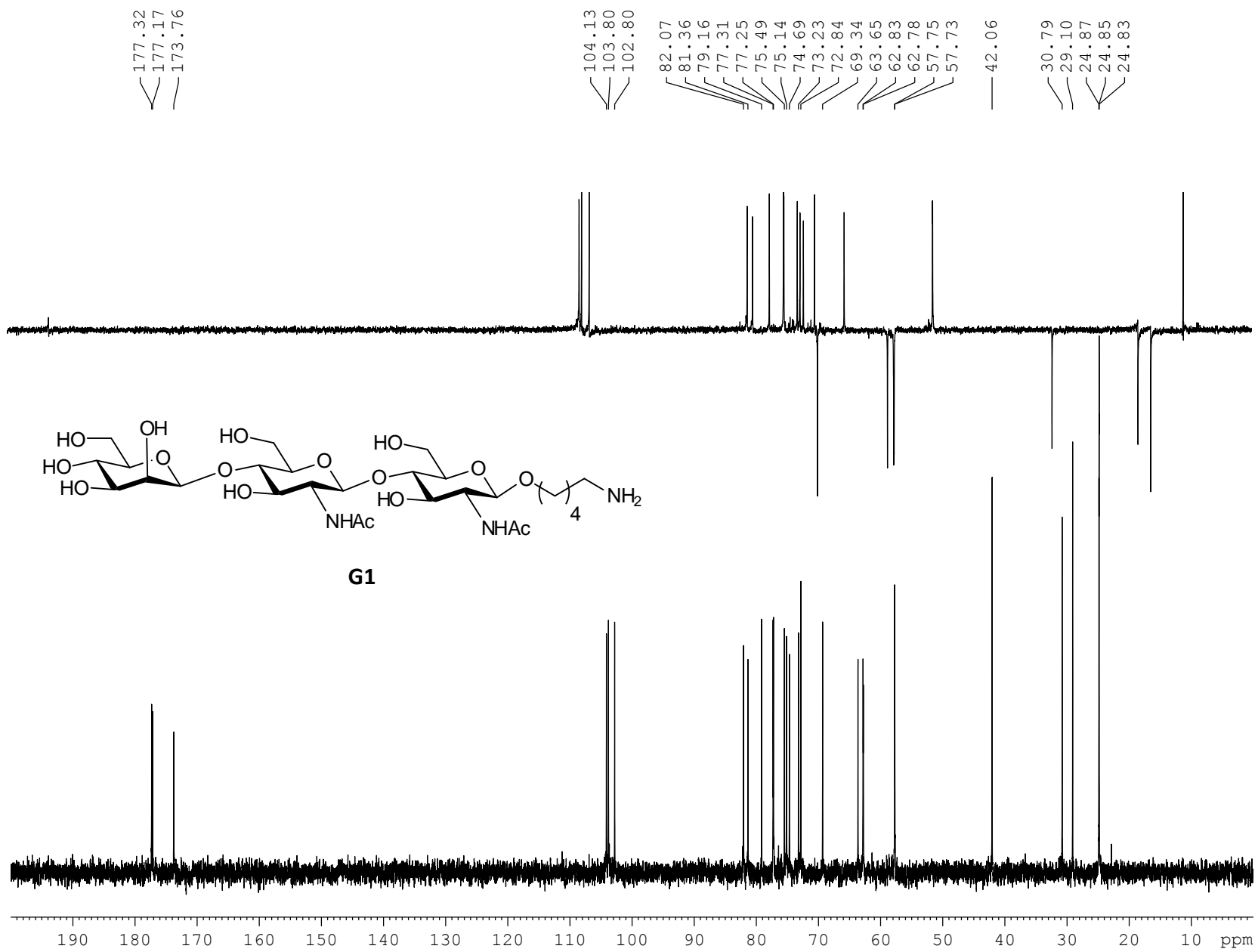
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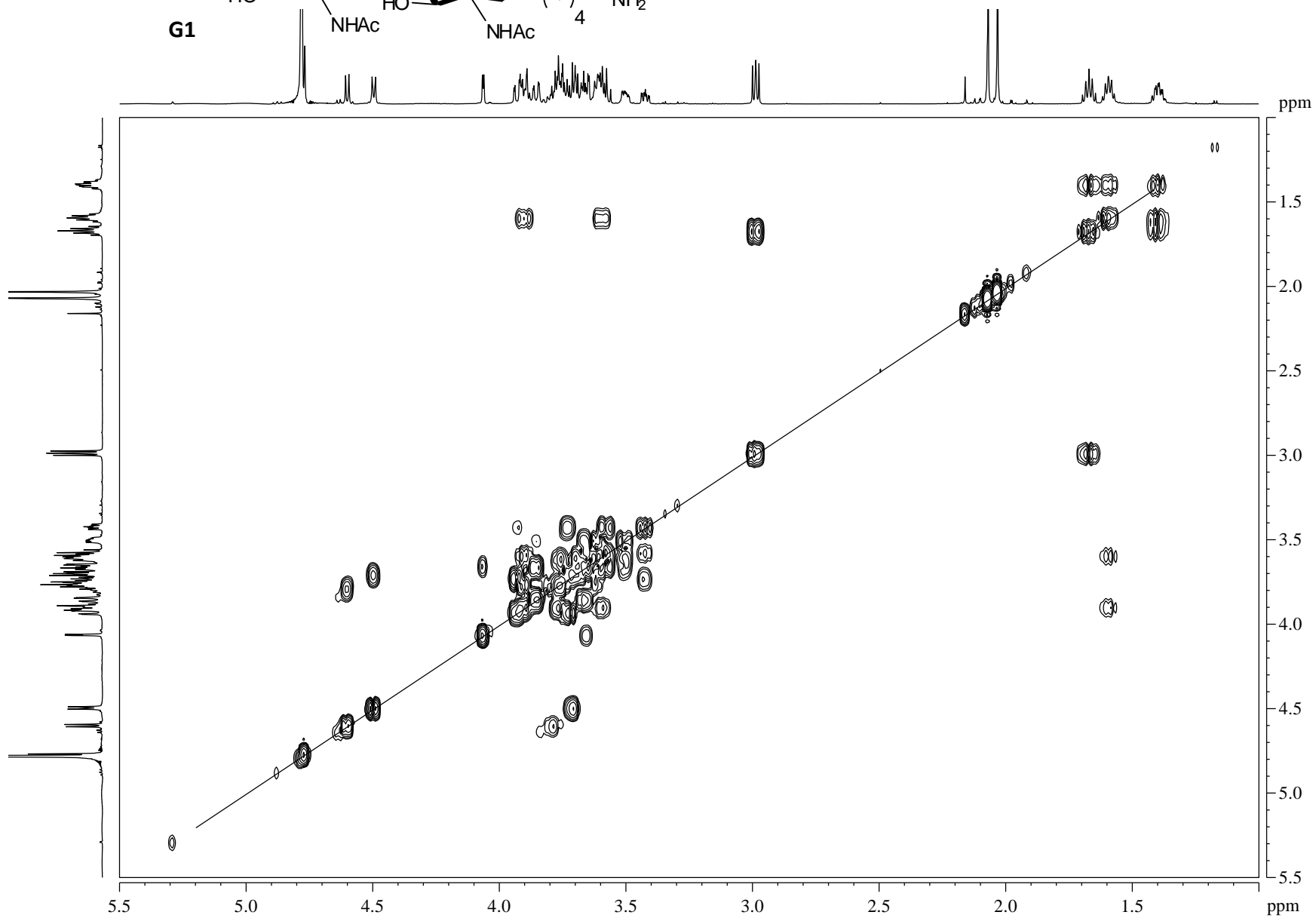
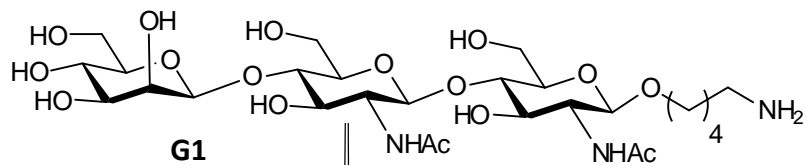
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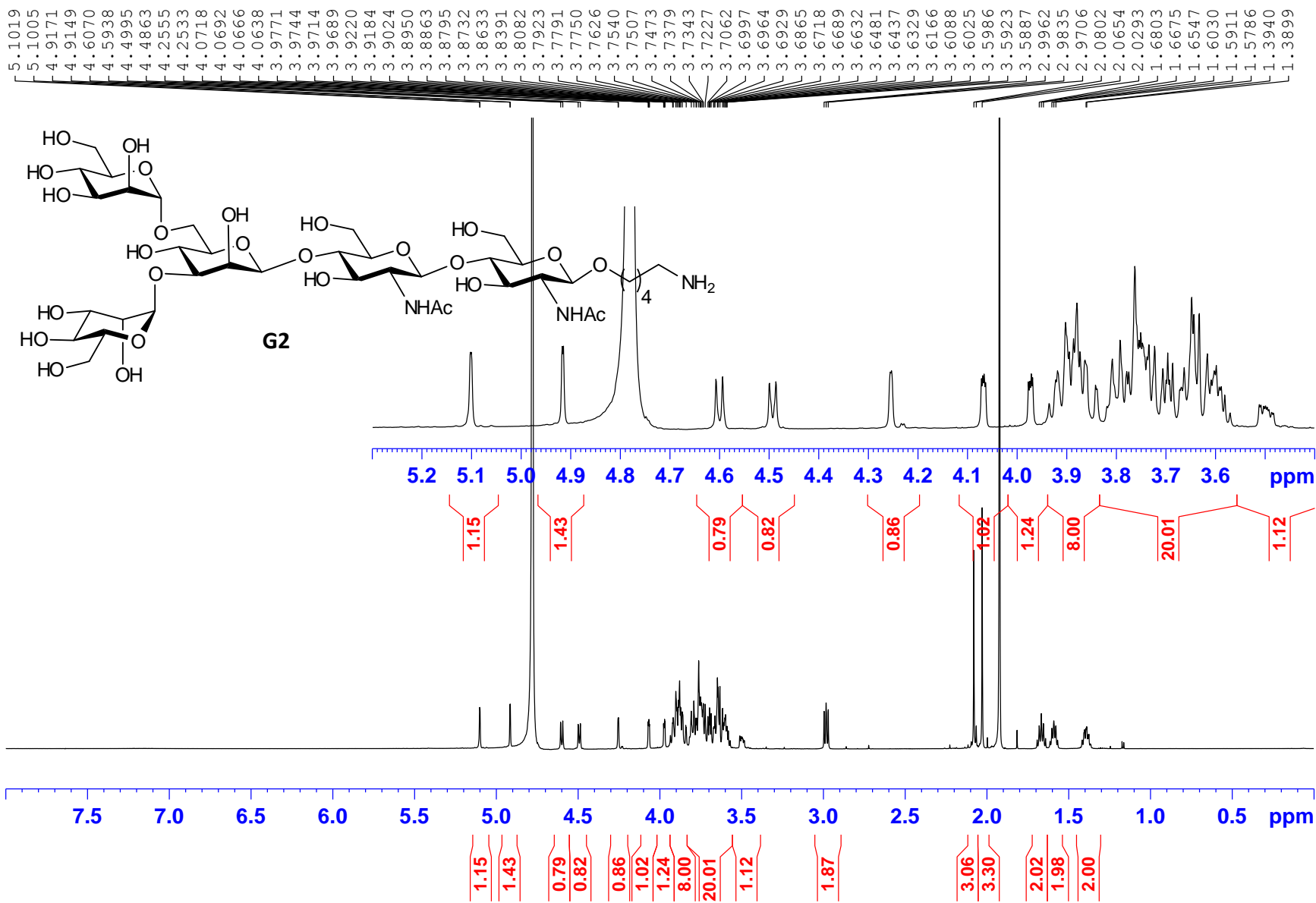


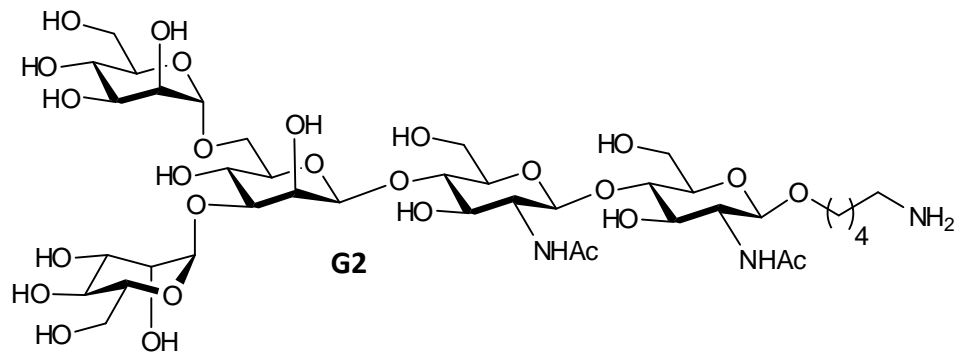
Characterization data of structures G15-17, G20-23, G26-28 and G32-33 was reported in *J. Am. Chem. Soc.* **135**(41), 15382-15391, (2013). The data for remaining structures is listed bellow.





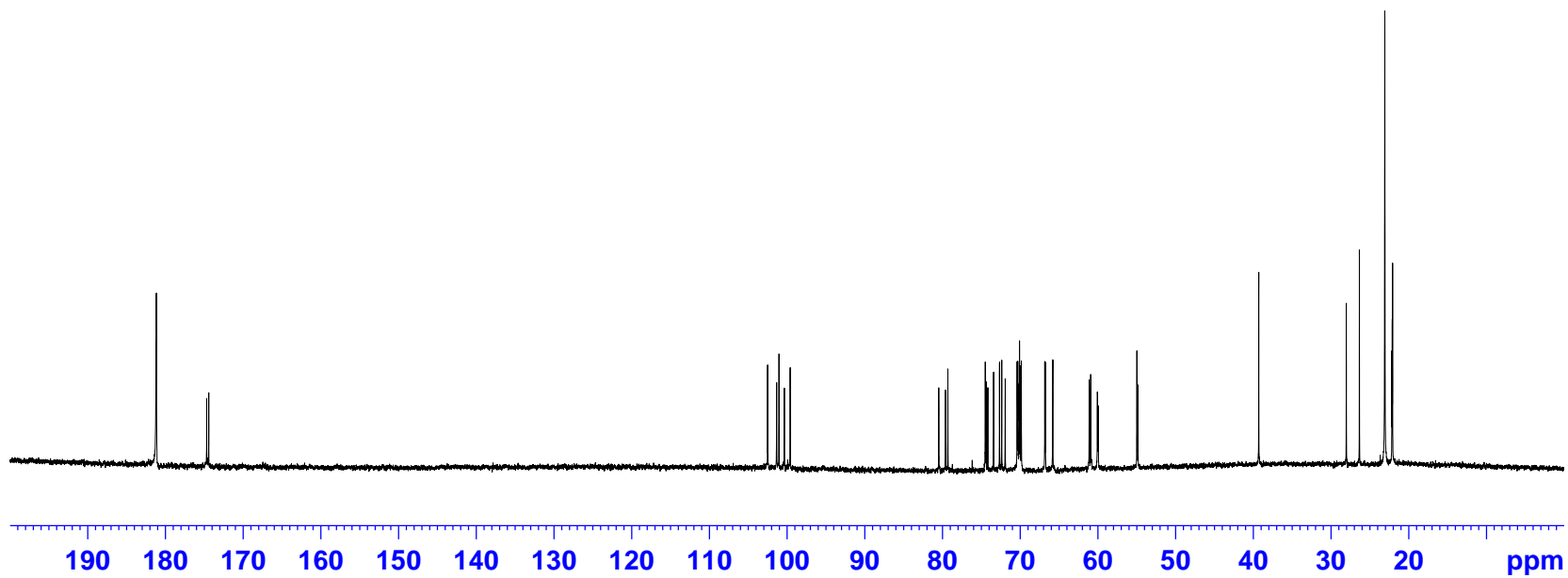


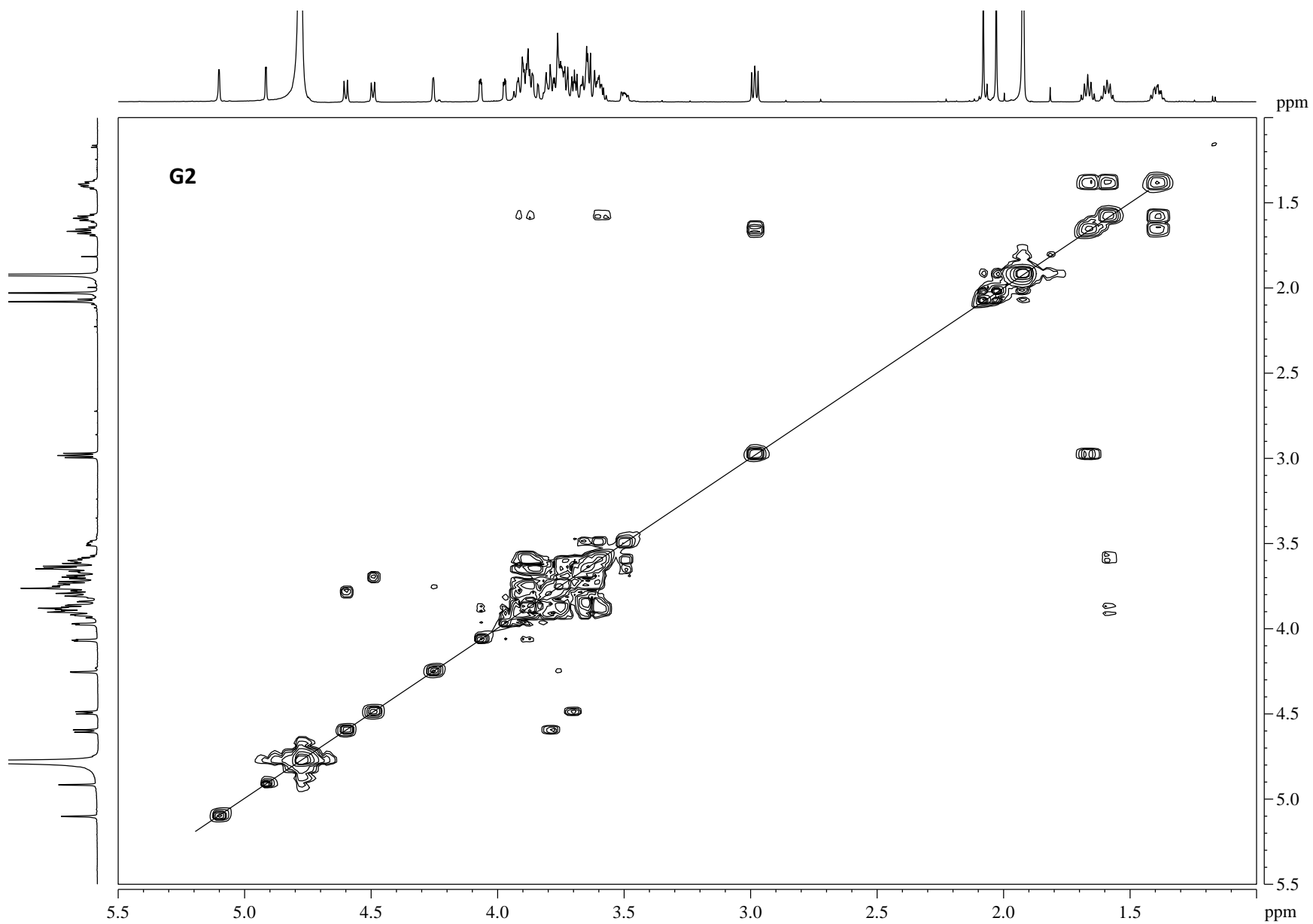


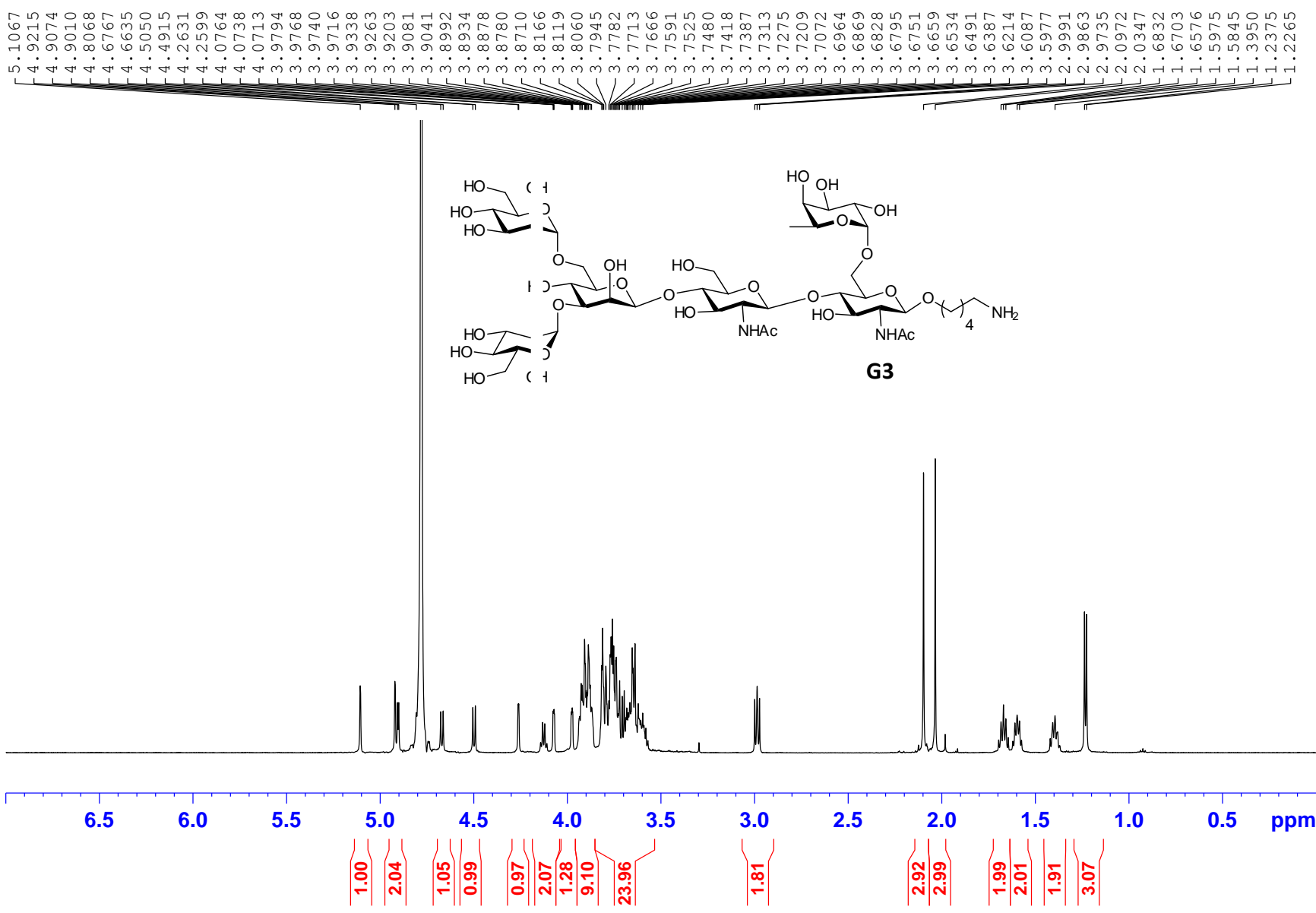


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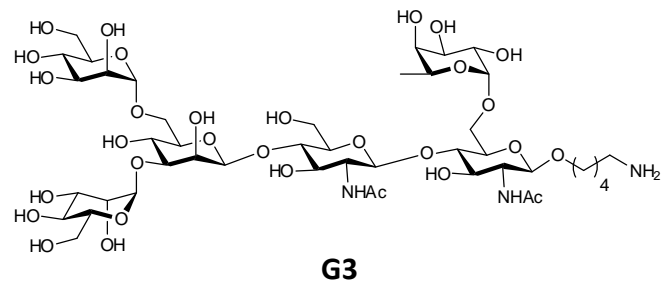
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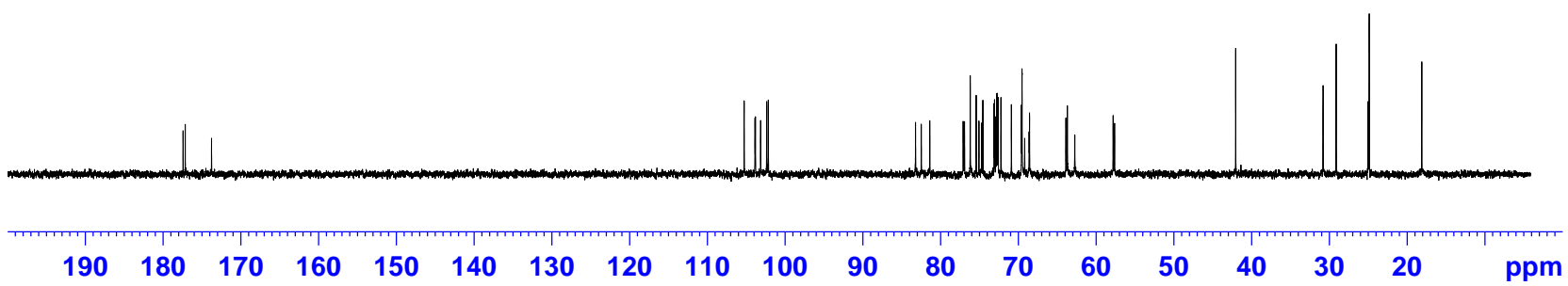


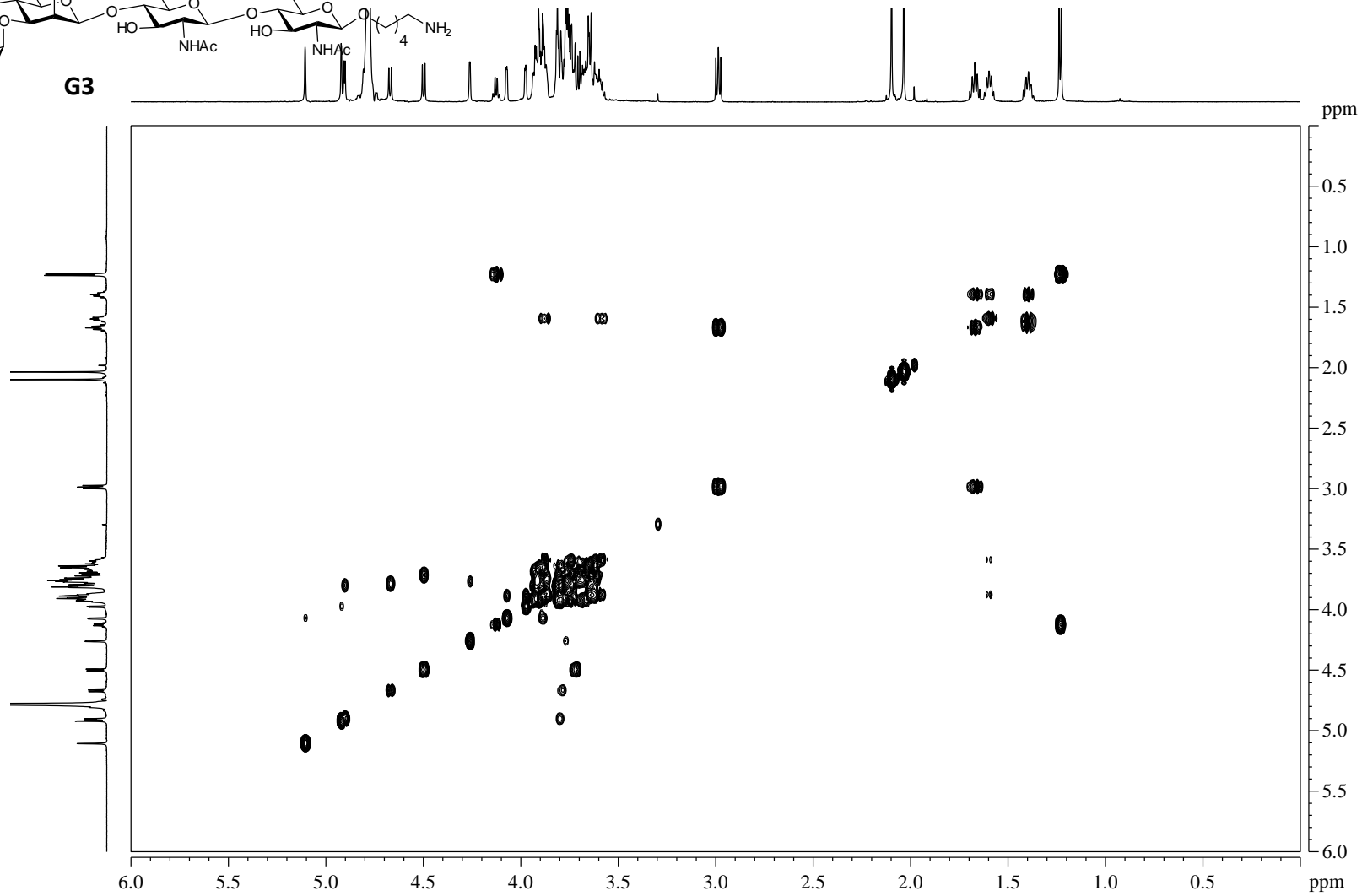
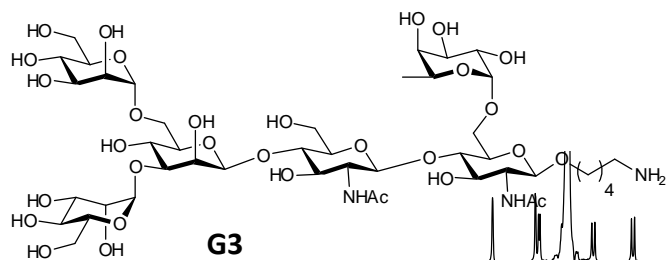
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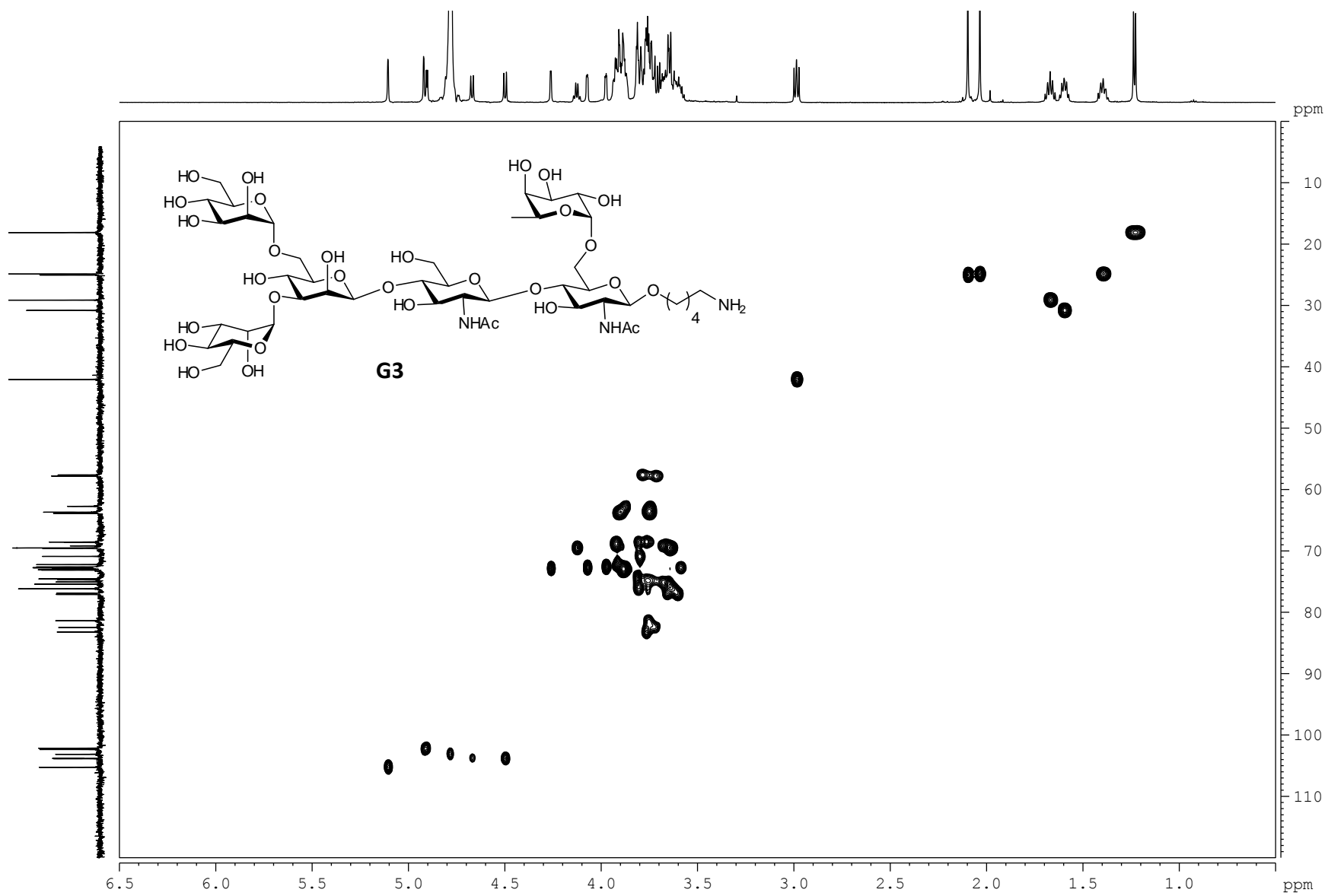


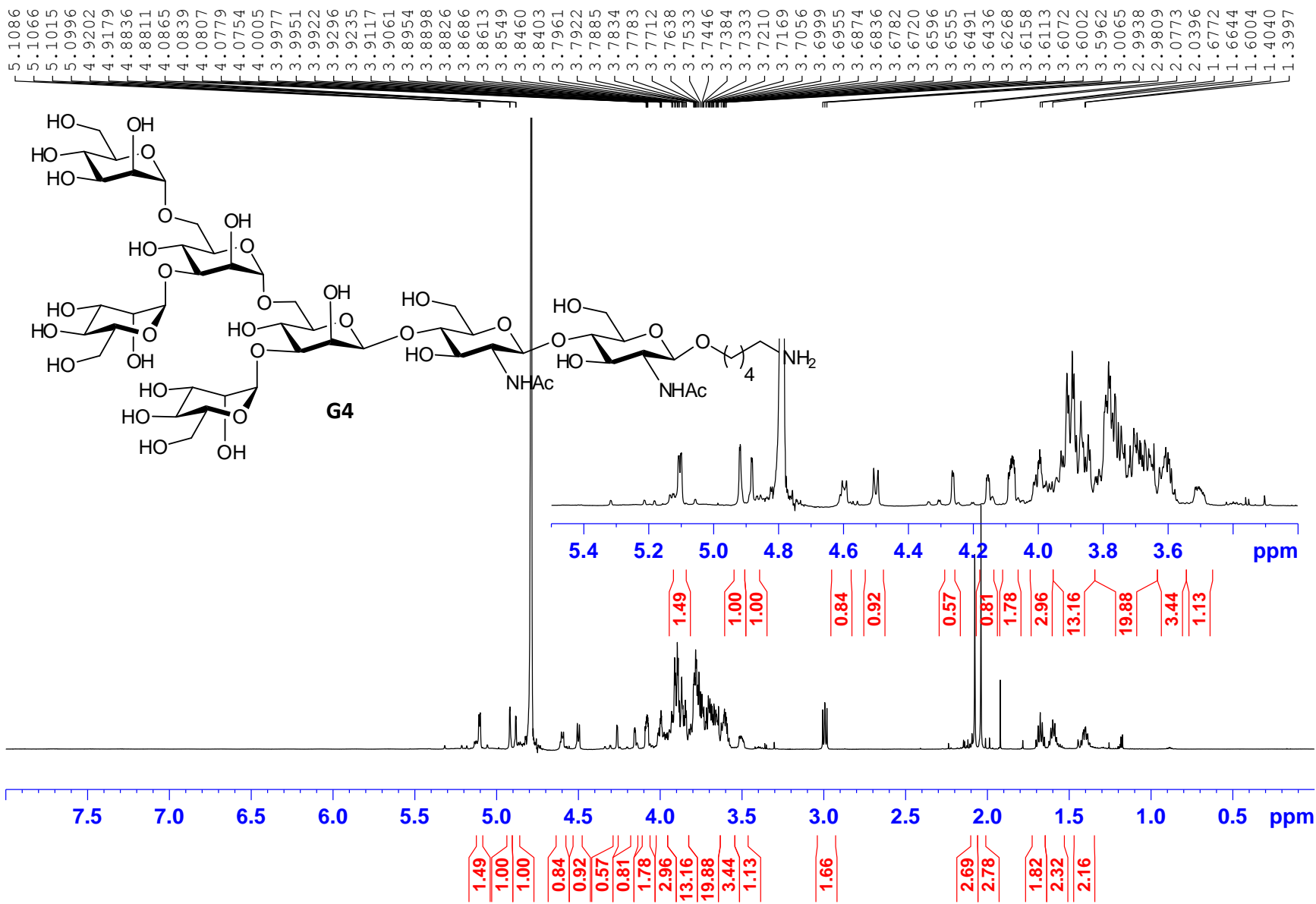
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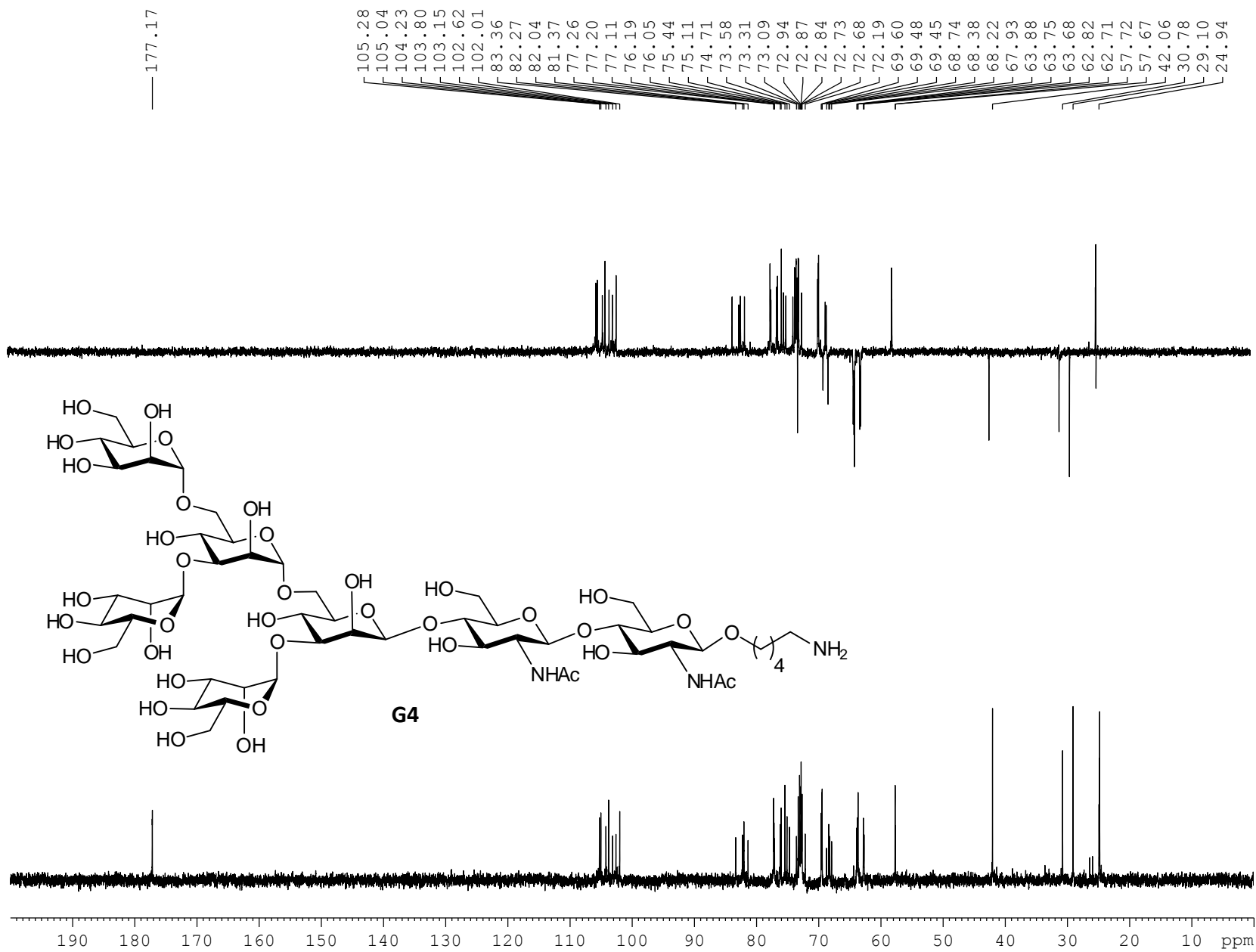
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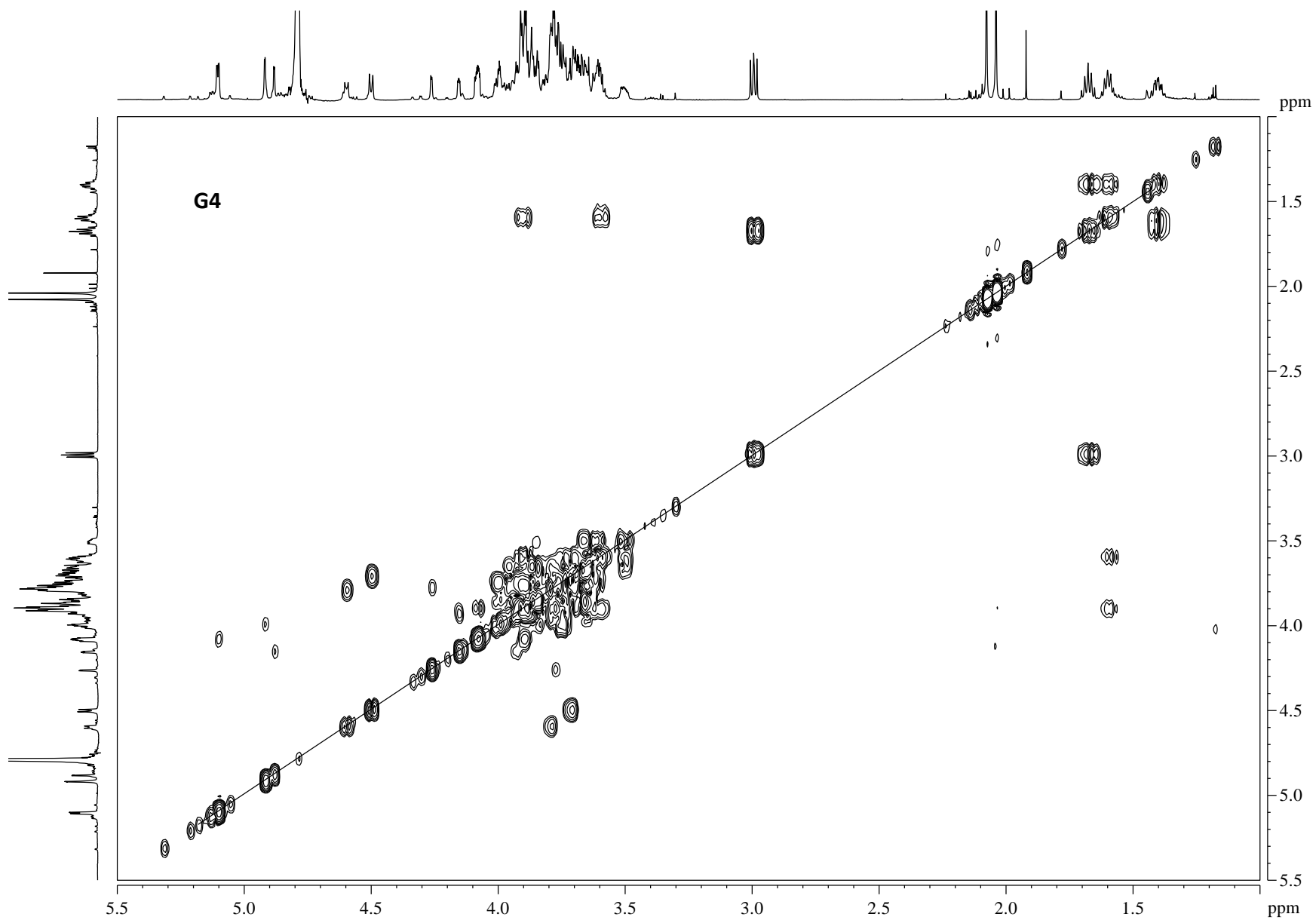


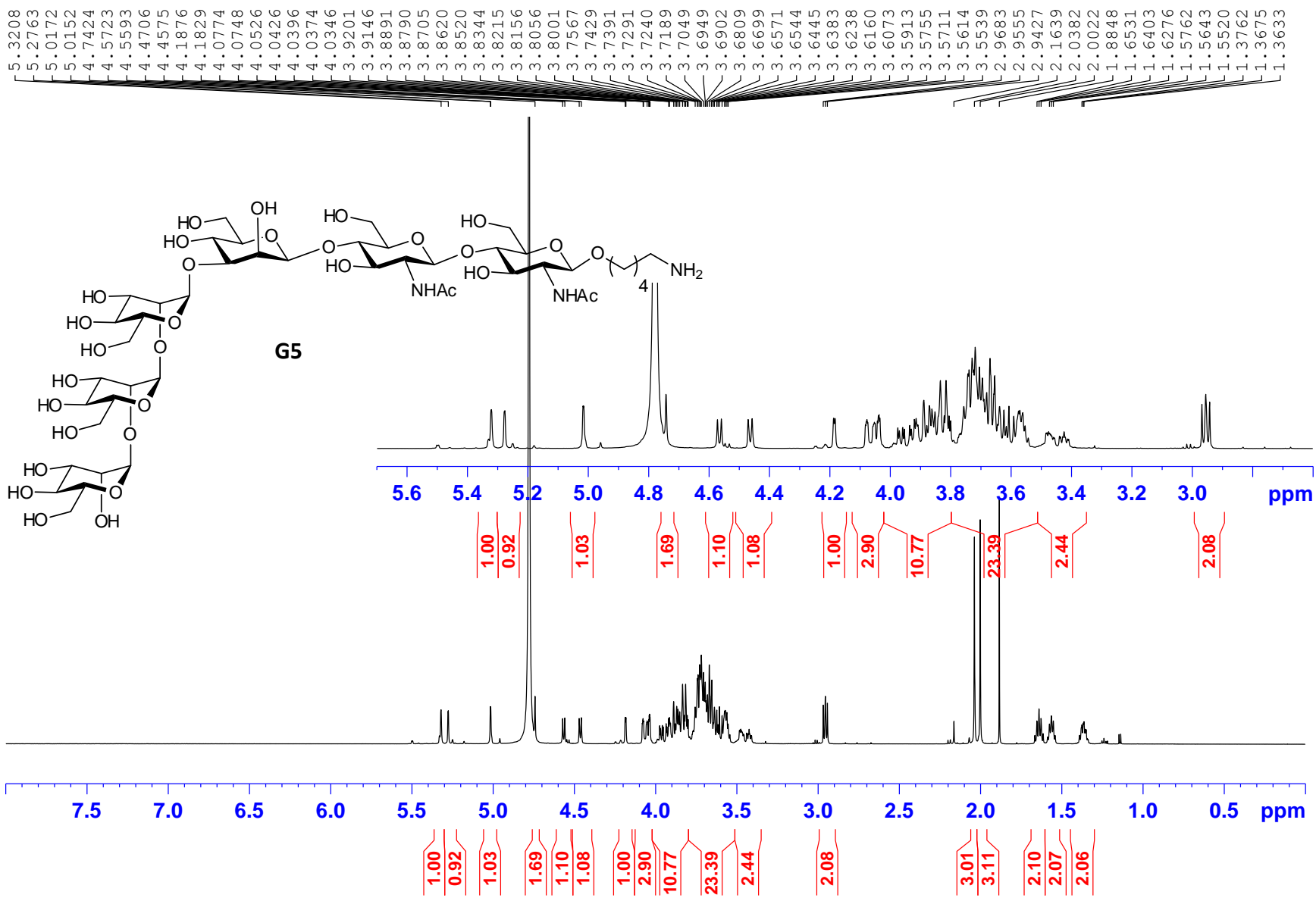


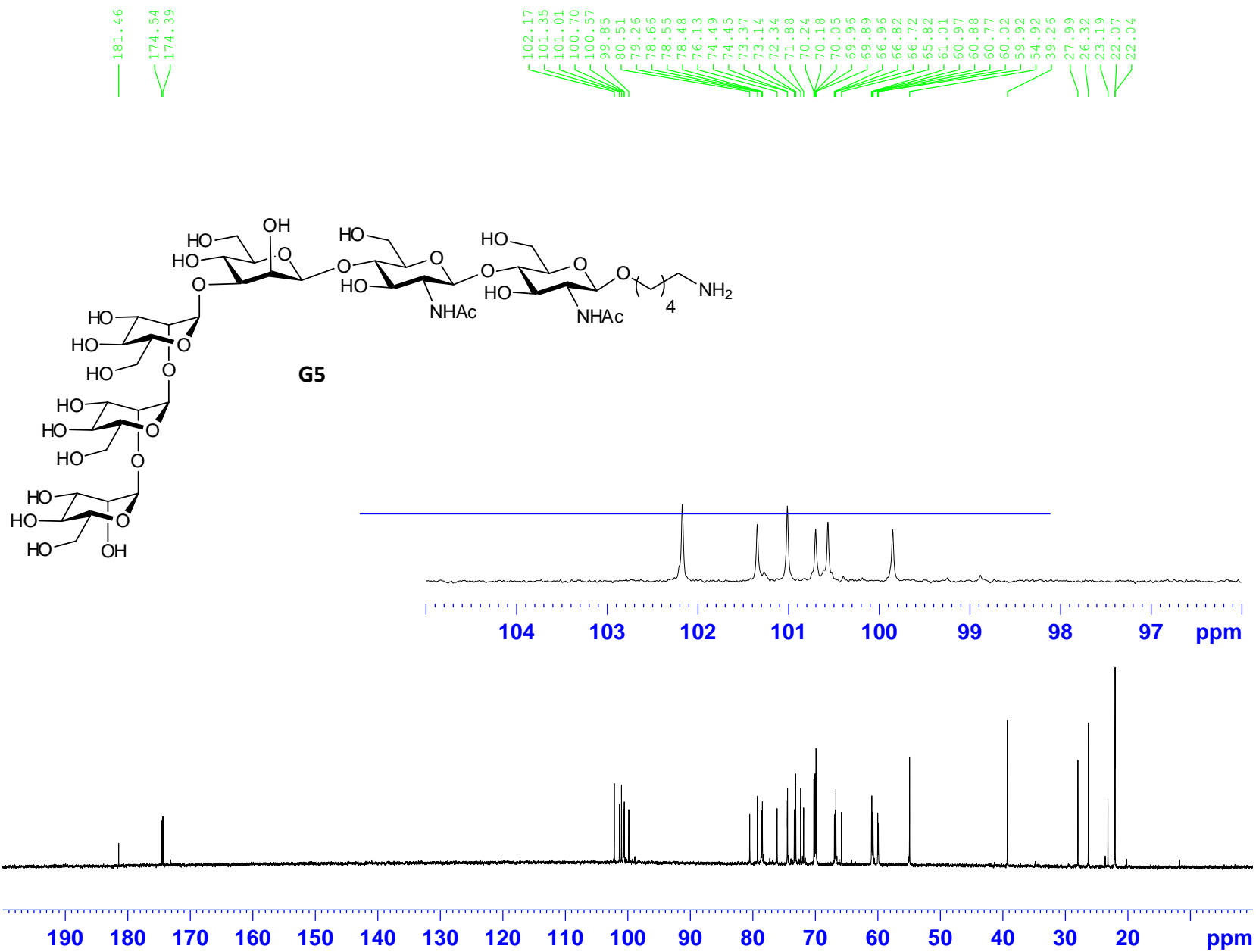


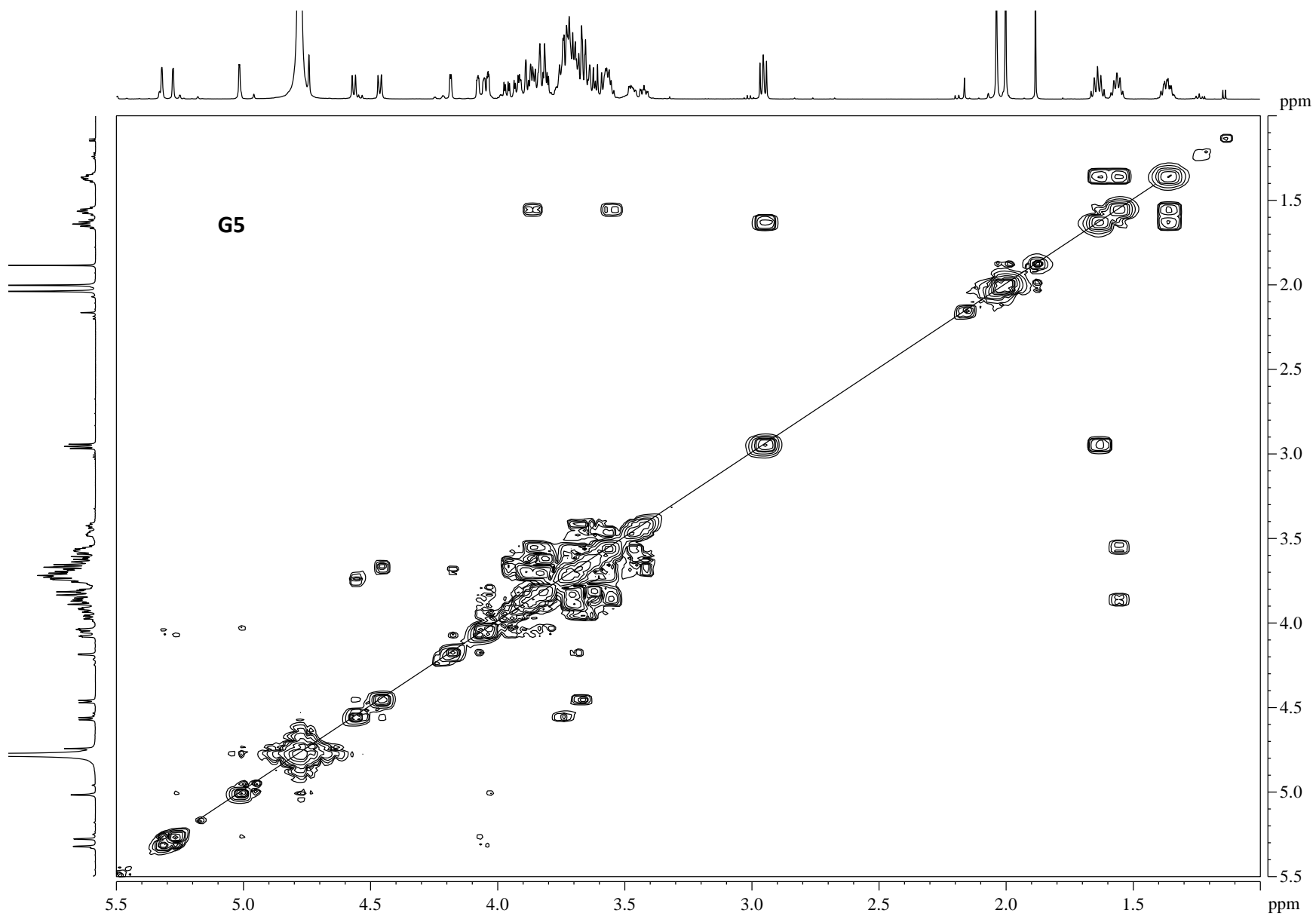


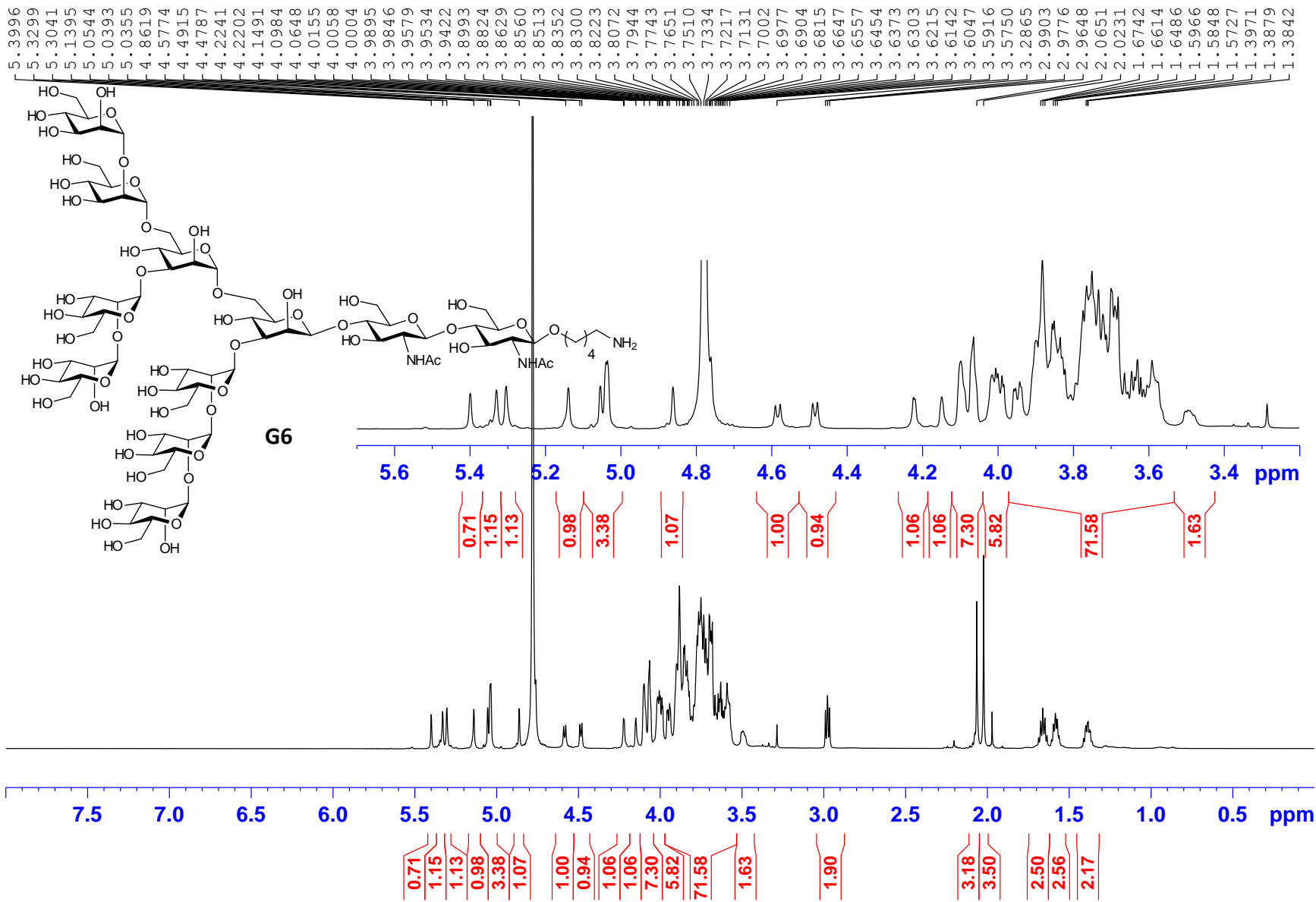


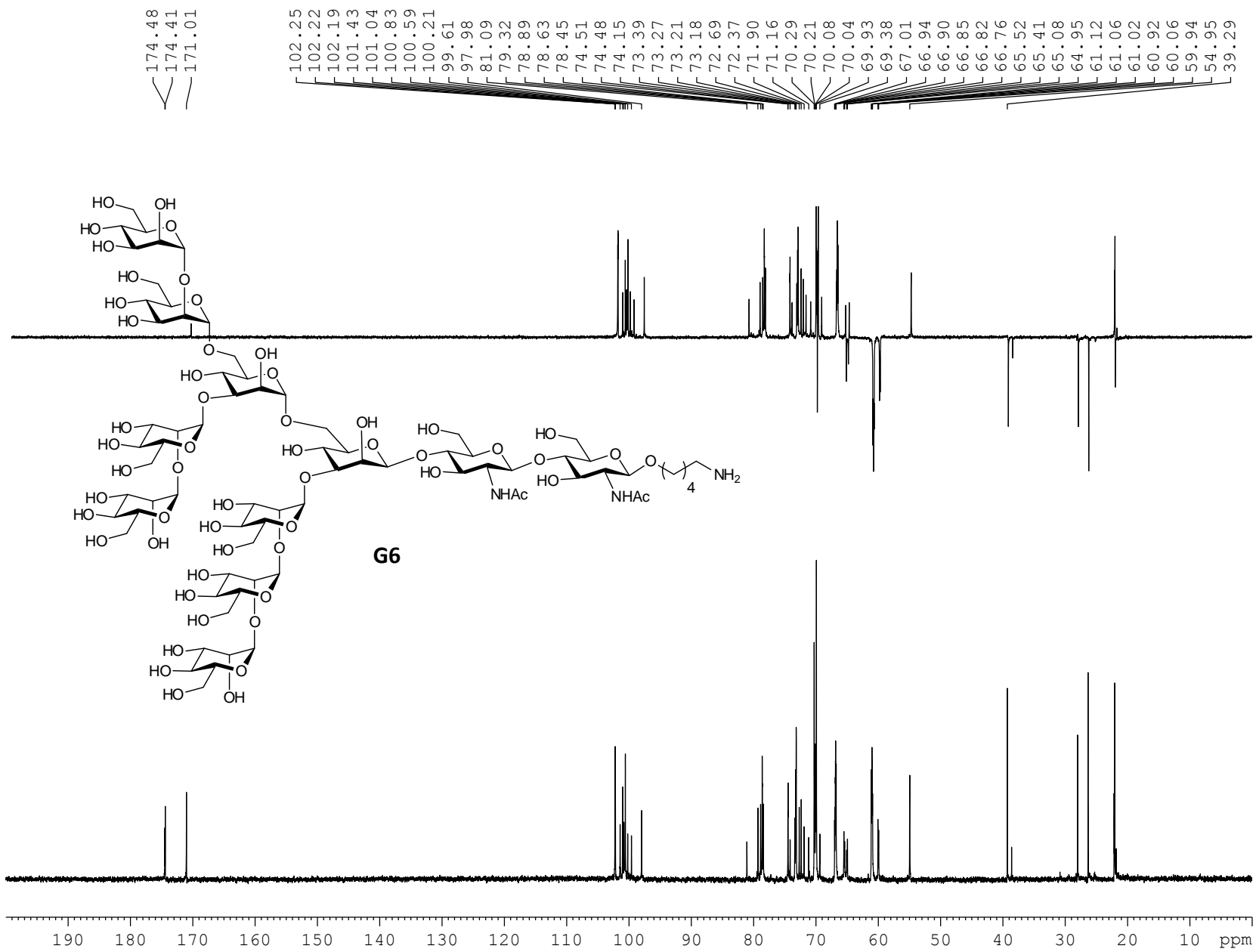


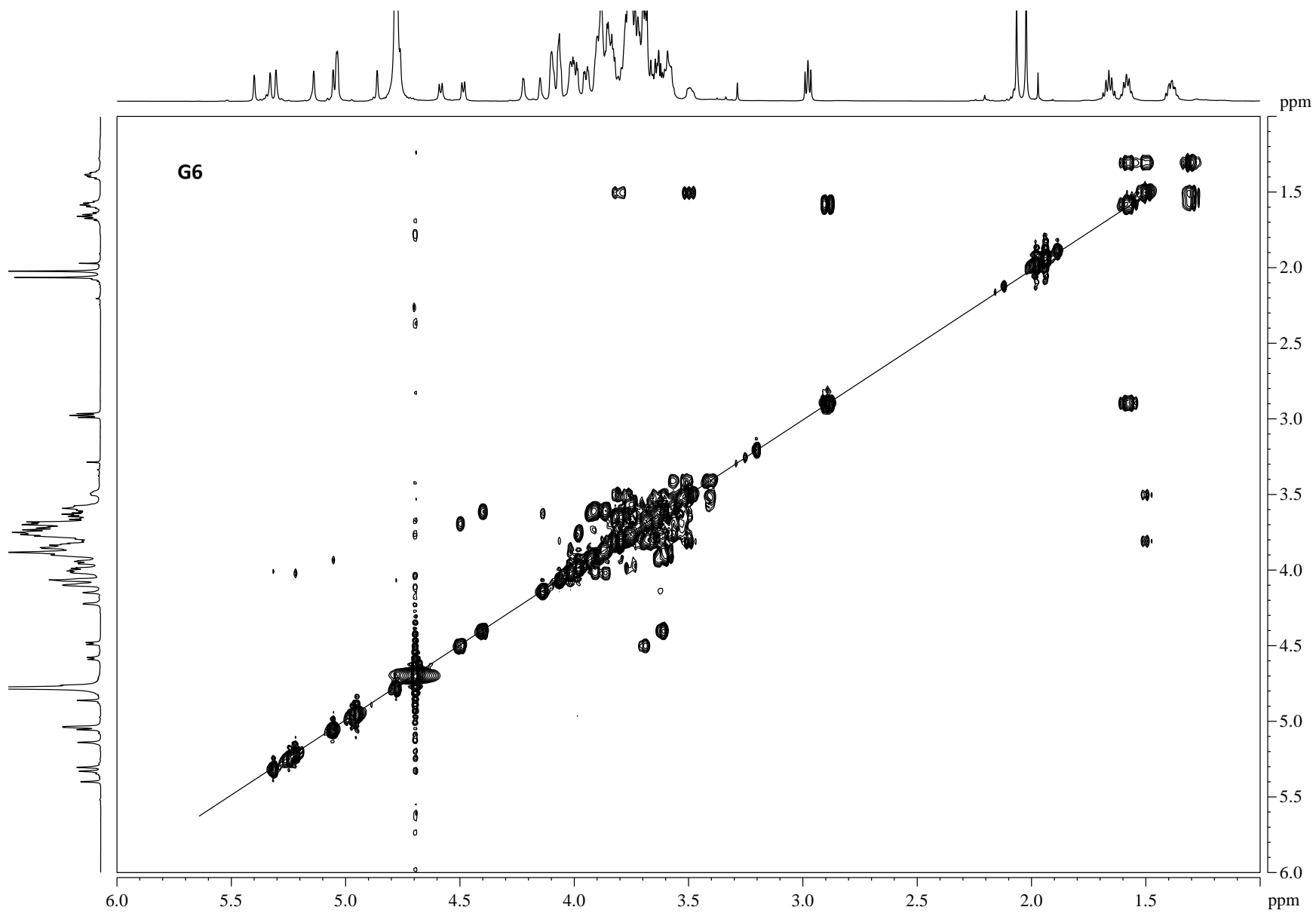


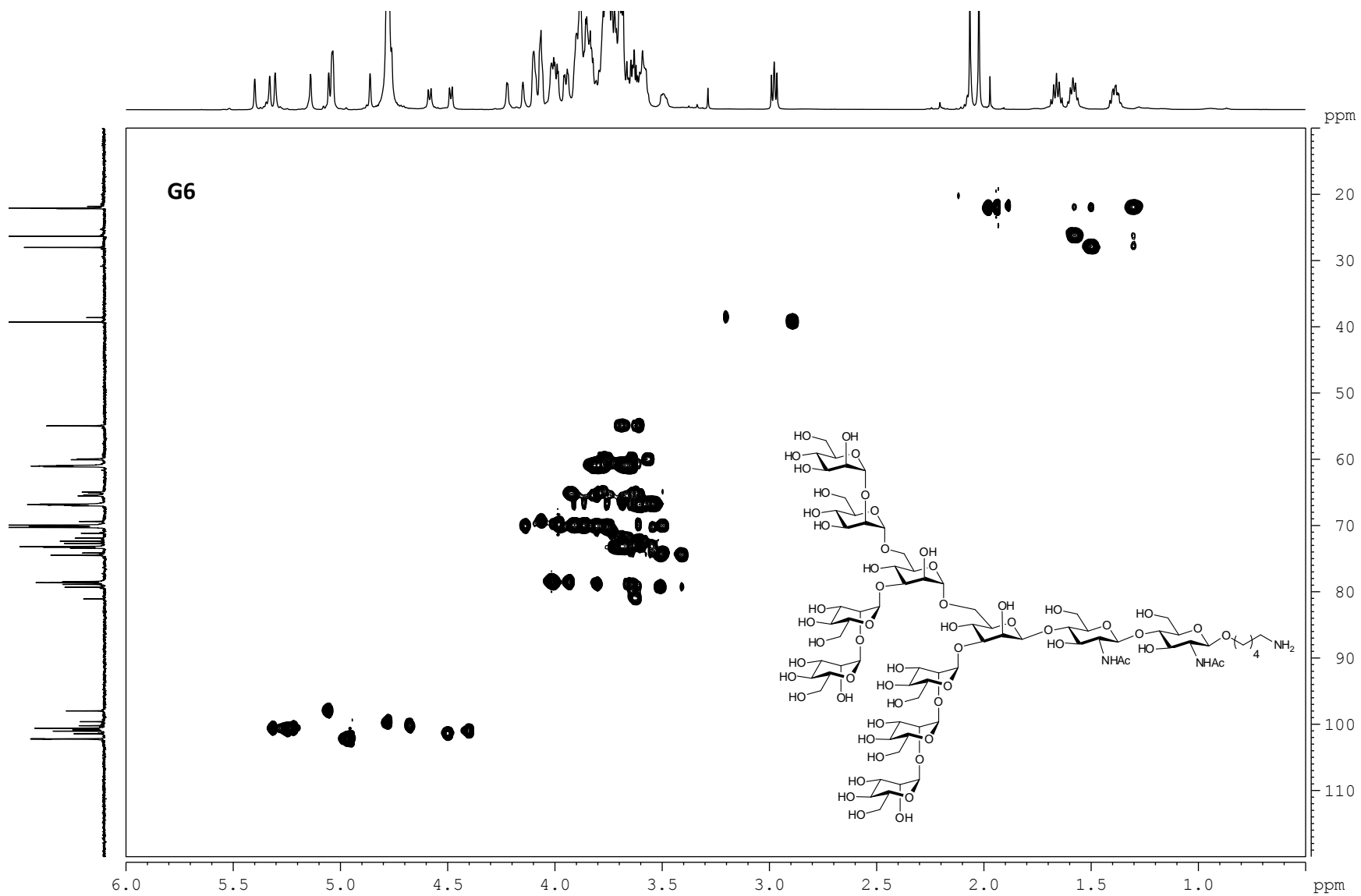


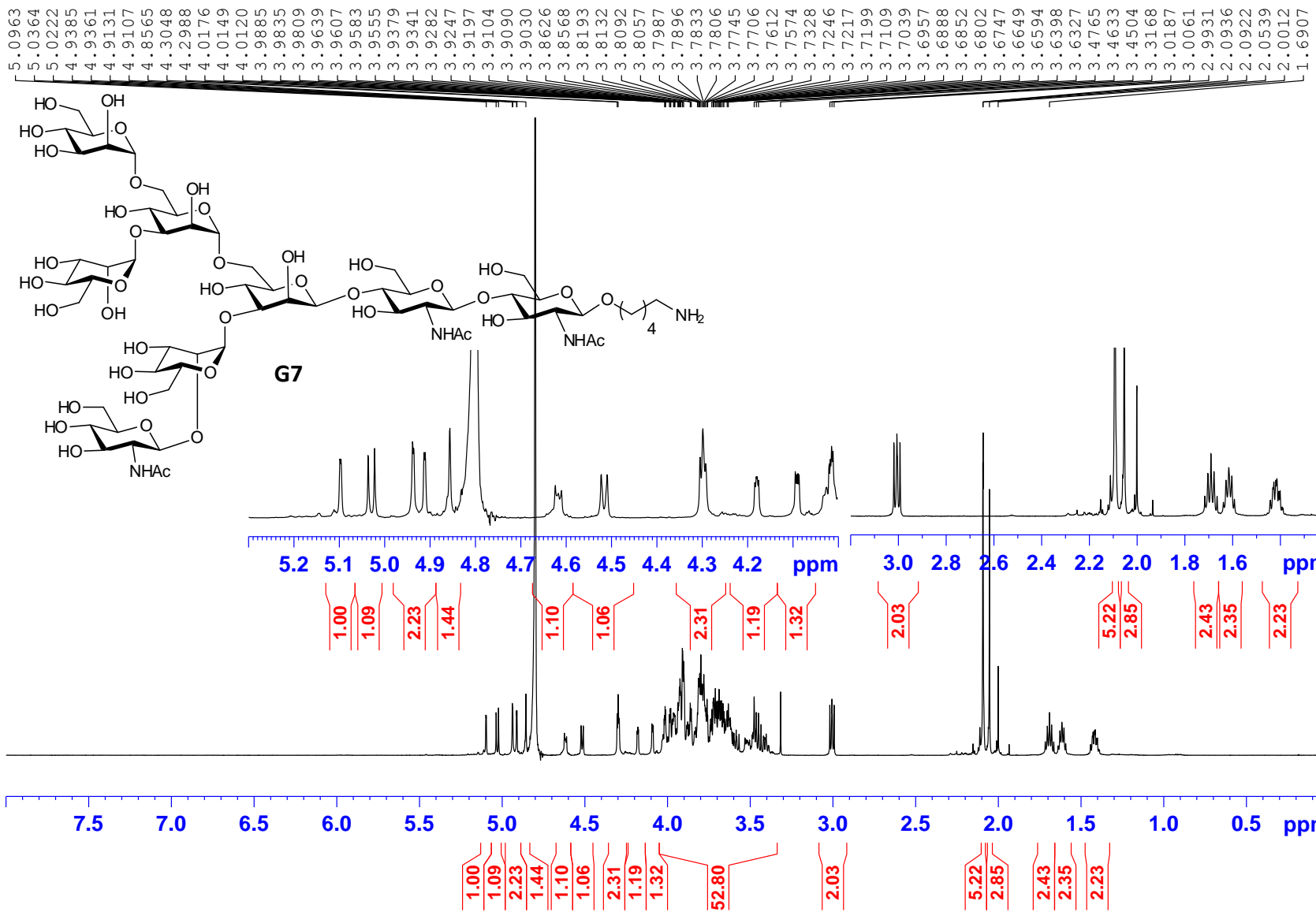


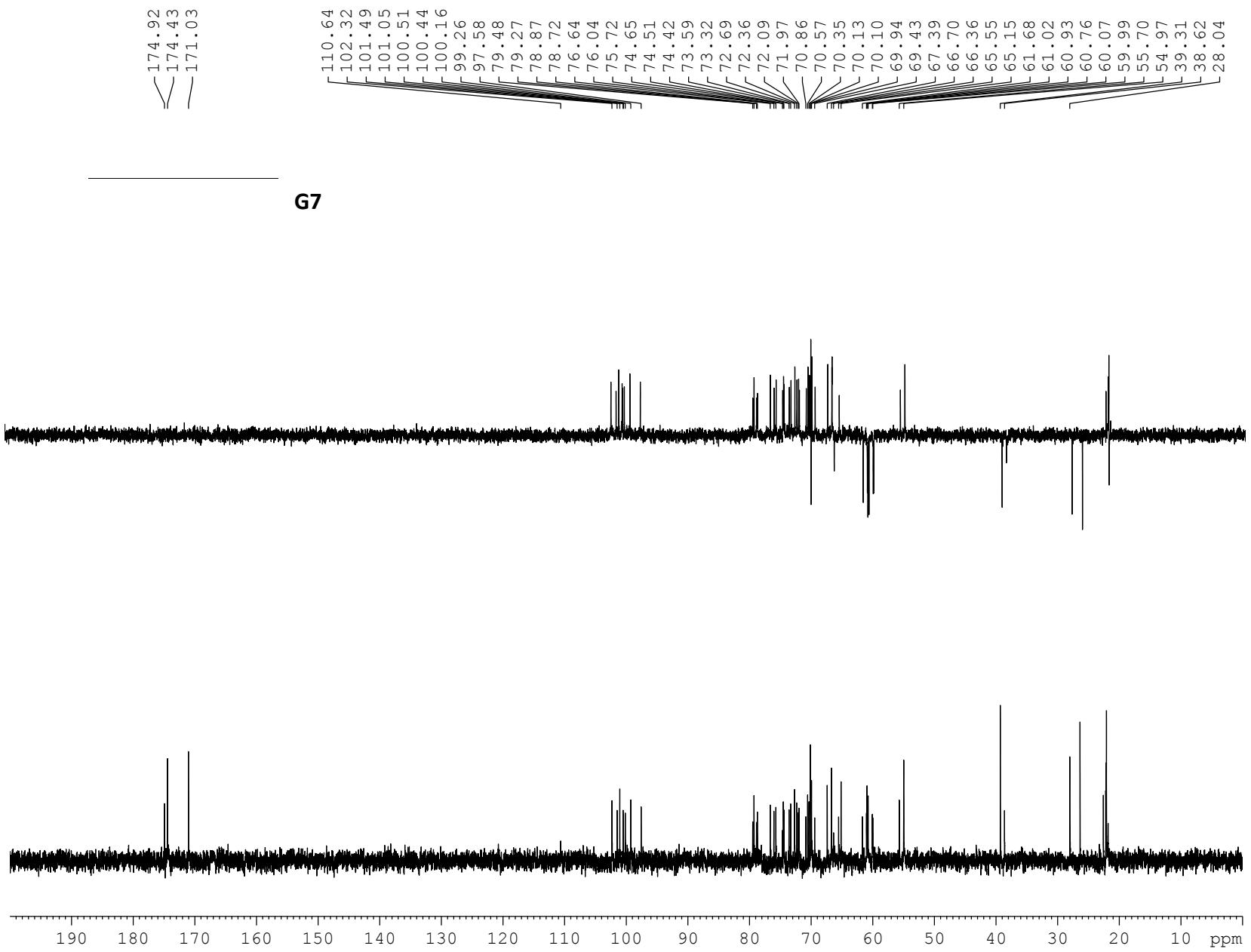


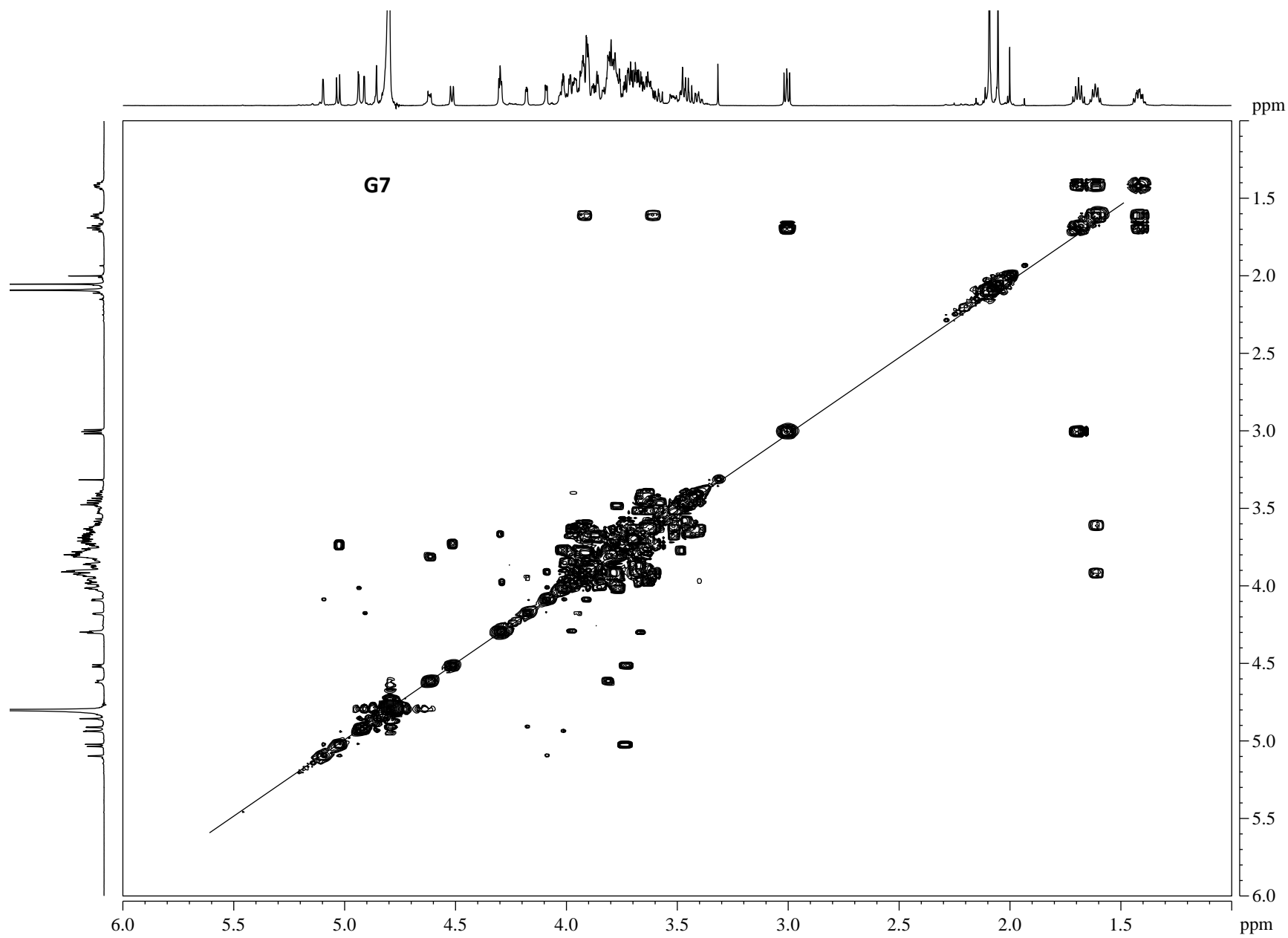


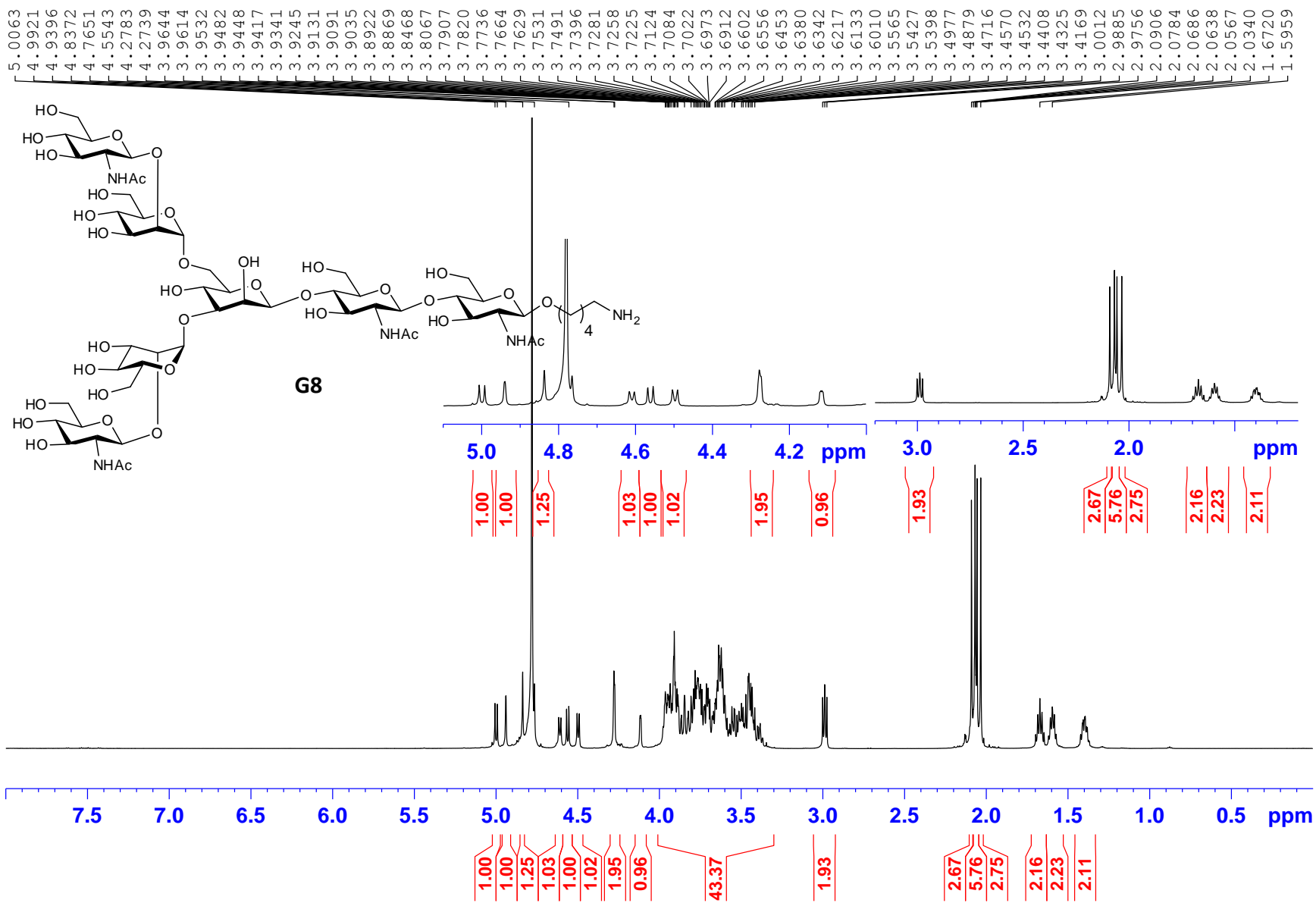


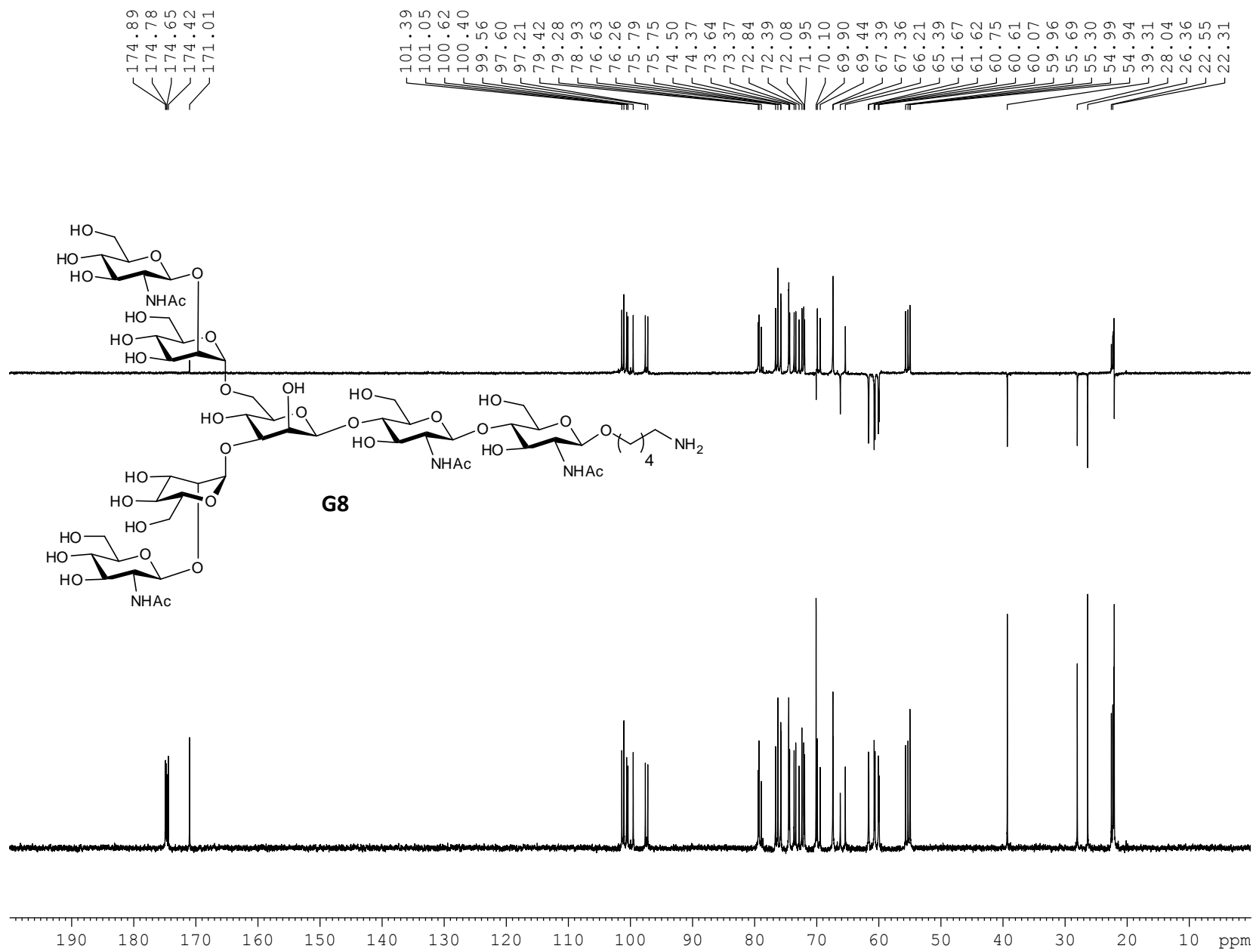


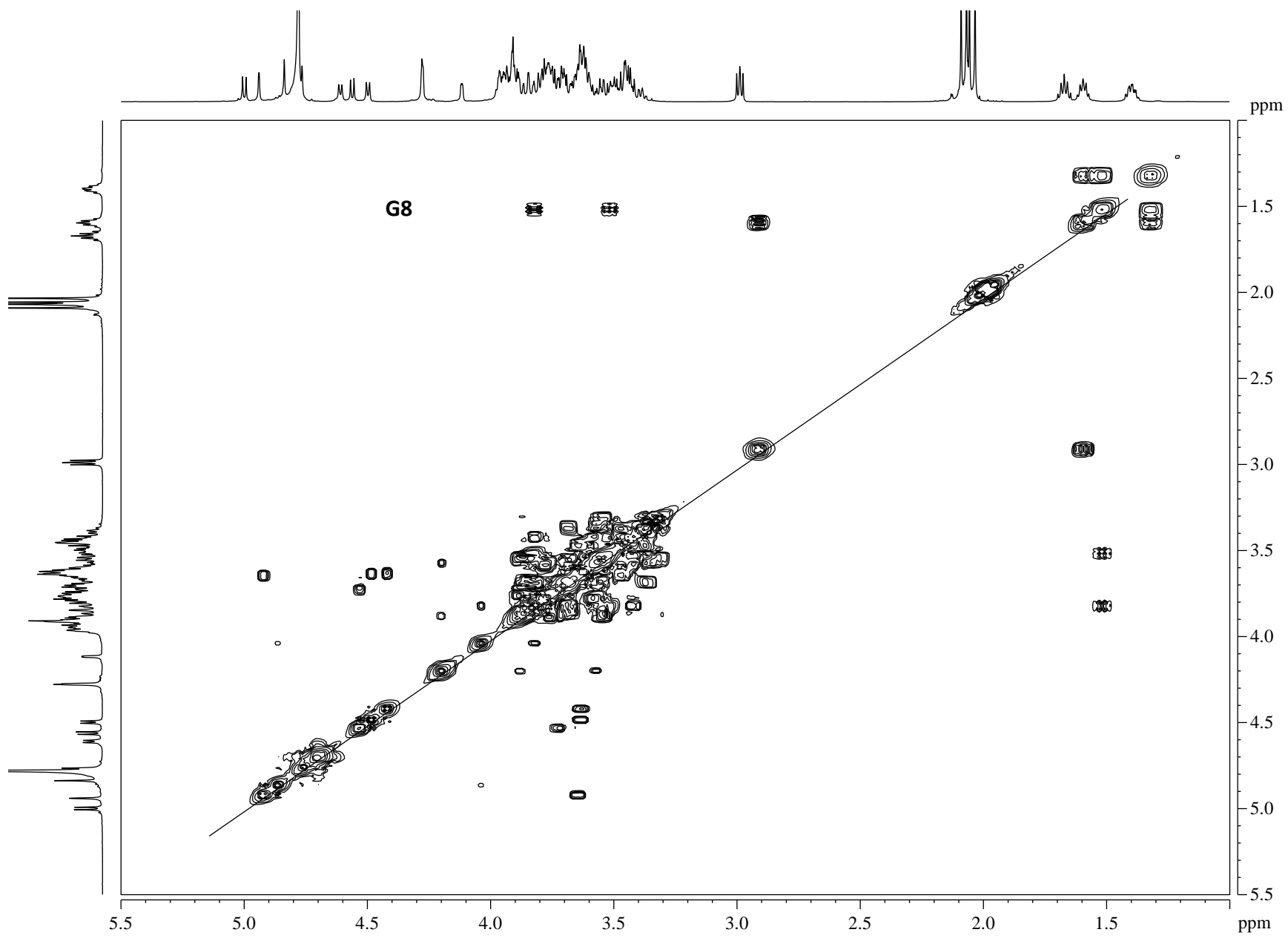


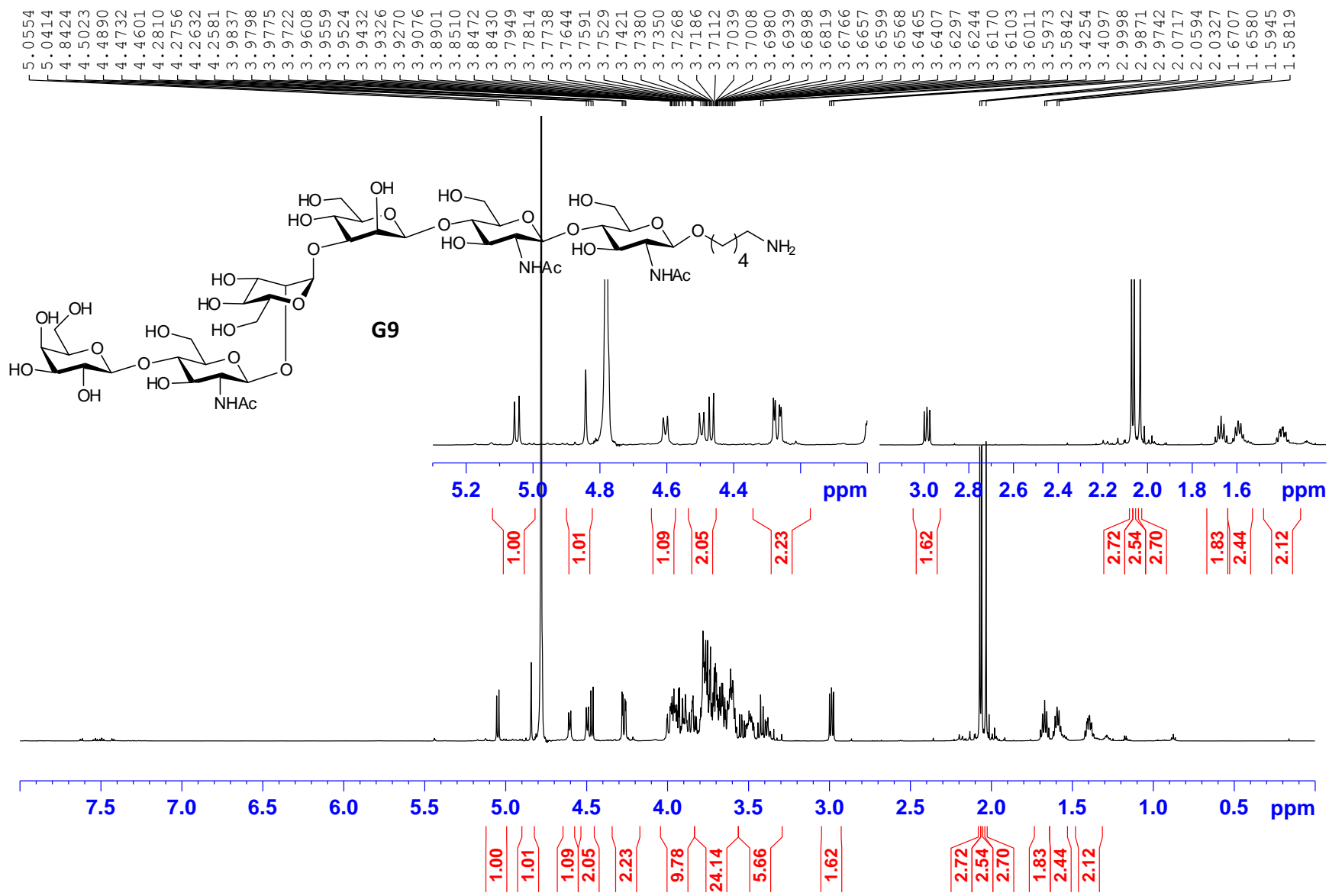


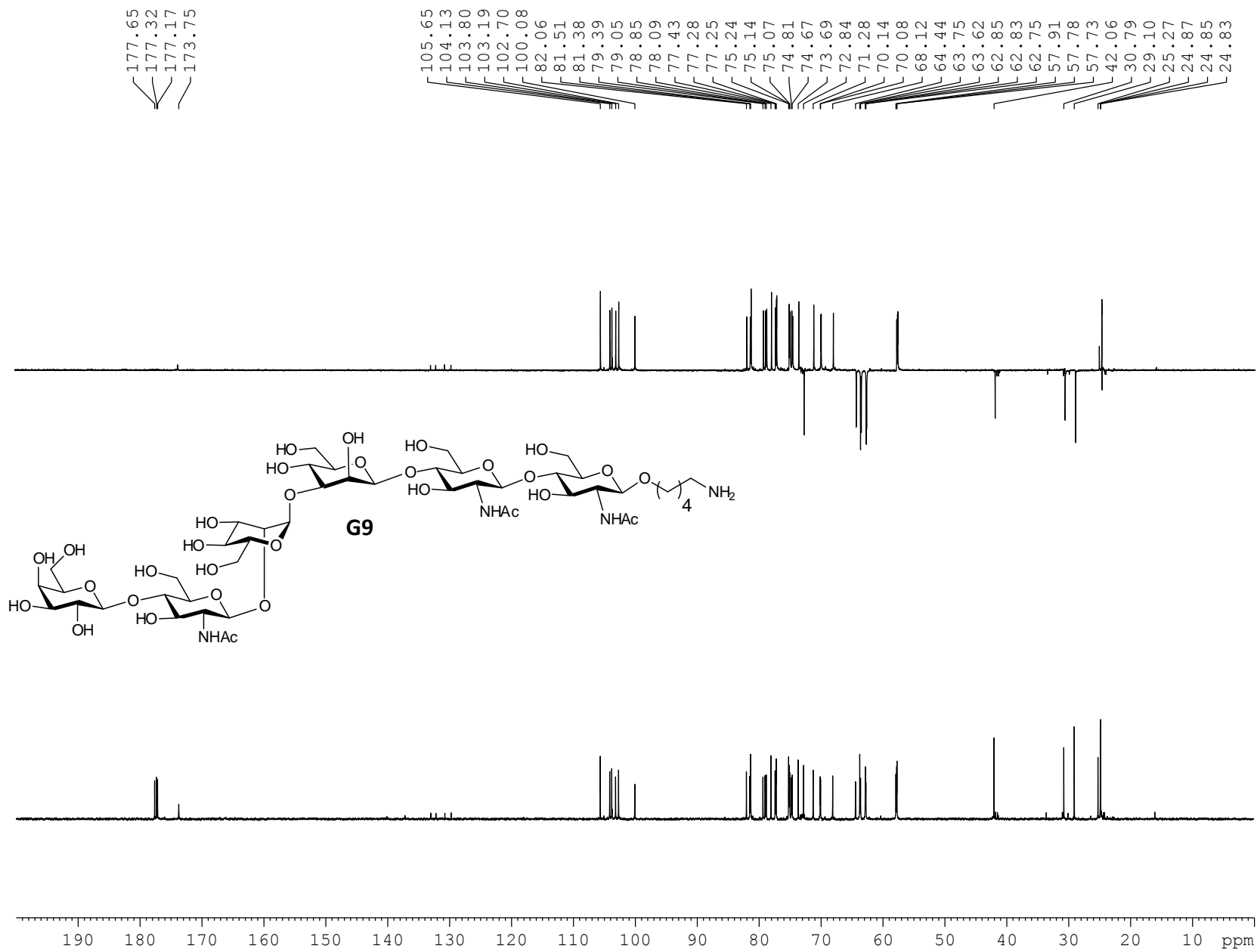


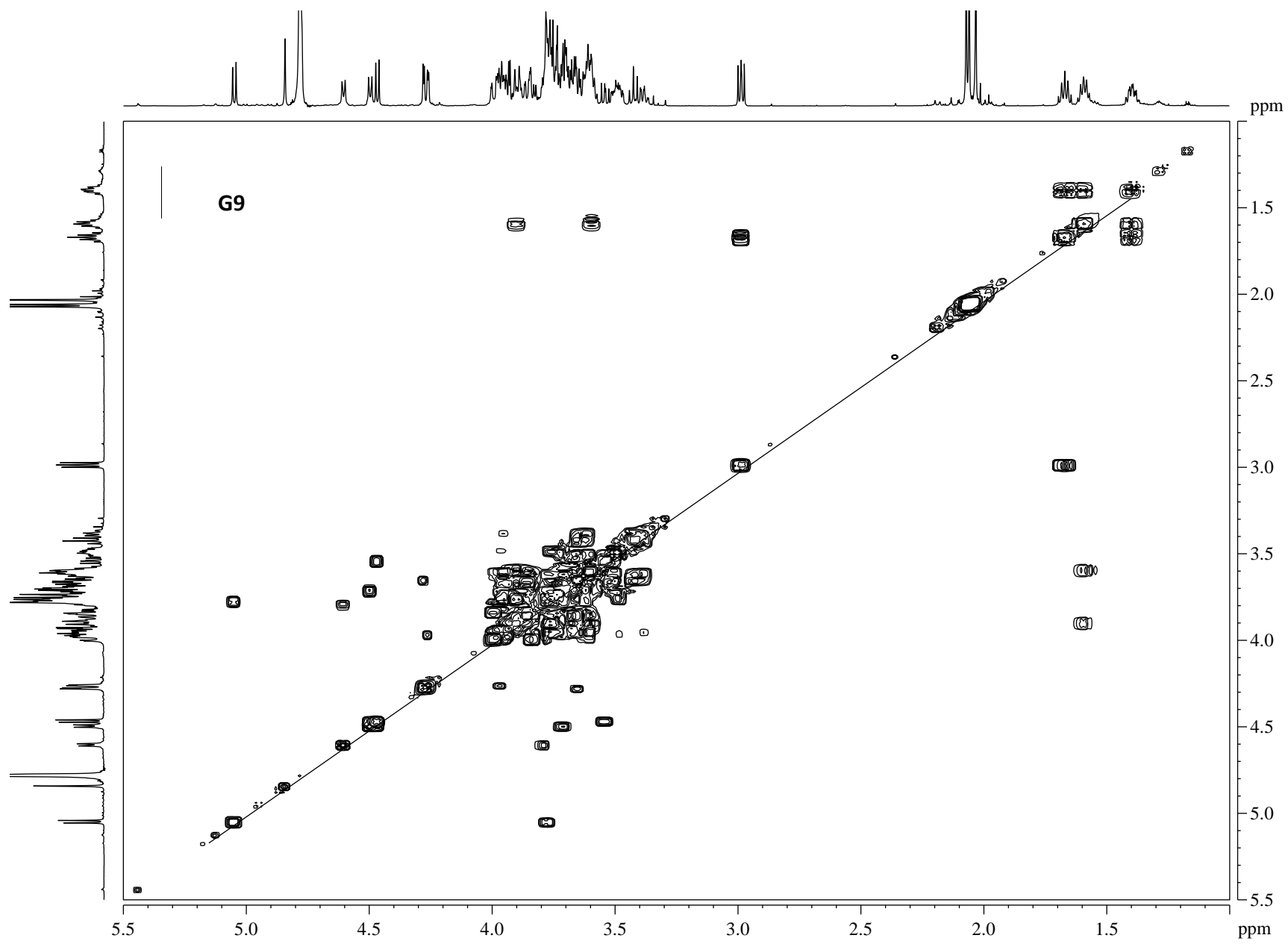


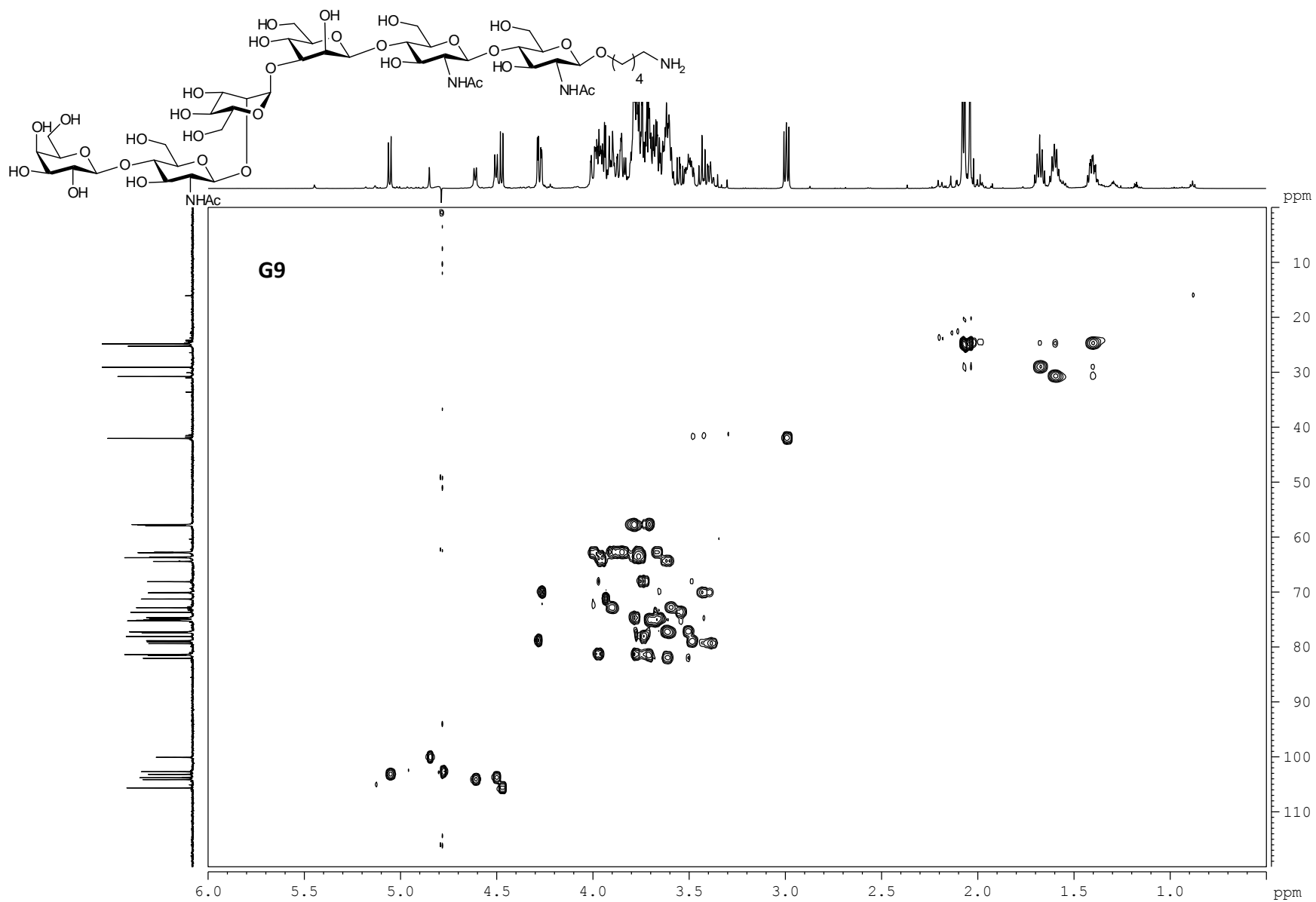


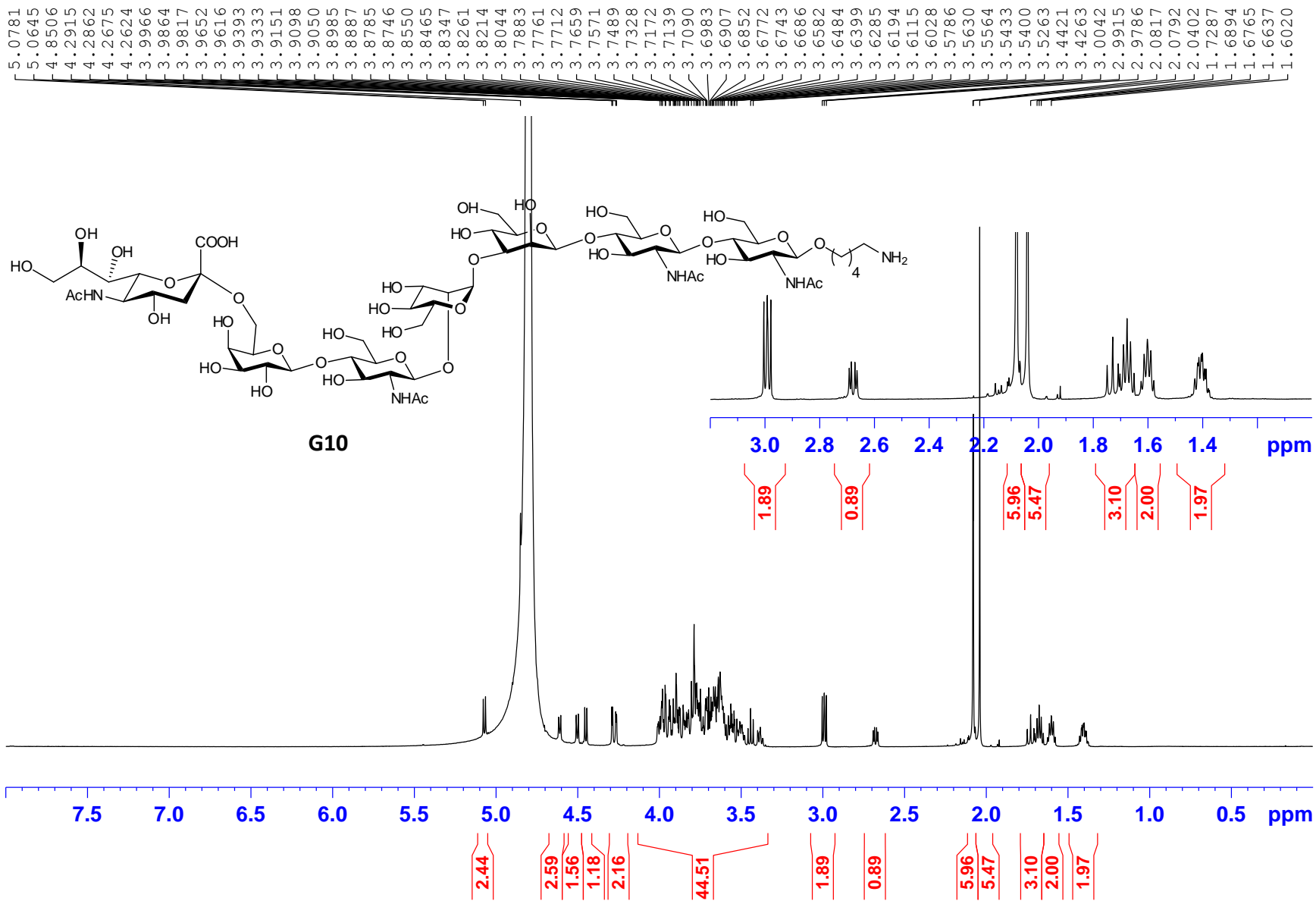


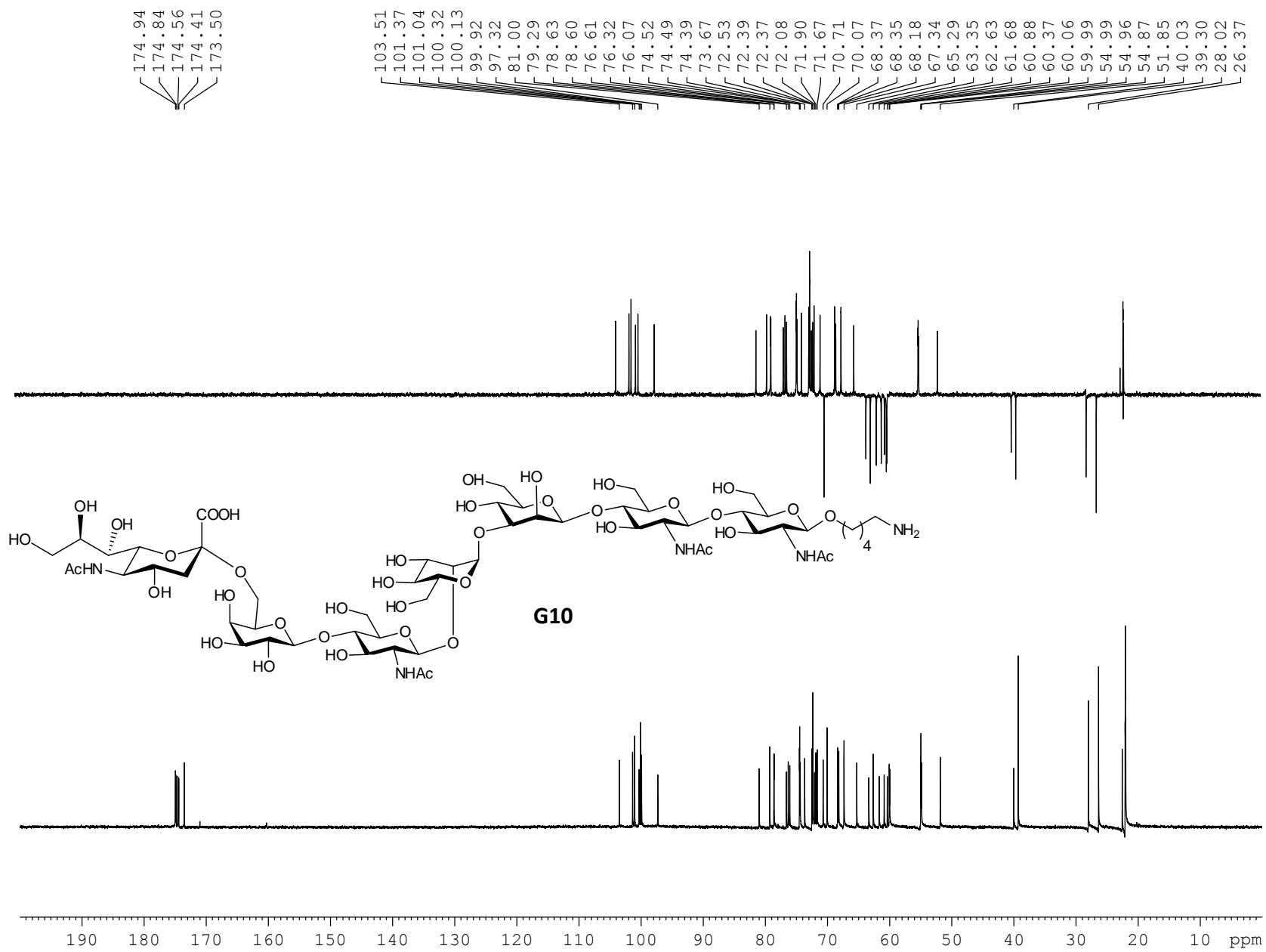


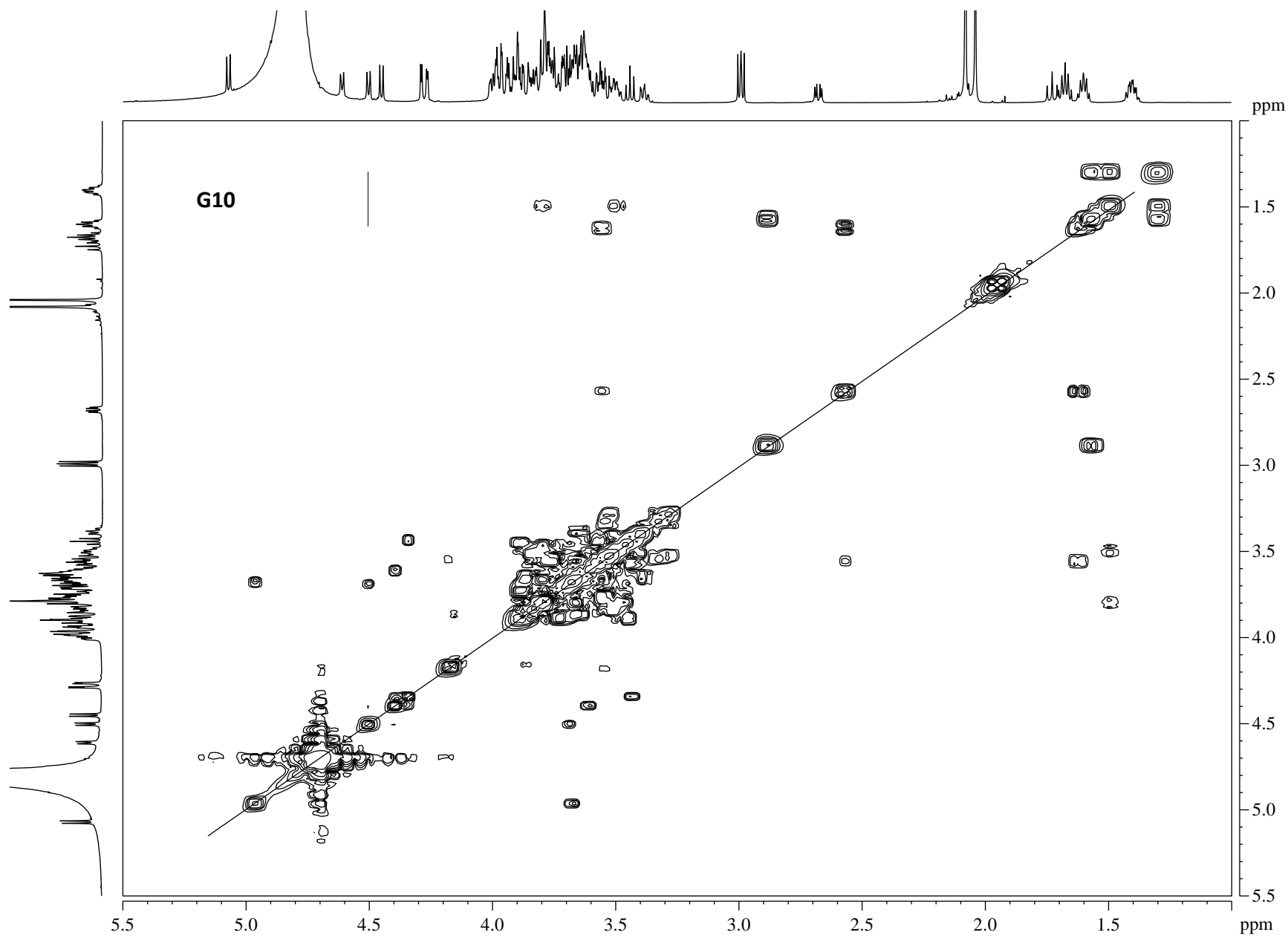


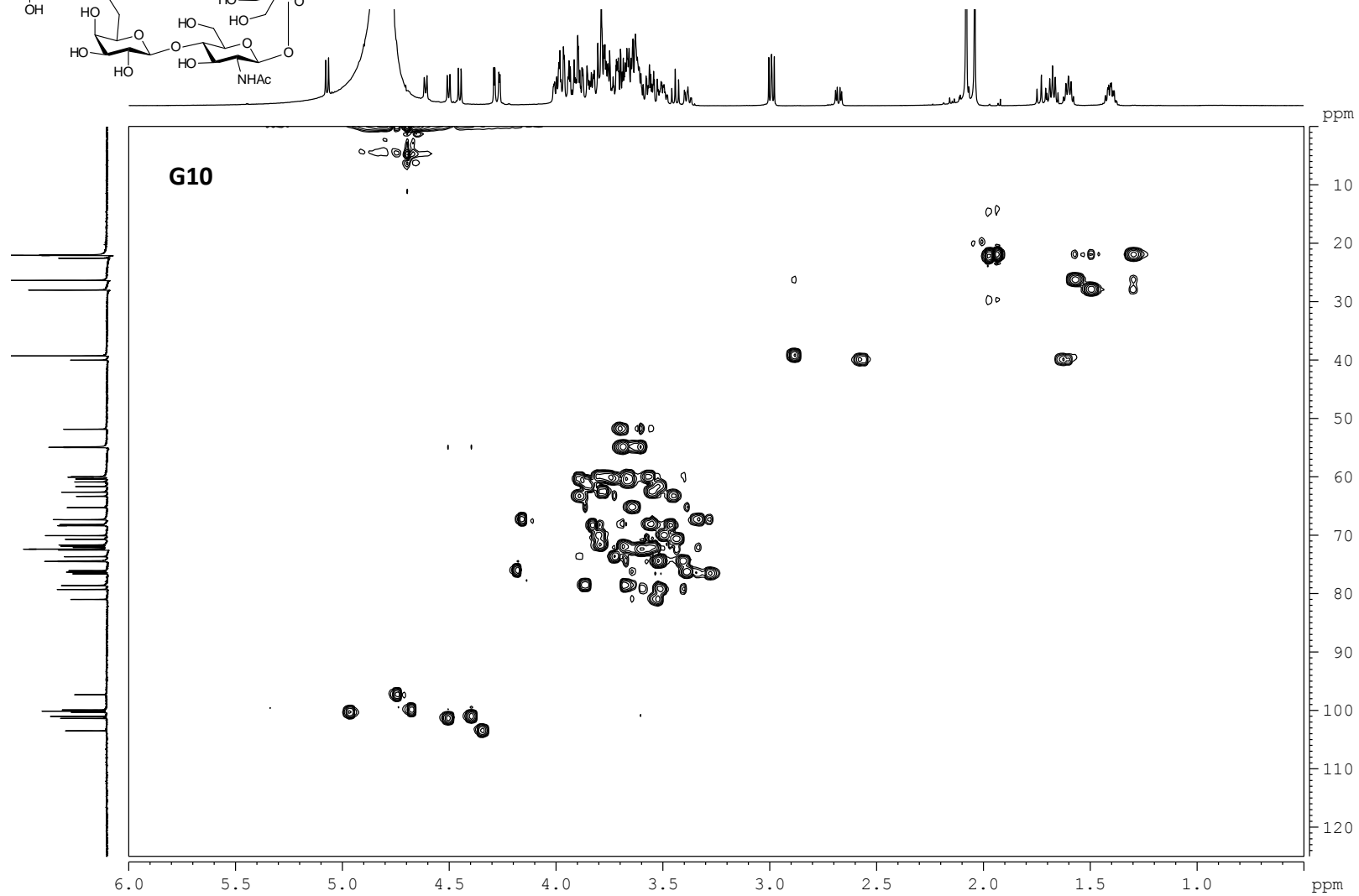
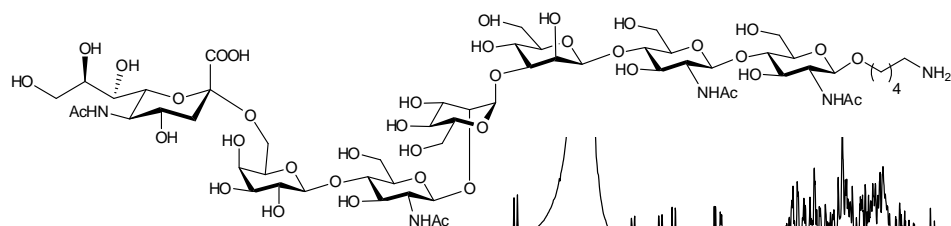


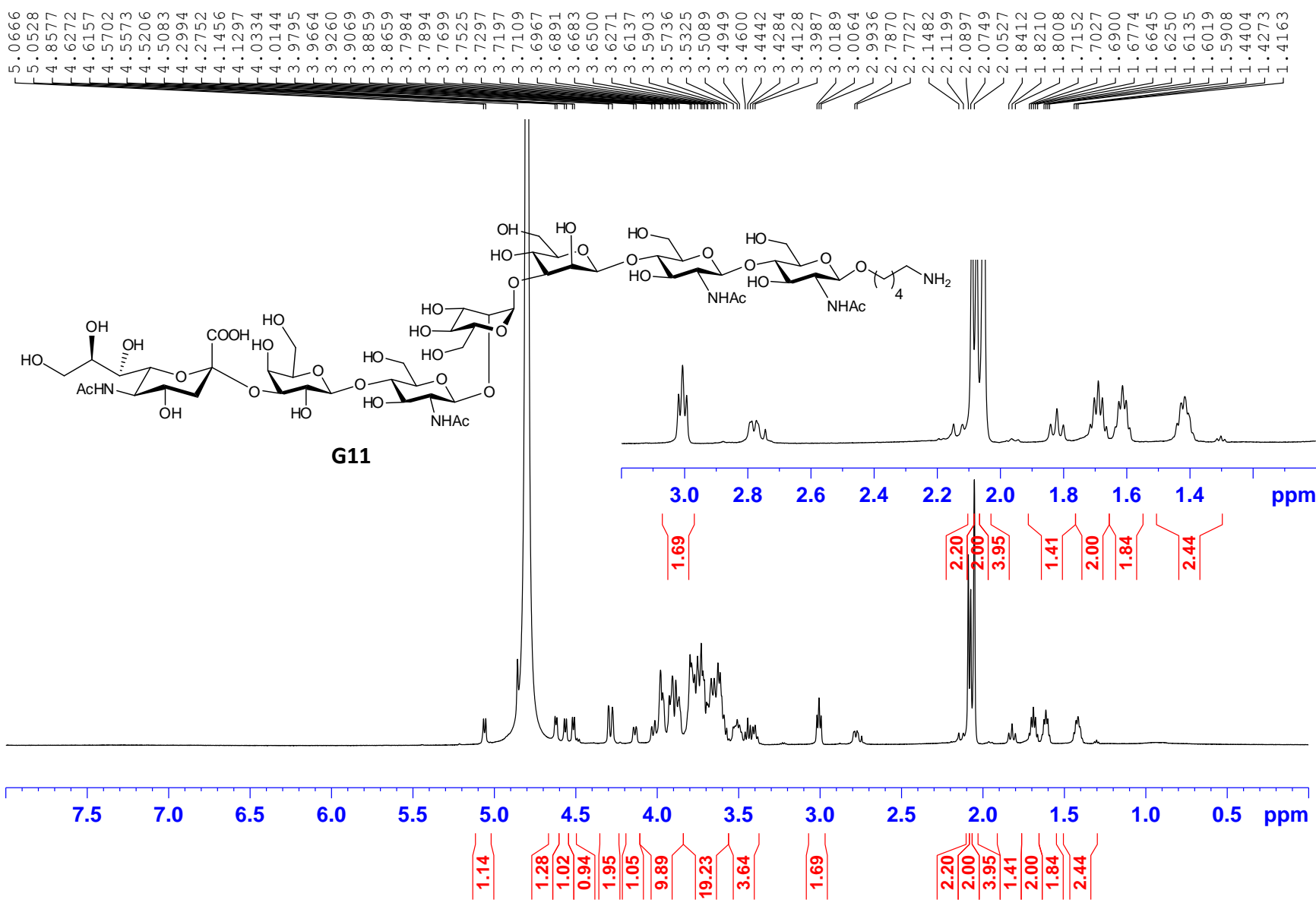


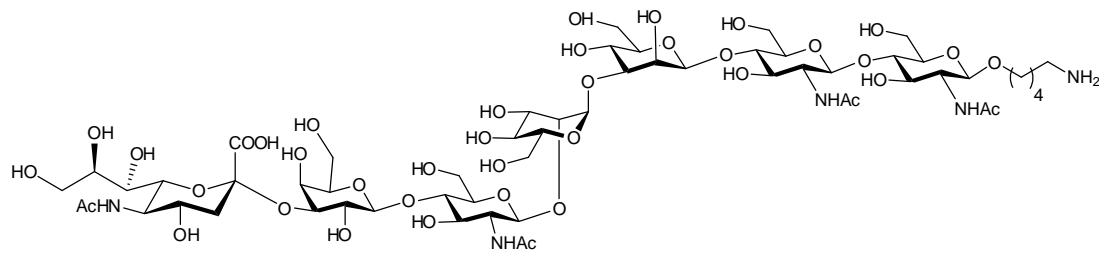
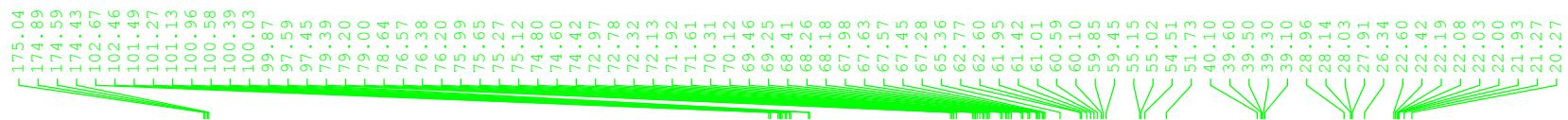




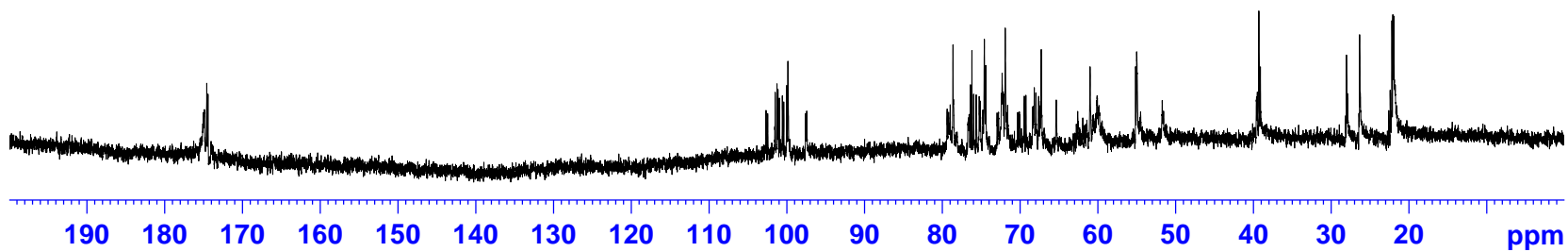


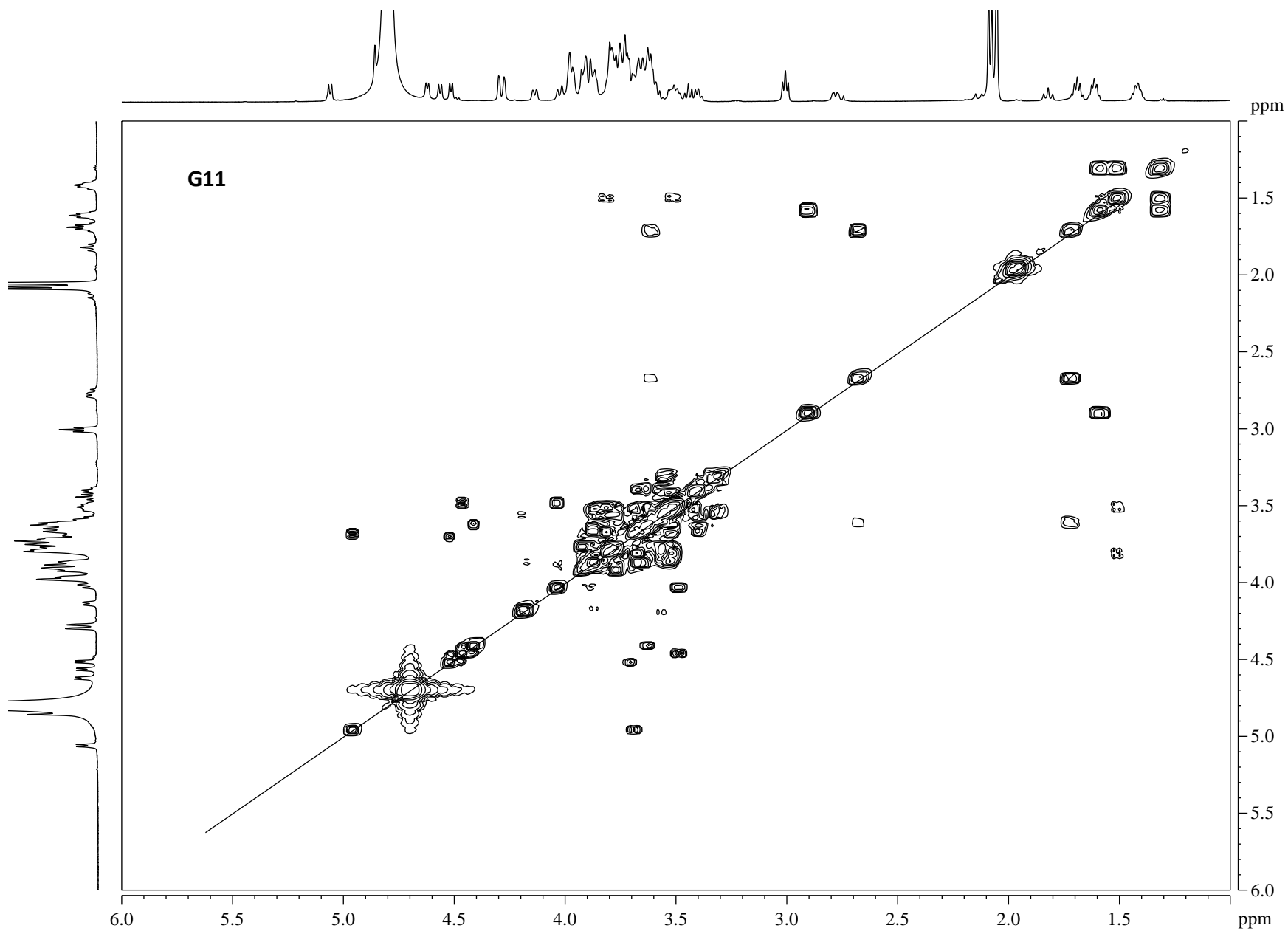


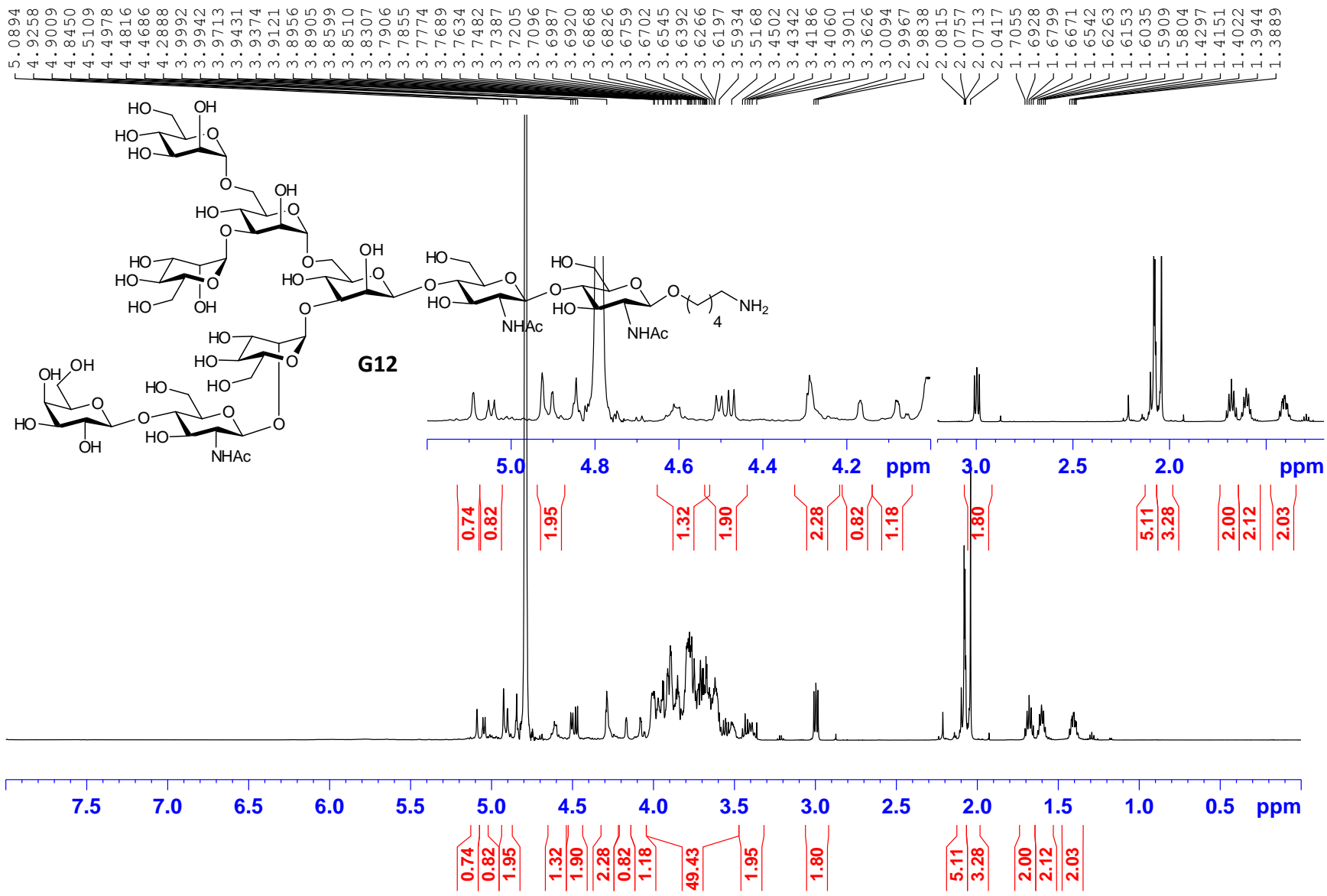


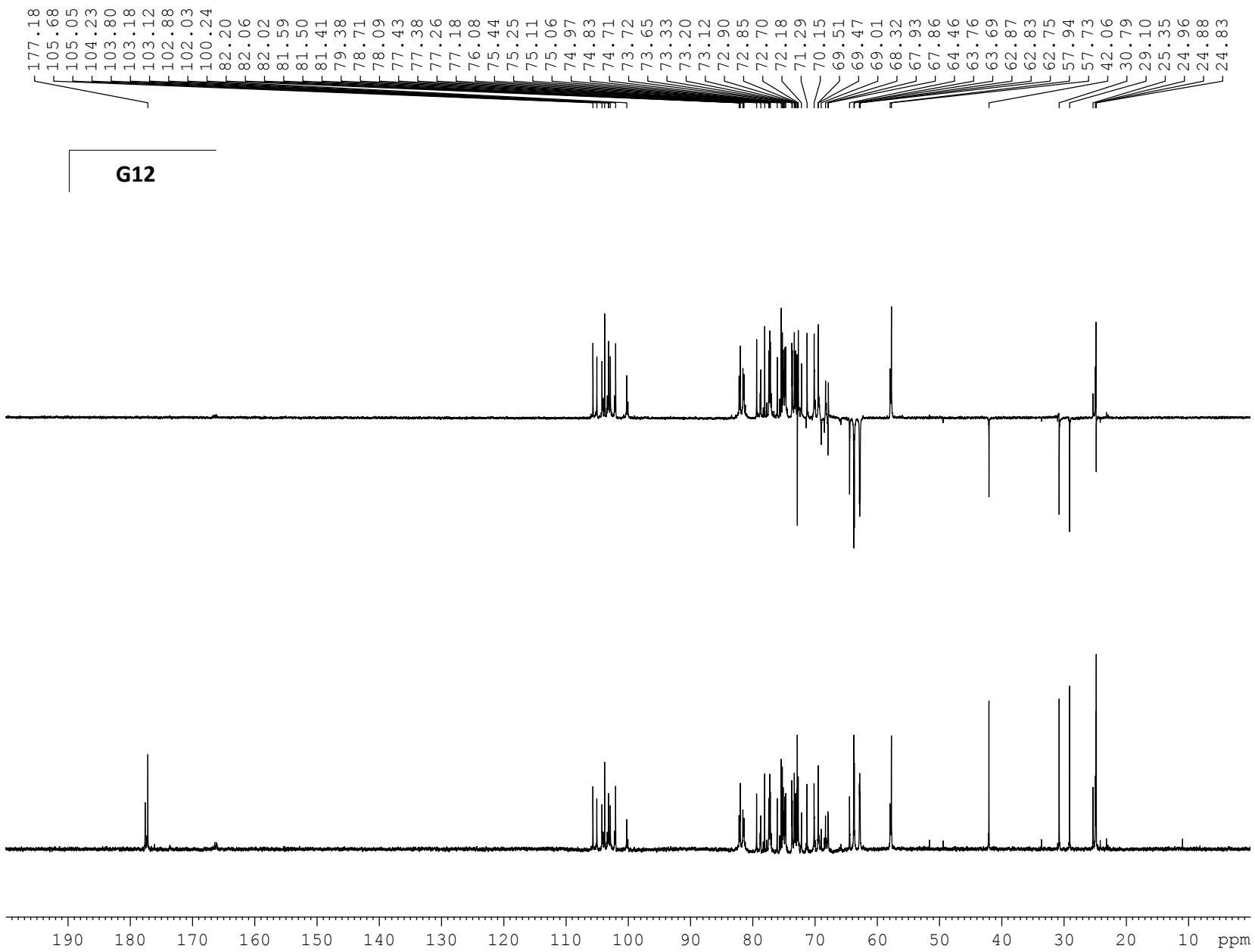


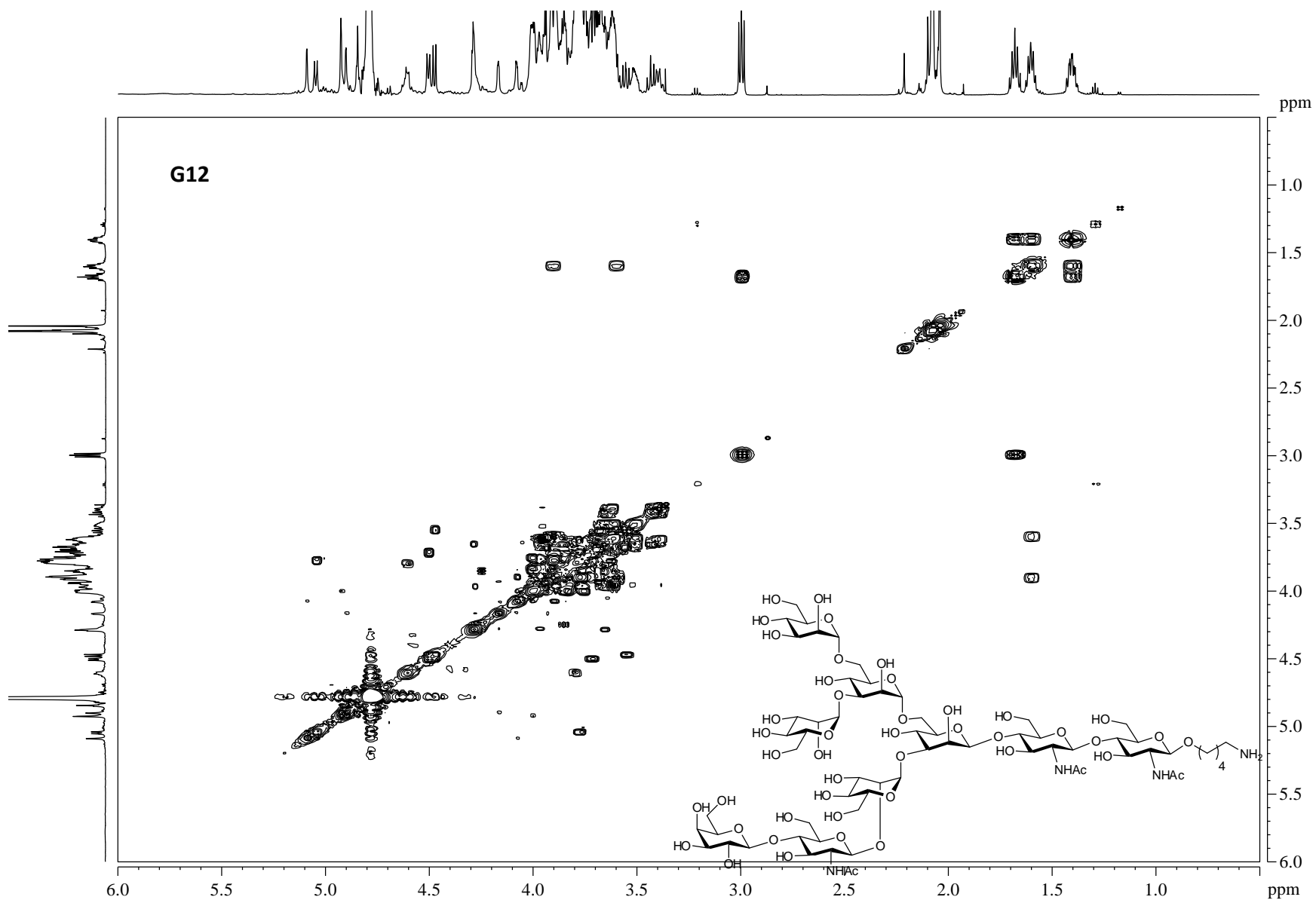
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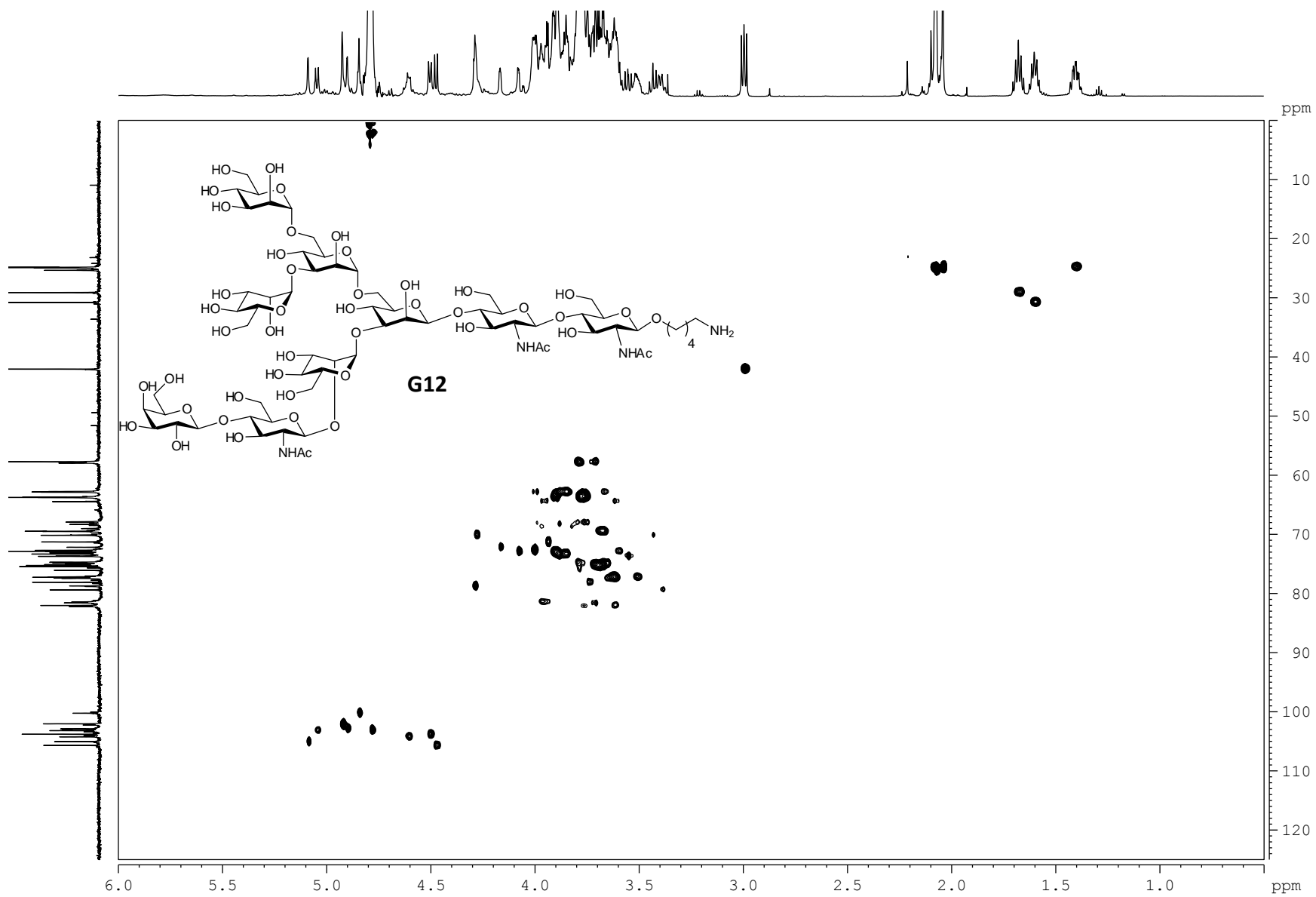


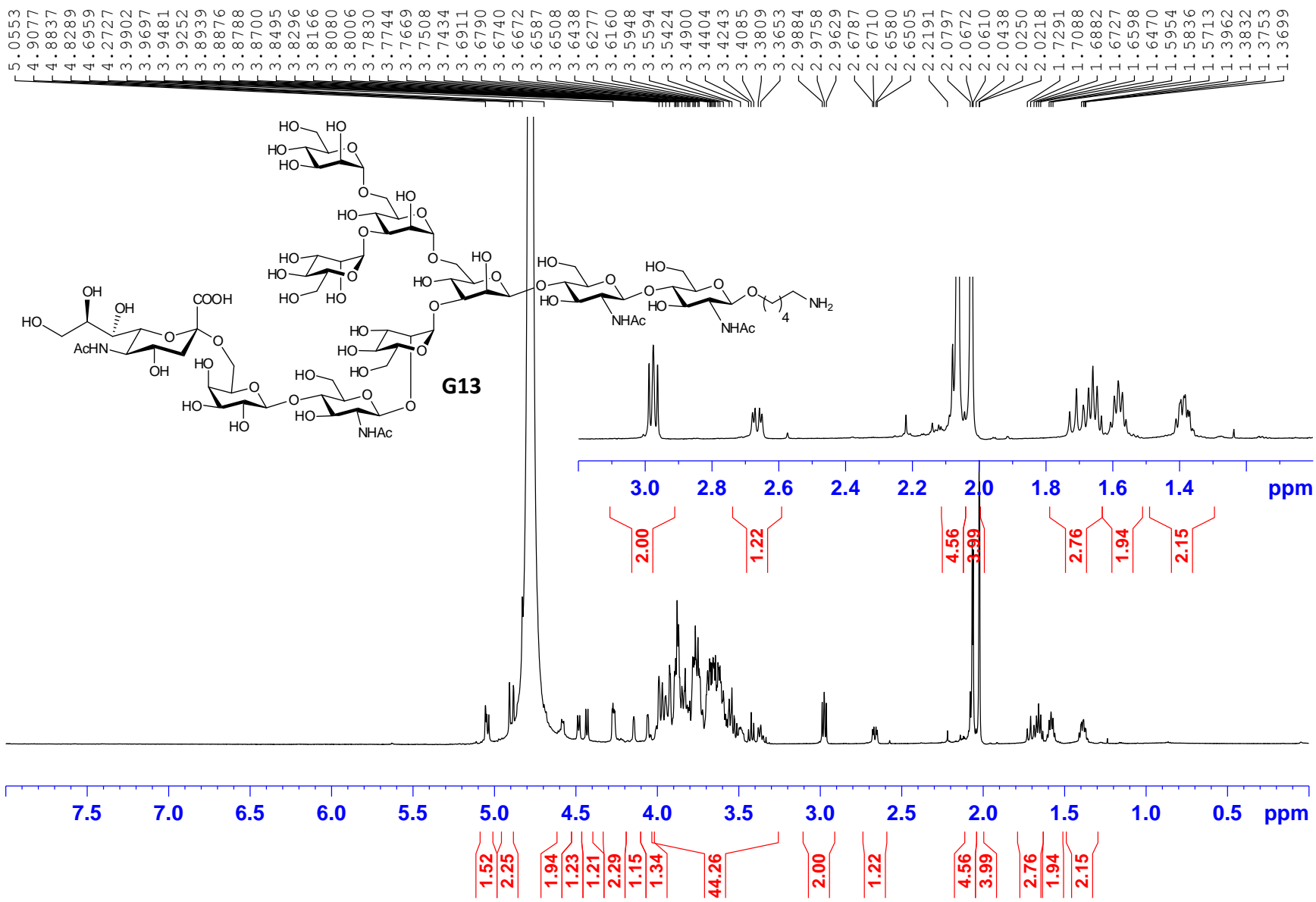


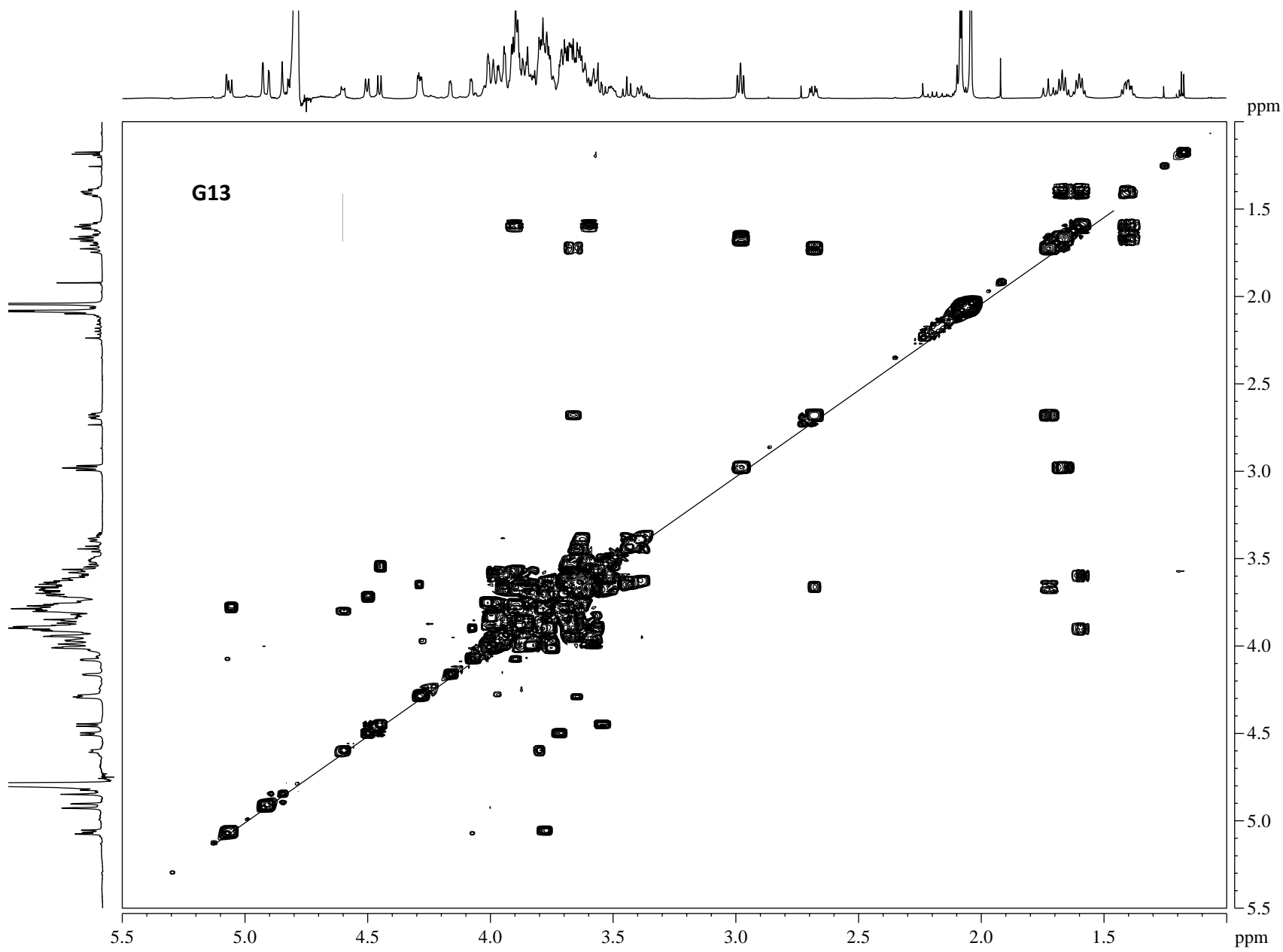


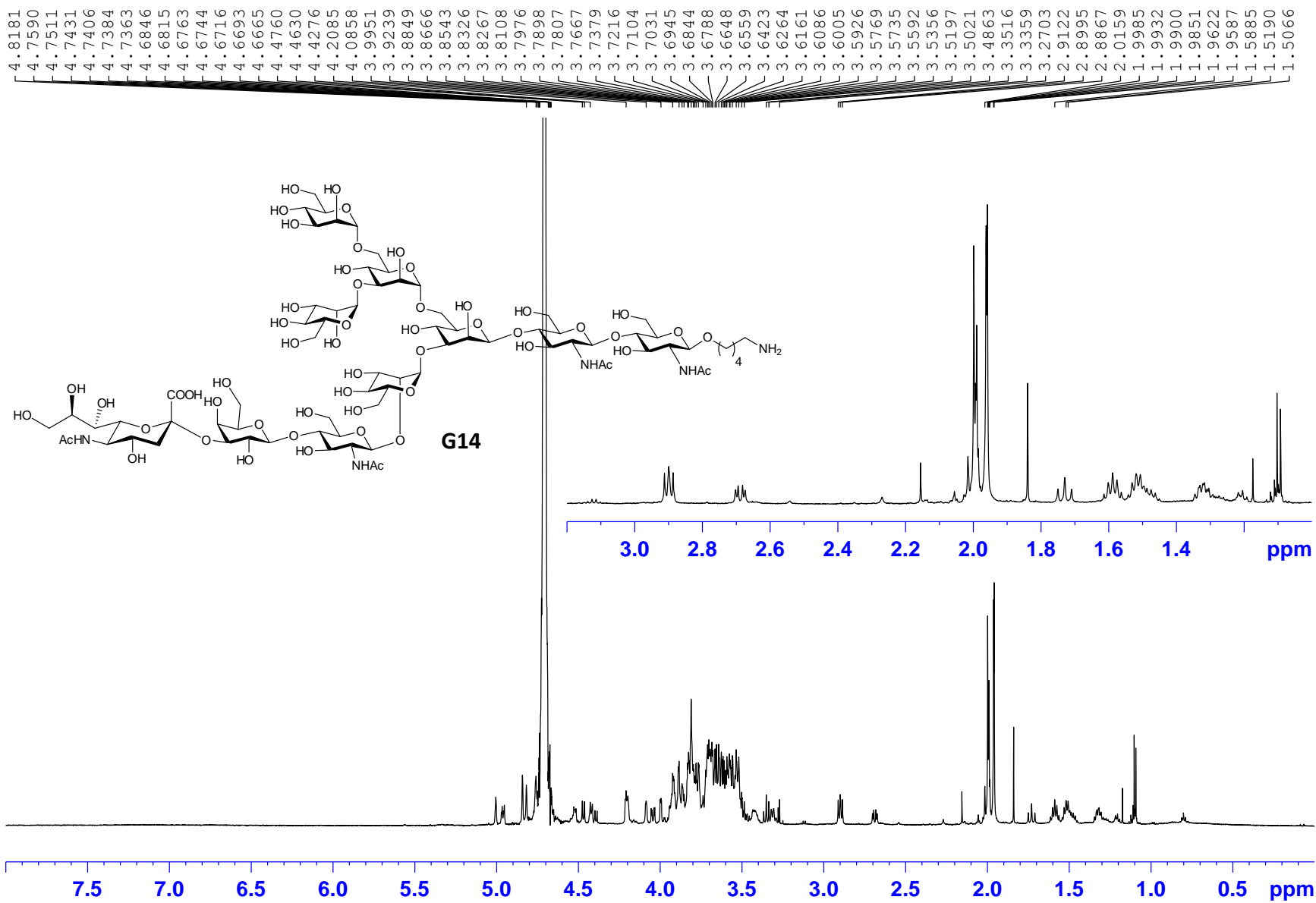


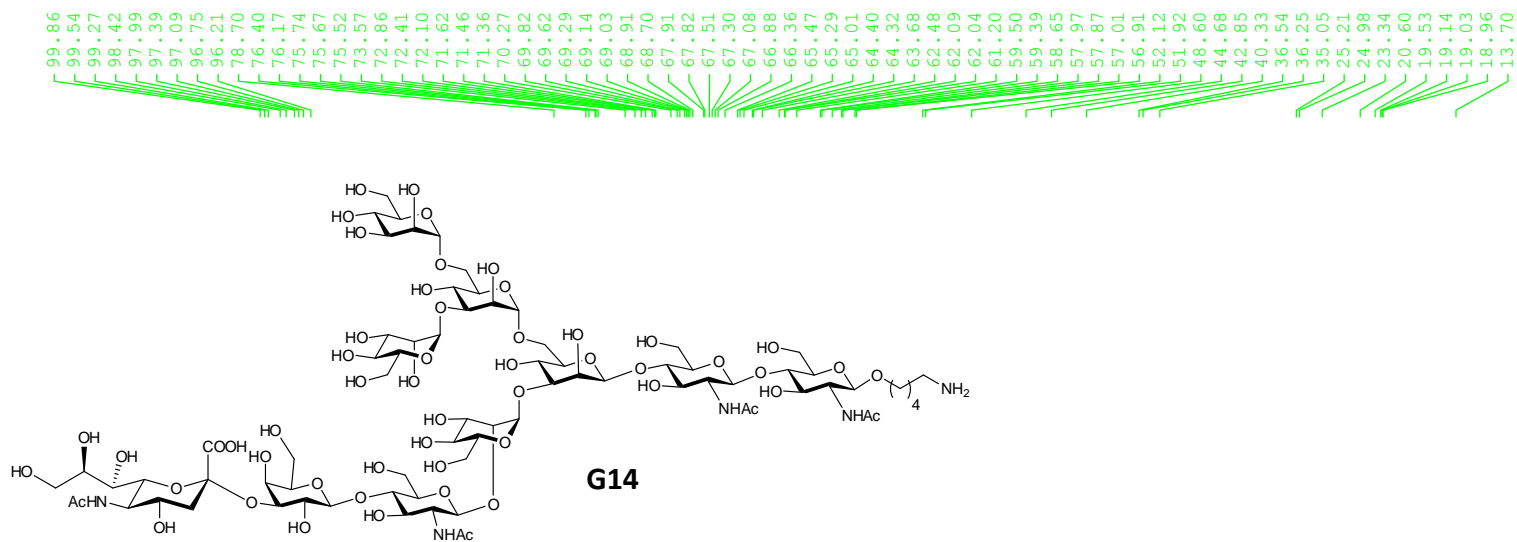




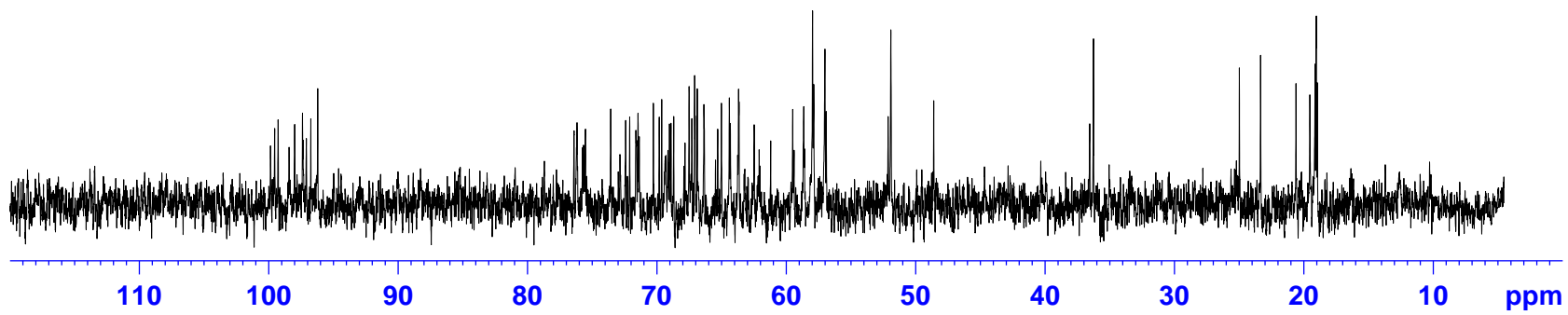


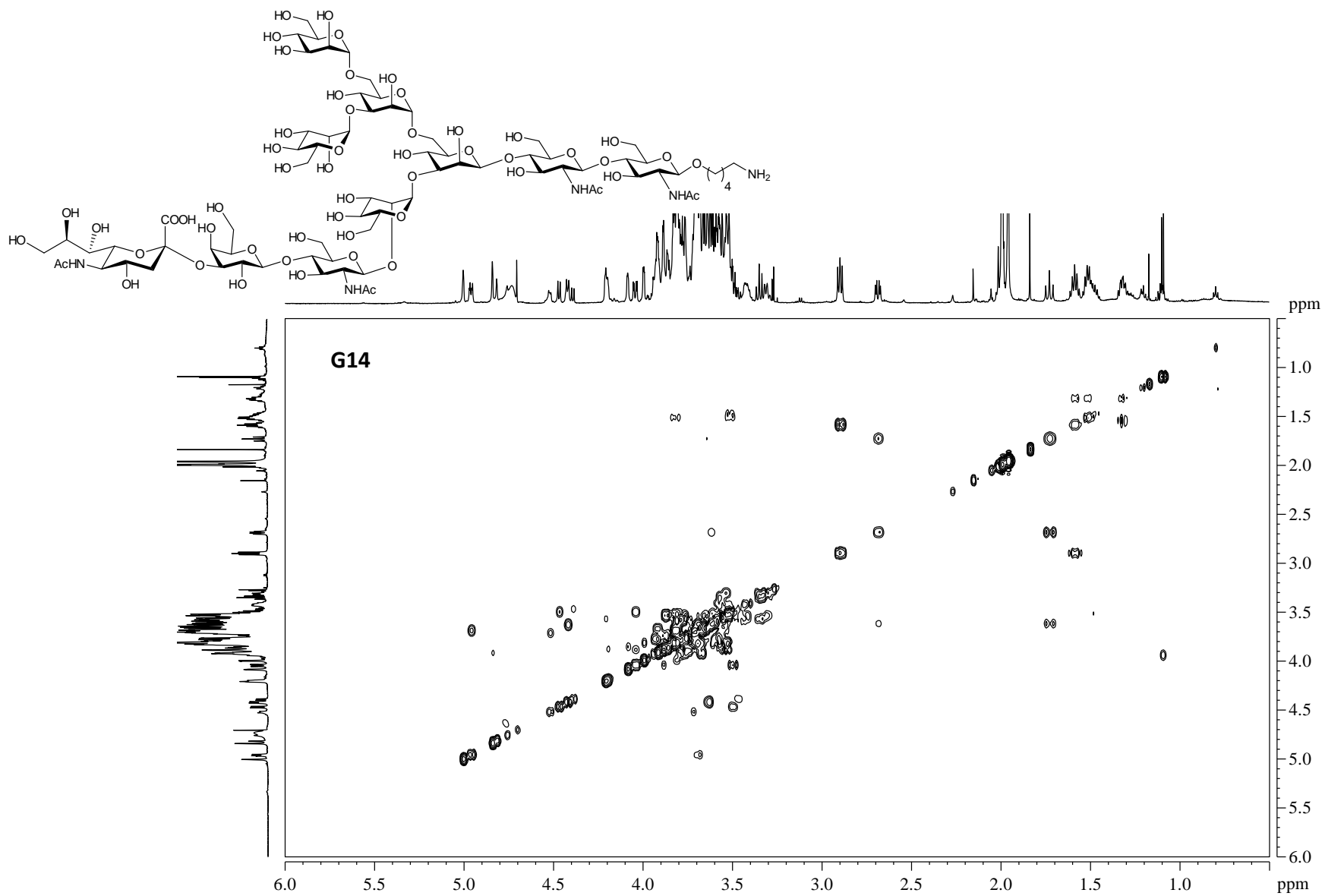


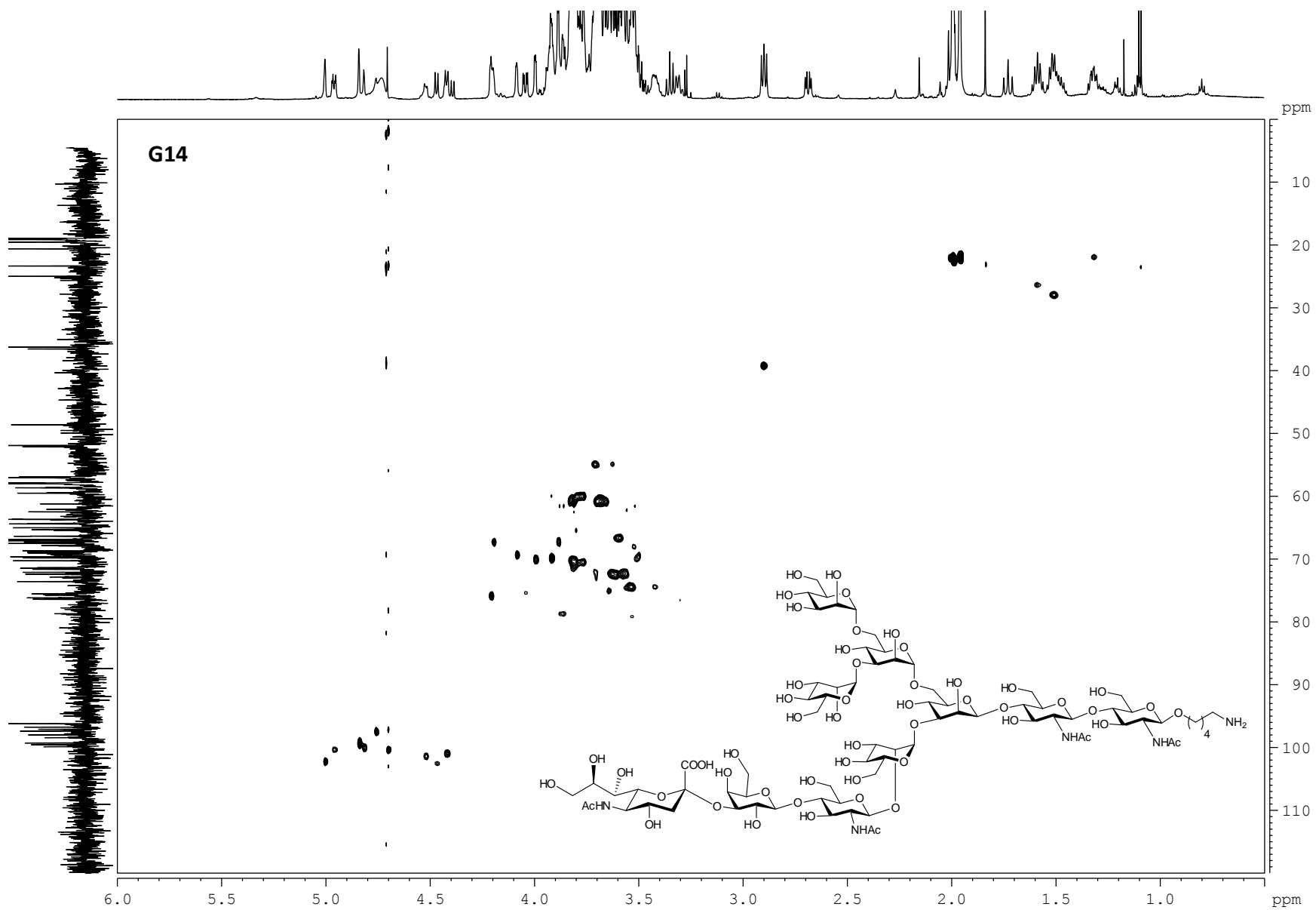


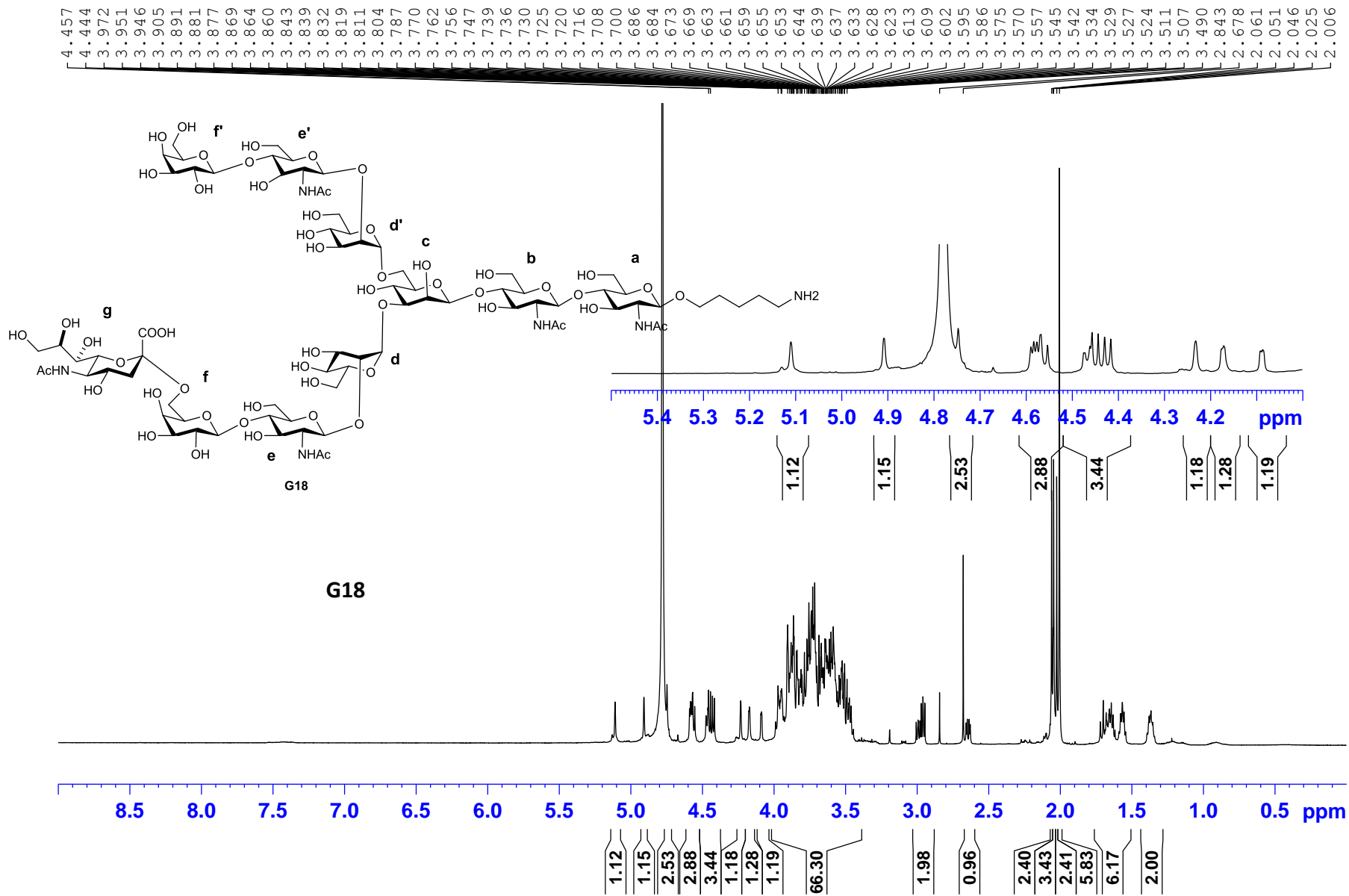


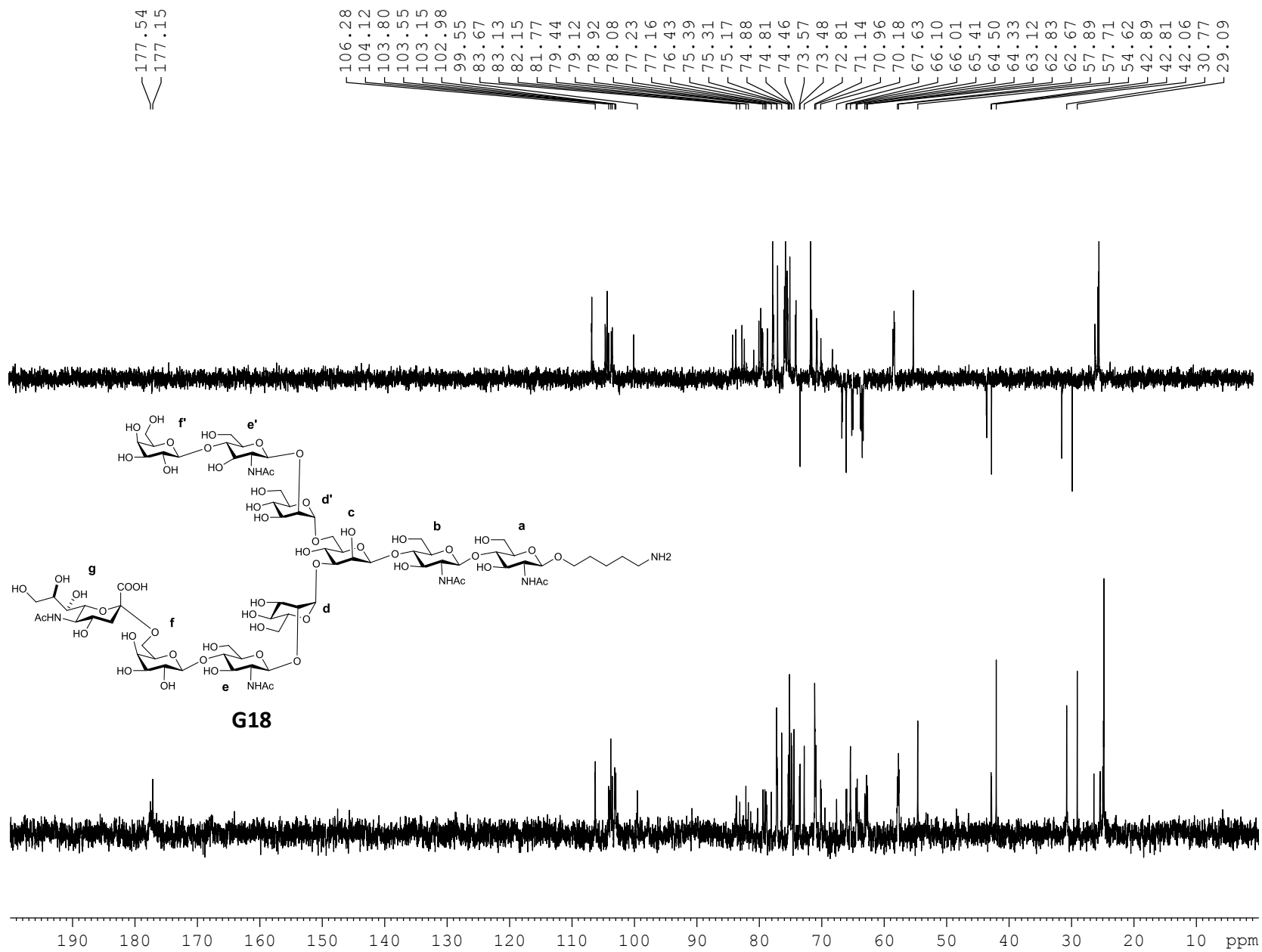
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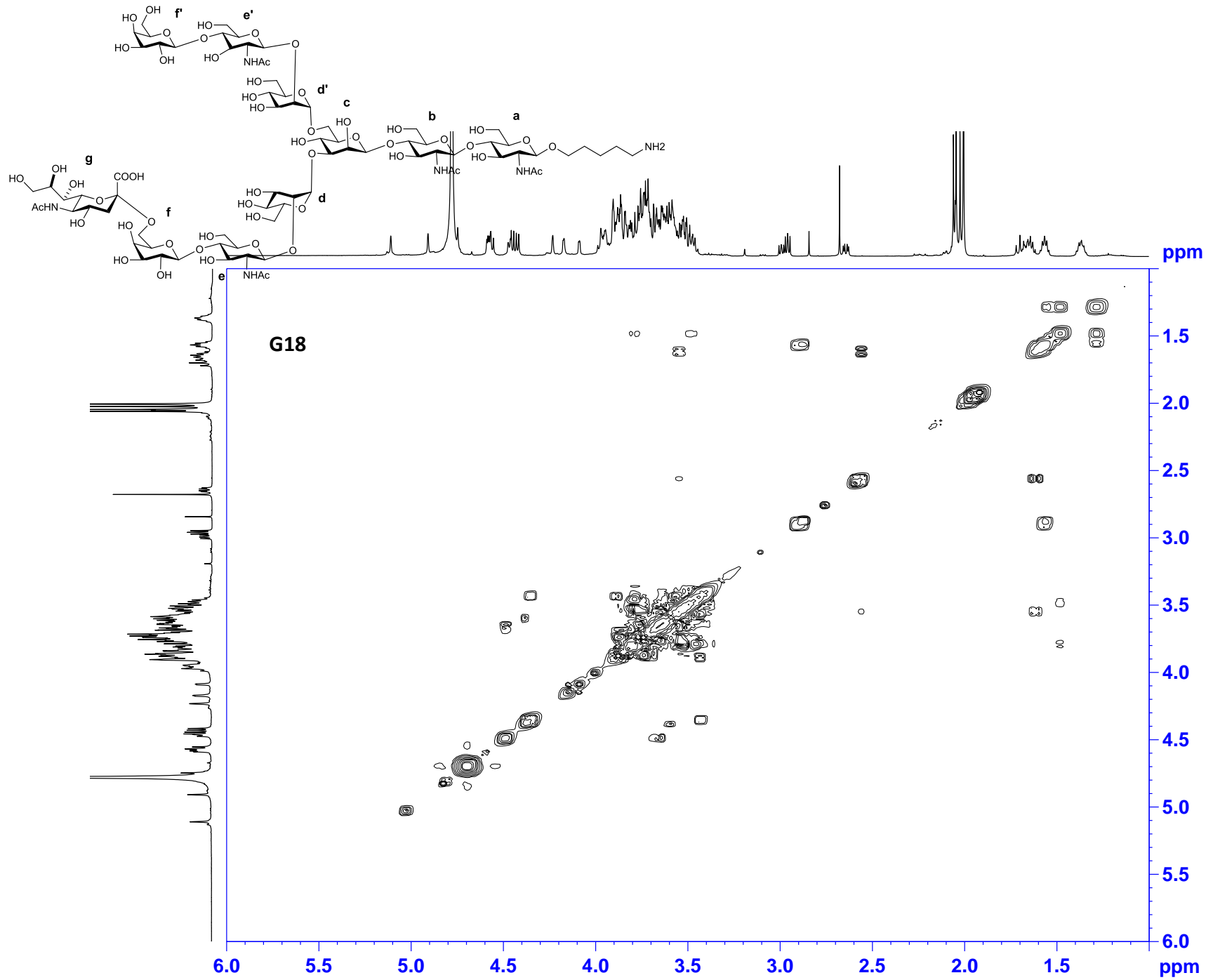


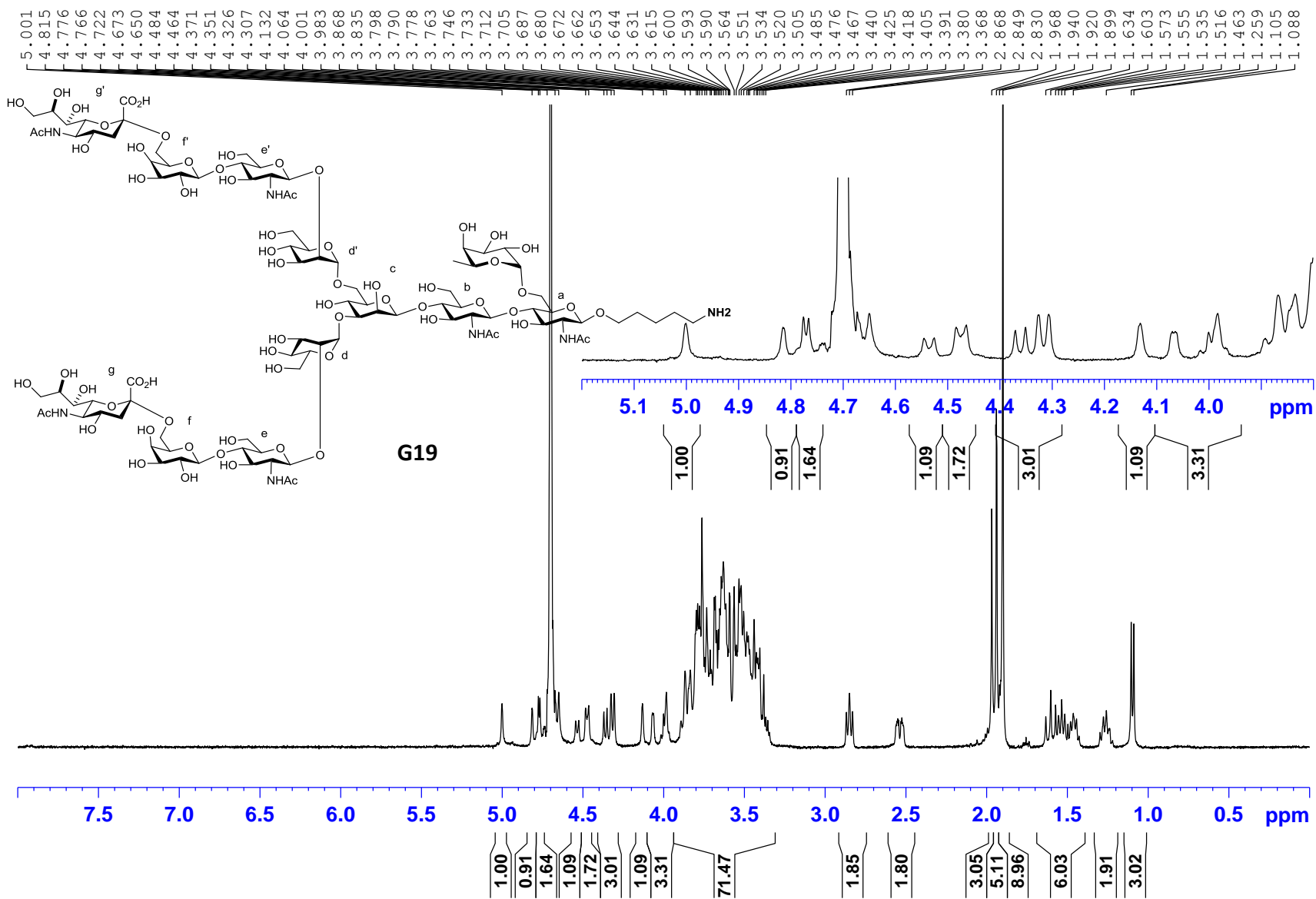


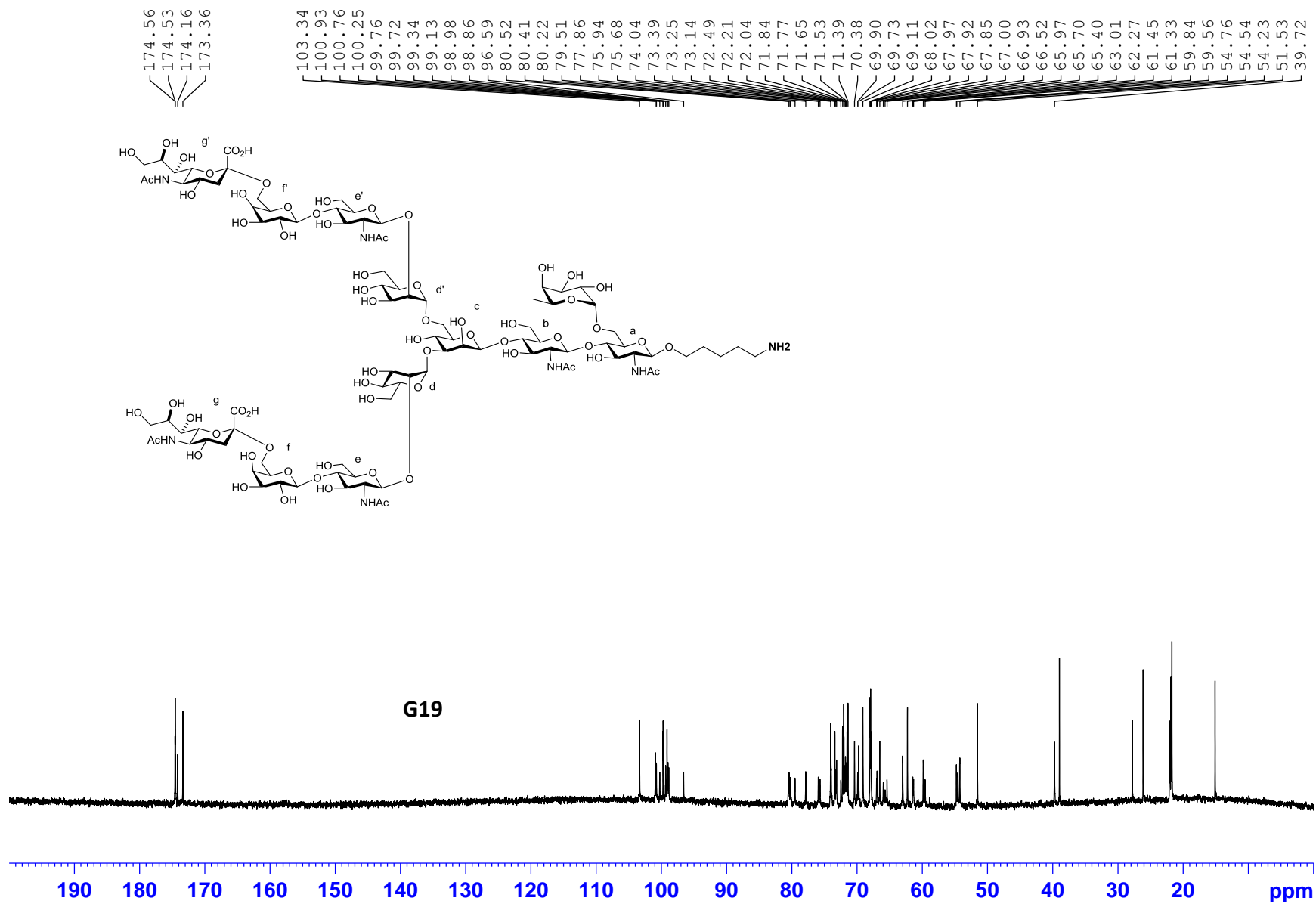


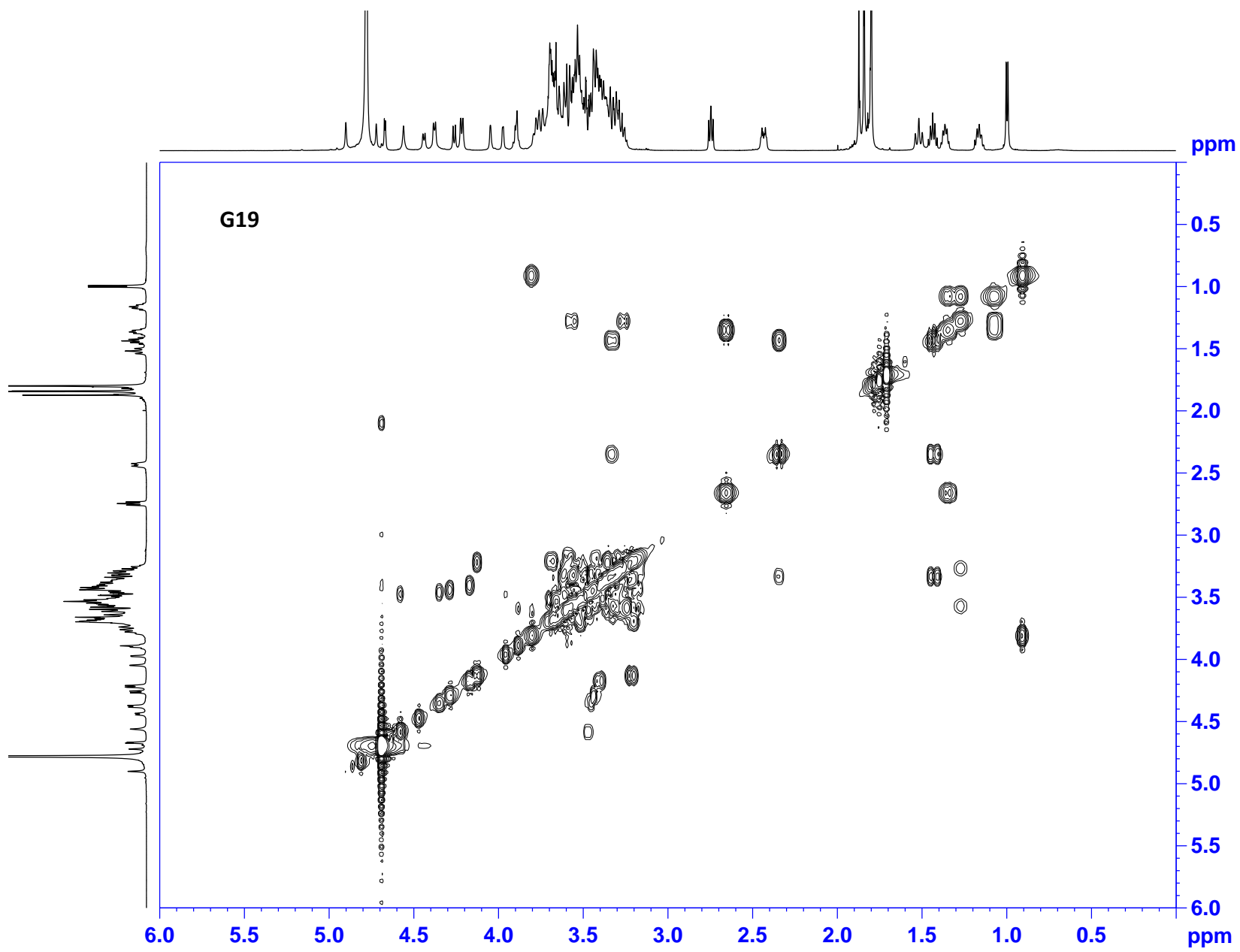


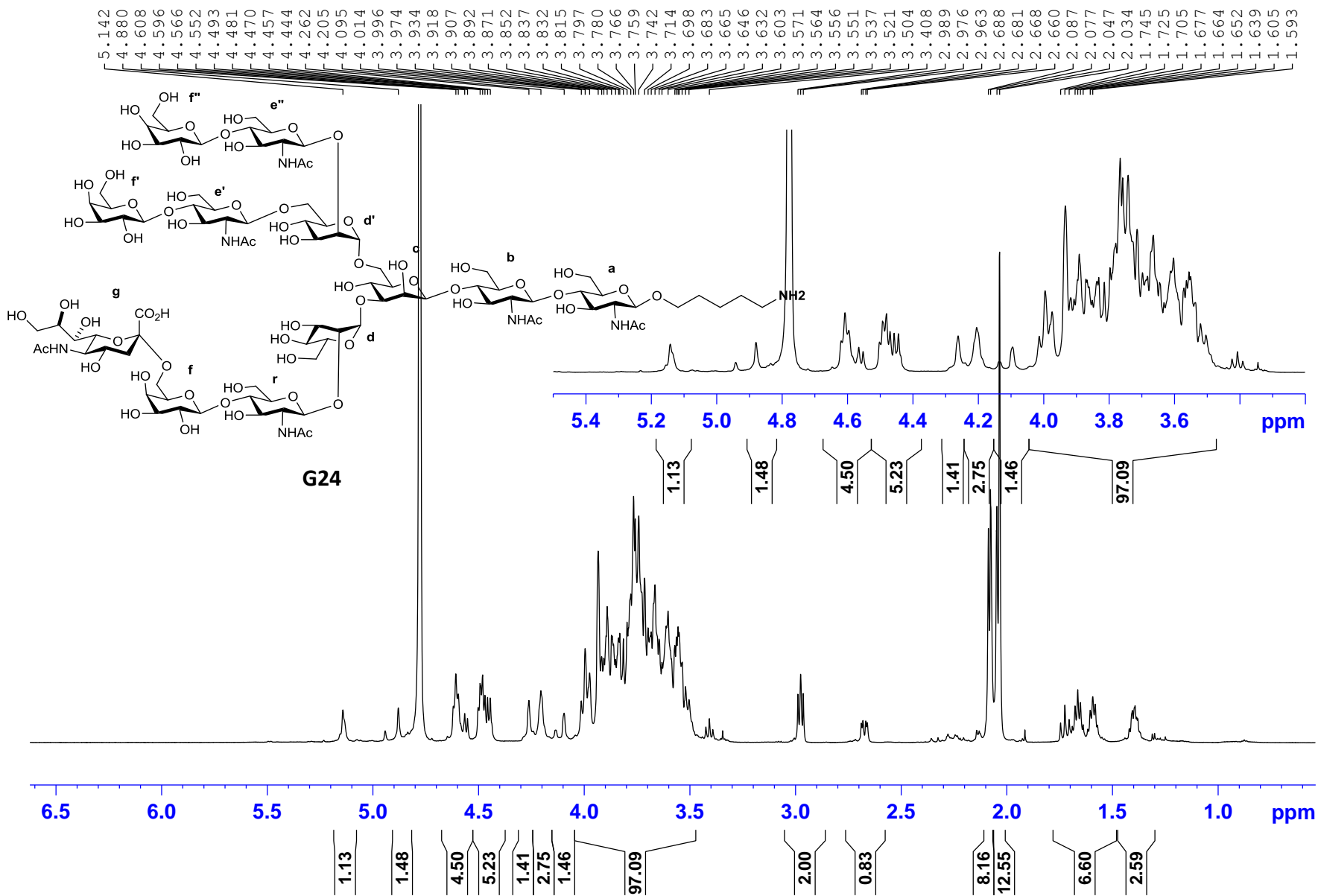




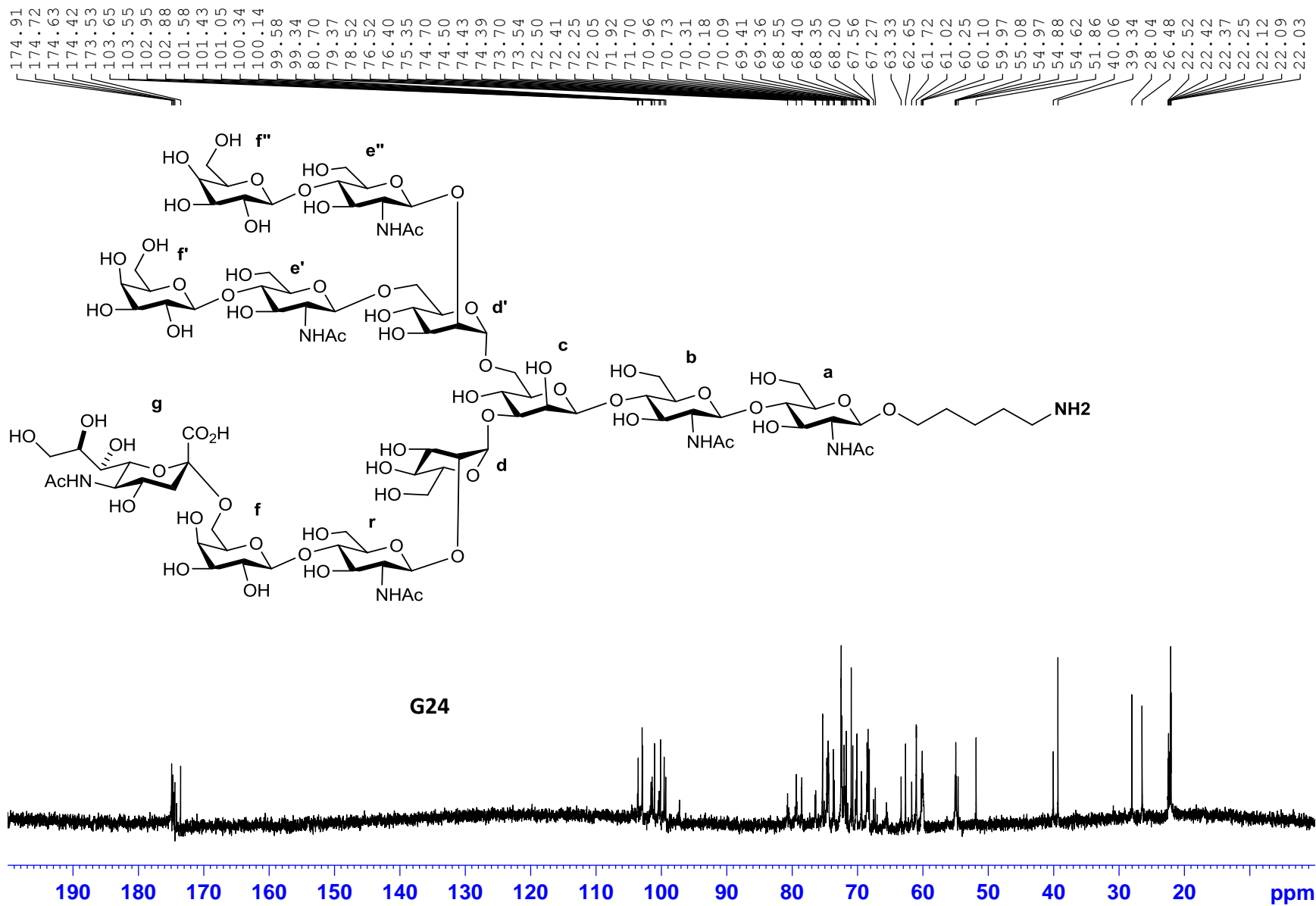


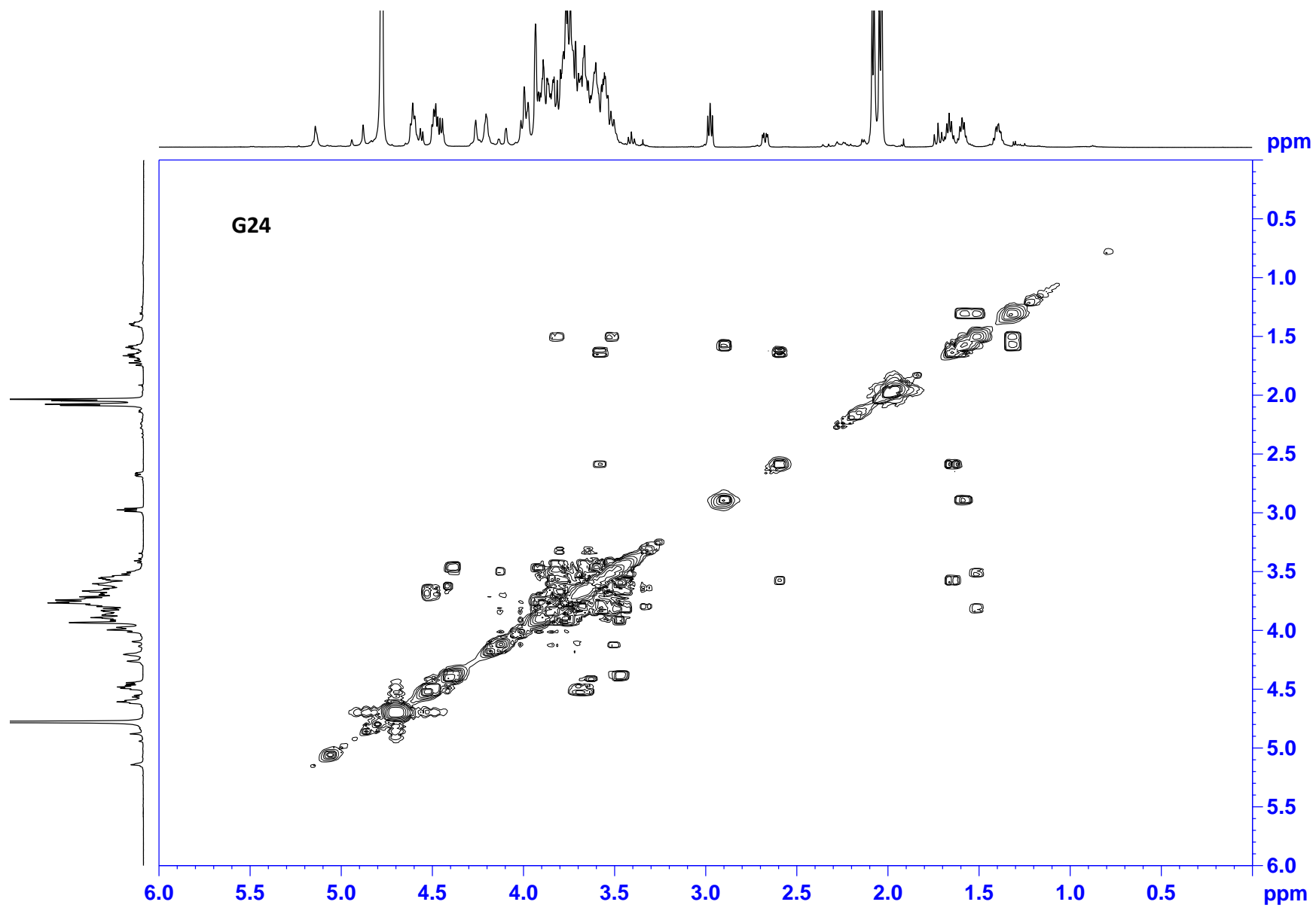


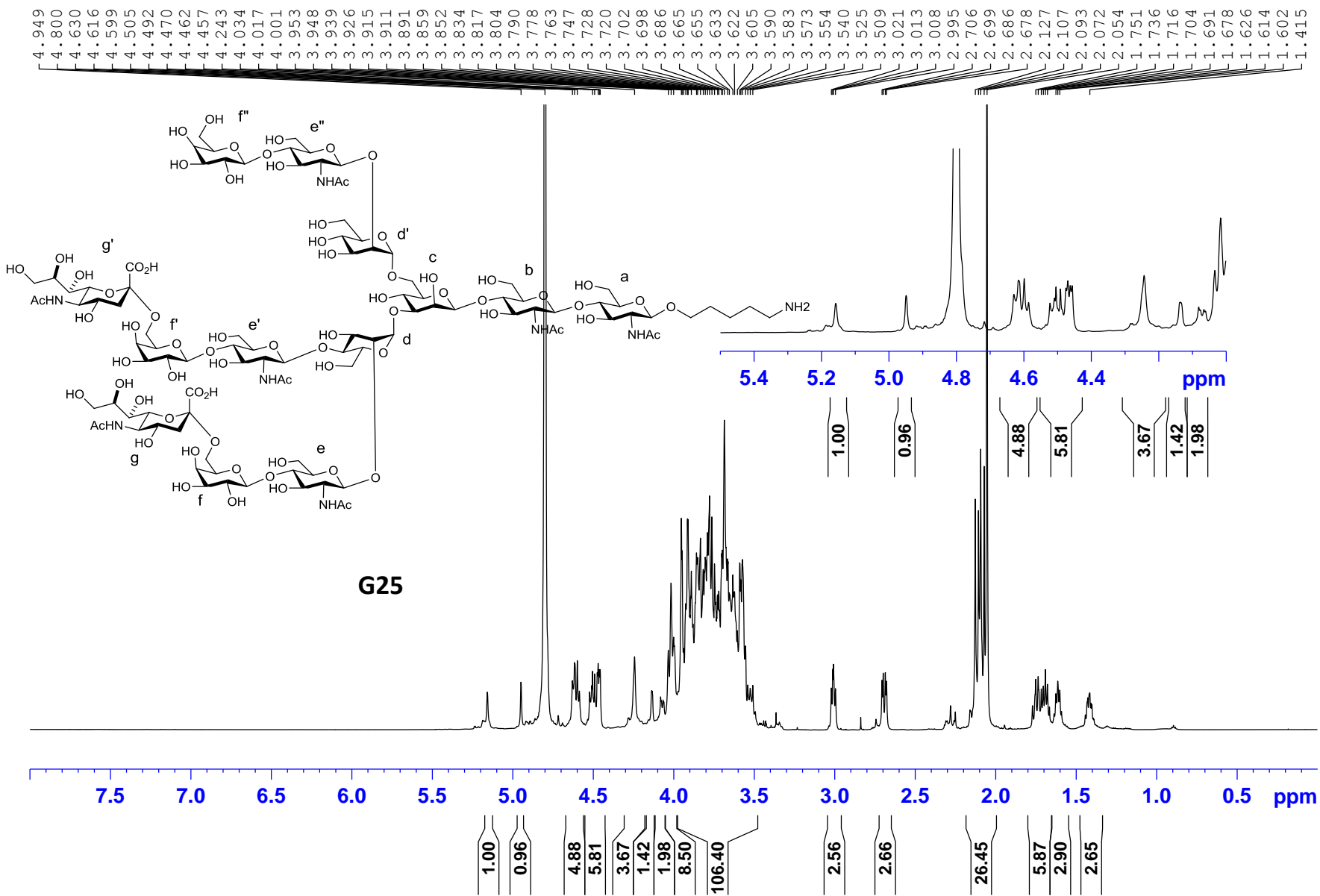


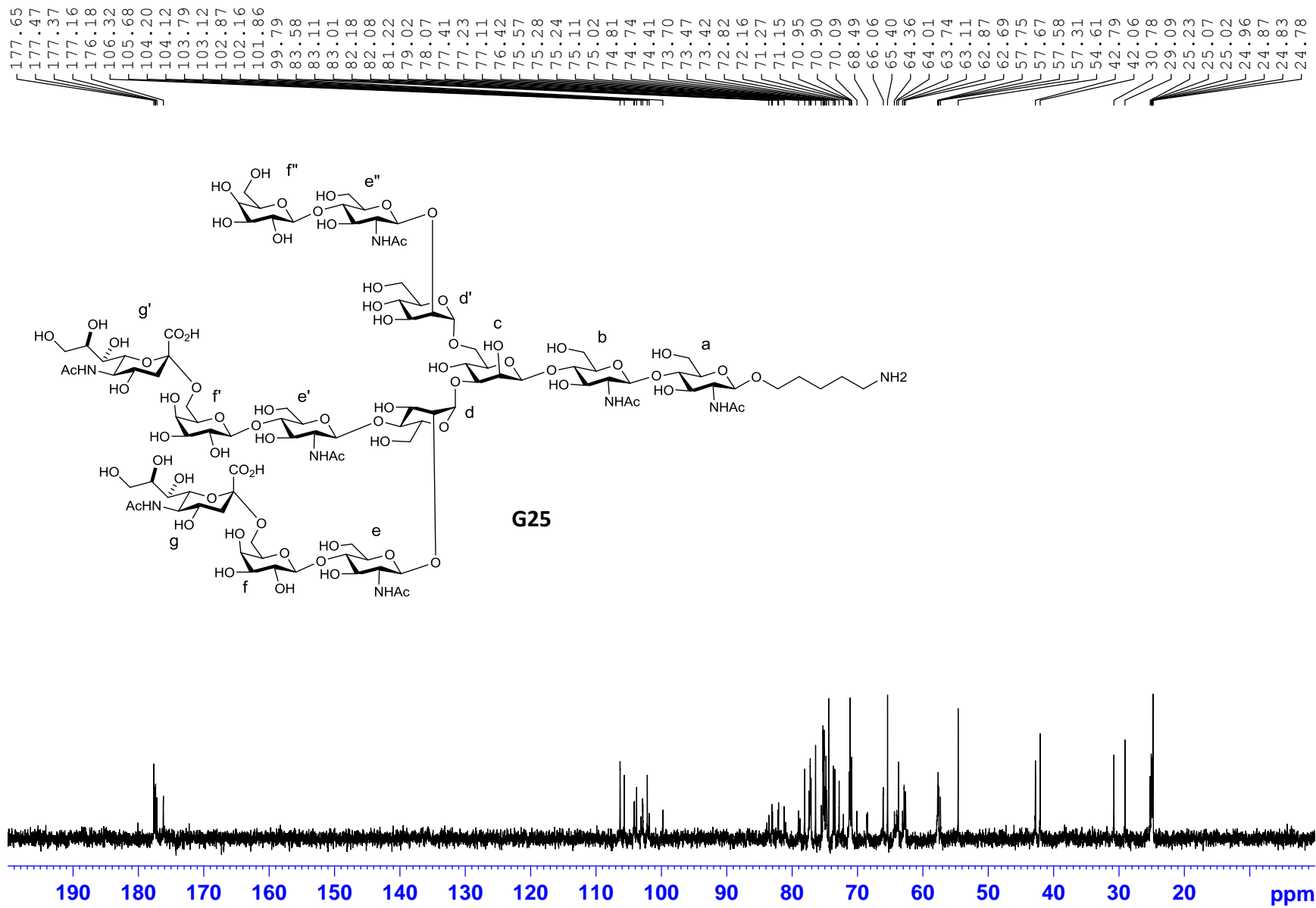


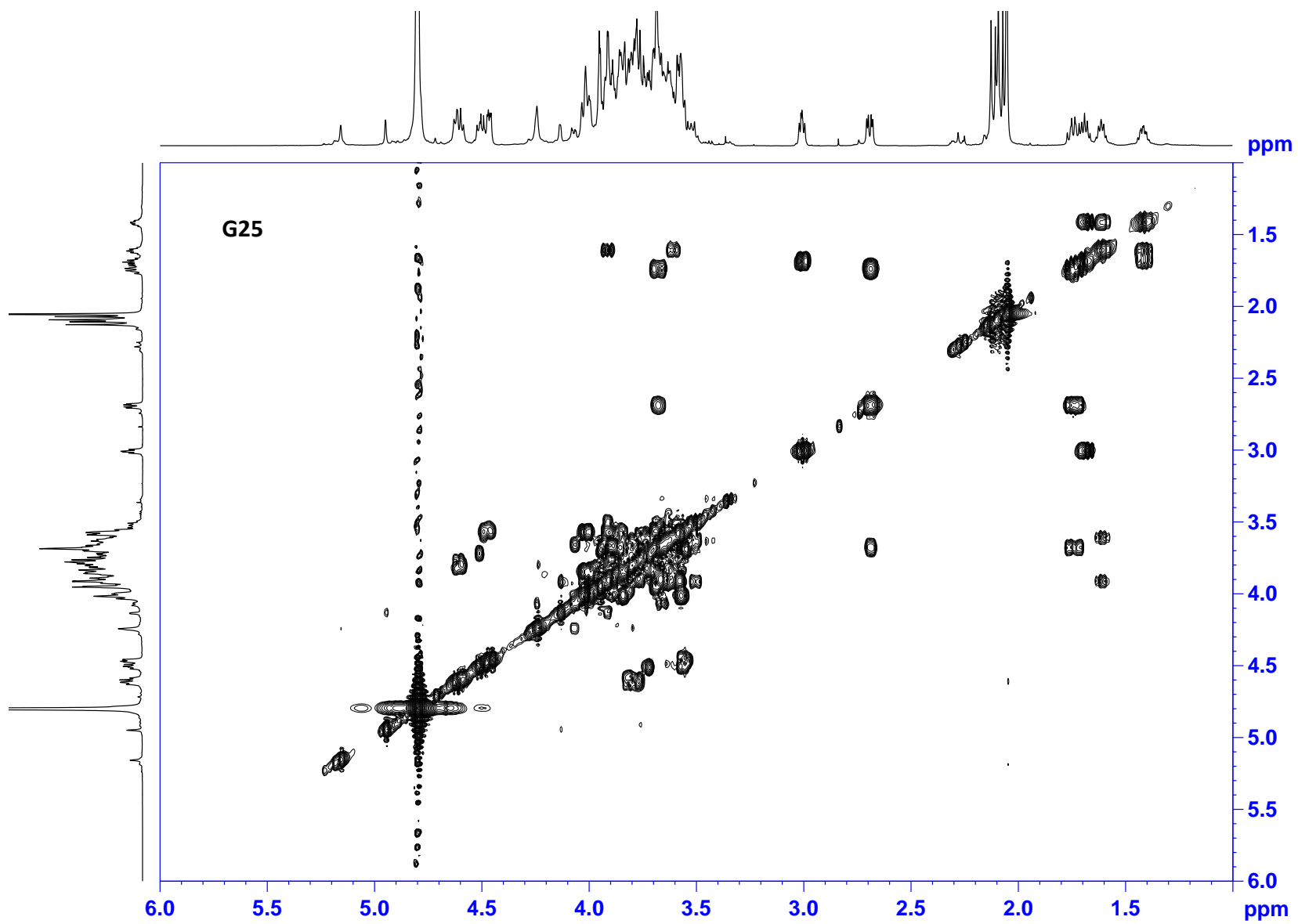
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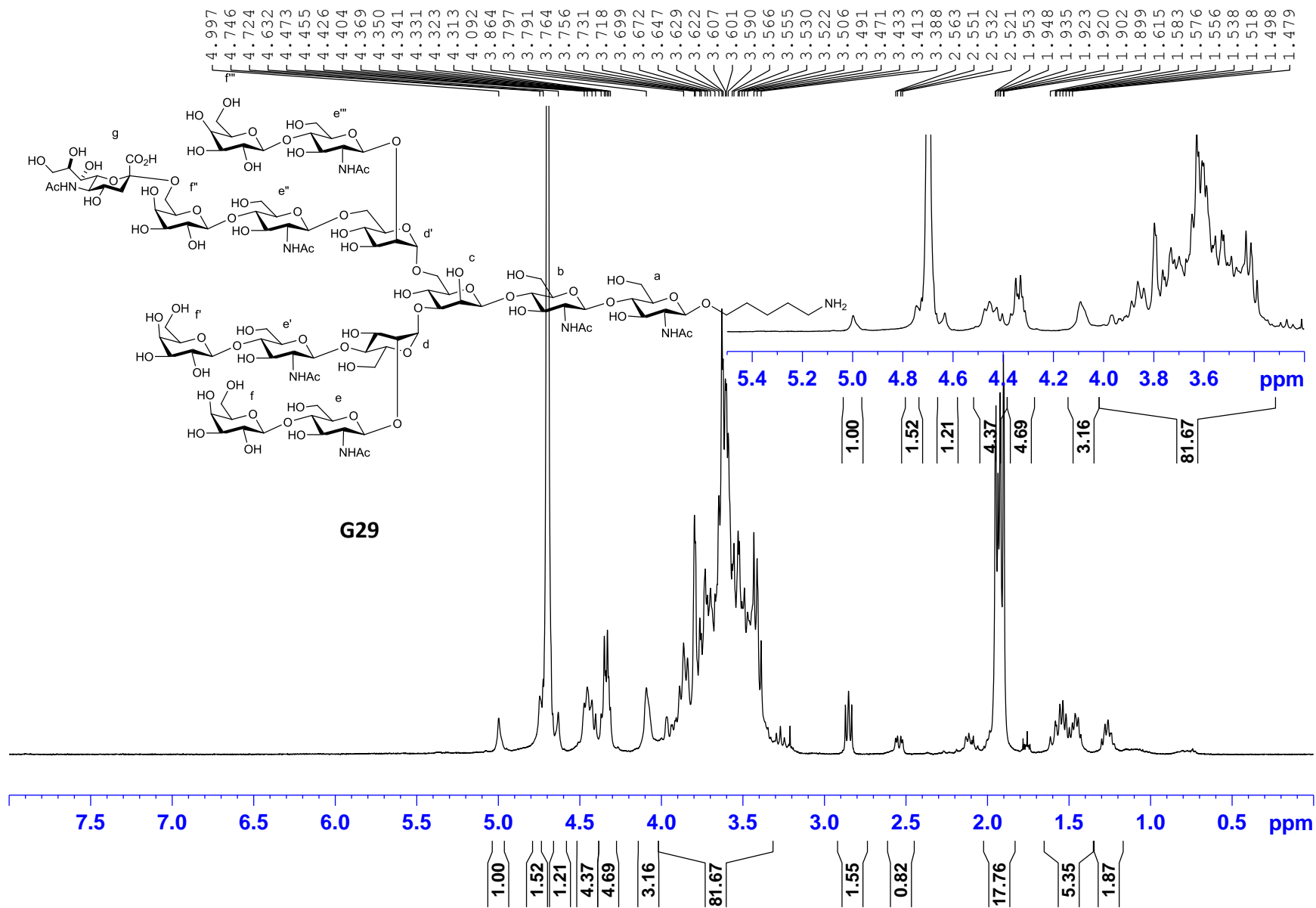


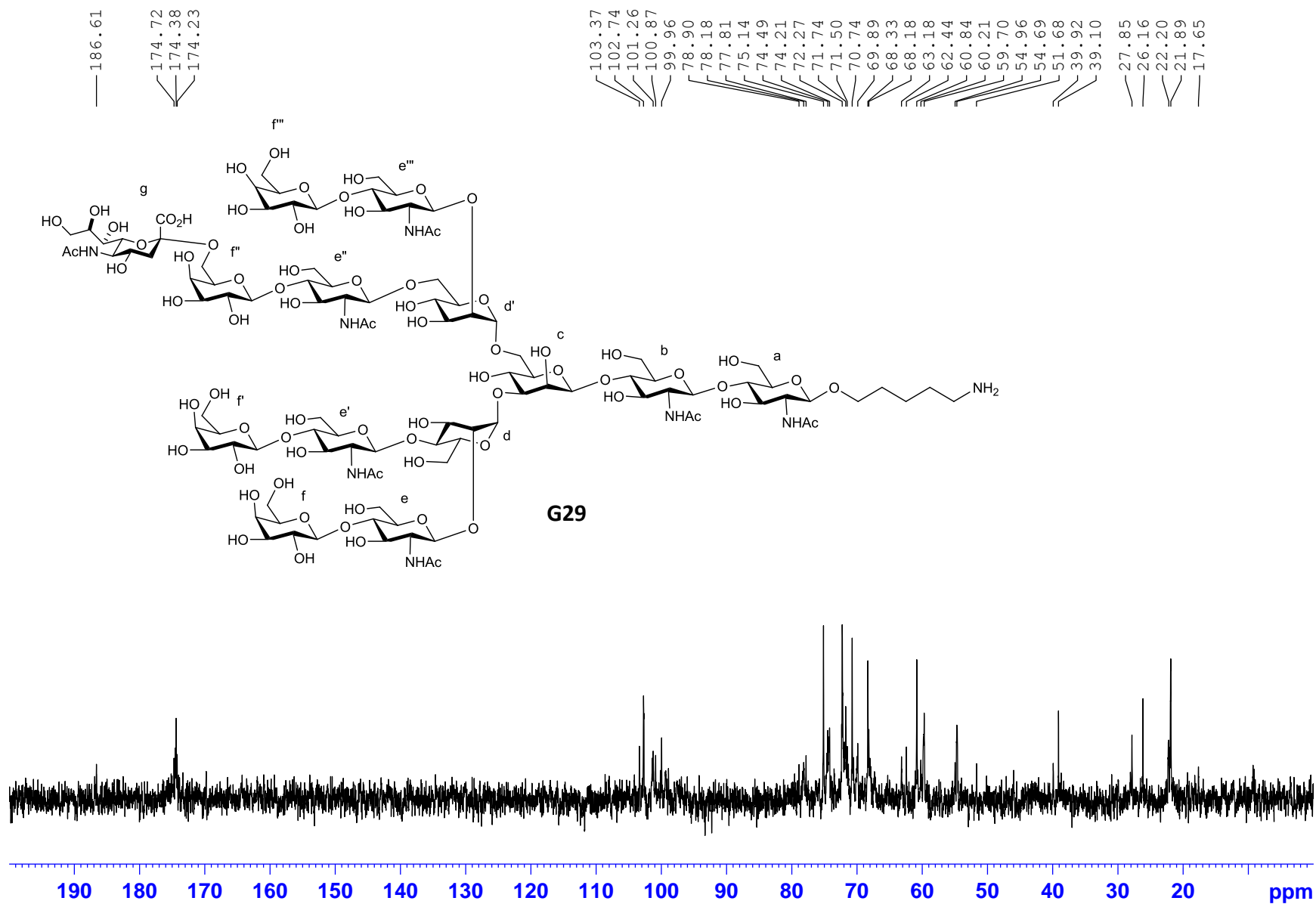


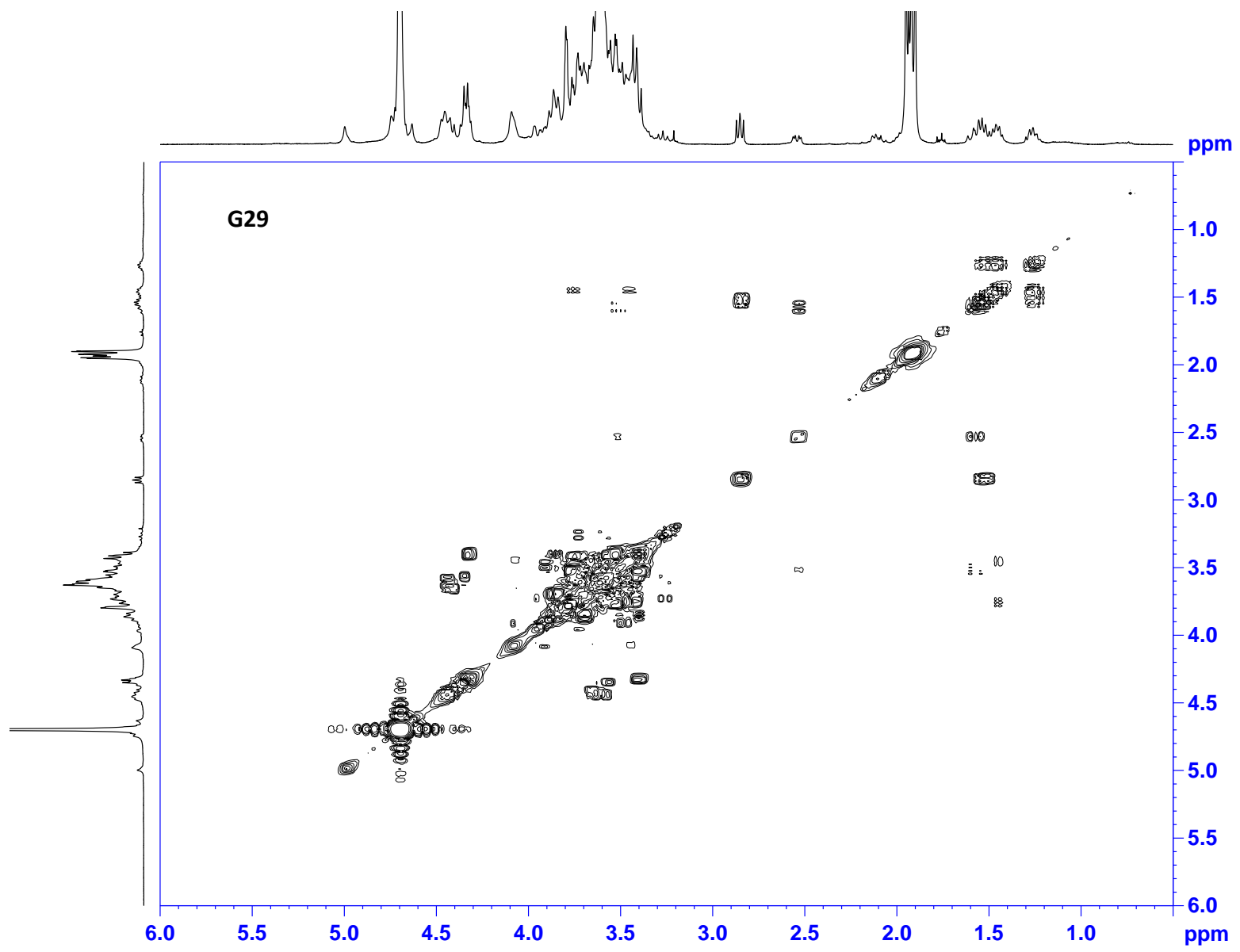


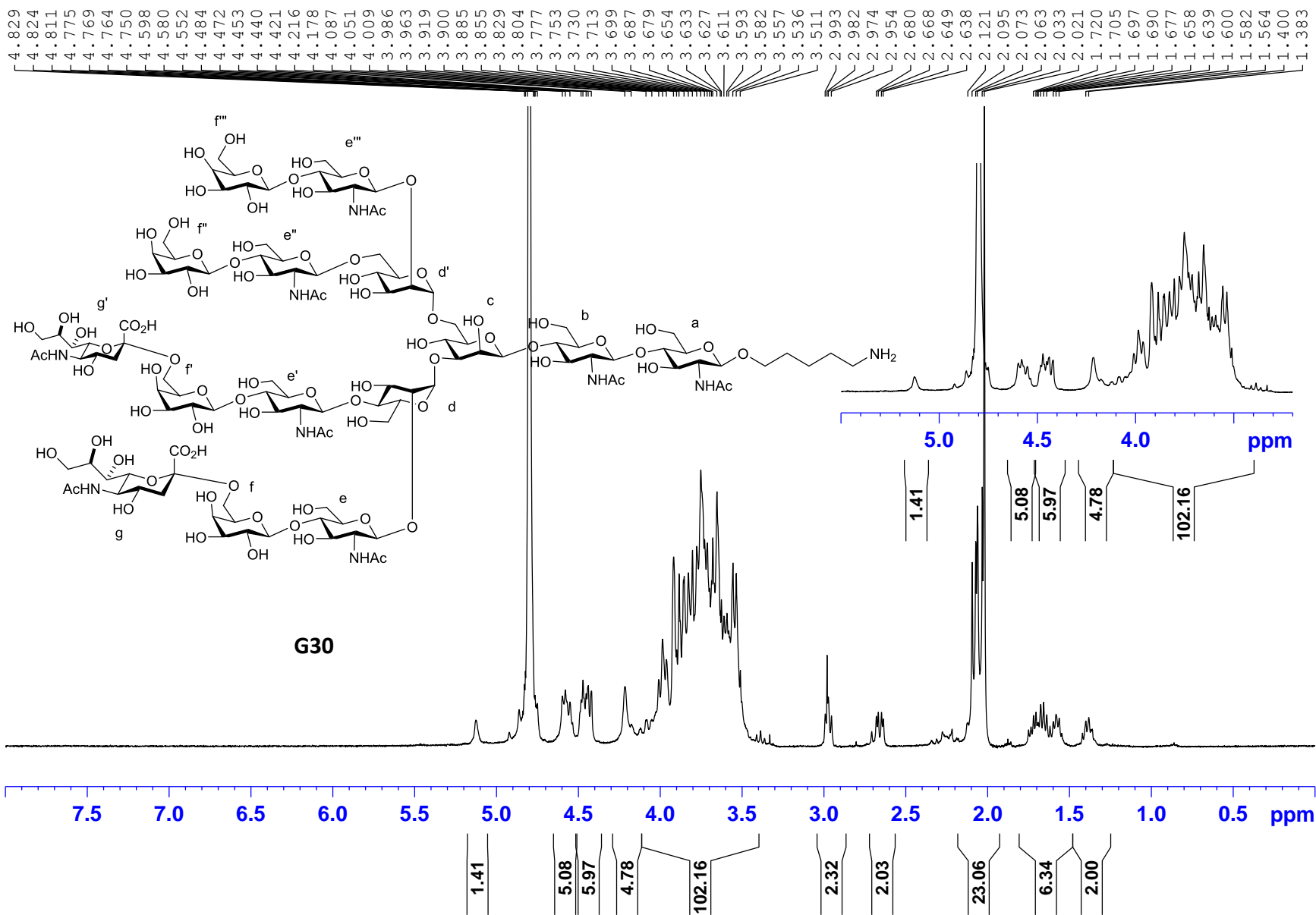


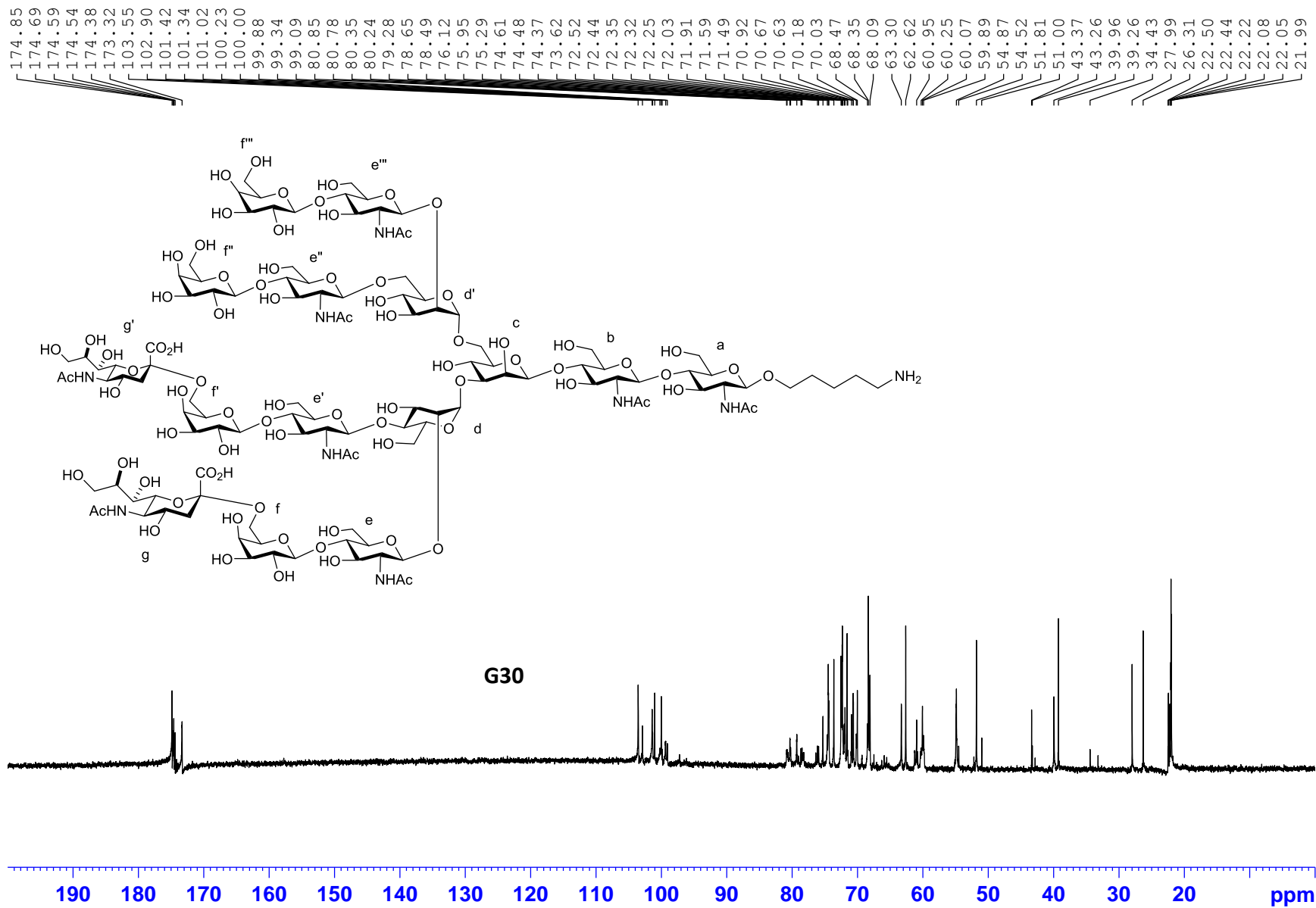


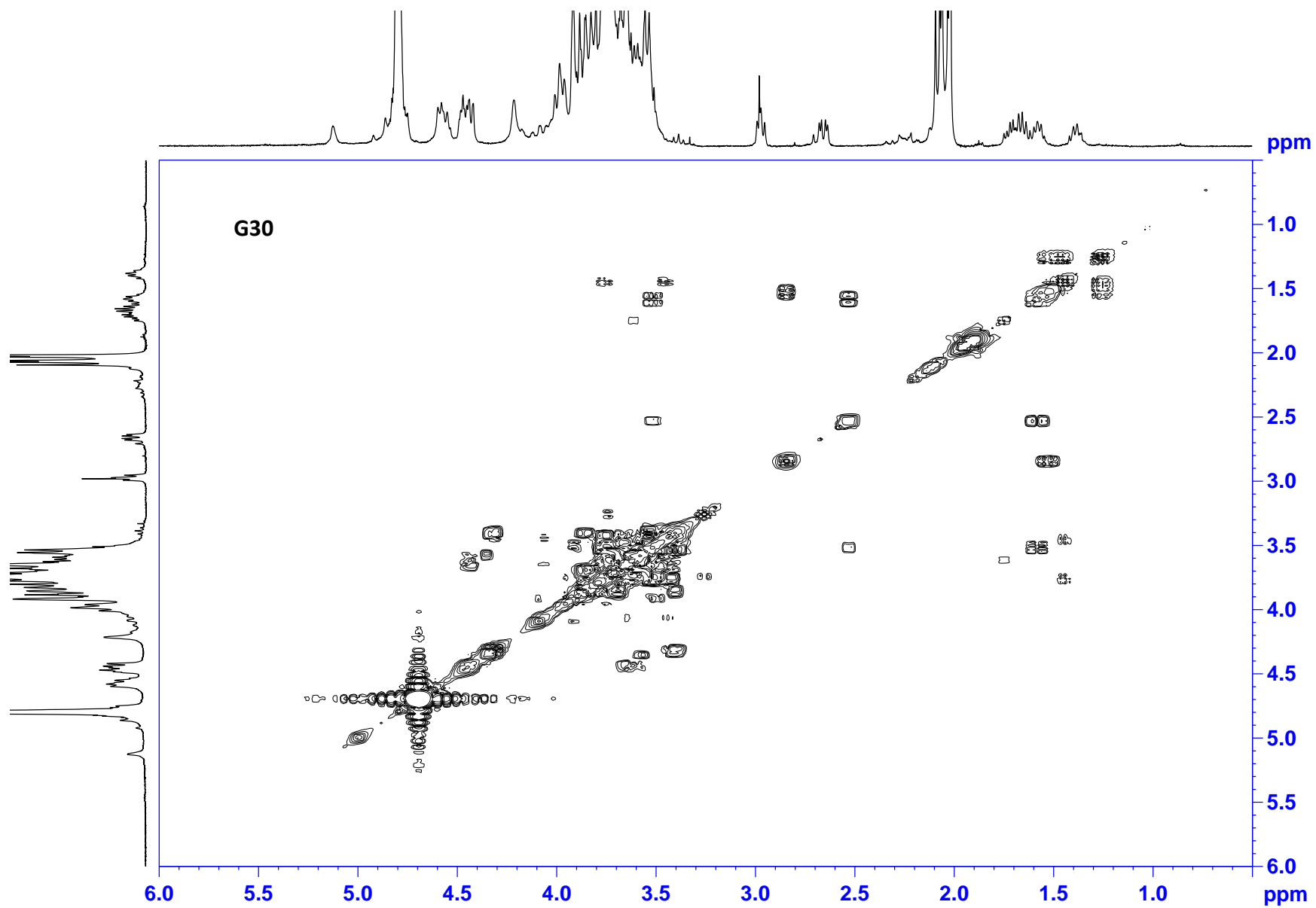


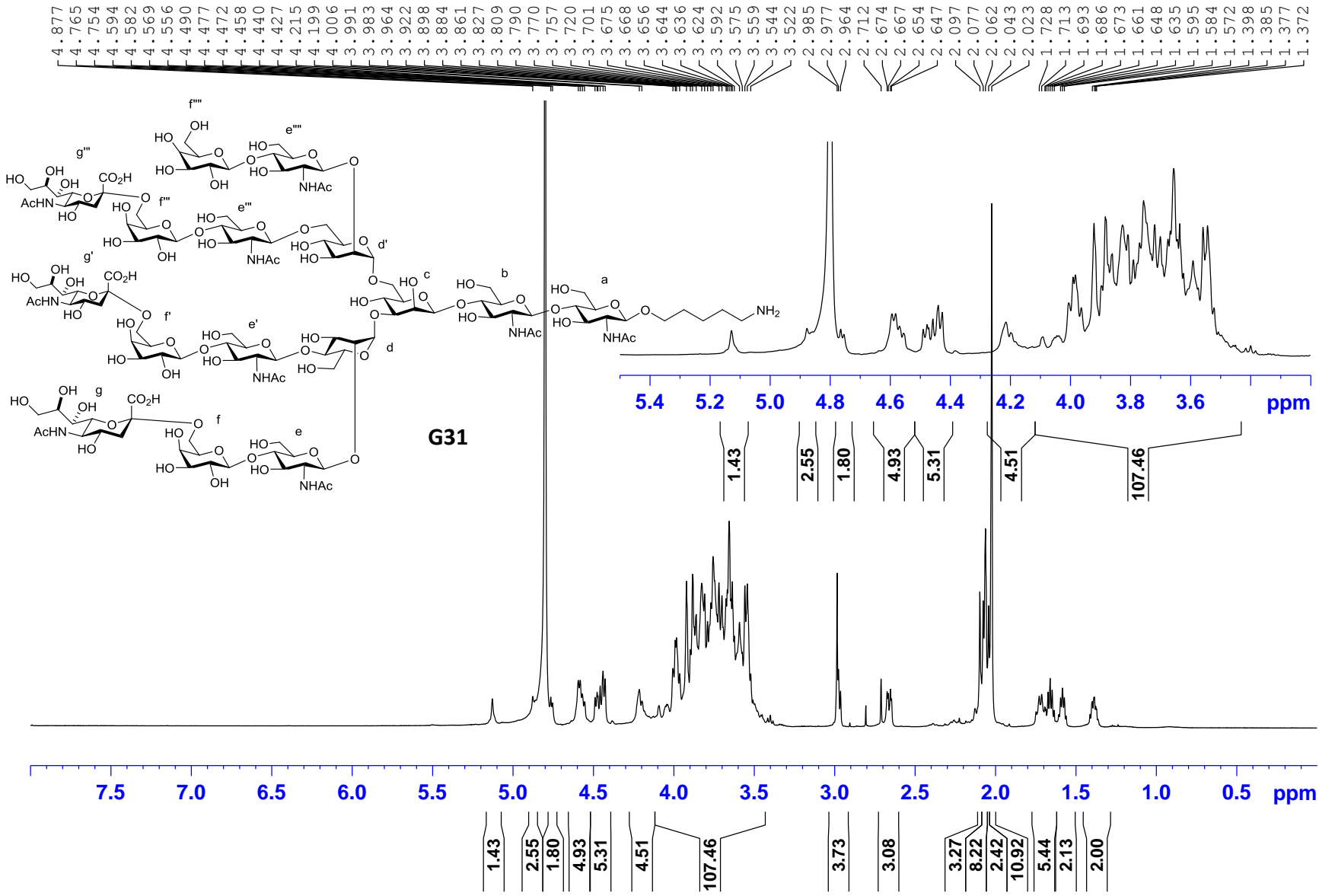






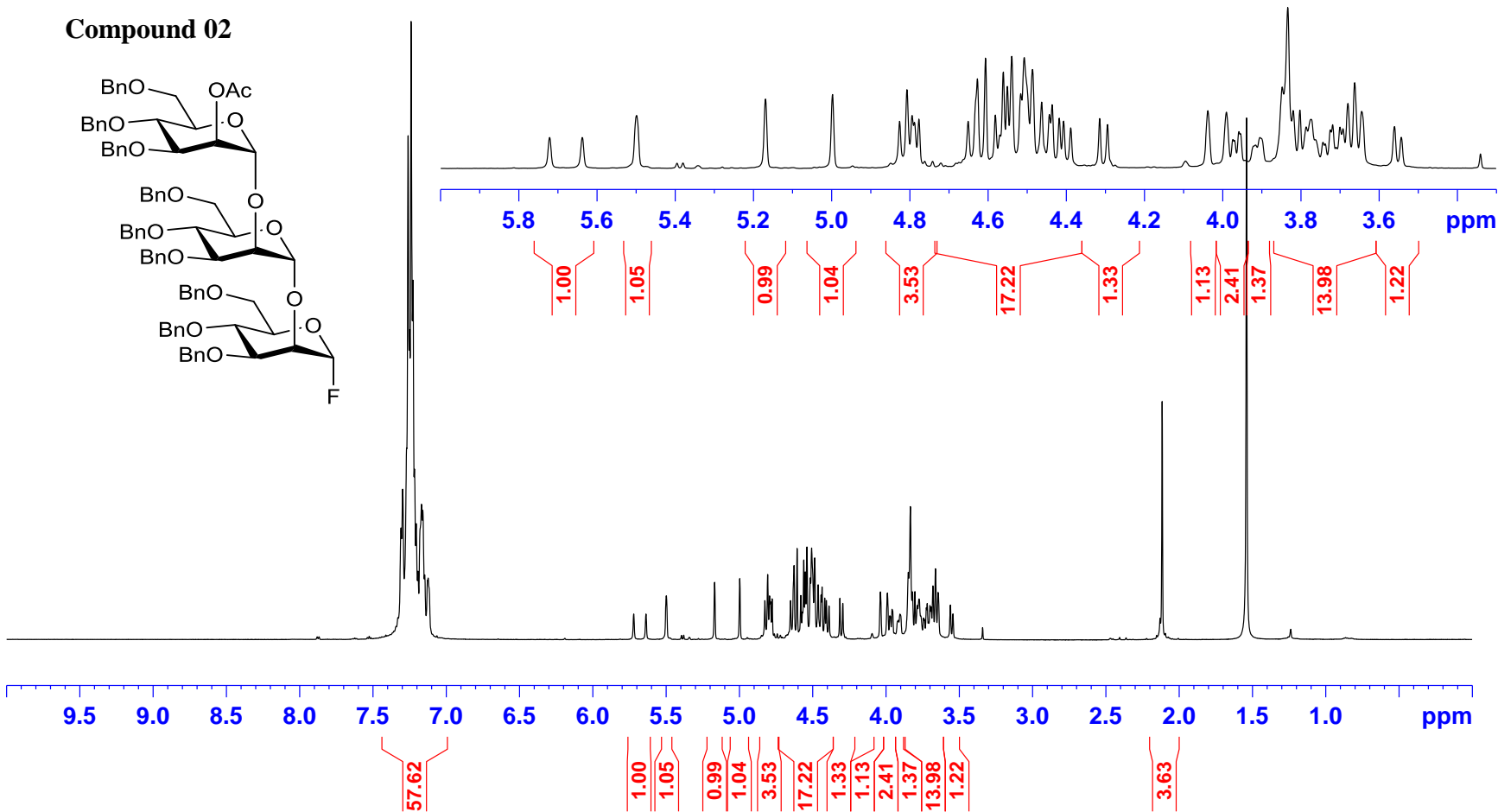
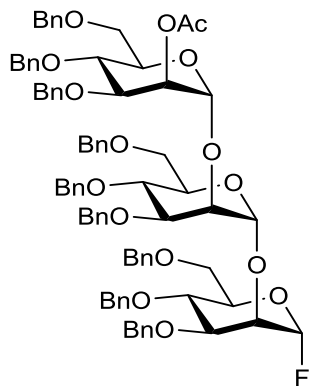




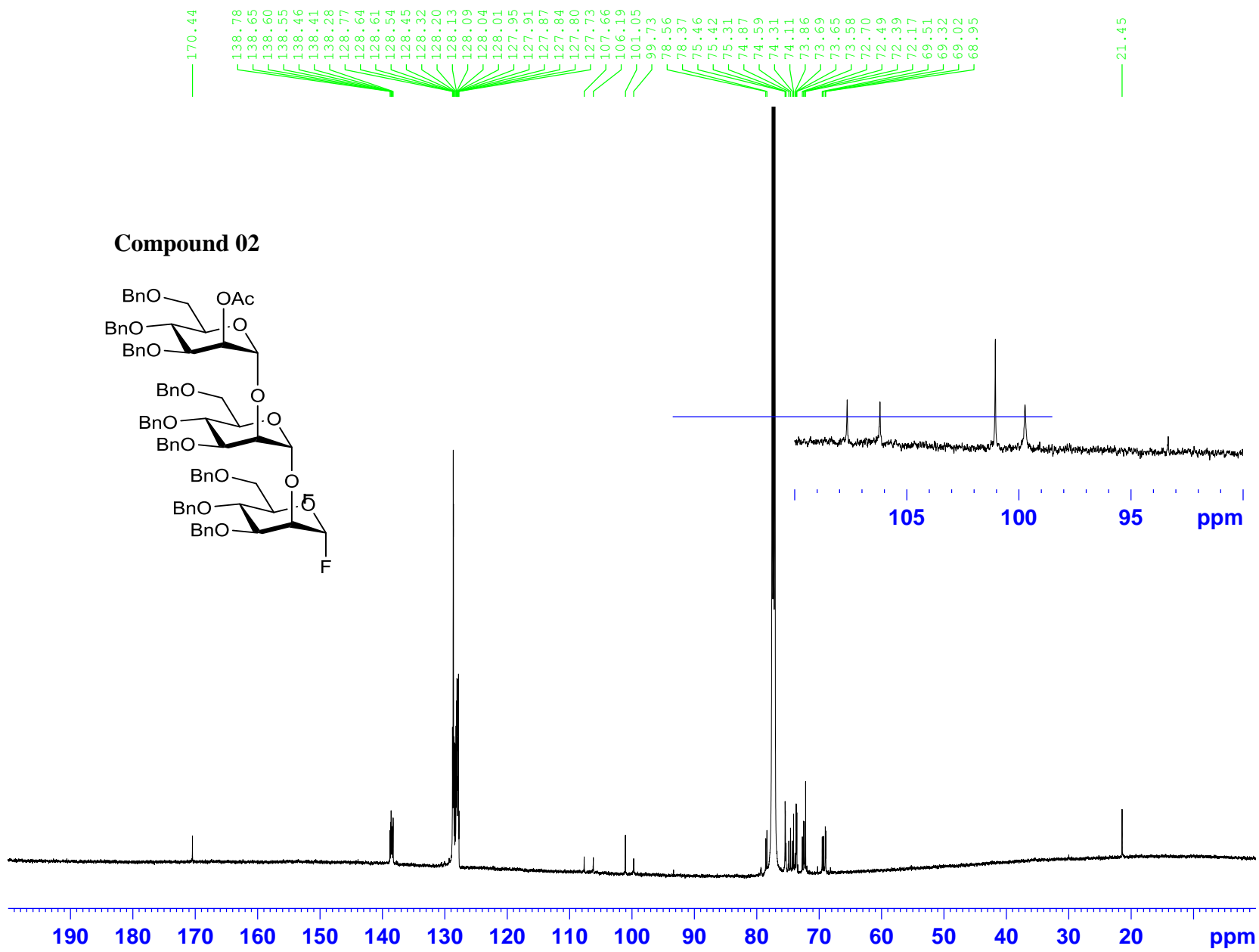
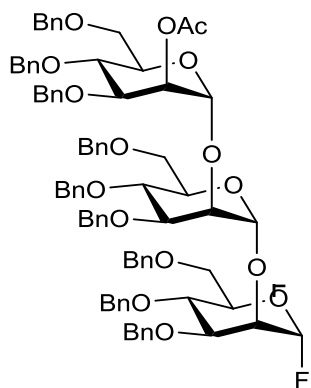


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Compound 02

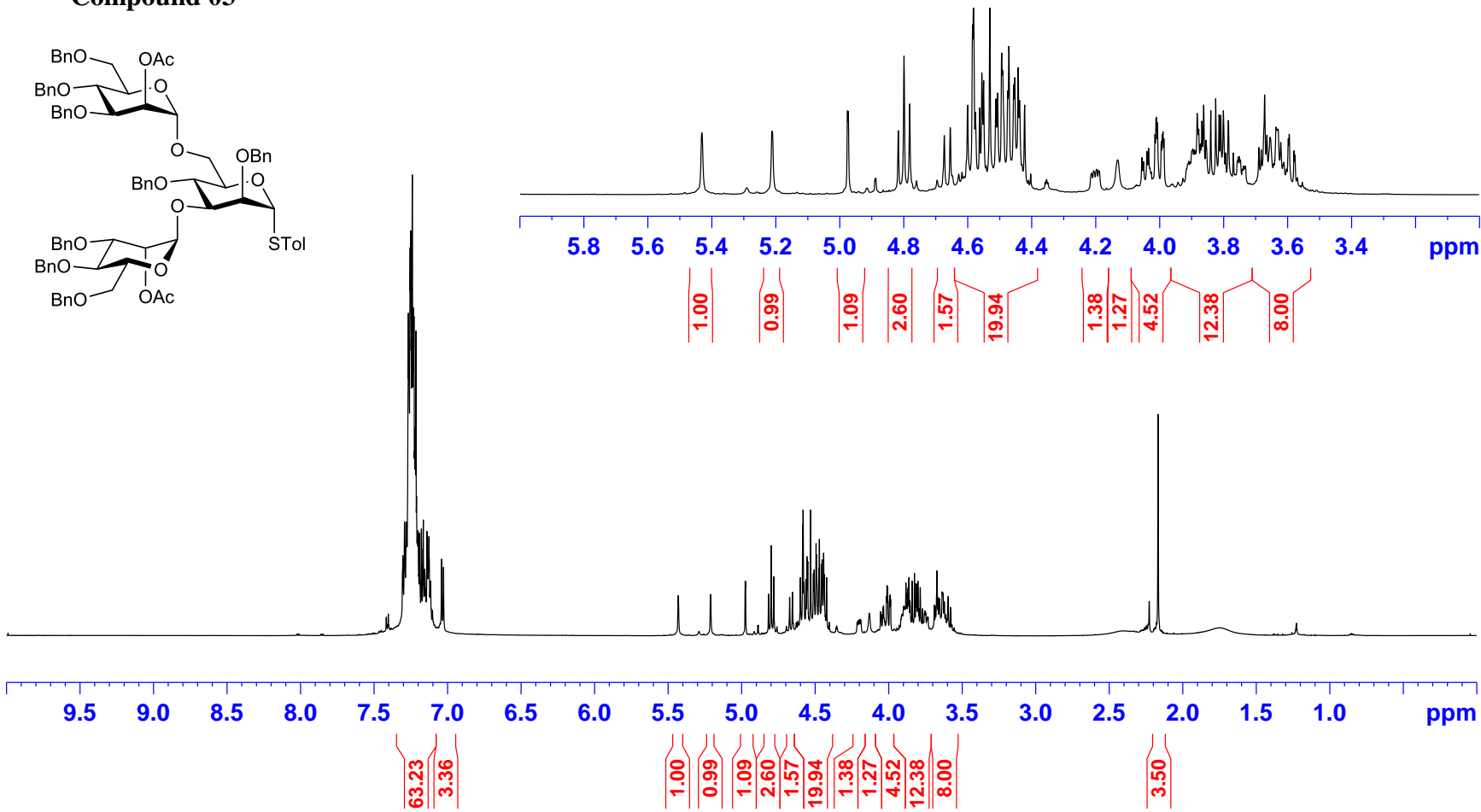
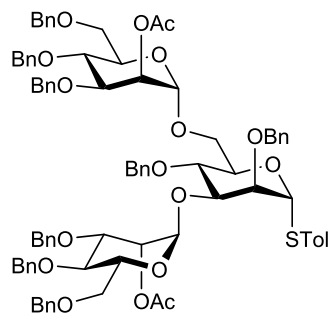


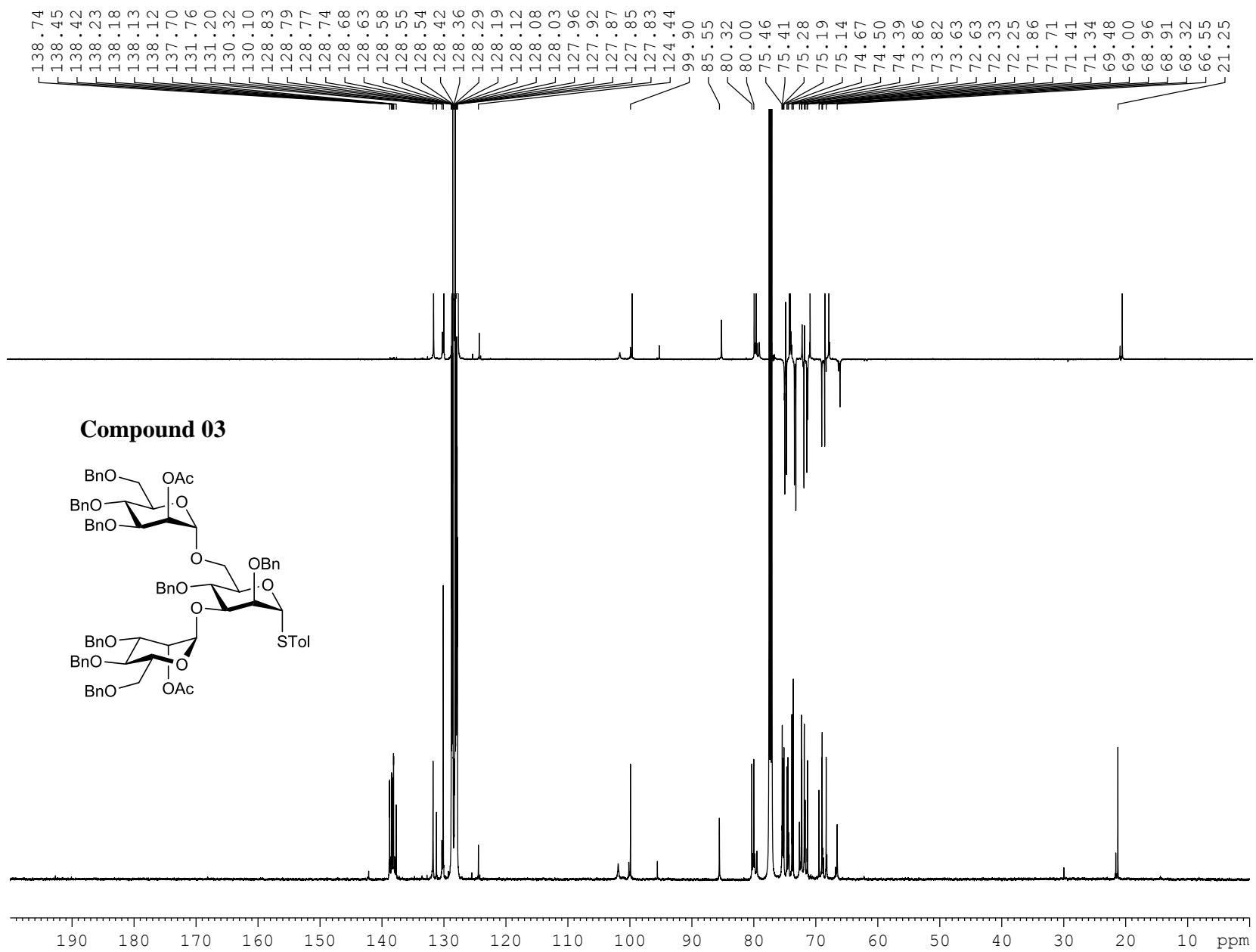
Compound 02



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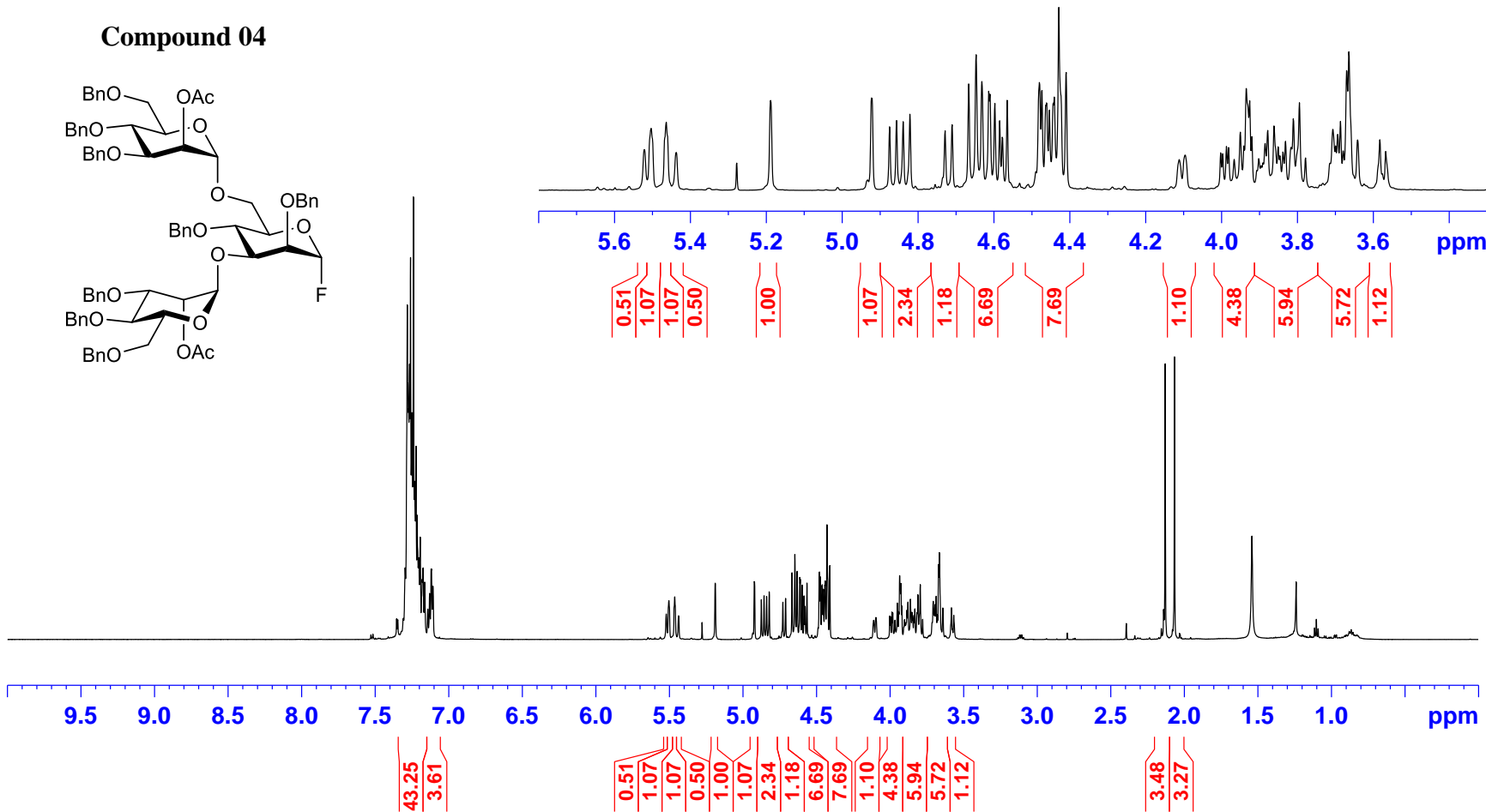
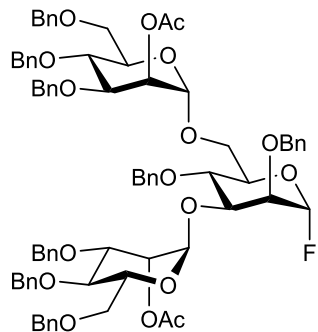
Compound 03





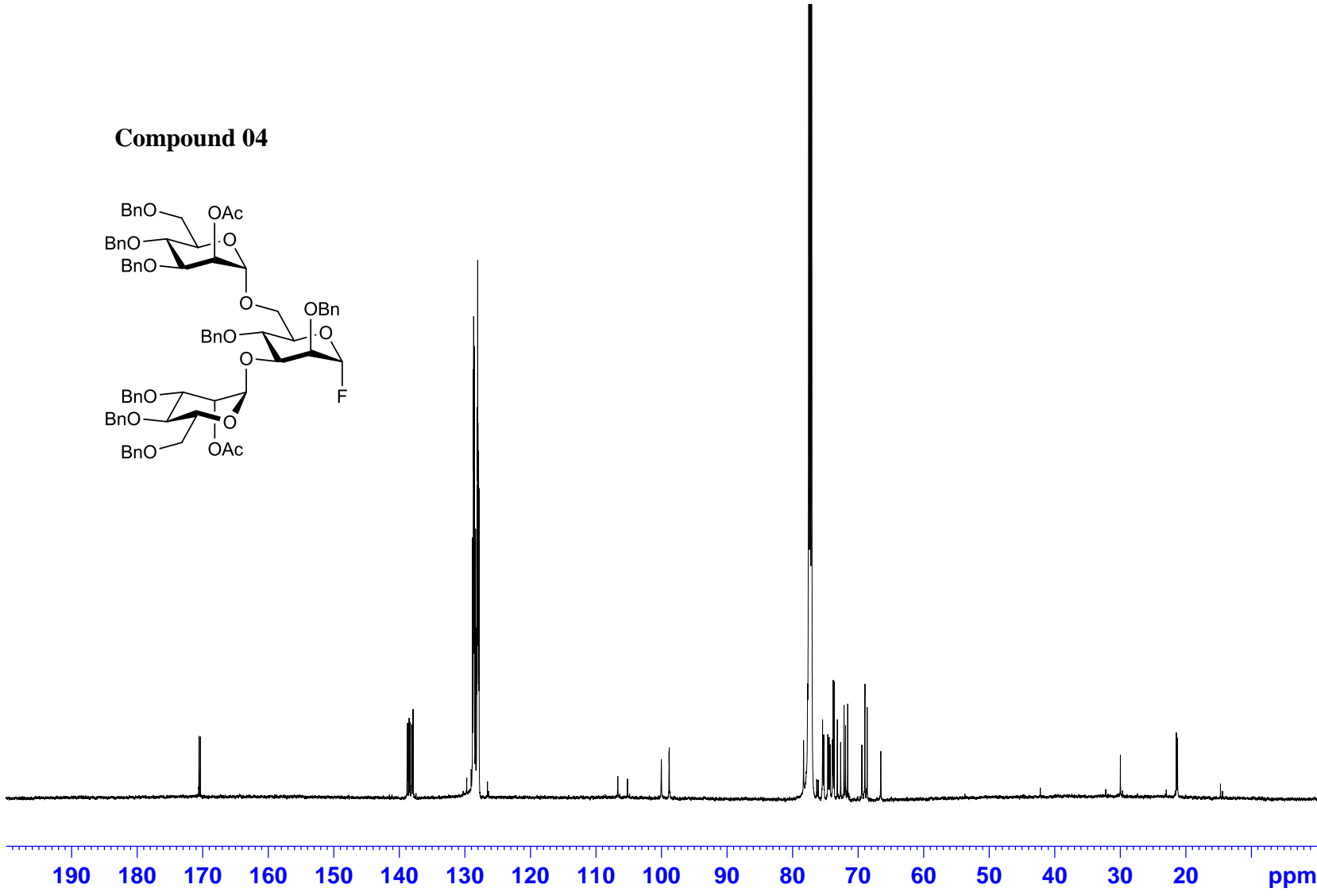
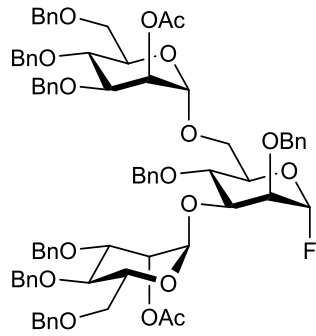
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Compound 04



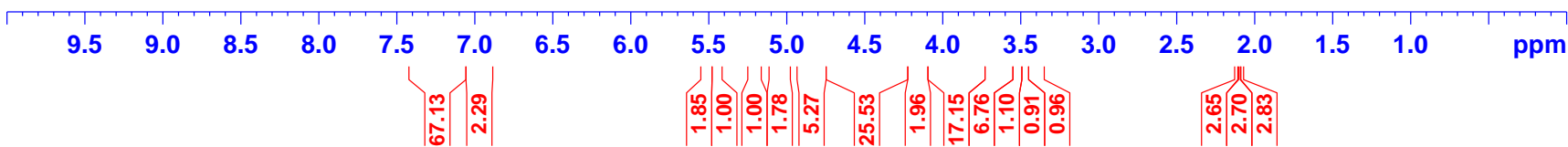
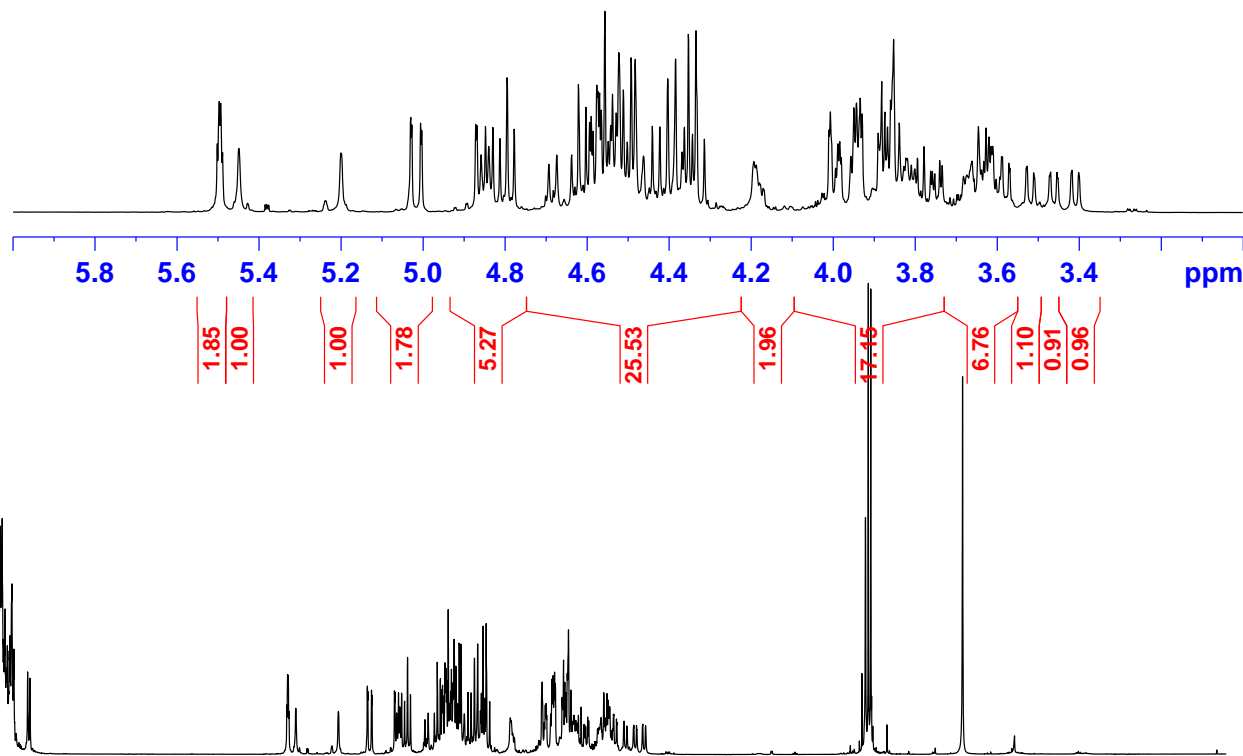
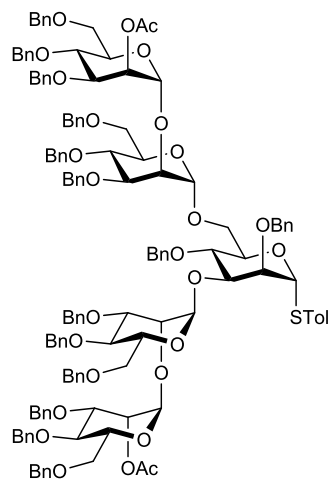
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Compound 04



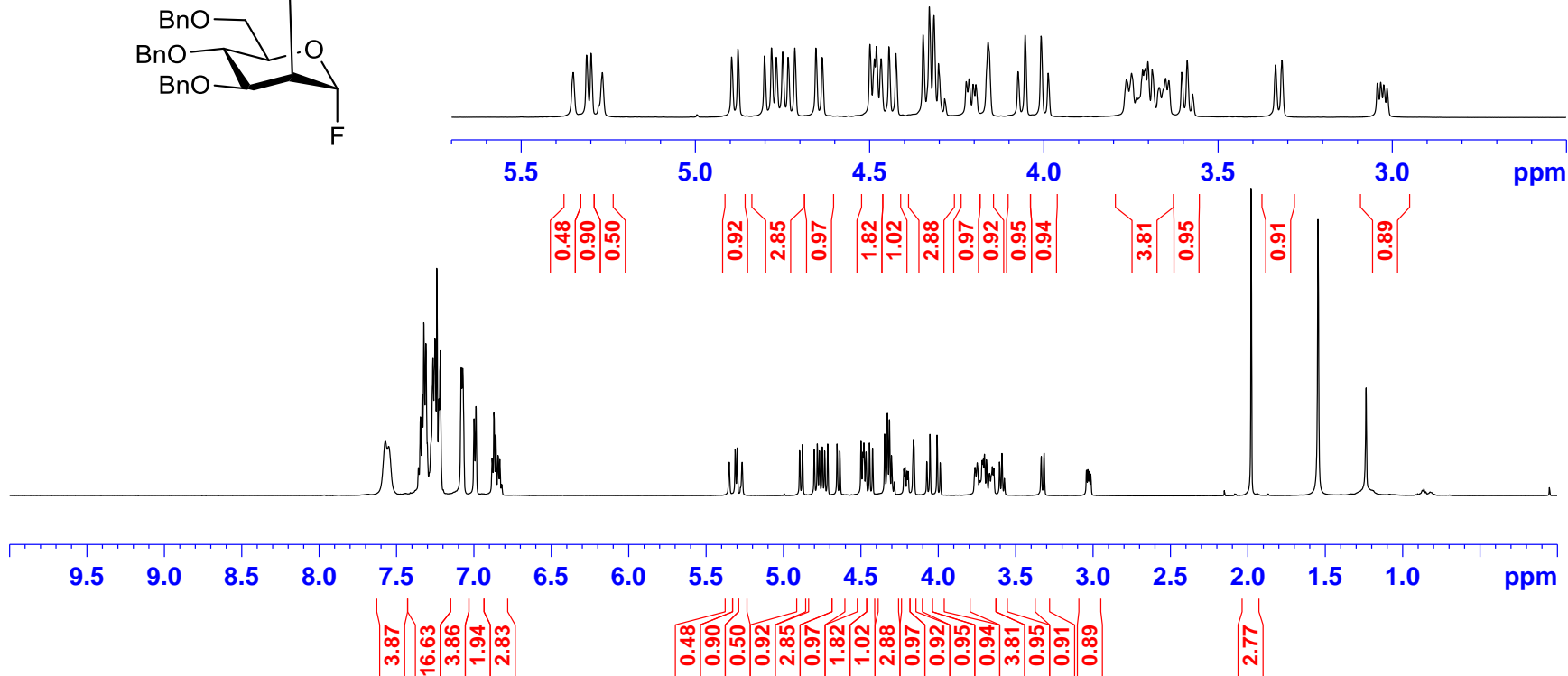
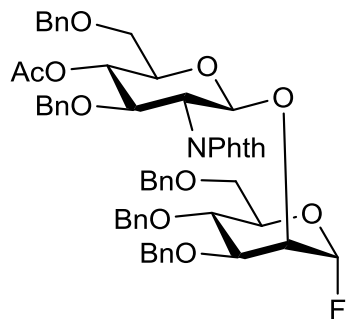
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Compound 05



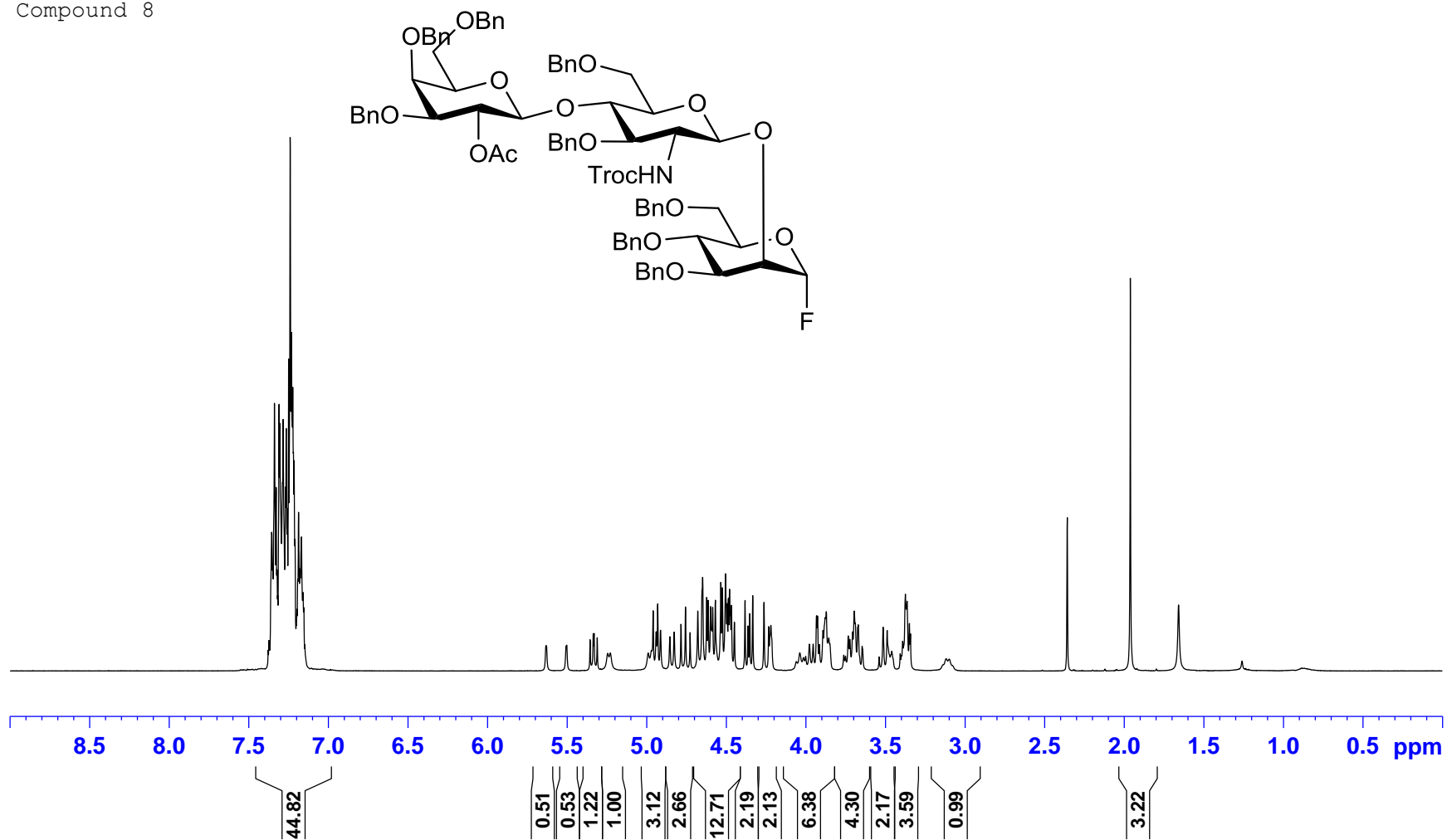
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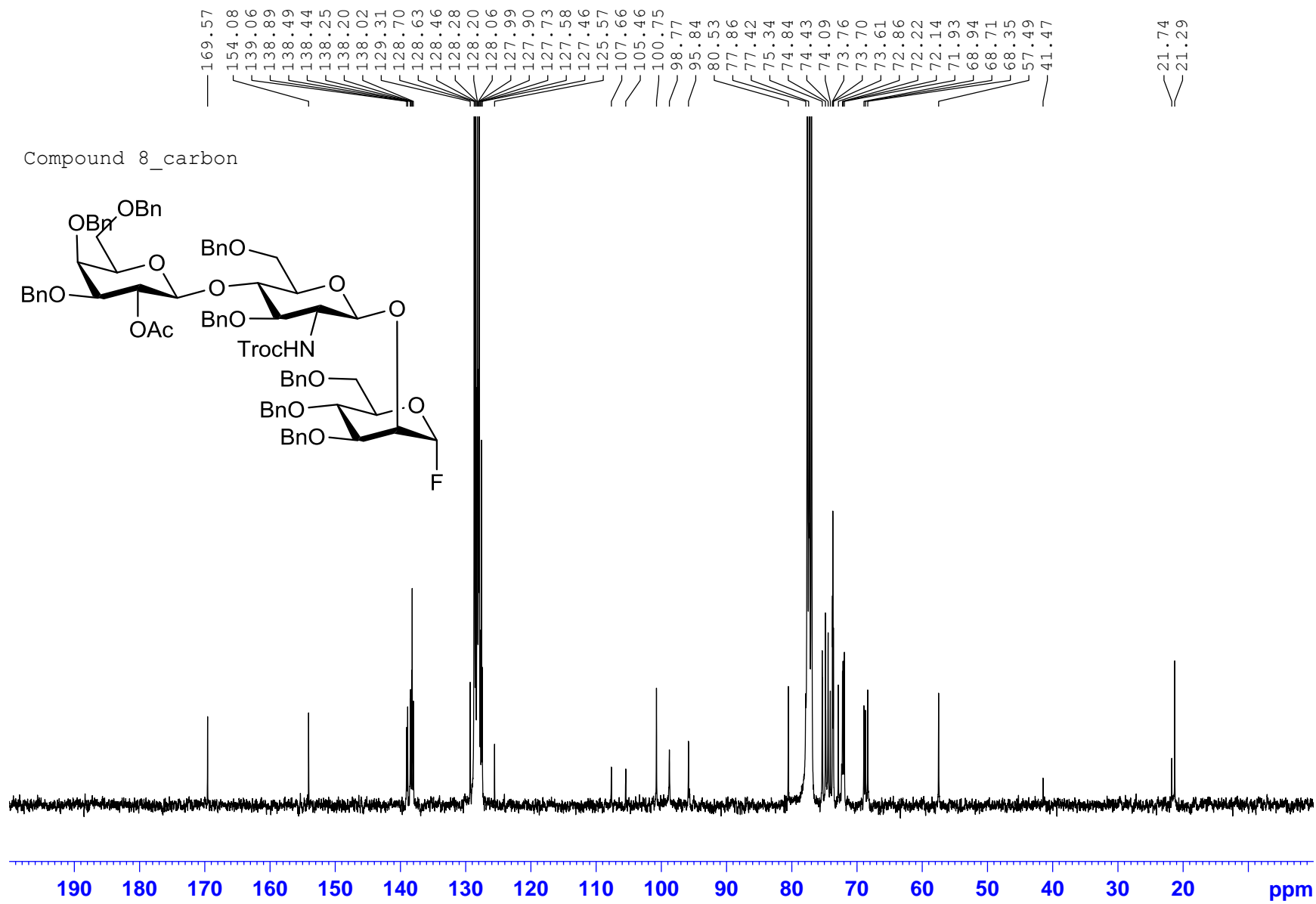
Compound 06



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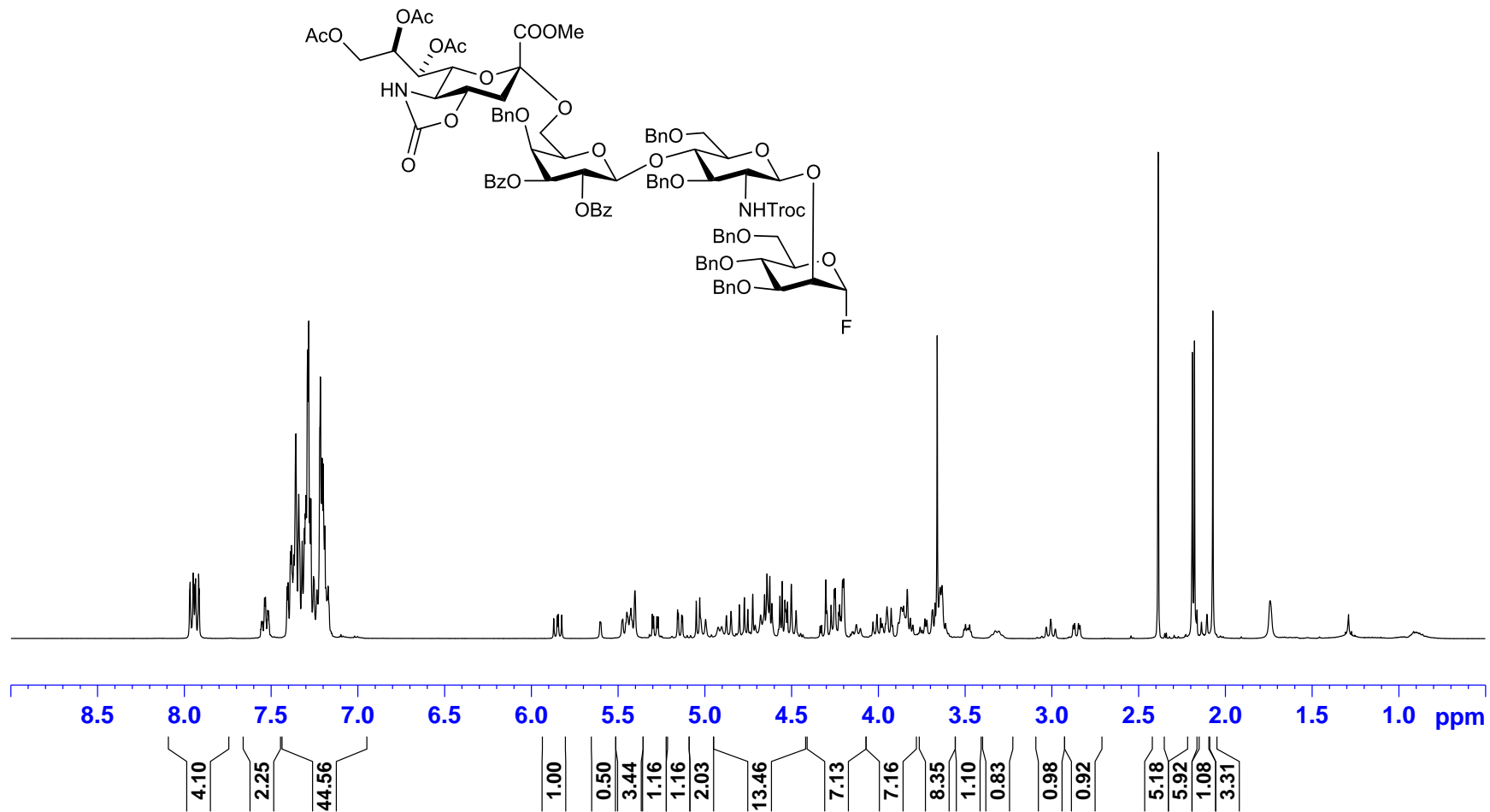
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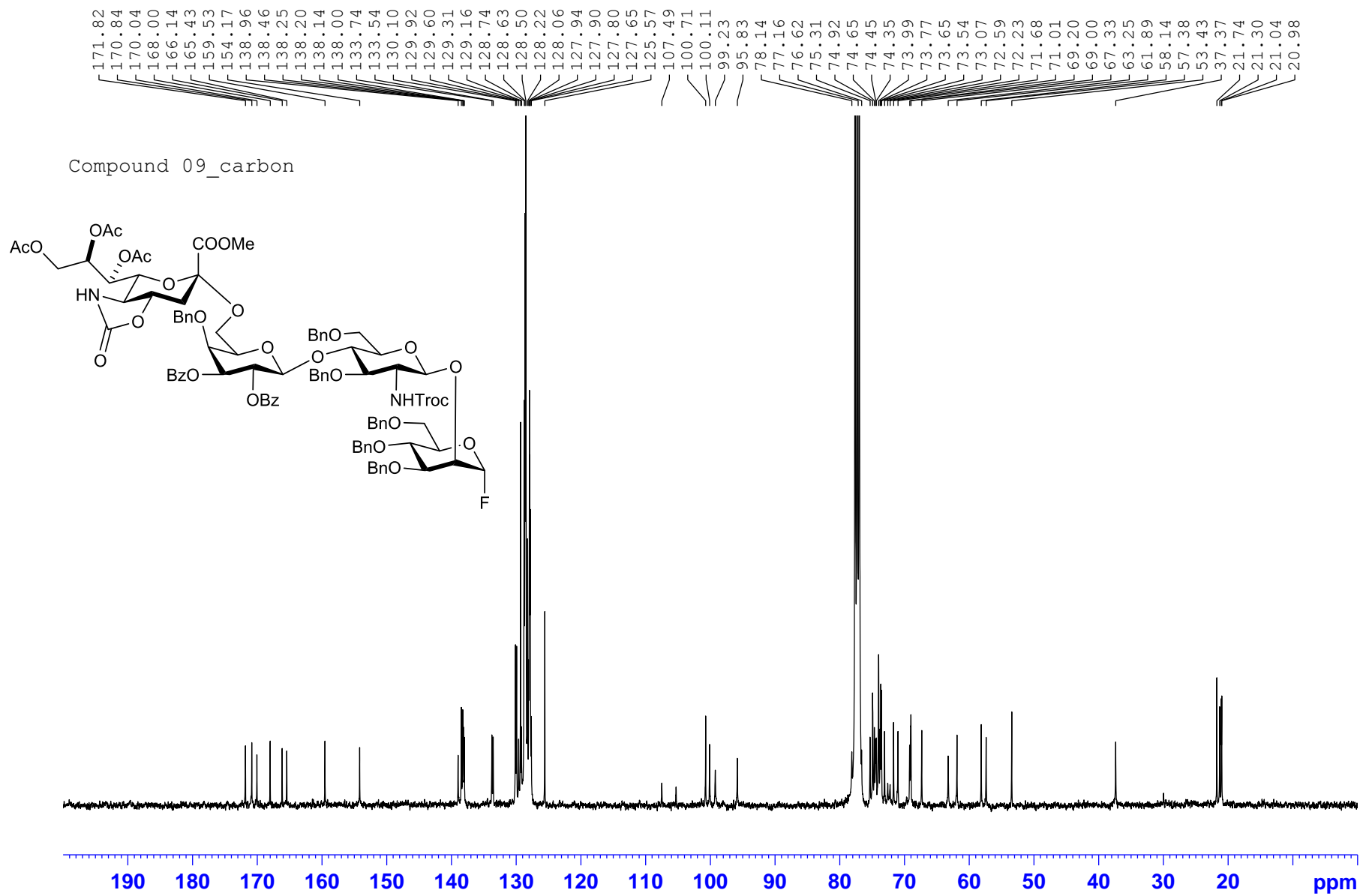




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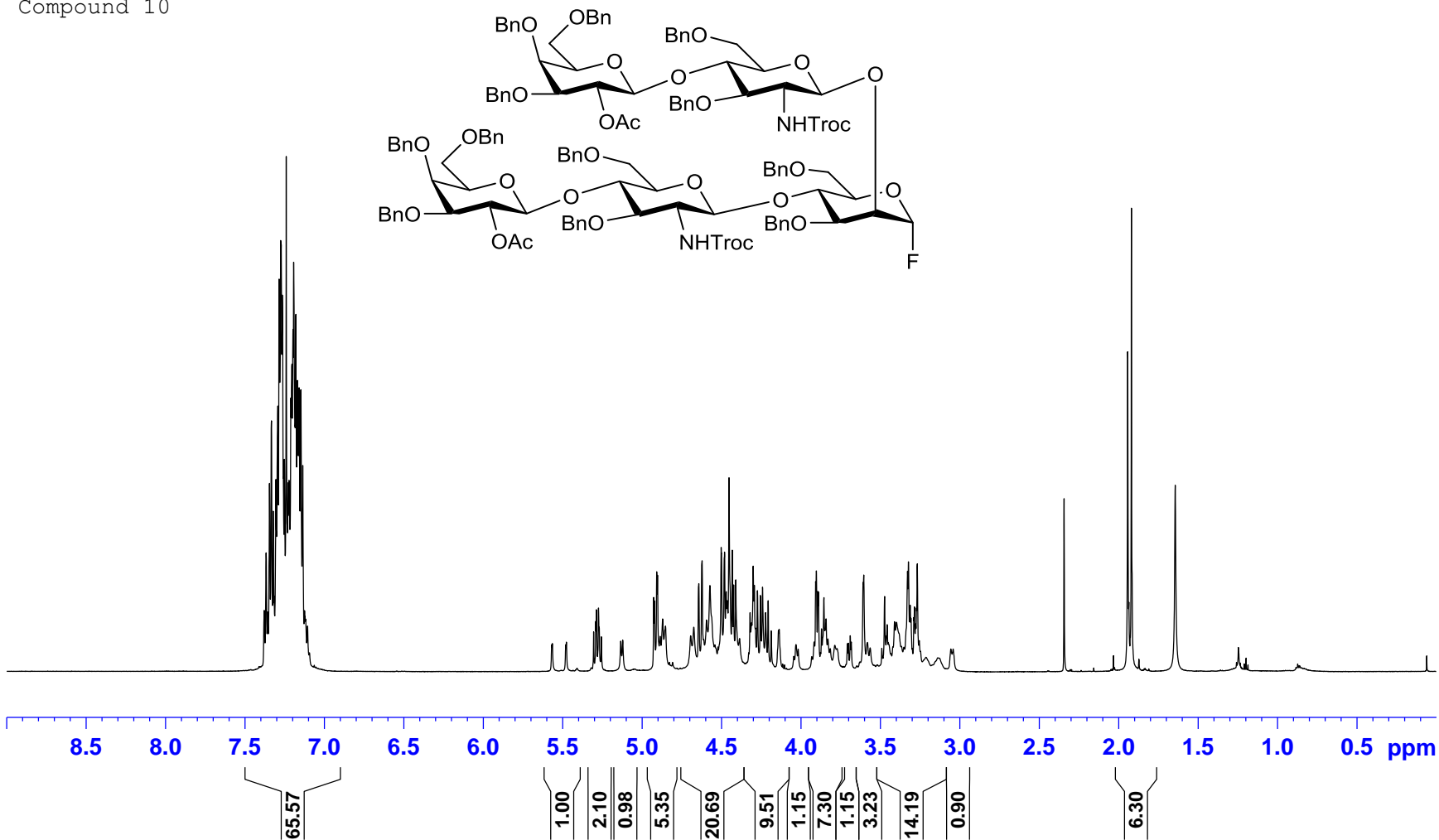
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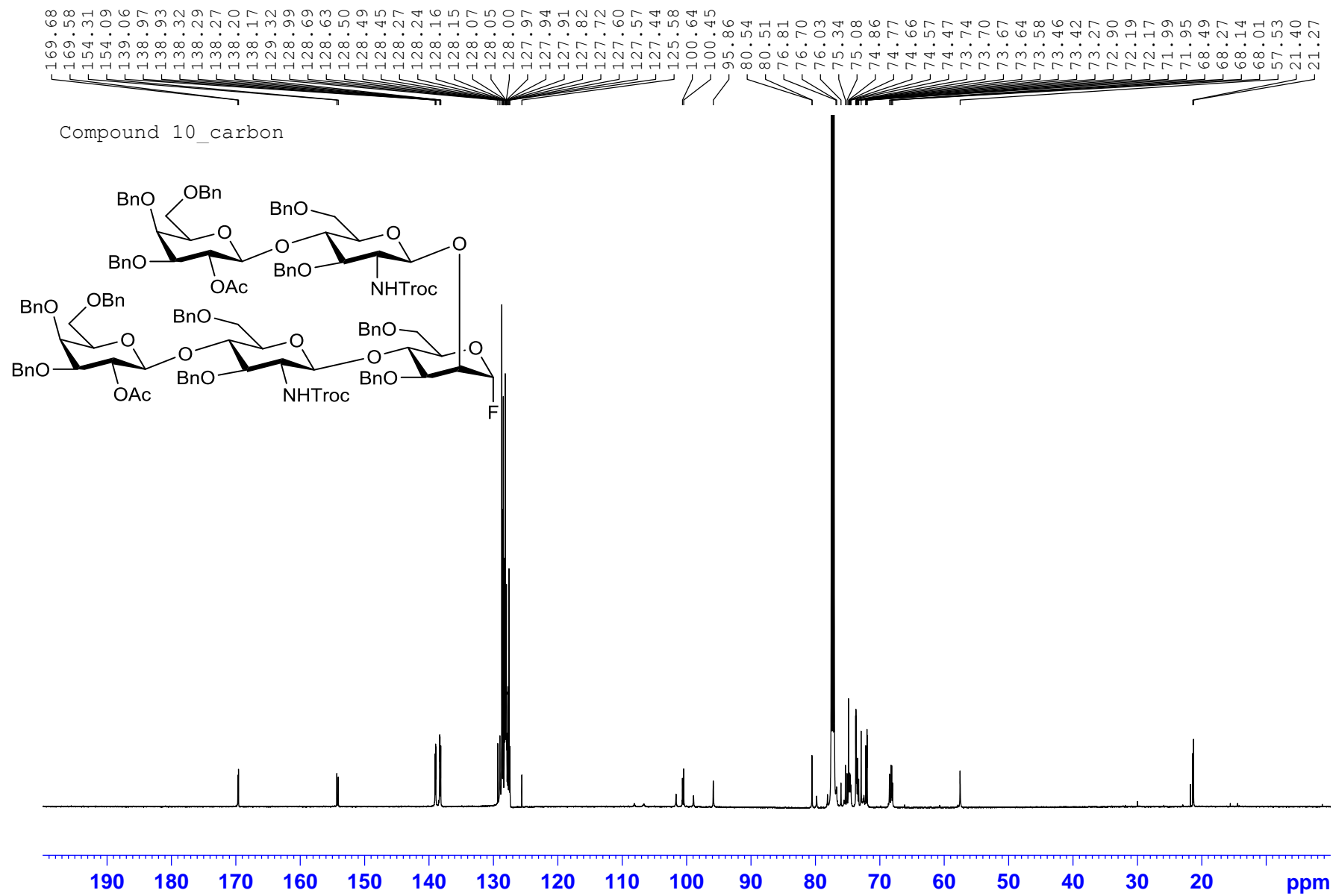


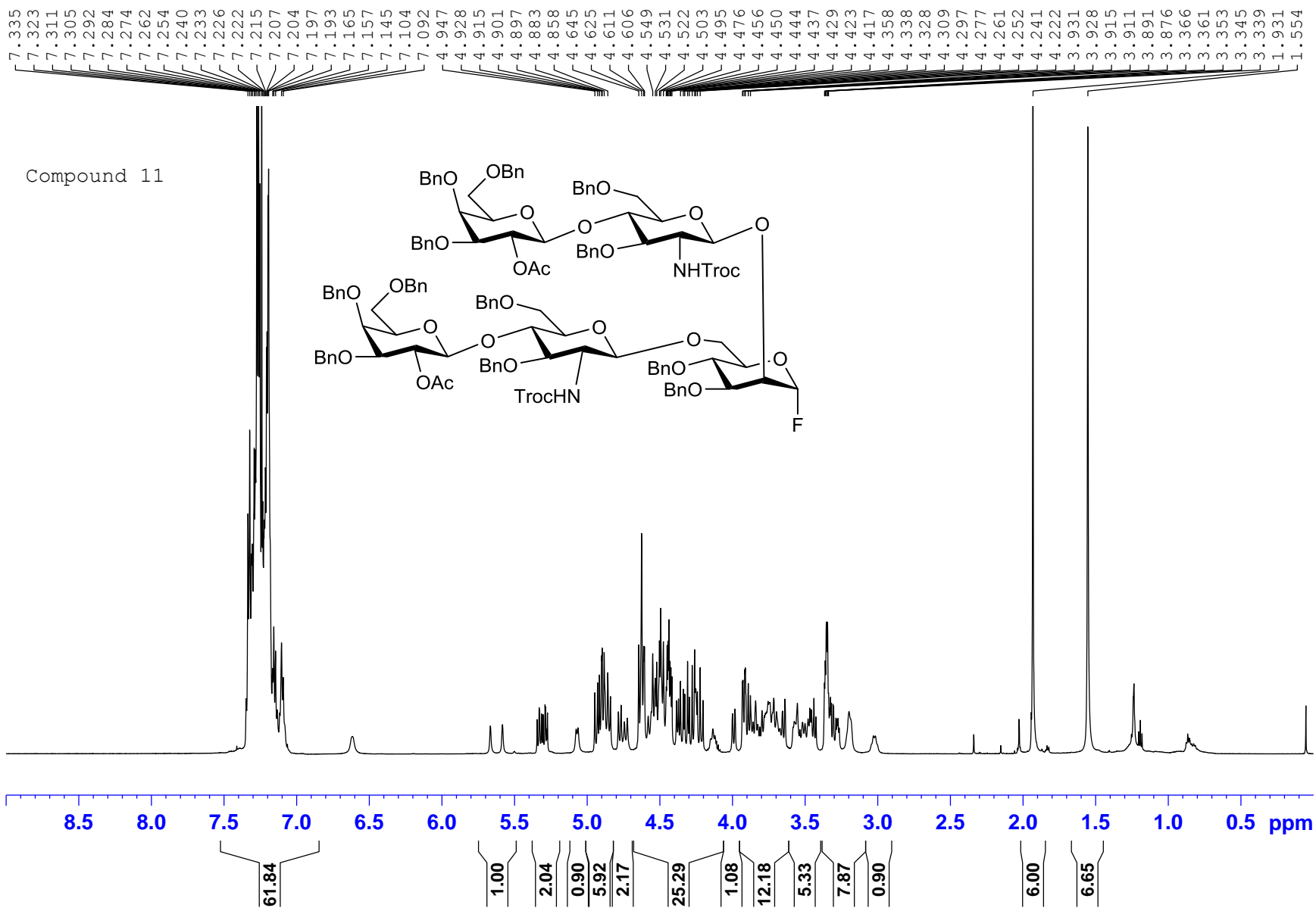


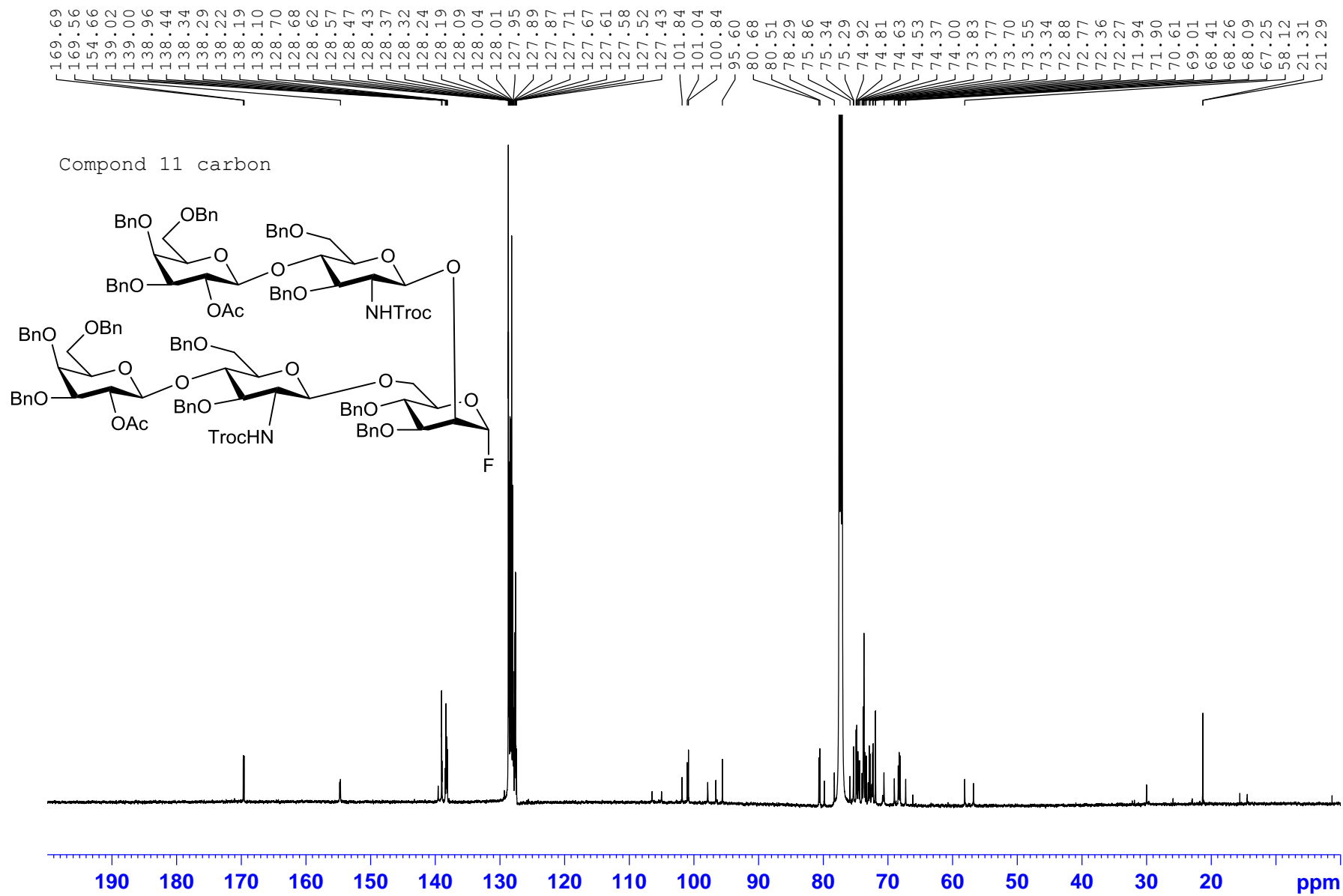
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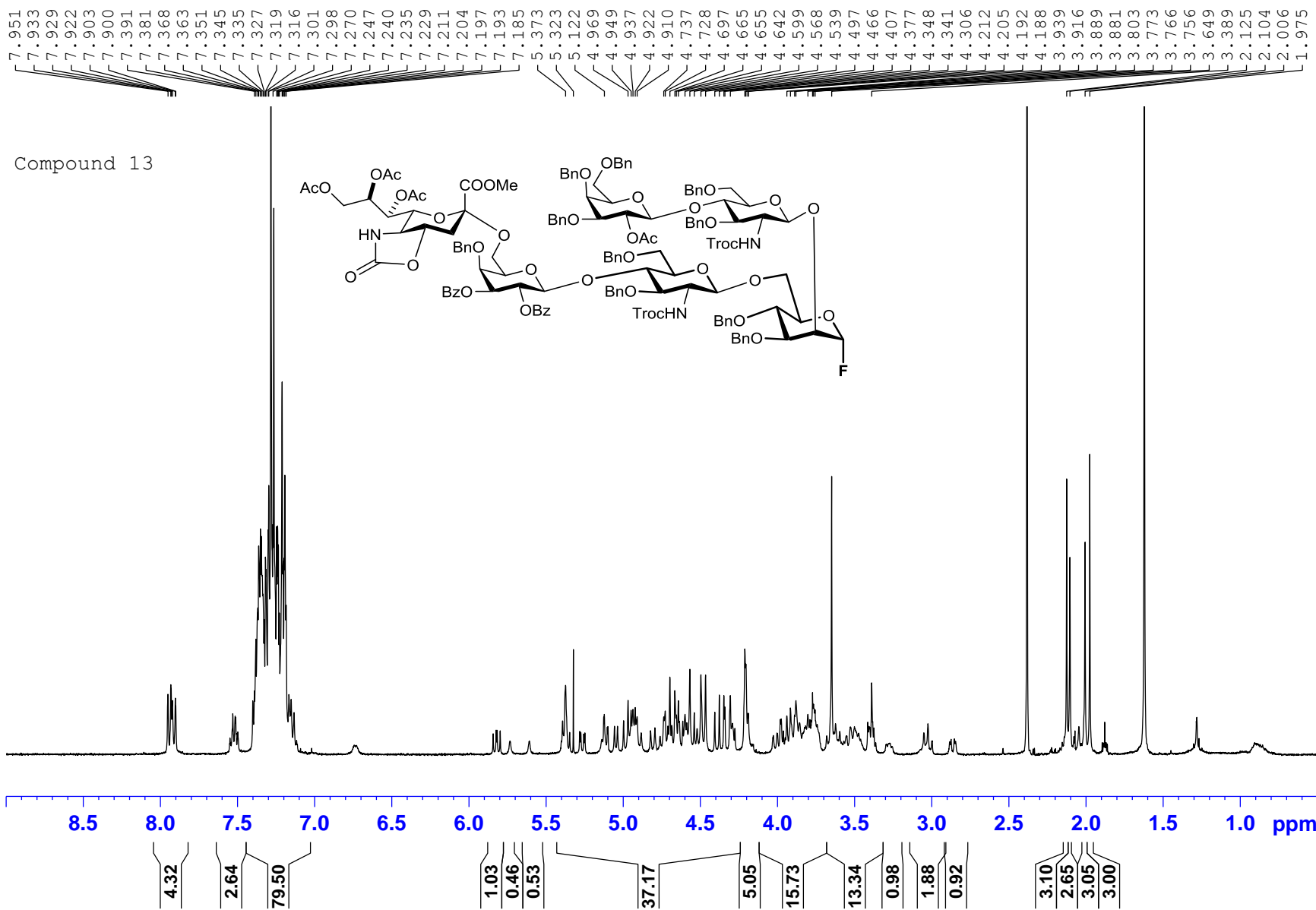
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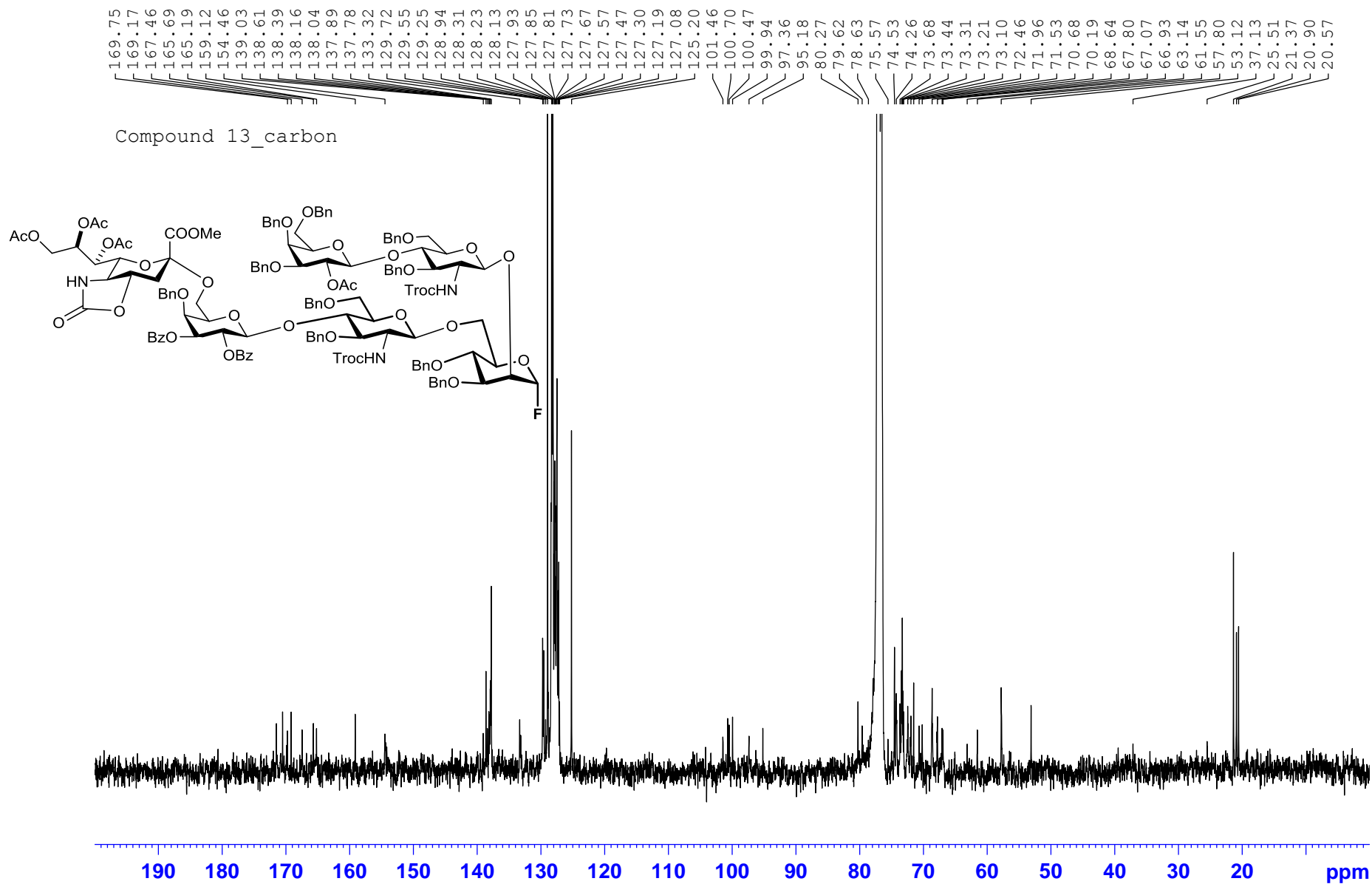






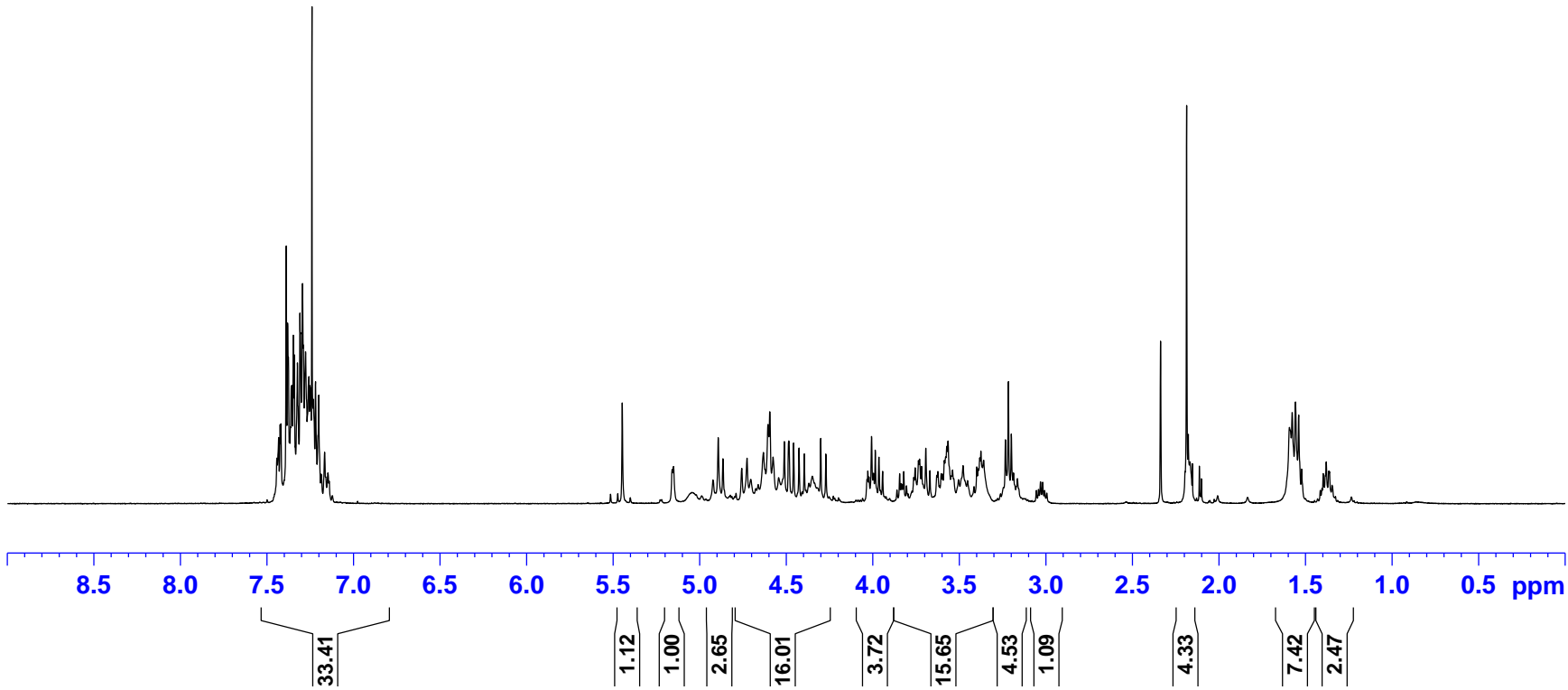
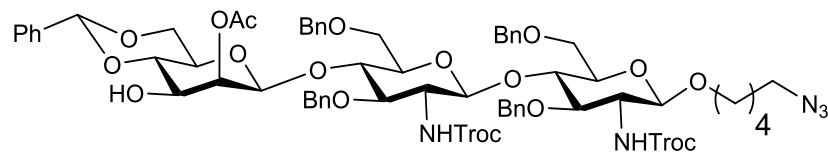




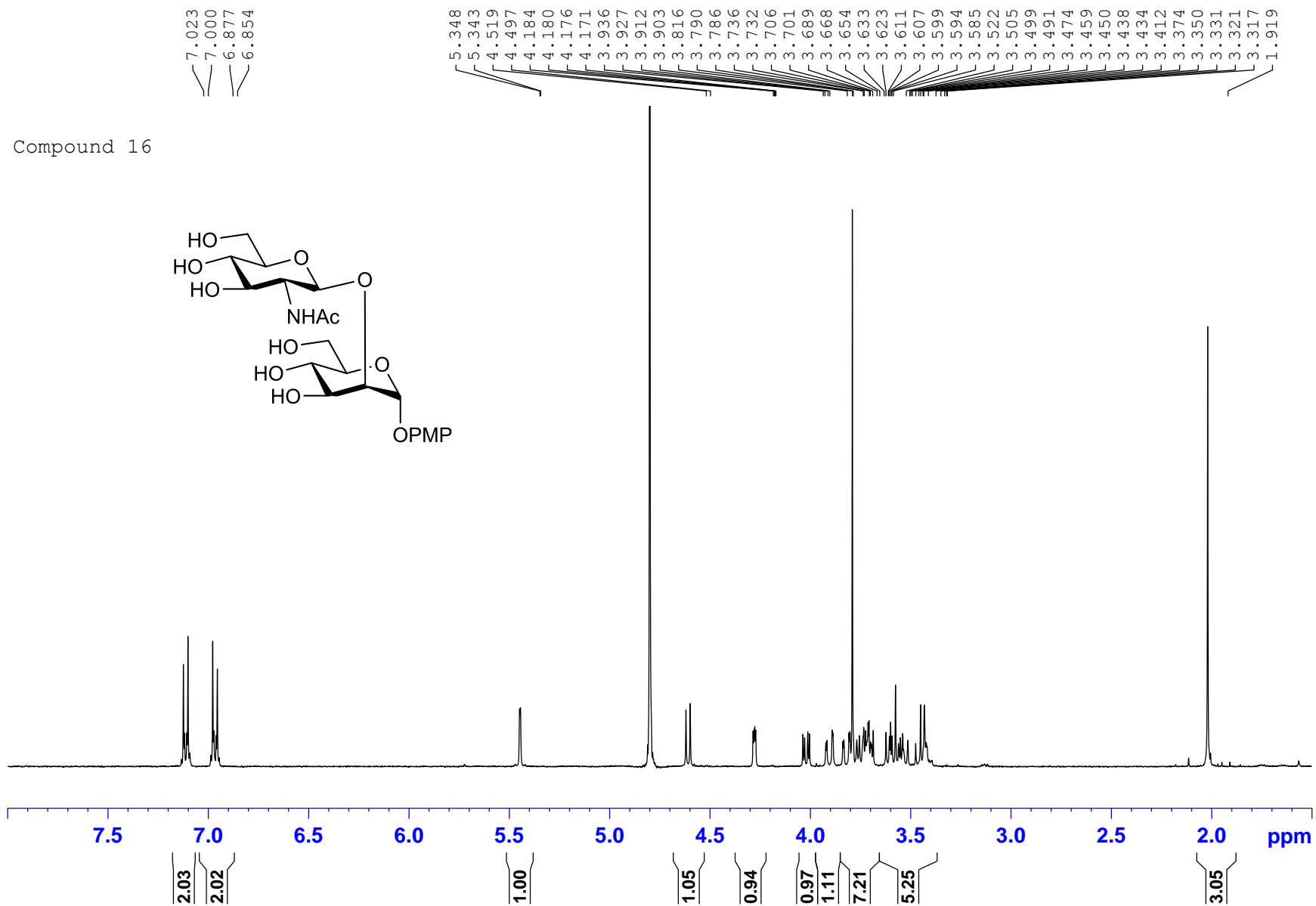
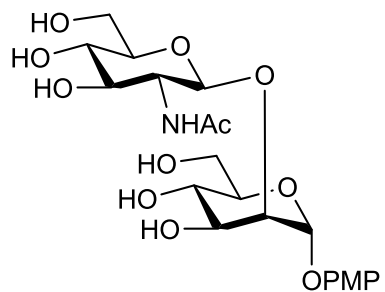


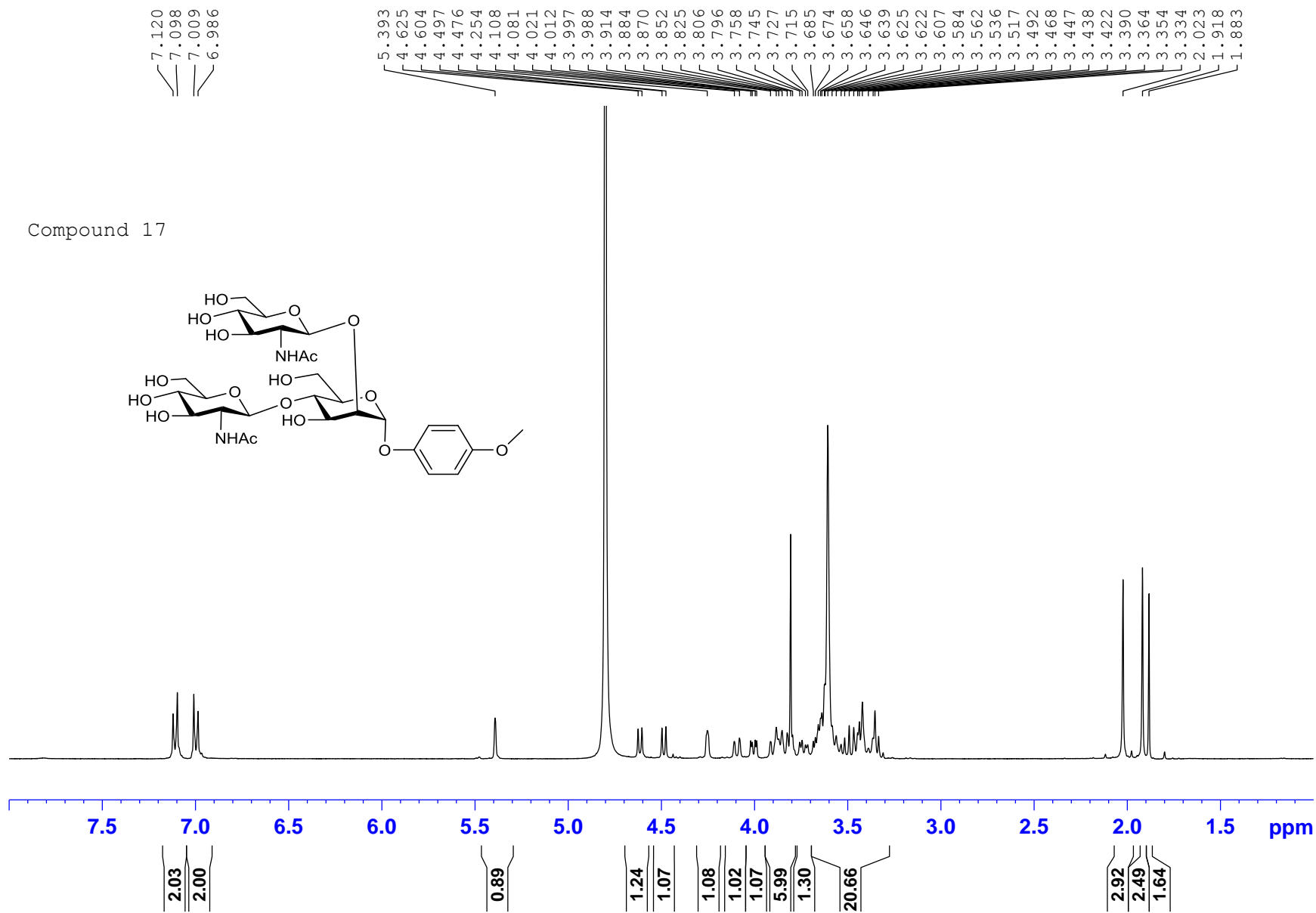
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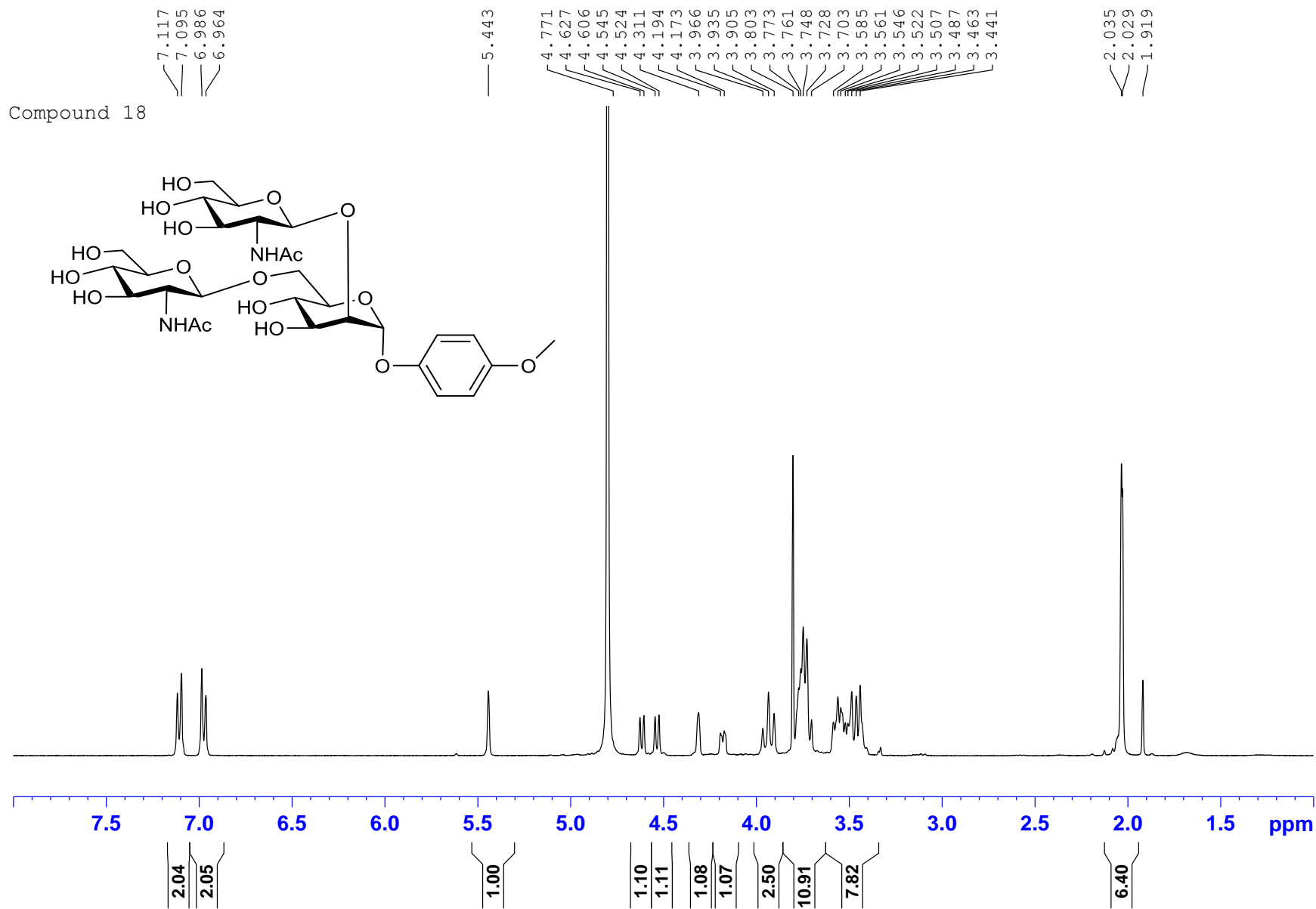
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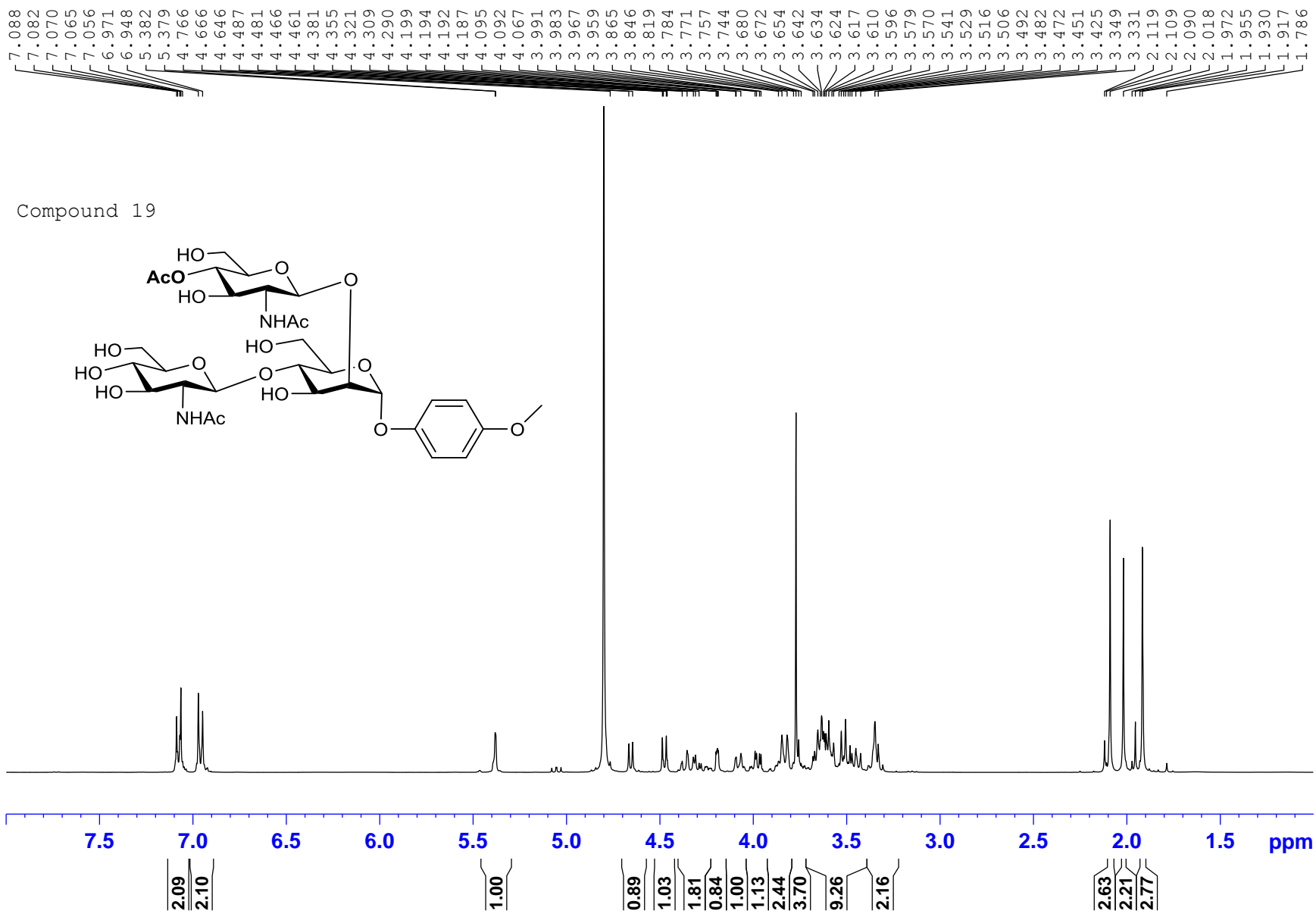


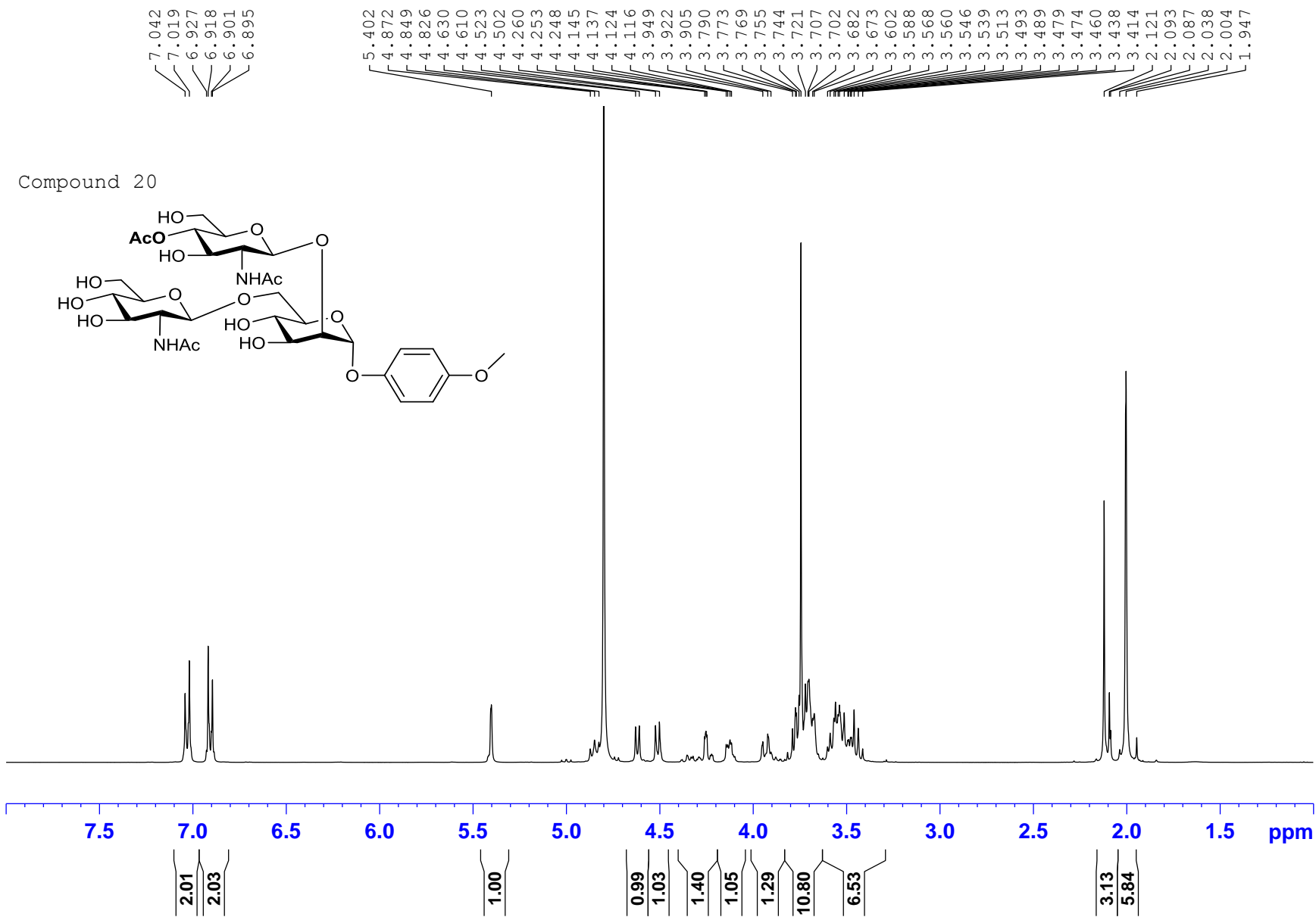
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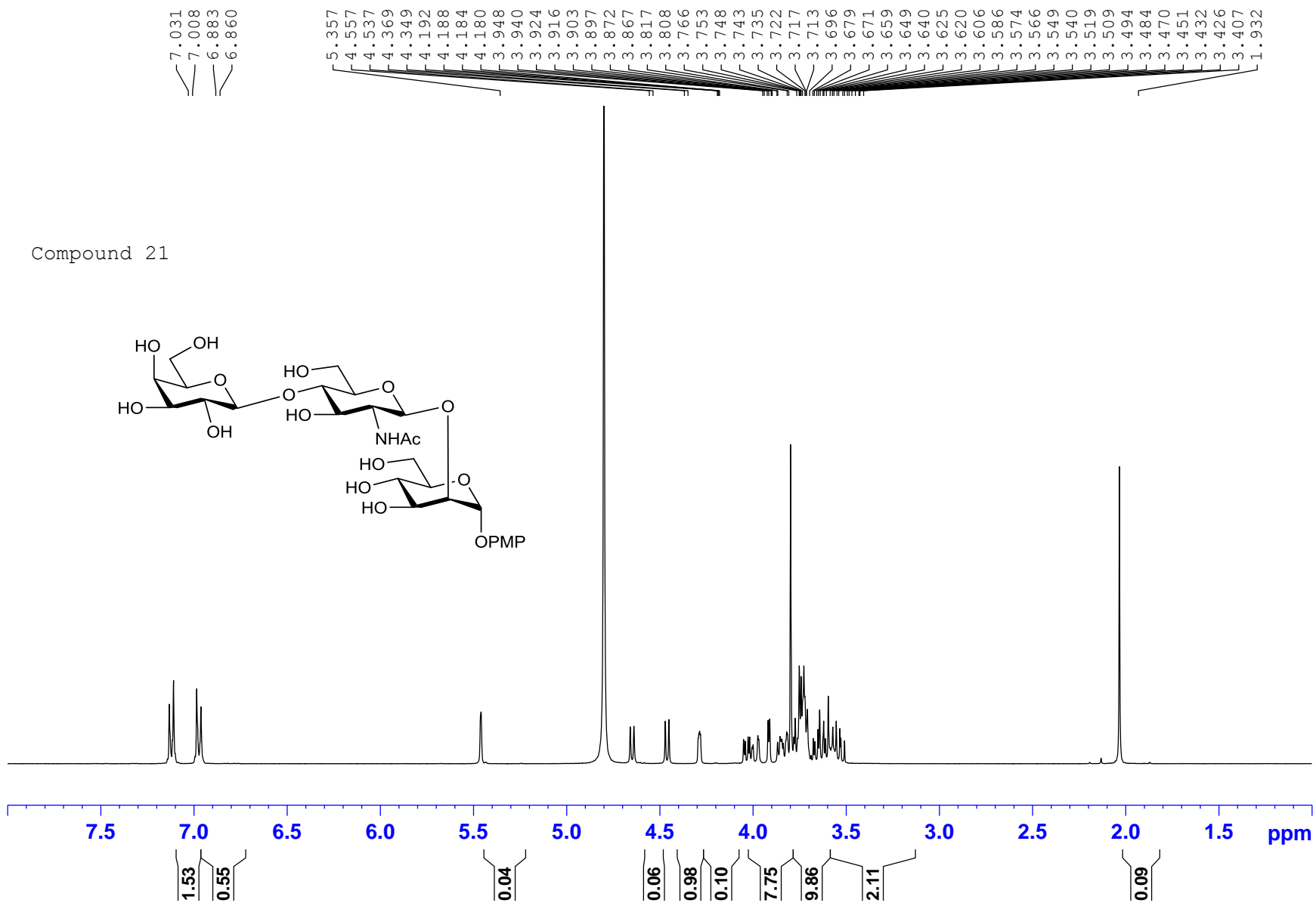
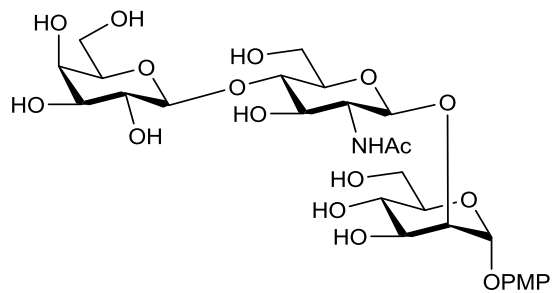


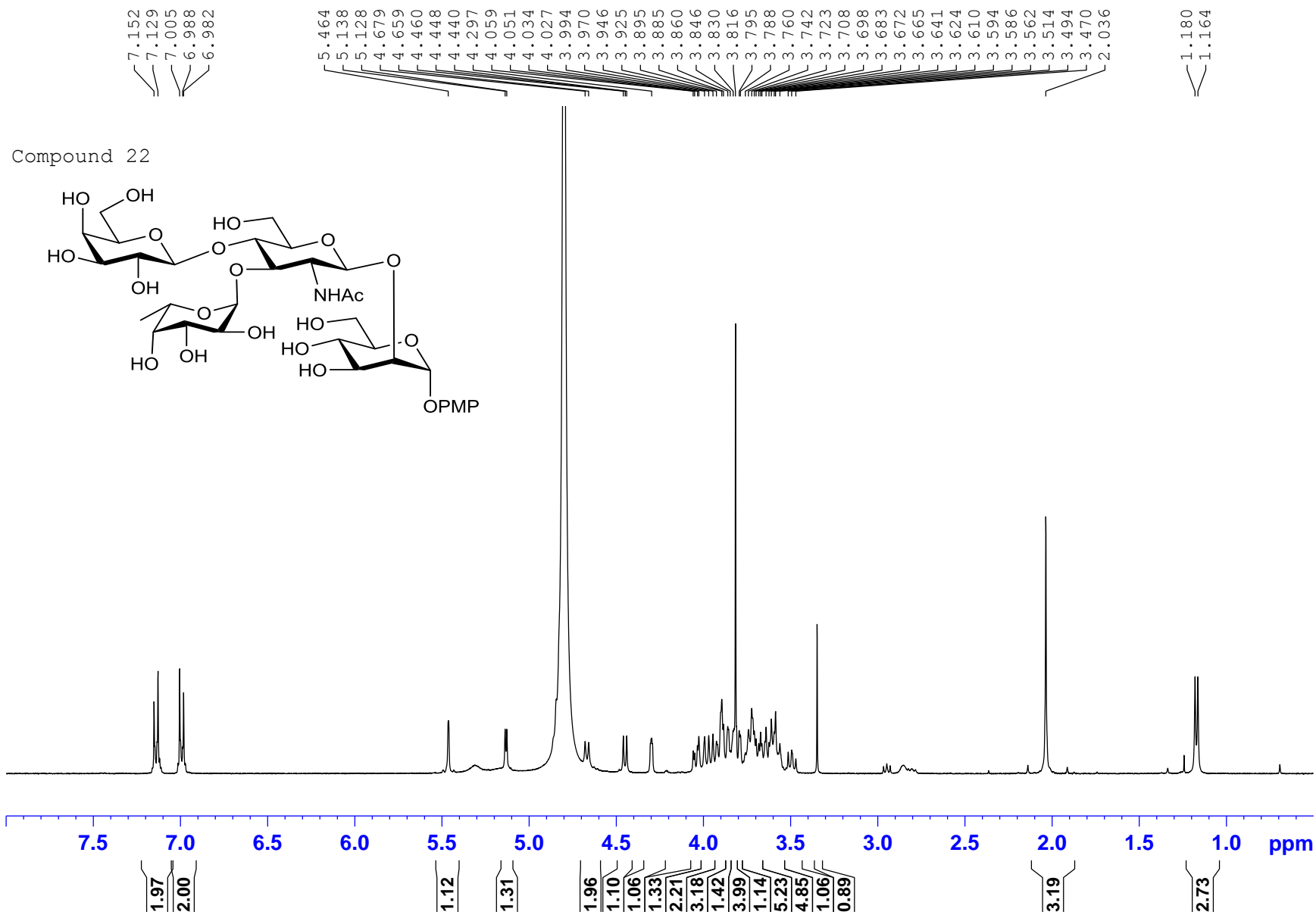


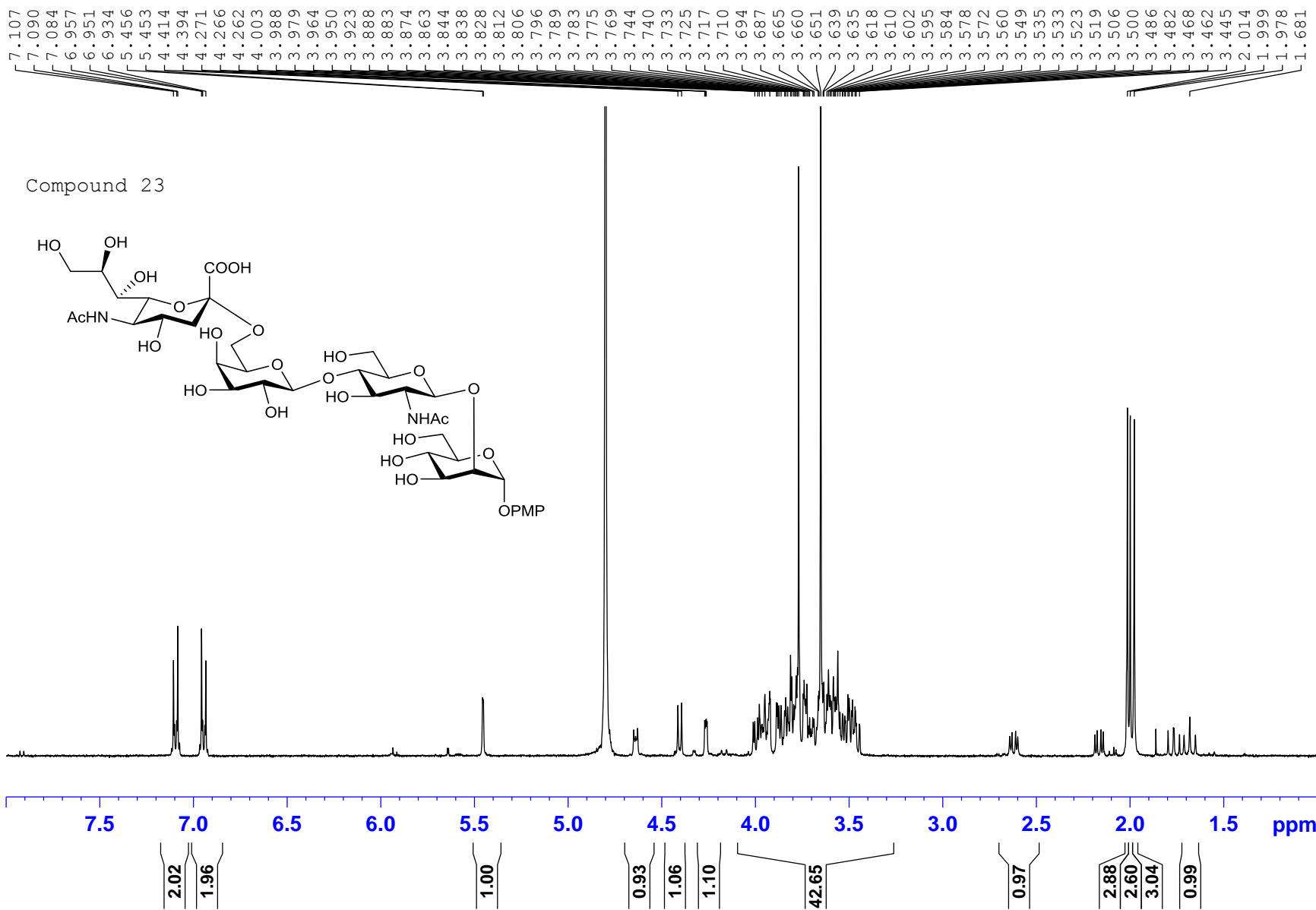


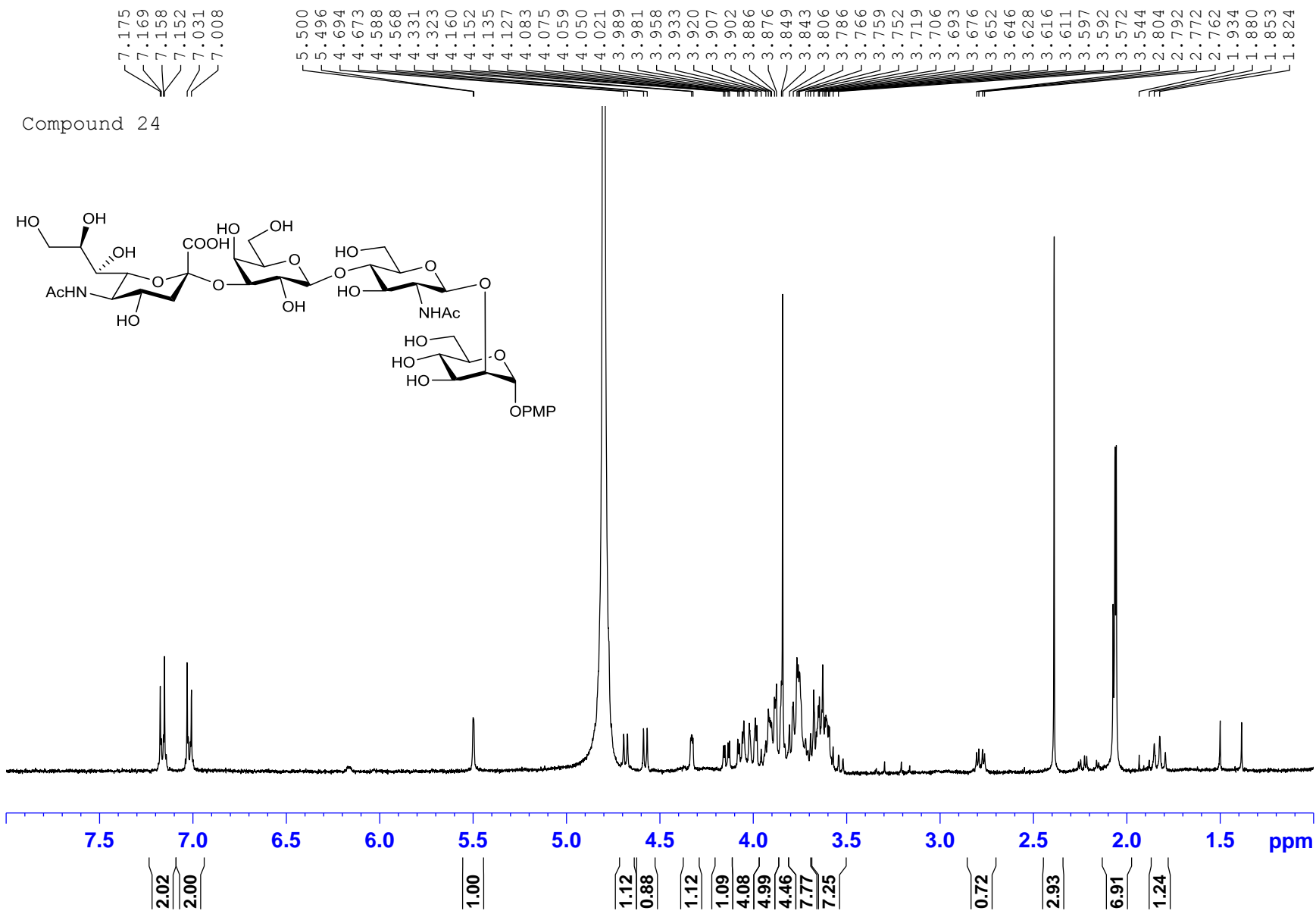


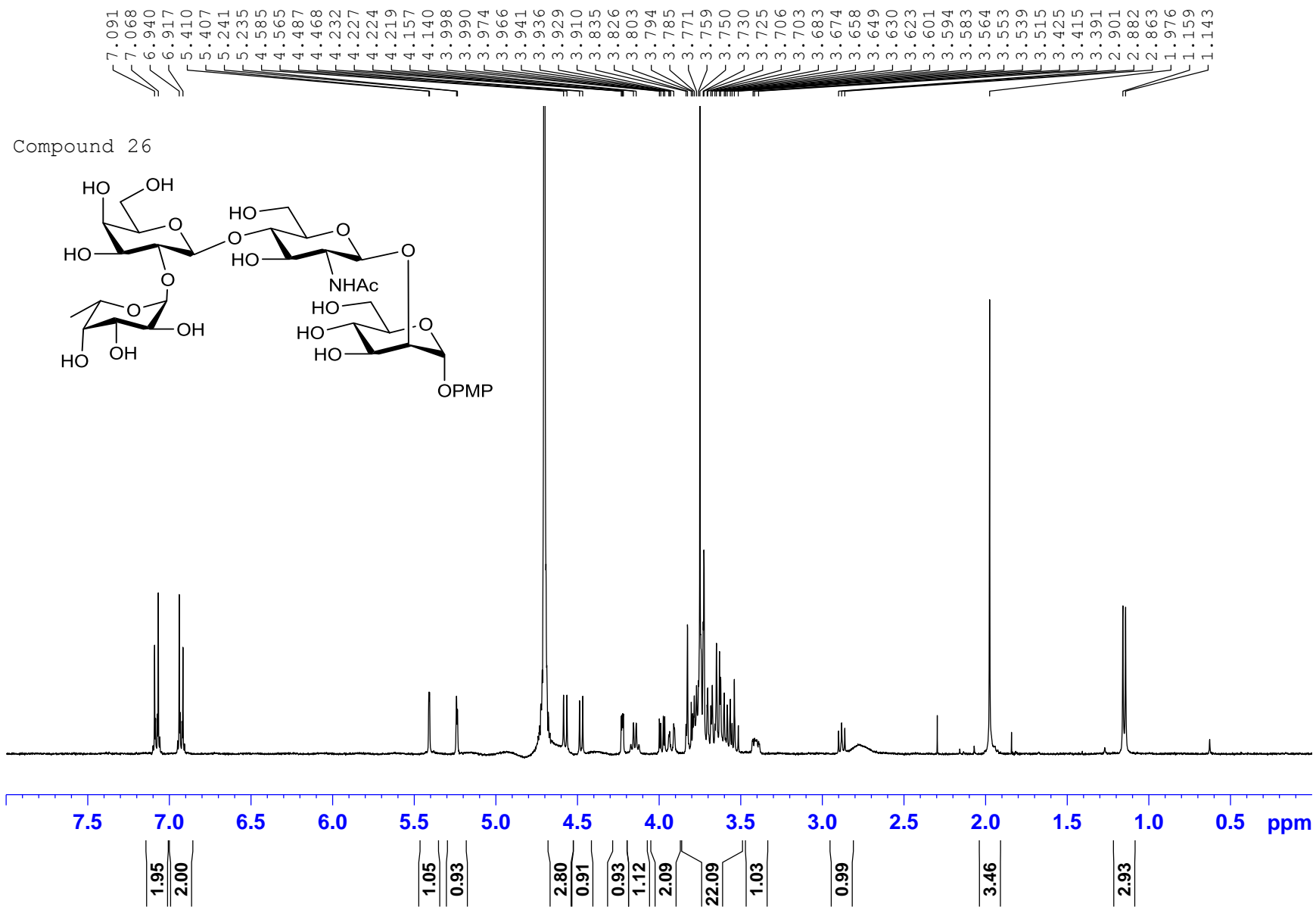
Compound 21

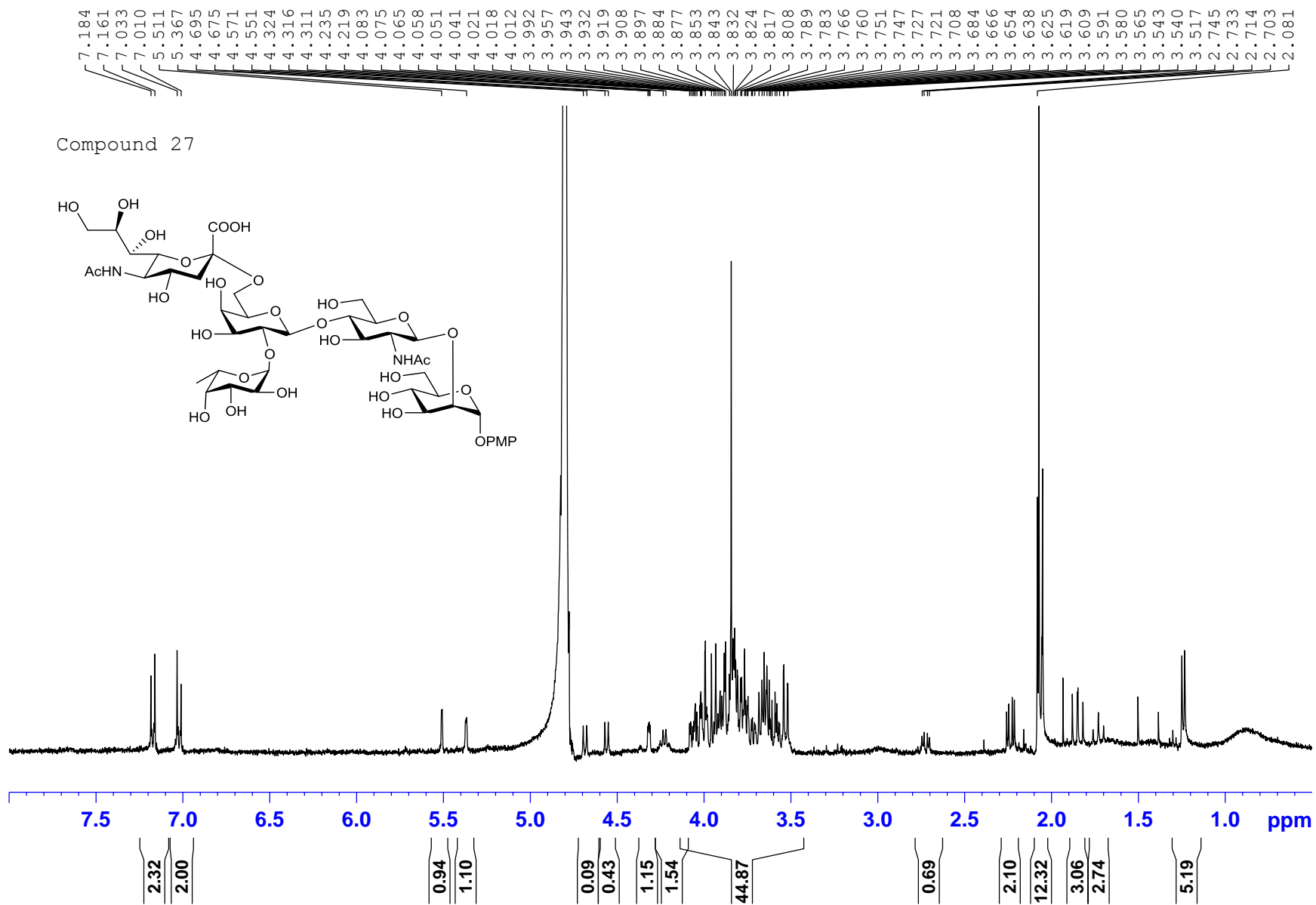


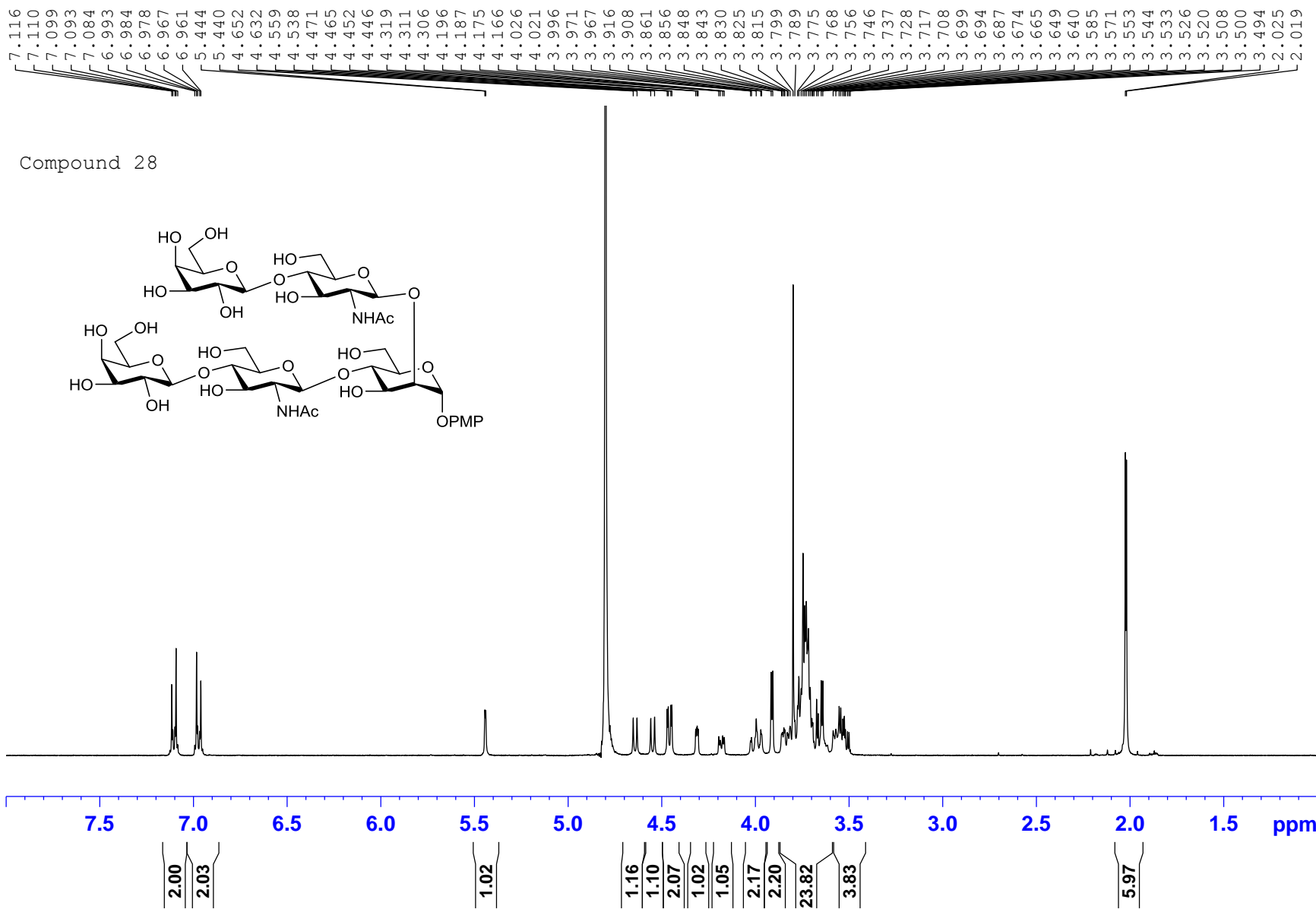


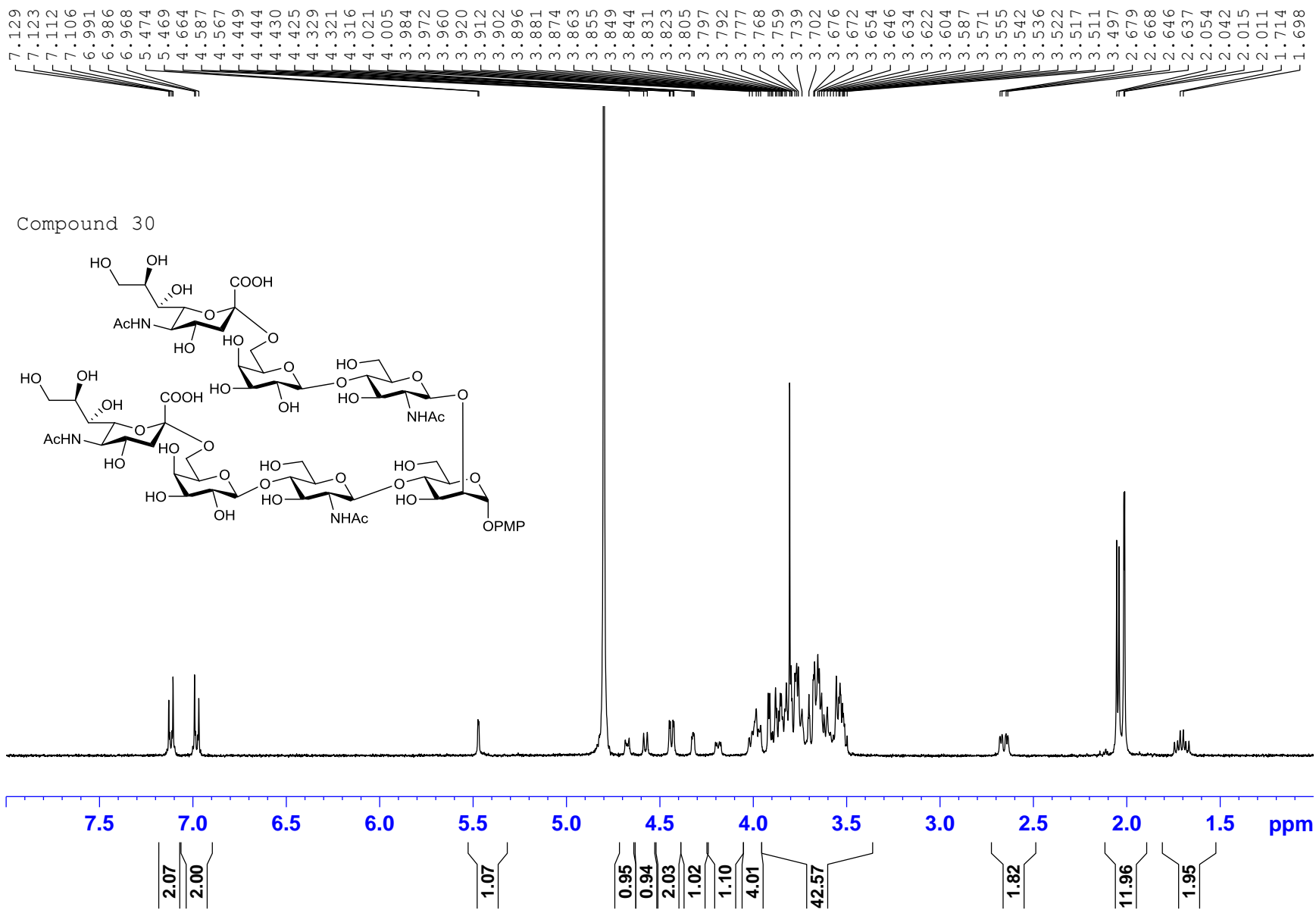


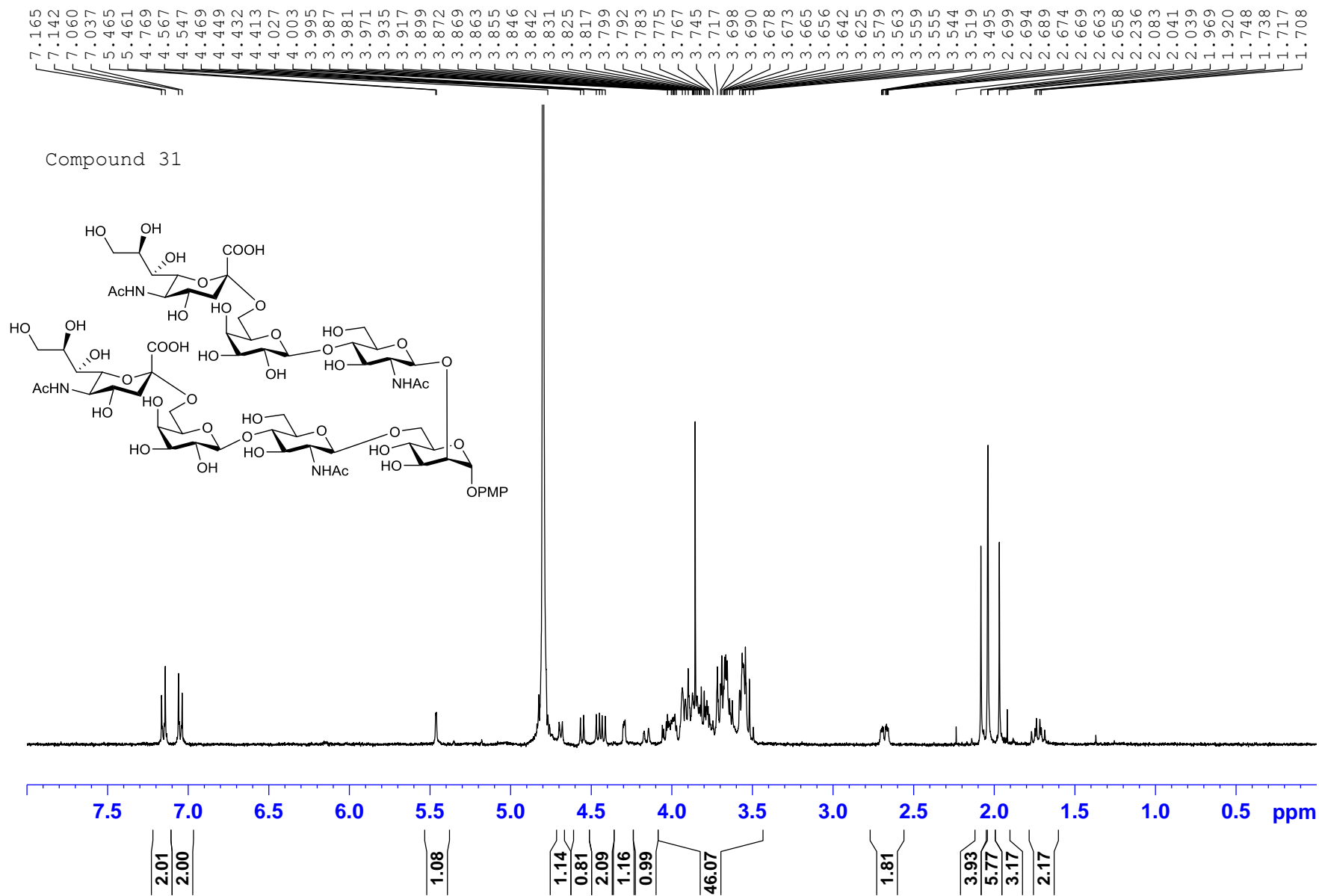


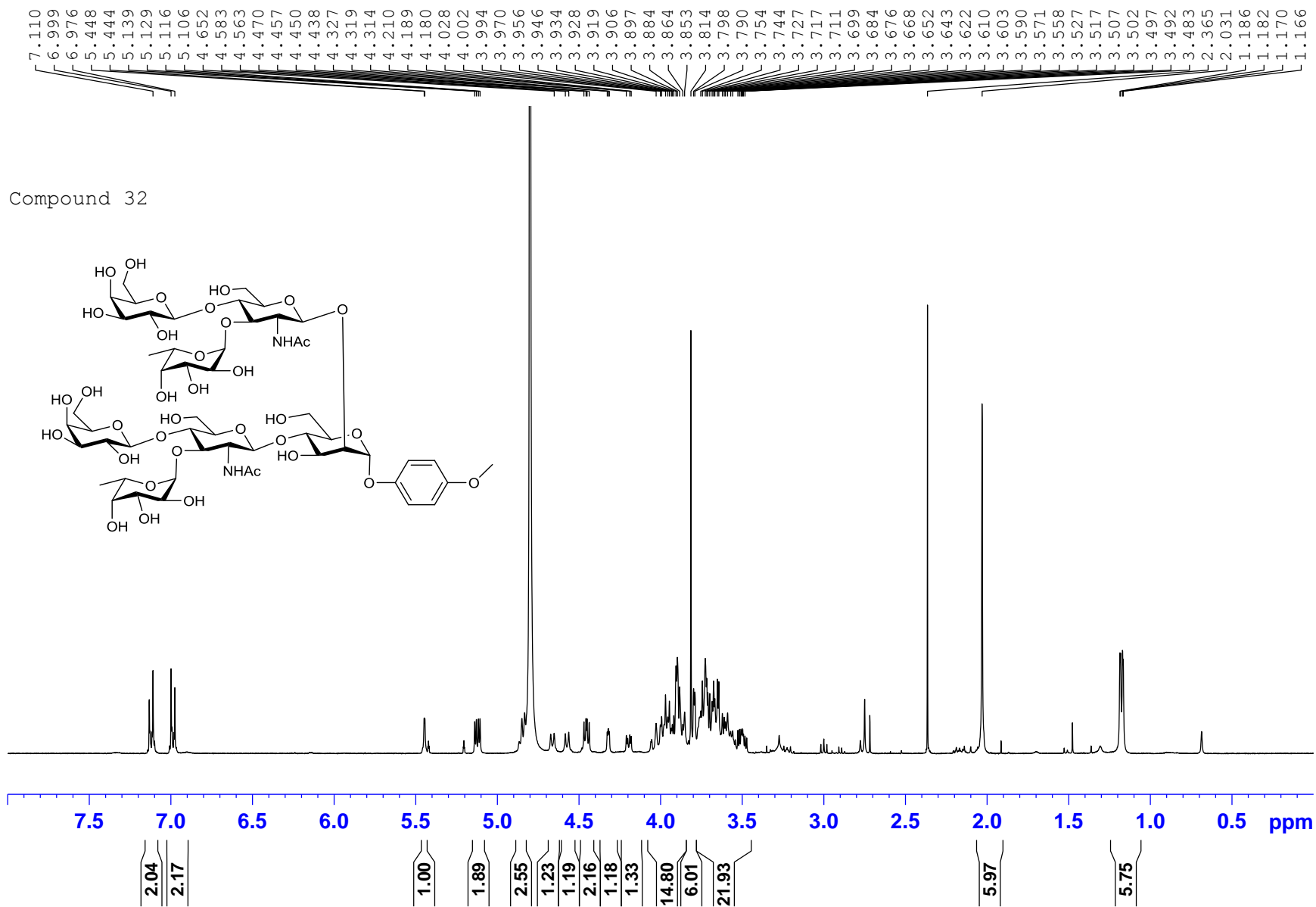


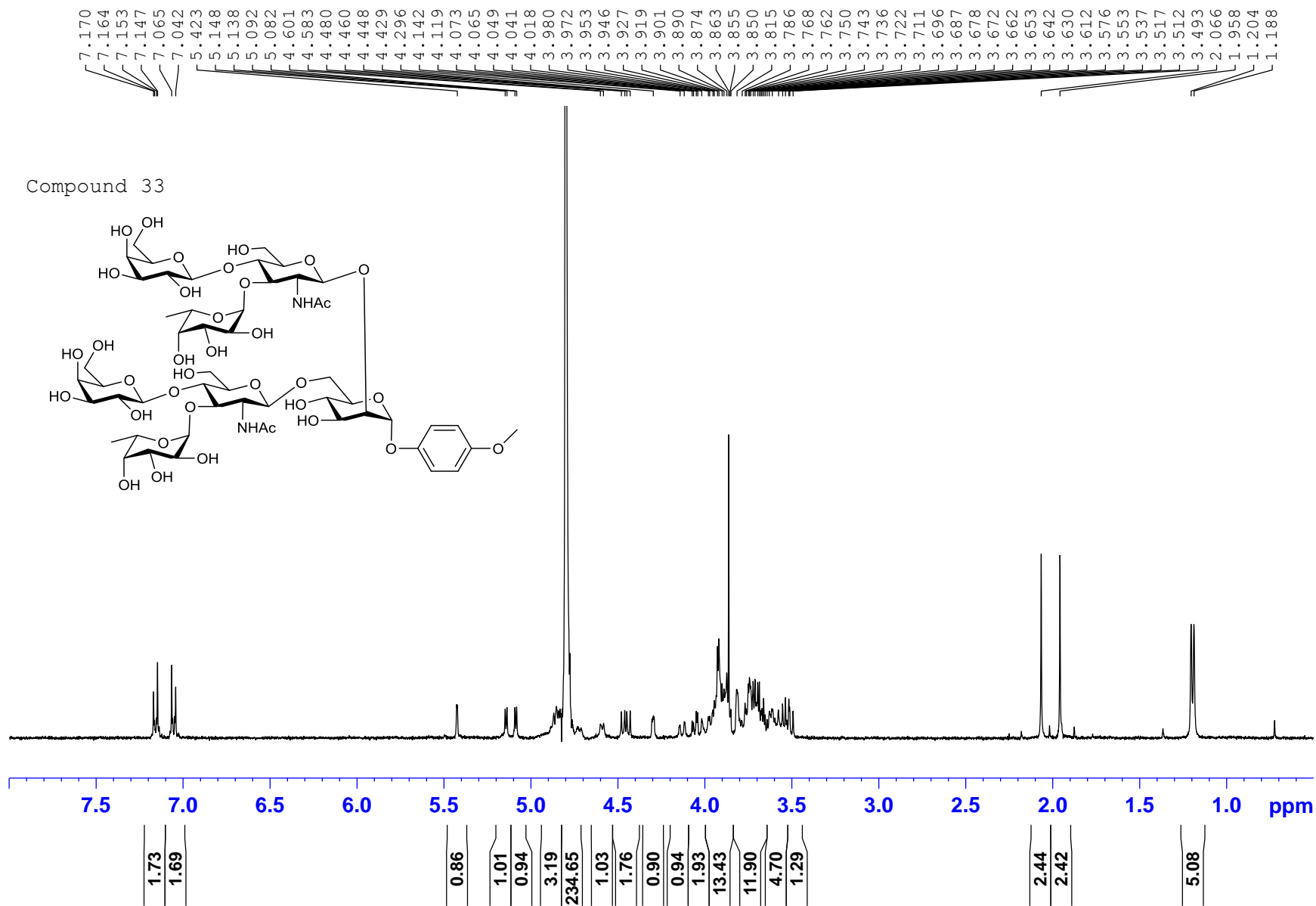


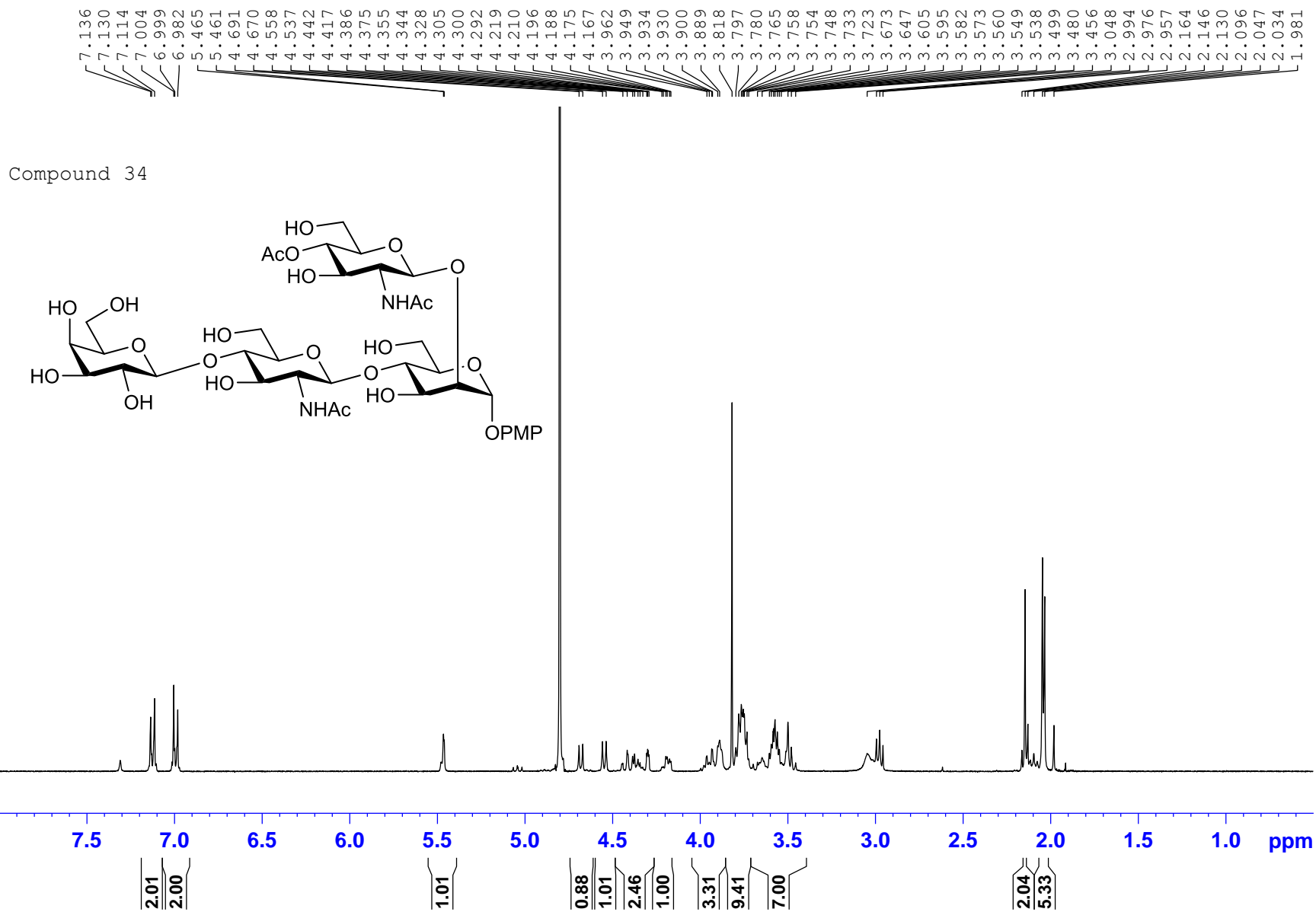


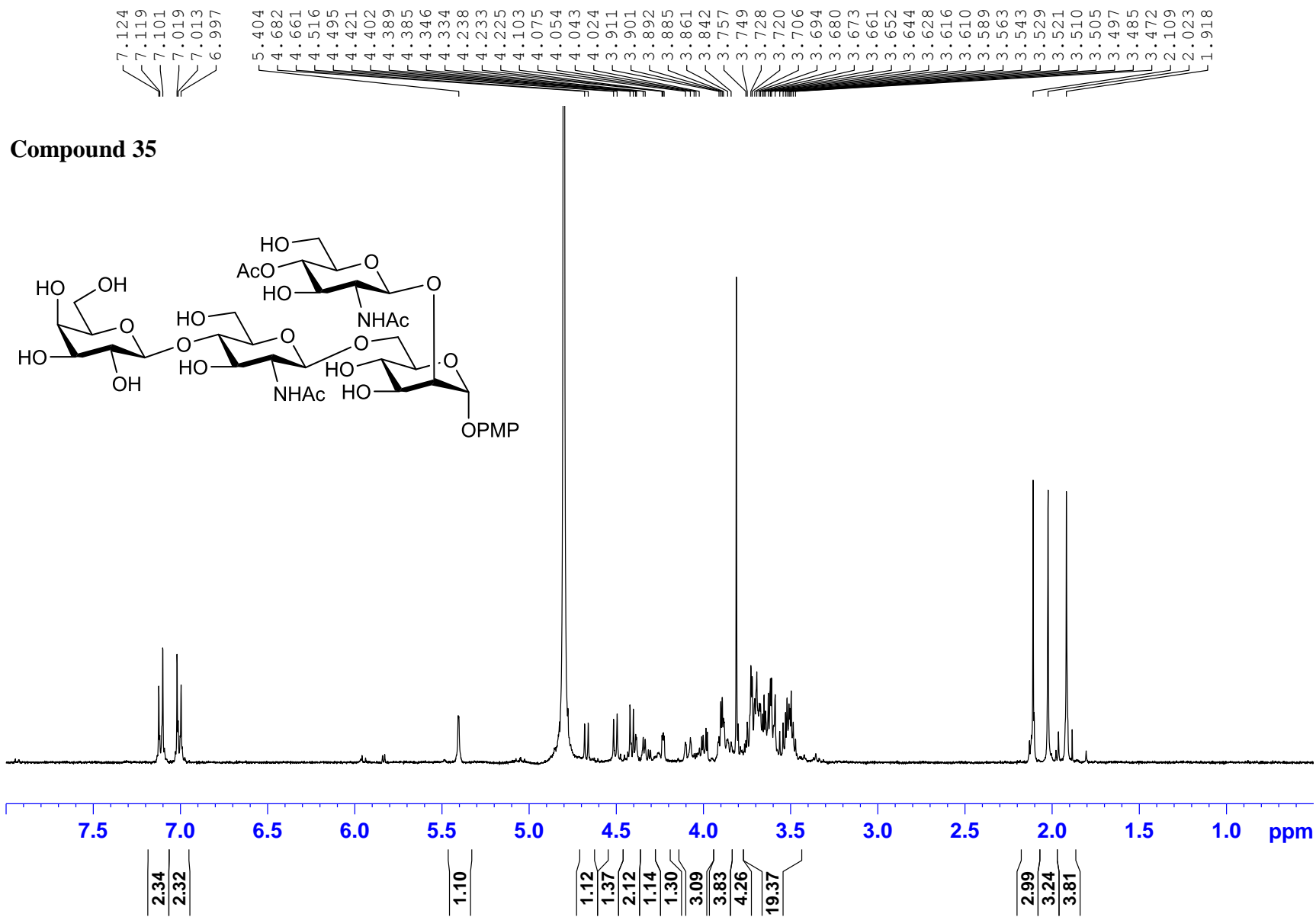


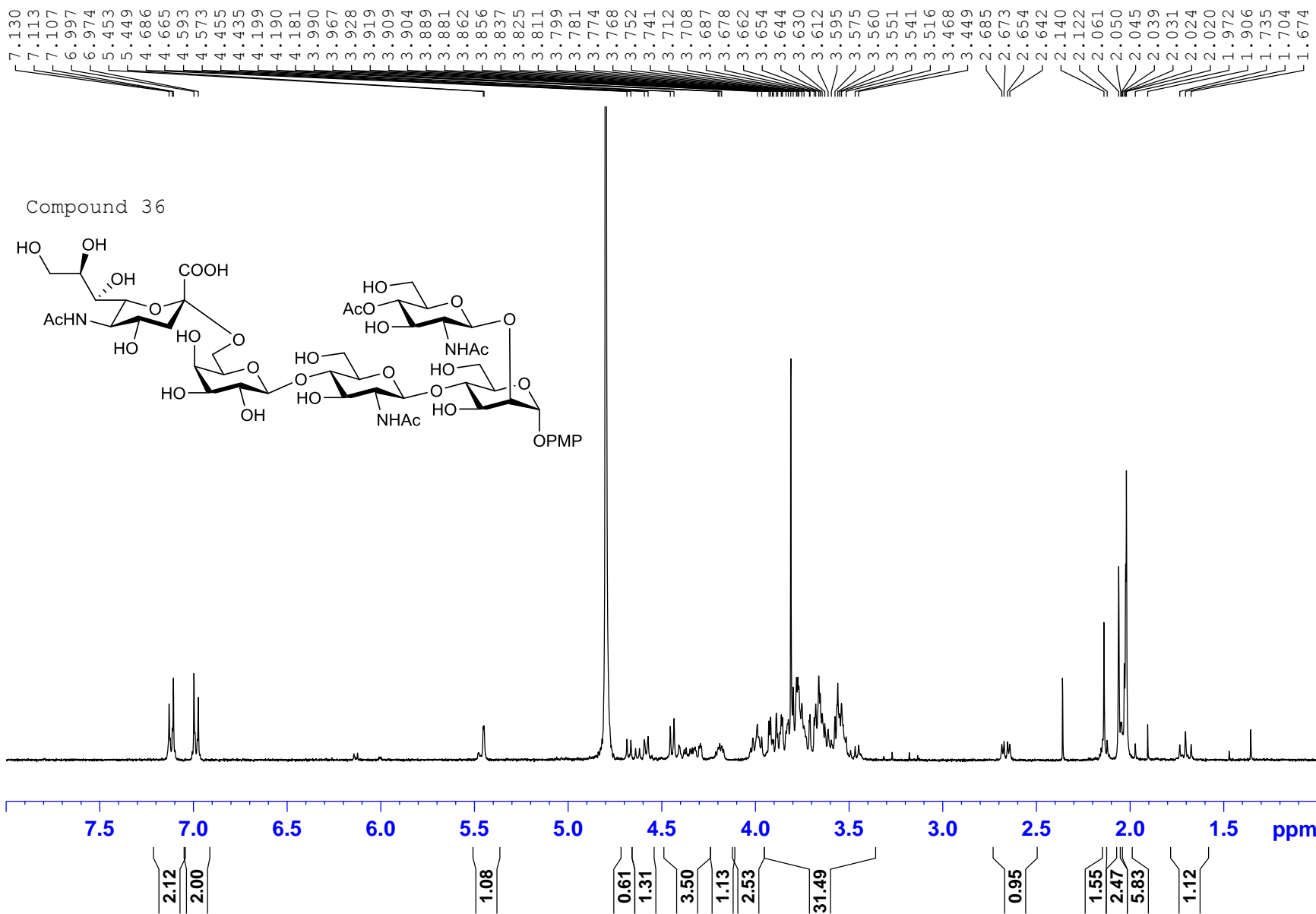


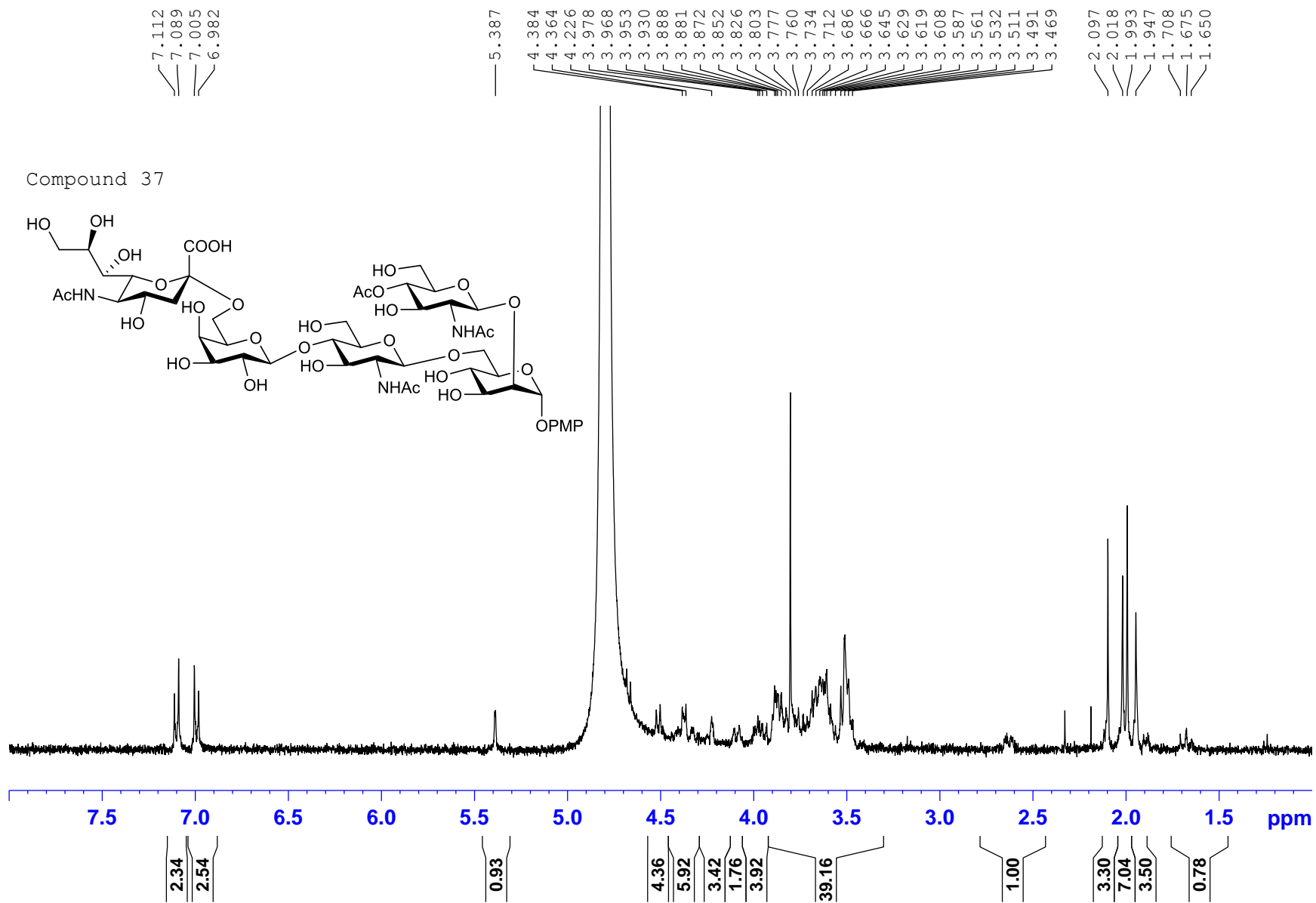


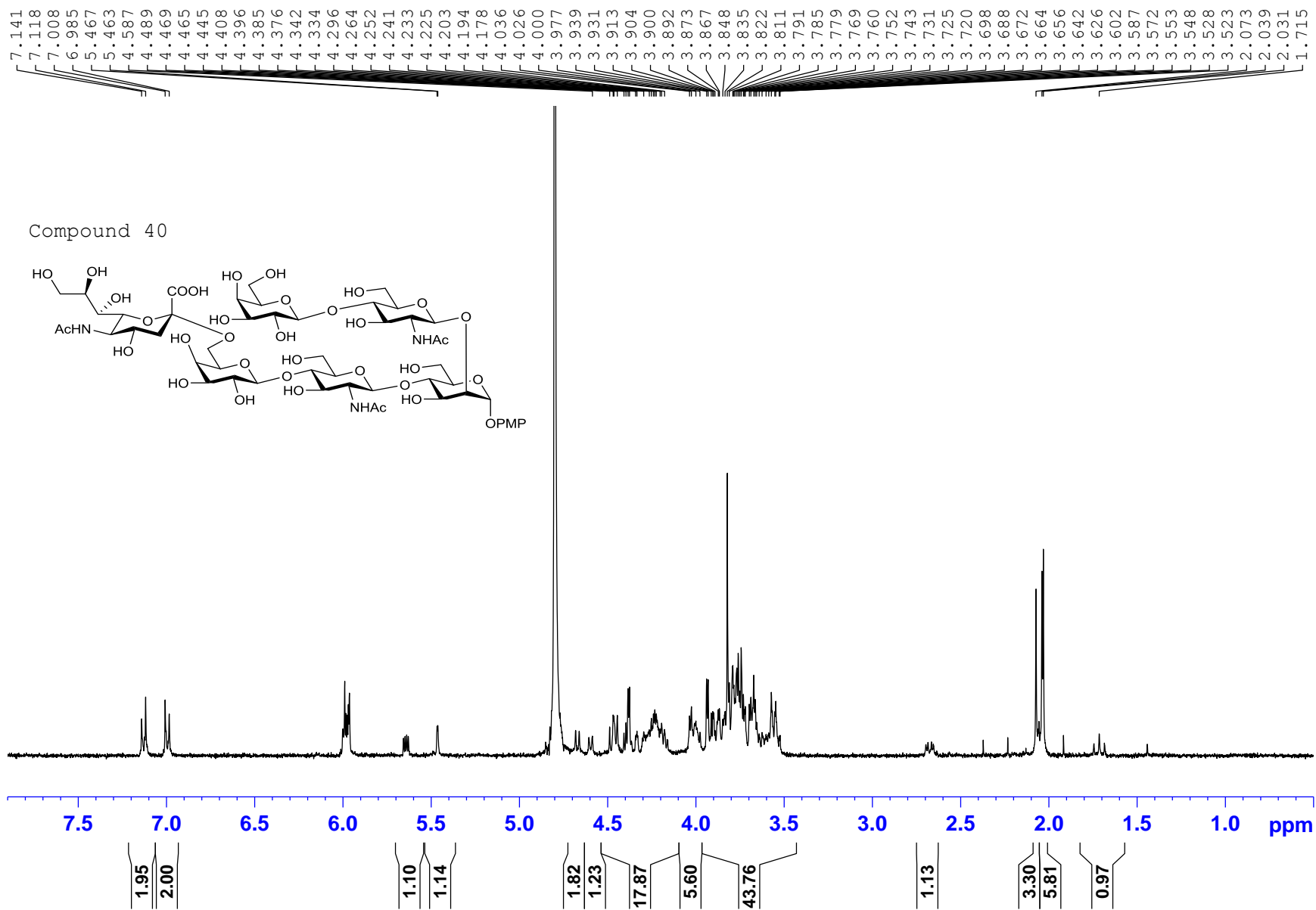


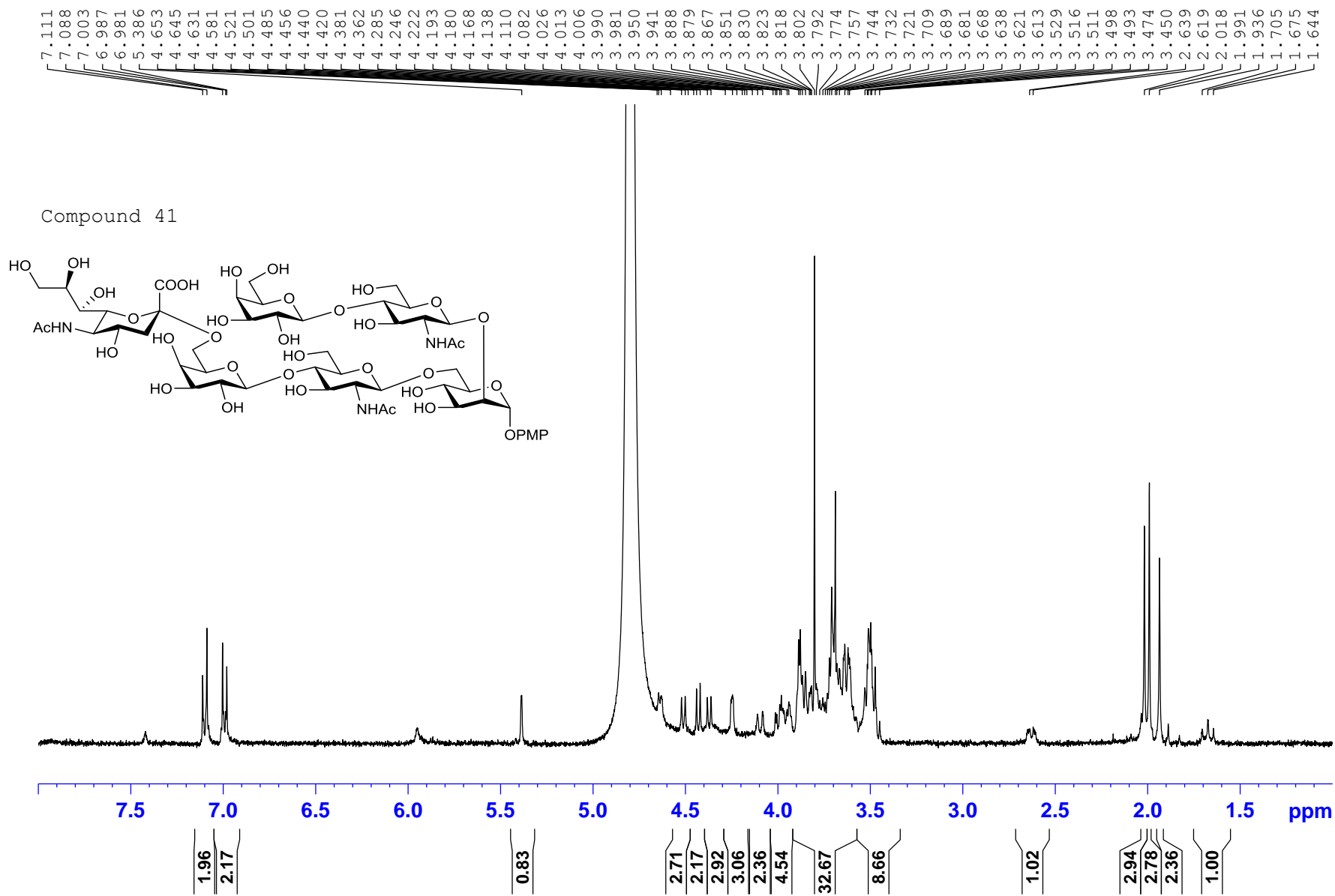


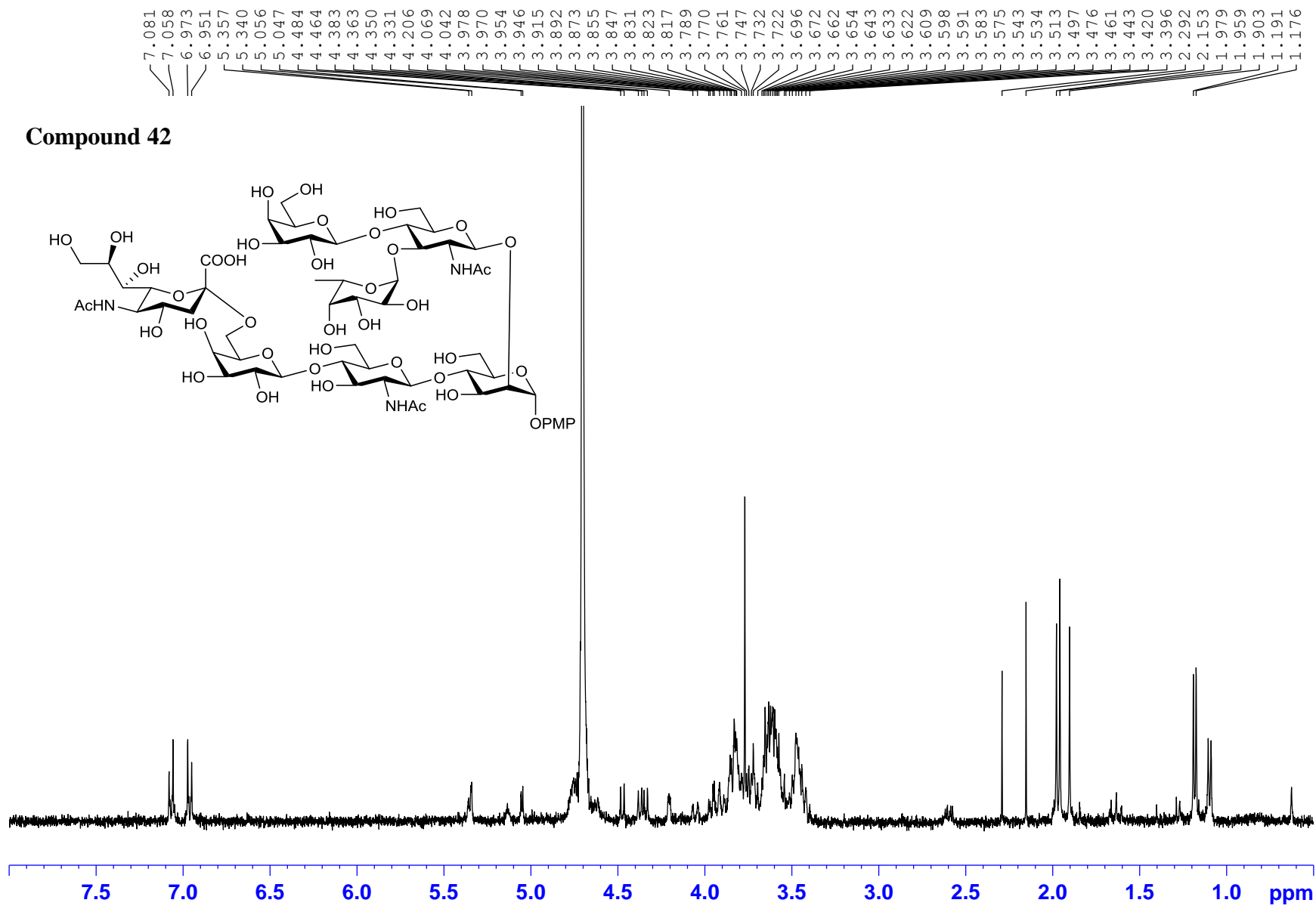




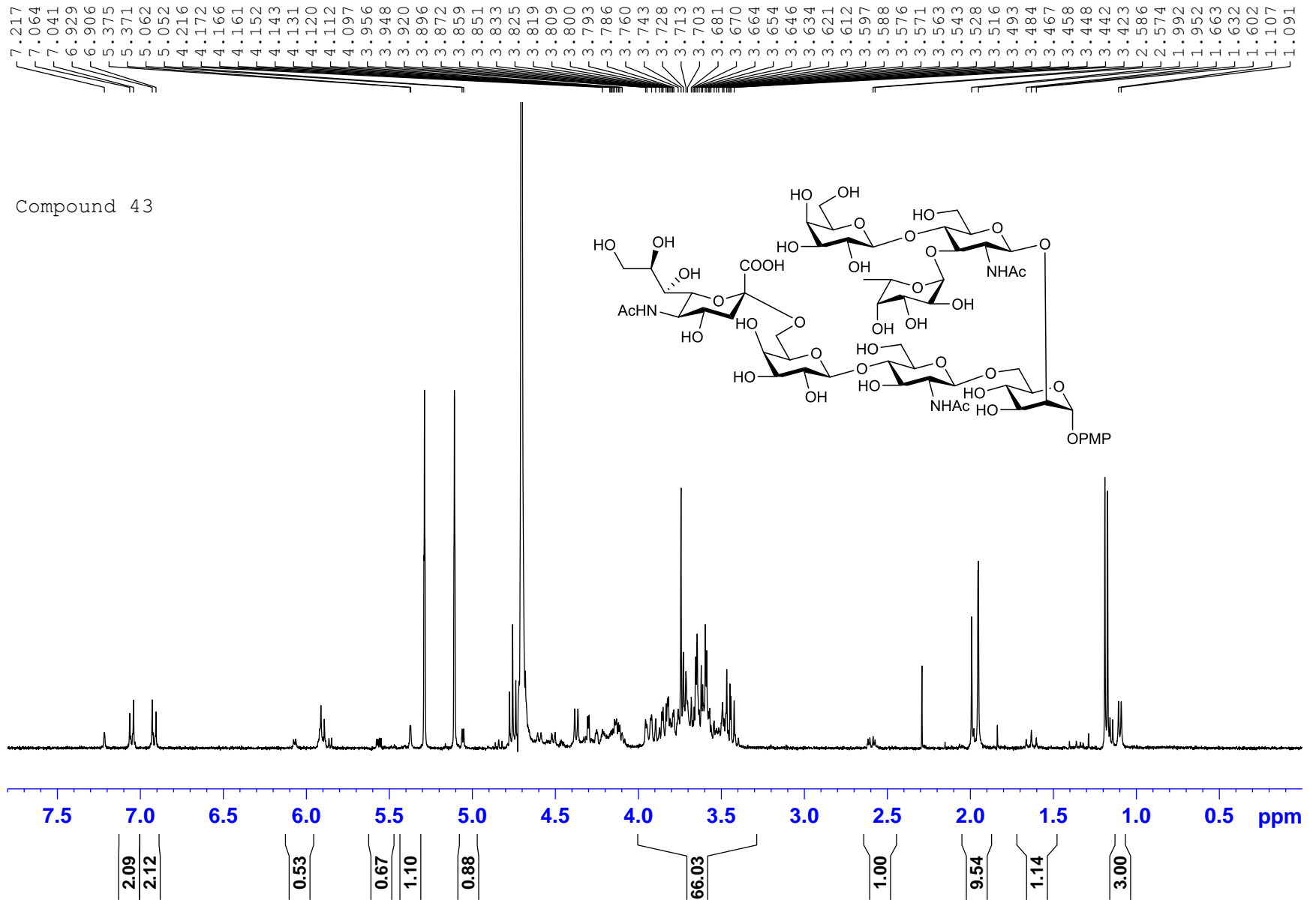


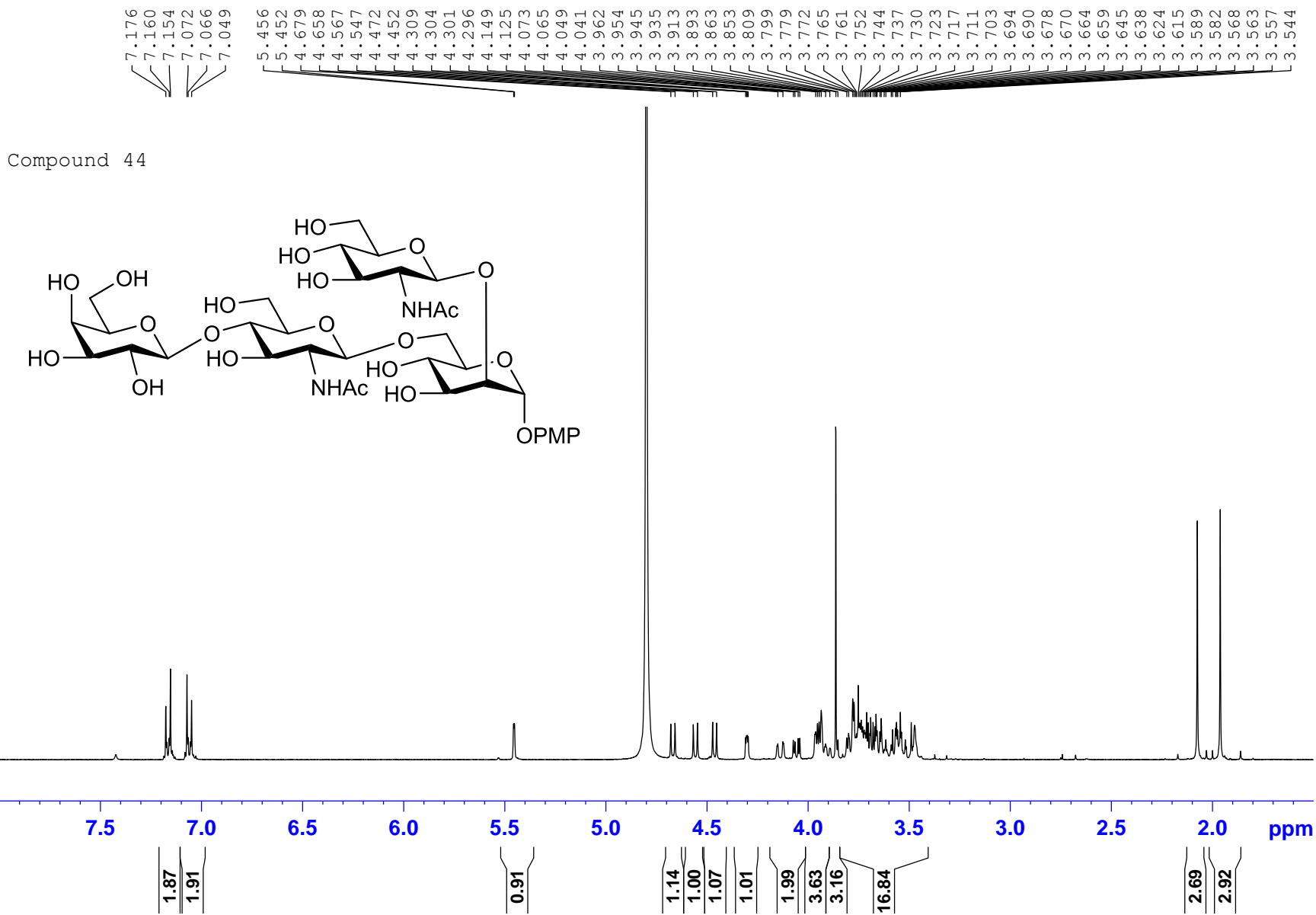


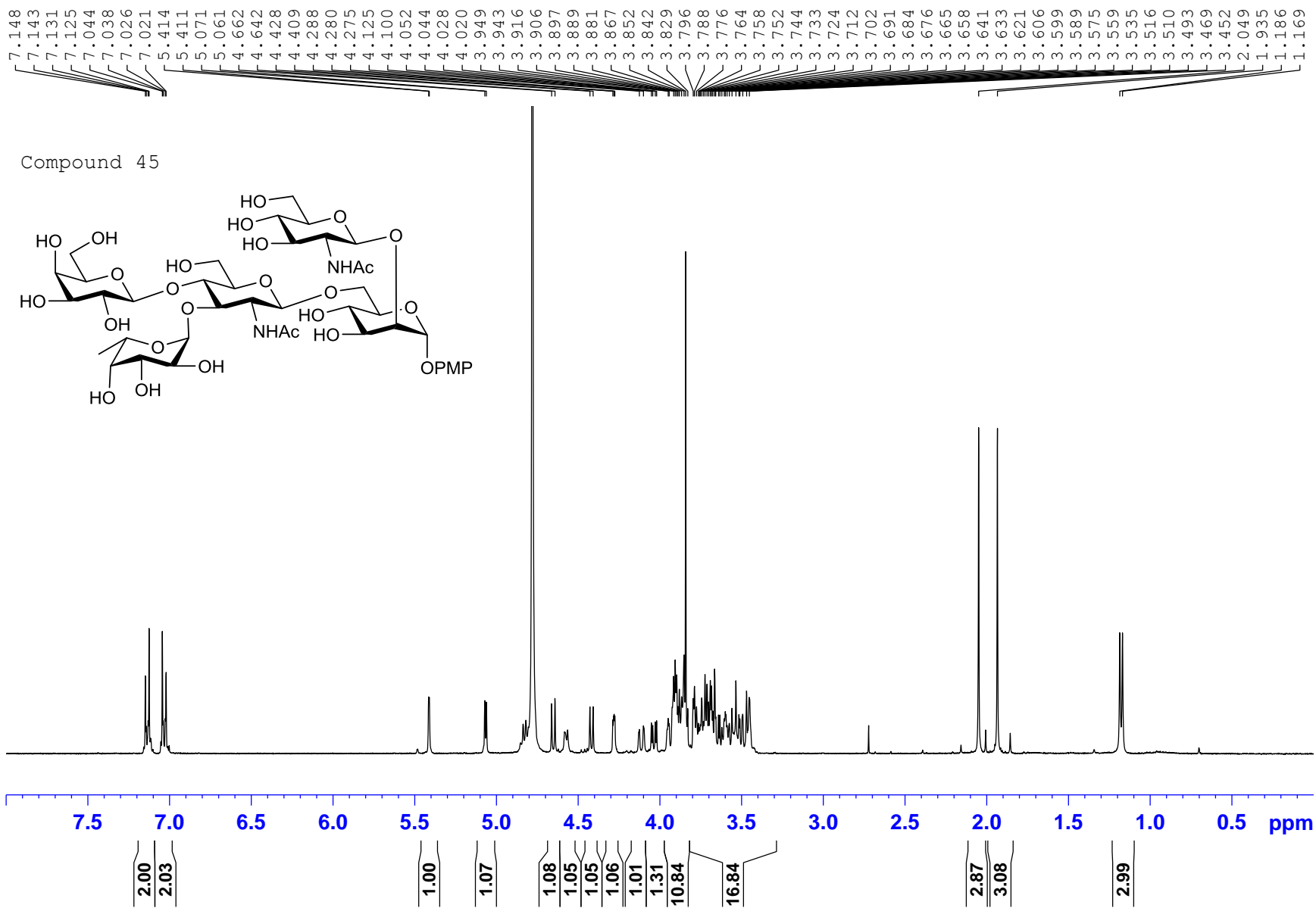


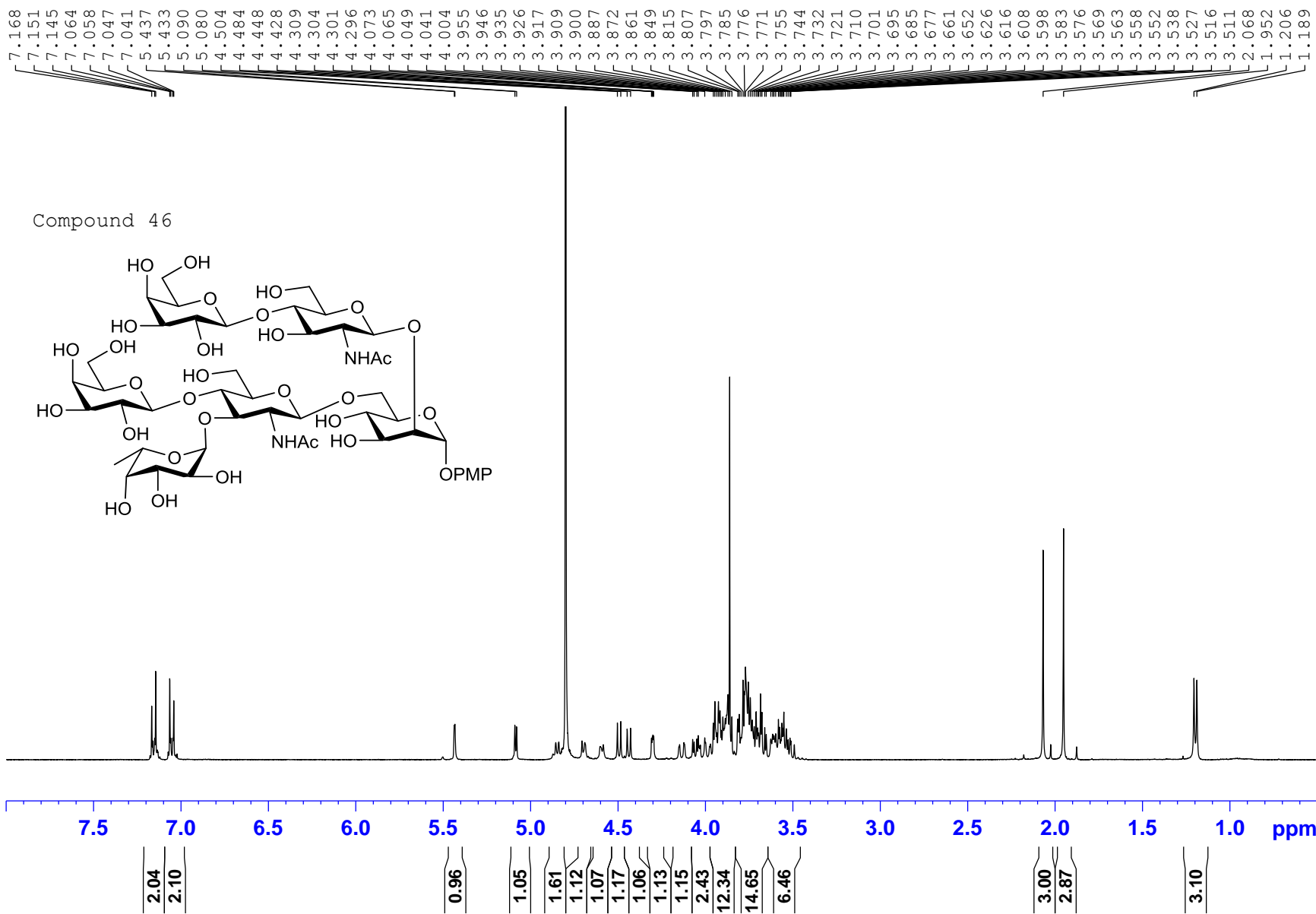


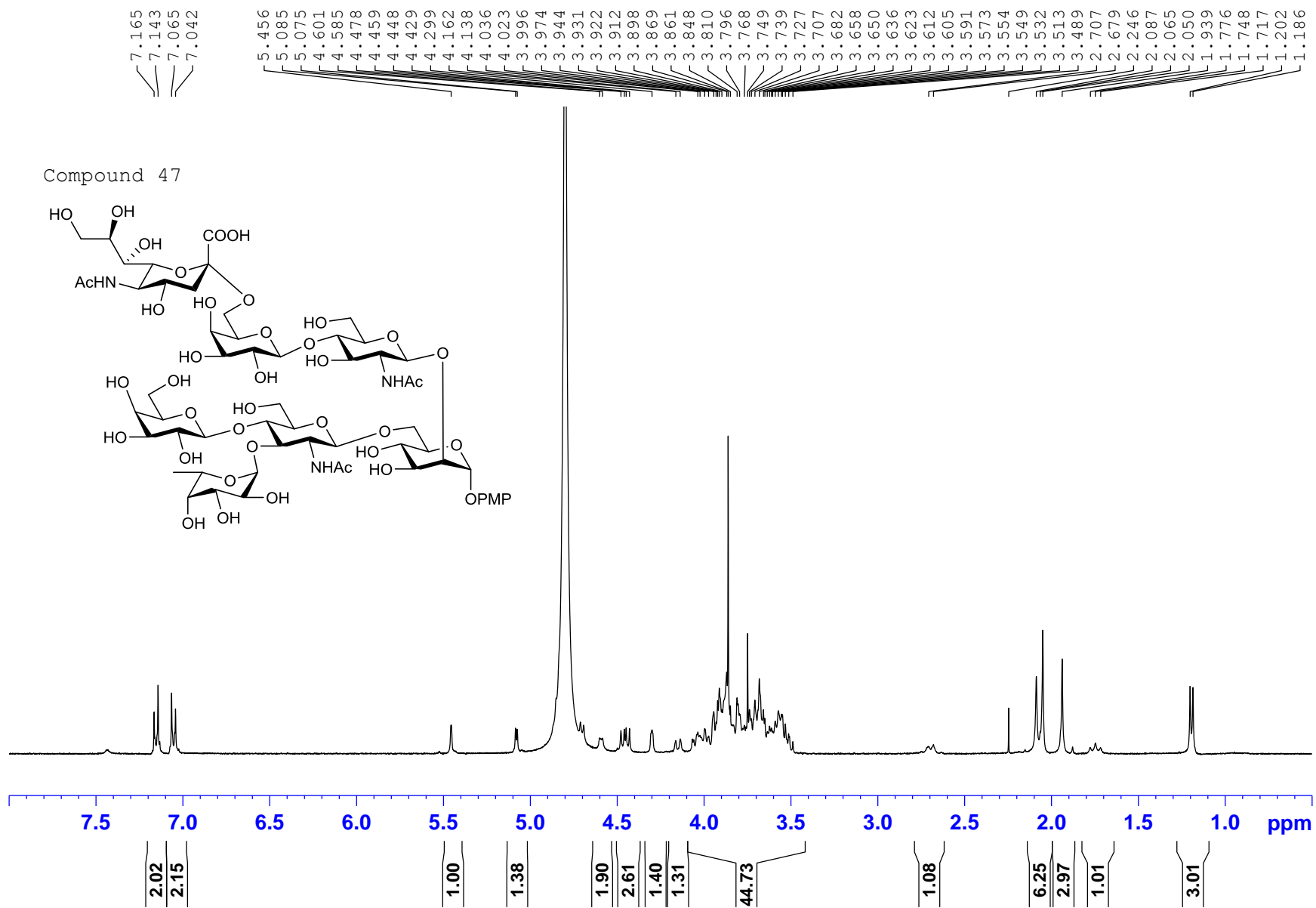
Compound 43

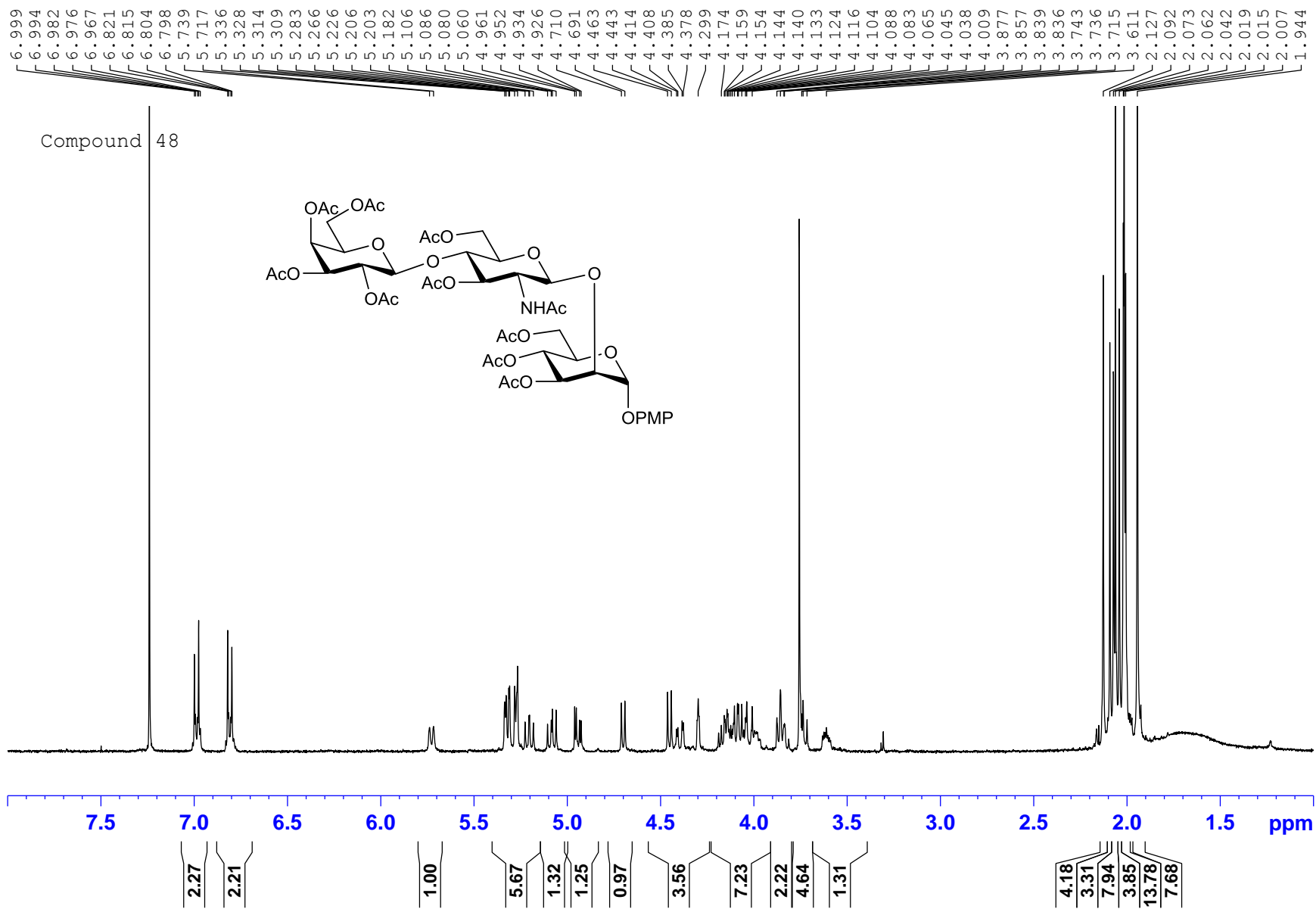


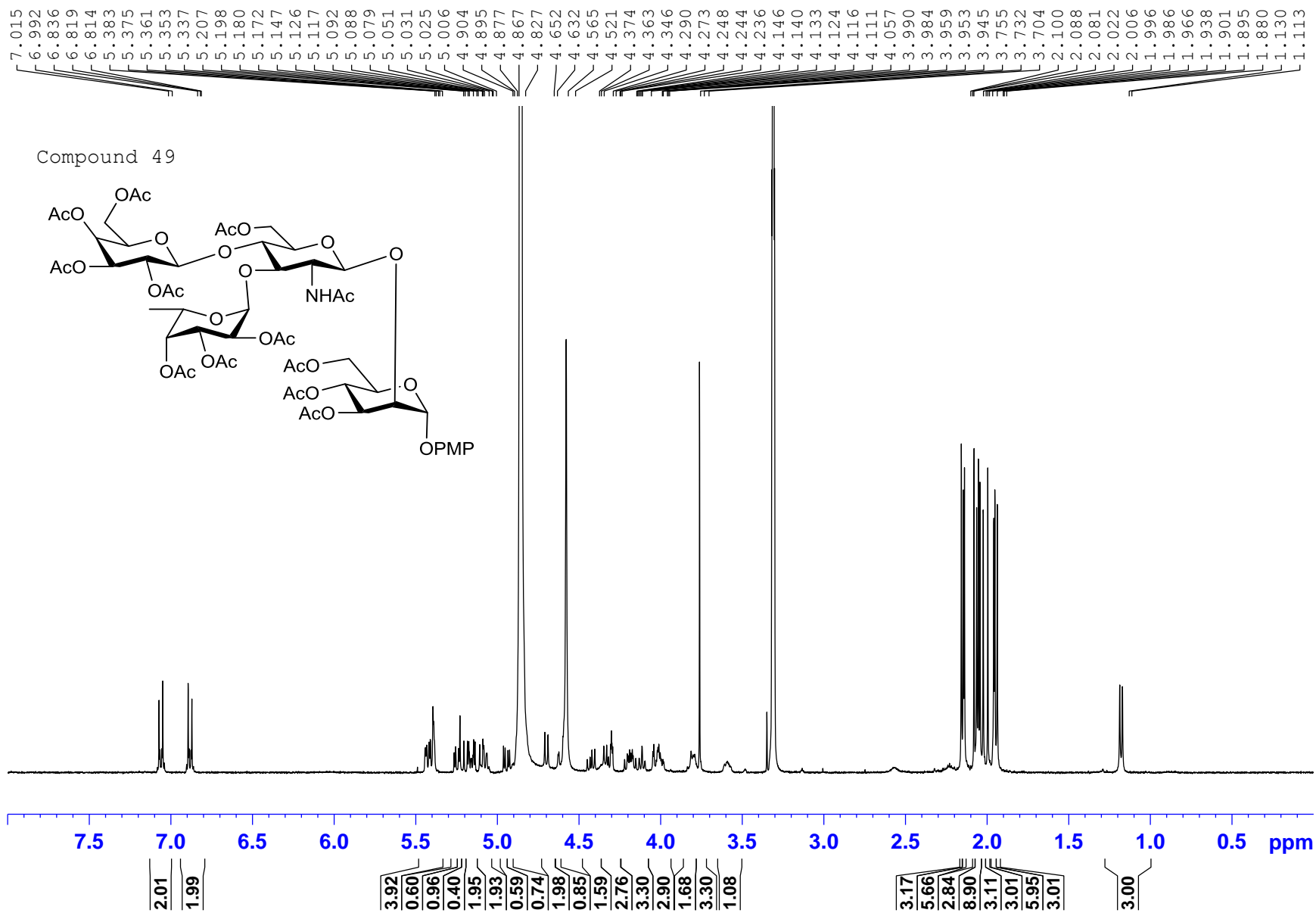






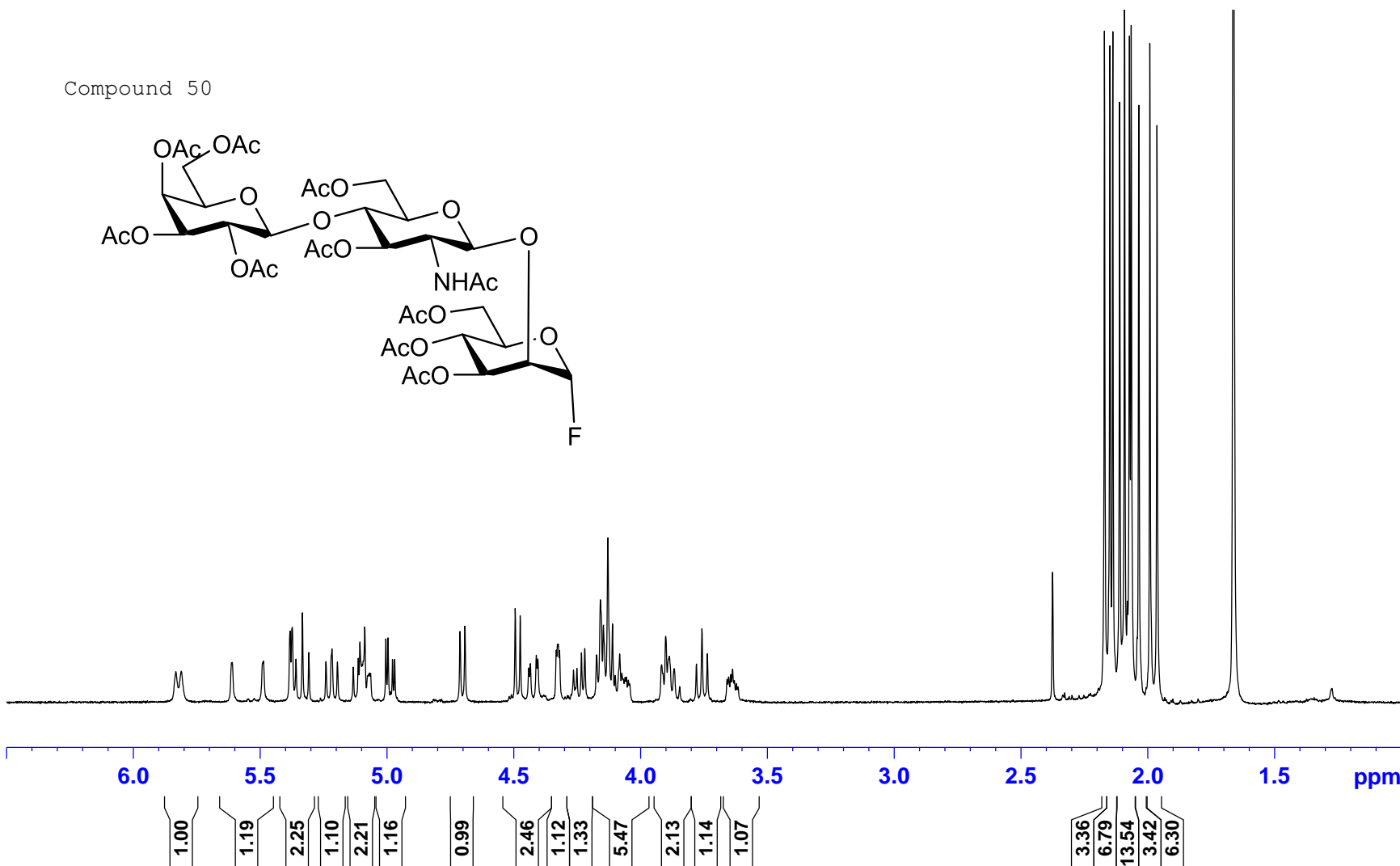
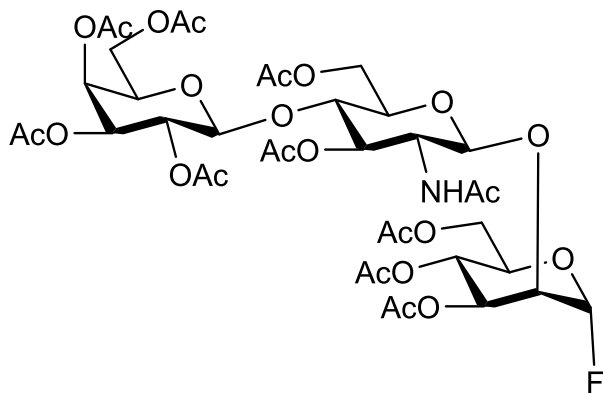


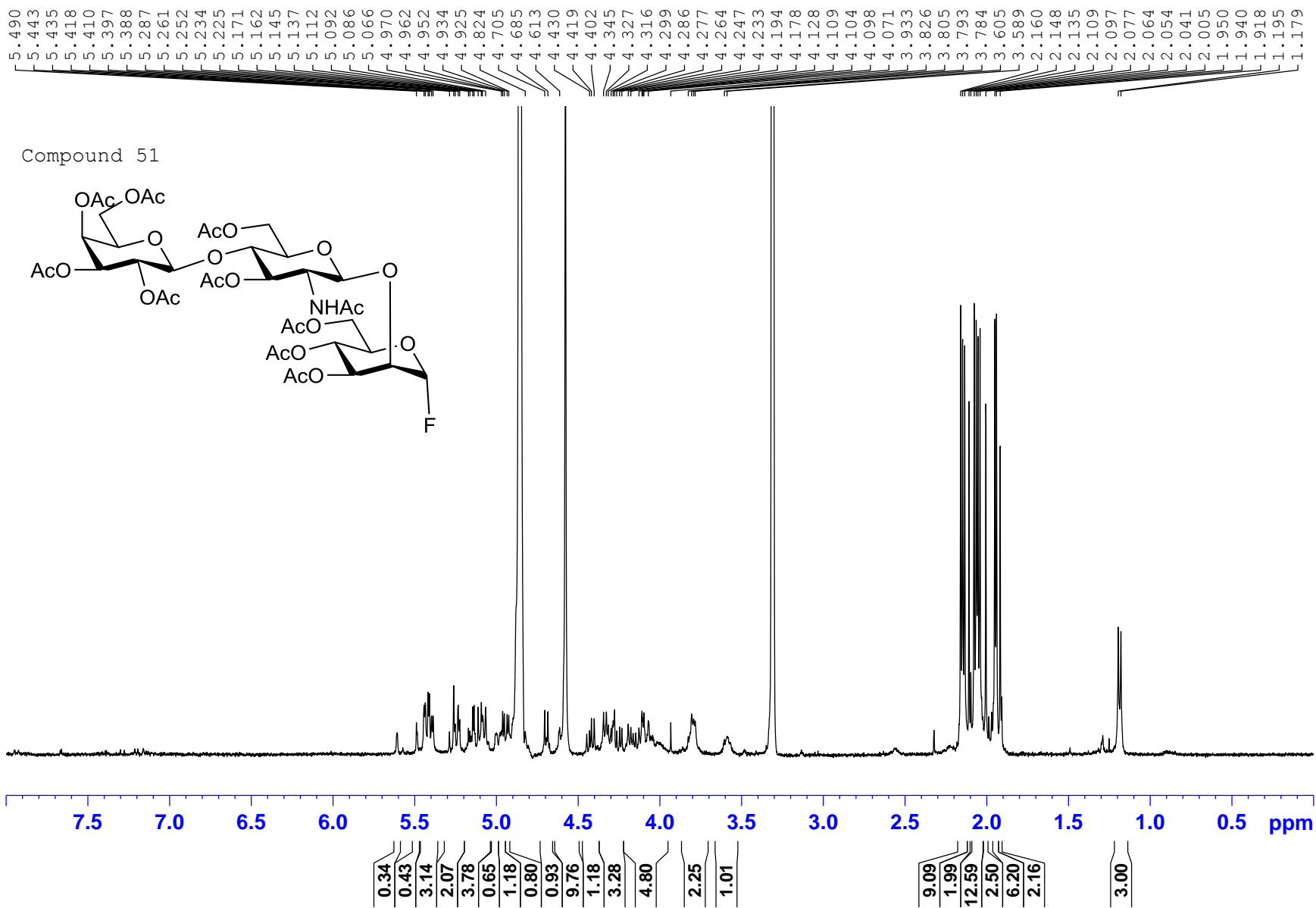


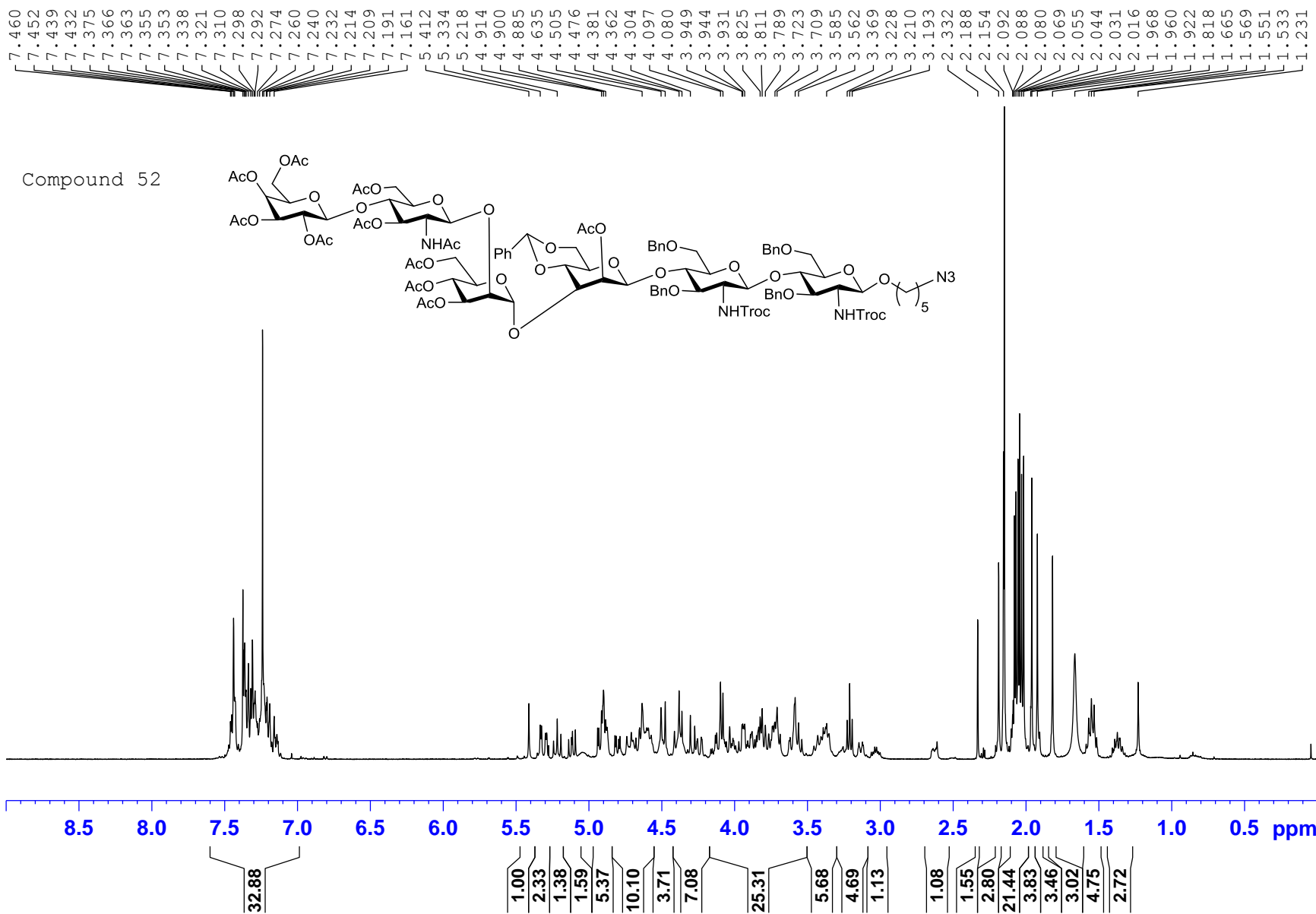


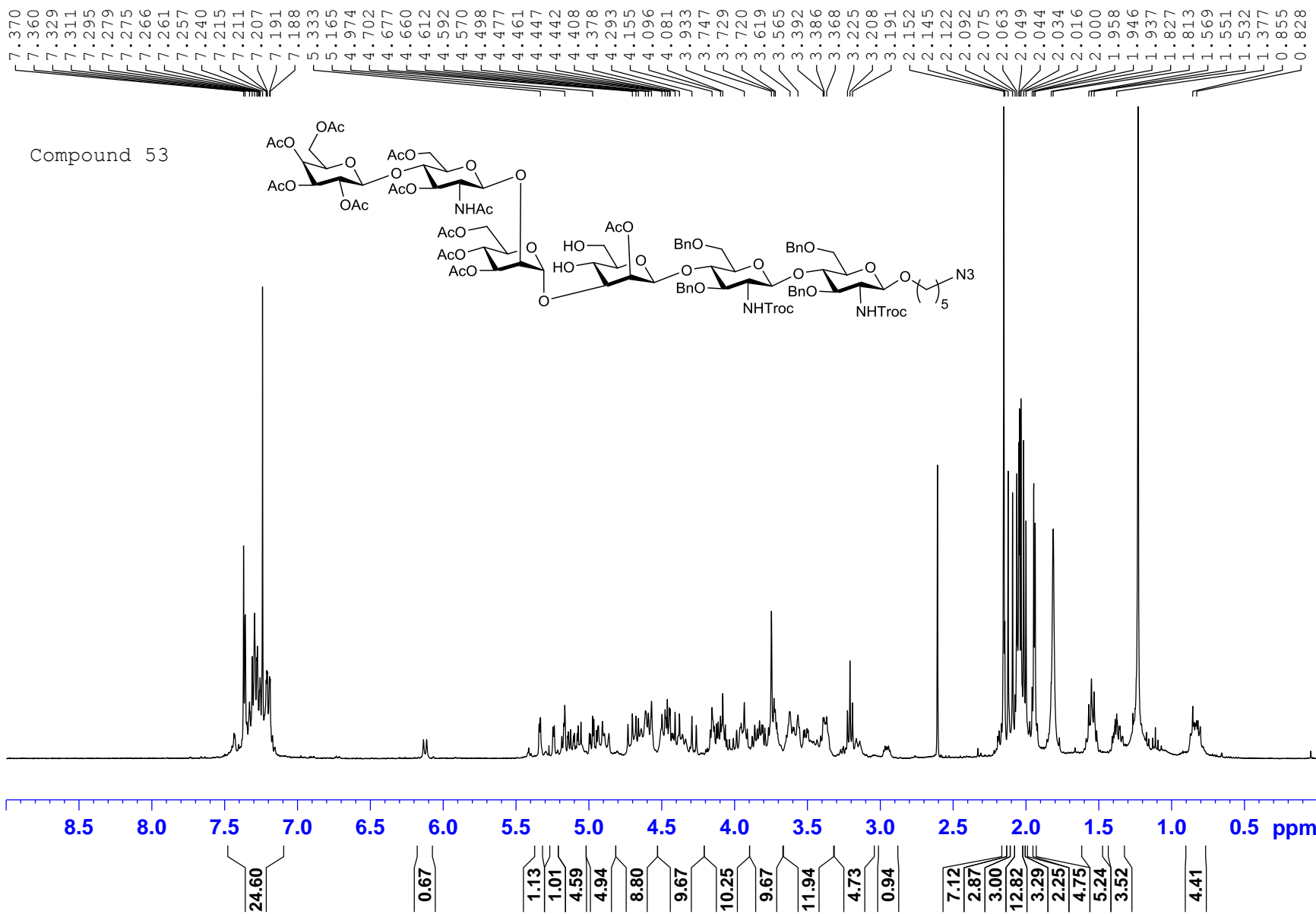
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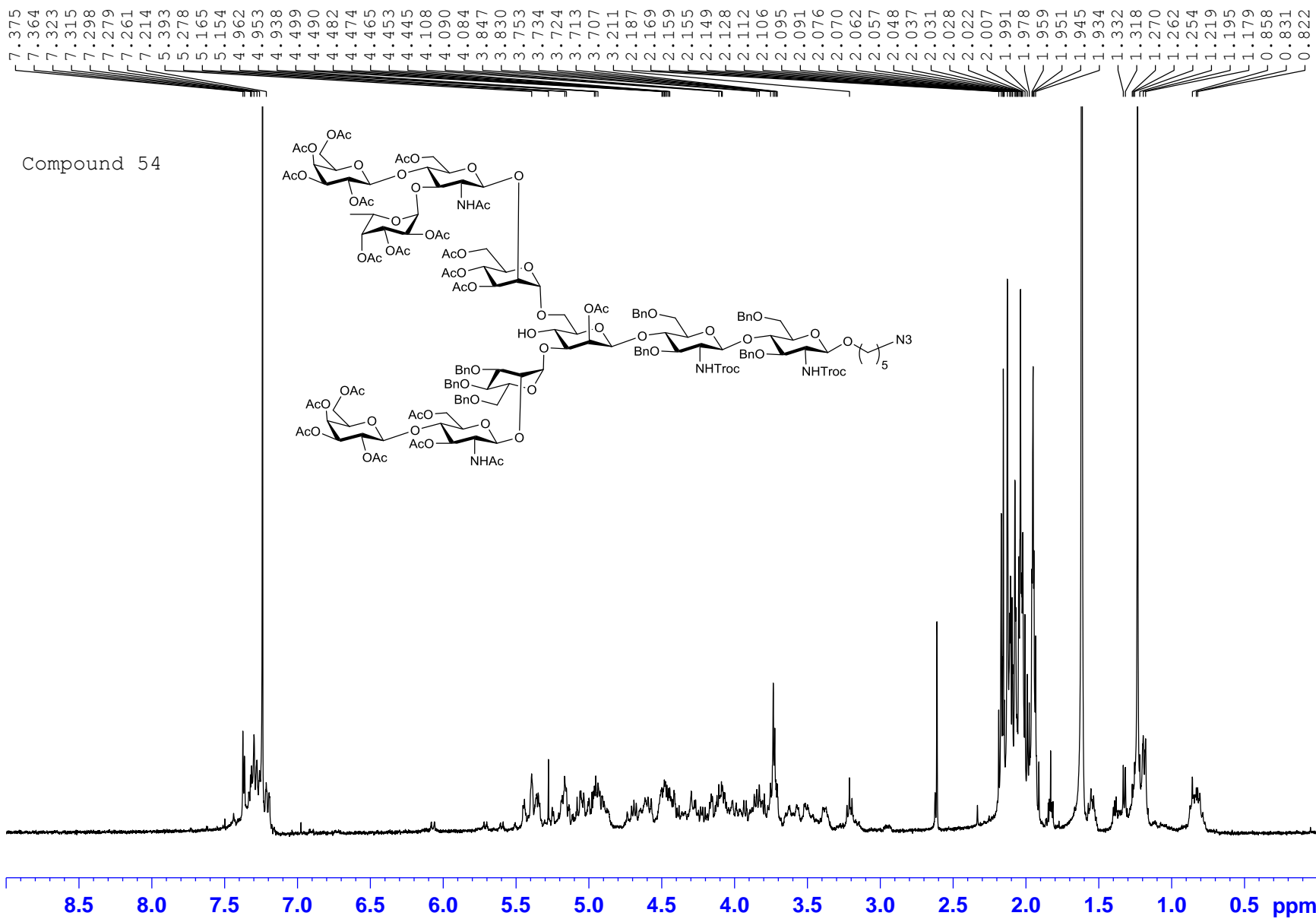
Compound 50

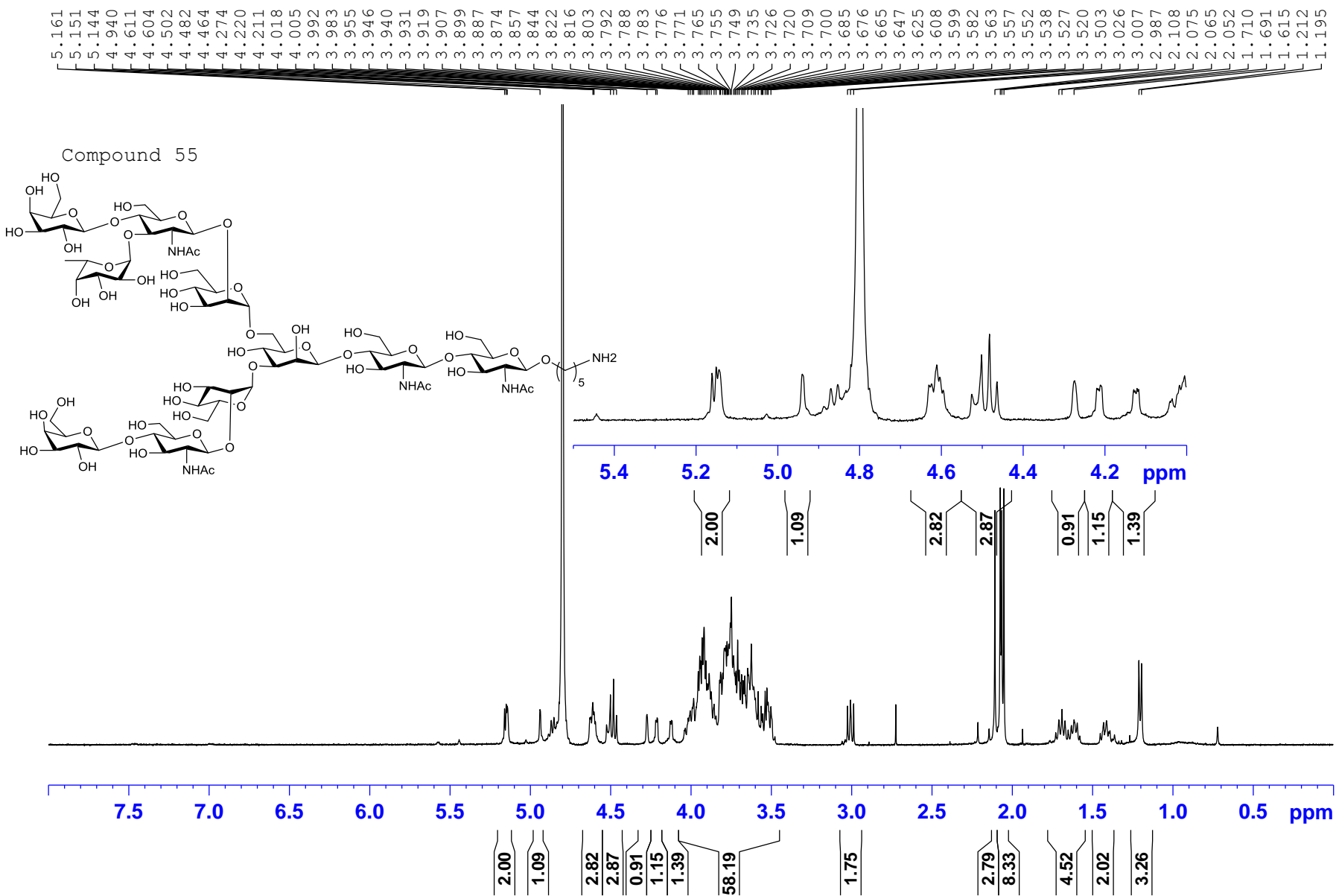












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22.39

Compound 55_carbon

