Supporting Information

Celastrol inhibits *Plasmodium falciparum* enoyl-acyl carrier protein reductase

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SI References

(1) Weisman JL, Liou AP, Shelat AA, Cohen FE, Kiplin Guy R, et al. (2006) Searching for New Antimalarial Therapeutics amongst Known Drugs. Chem Biol Drug Des 67: 409–416. doi:10.1111/j.1747-0285.2006.00391.

(2) Kapoor M, Jamal Dar M, Surolia A, Surolia N (2001) Kinetic Determinants of the Interaction of Enoyl-ACP Reductase from *Plasmodium falciparum* with Its Substrates and Inhibitors. Biochem Biophys Res Commun 289: 832–837. doi:10.1006/bbrc.2001.6061.

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Name	PDB ID	Score	Structure	
		(kcal/mol)		
Celastrol	20L4:A	-12.8	но н	
Aclarubicin	1UH5:B	-12.6		
Gambogic acid	1ZSN:B	-12.6		
Aclacinomycin Y	20L4:A	-12.3		
Salinomycin	1UH5:B	-11.9		
Lasalocid	202Y:D	-11.8		
Dequalinium	1VRW:B	-11.5	H_2N	

Emetine	200S:B	-11.5	
Tannic acid	1NNU:A	-11.4	
Aklavine hydrochloride	1V35:B	-11.2	
Nicergoline	1VRW:B	-11.2	$Br \underbrace{\downarrow}_{N} O \underbrace{\downarrow}_{$
Monensin sodium	2NQ8:B	-11.2	
Methylergonovine	202Y:C	-11.1	
Puromycin	2OL4:B	-11.1	$ \begin{array}{c} $
Hydroxyprogesterone	200S:B	-10.7	
Mefloquine	1VRW:A	-10.6	

Tetrandrine	20L4:A	-10.6	
			N N N N N N N N N N N N N N N N N N N
Methotrexate	20L4:B	-10.6	O O OH
			$ \begin{array}{c c} & & & \\ & & & \\ & & $
Homidium	20L4:B	-10.4	H ₂ N - NH ₂
lvermectin	20L4:A	-10.2	
Deoxygeduniun	1VRW:A	-10.1	
Thioridazino	2000.0	10.1	
Thioridazine	2000.0	-10.1	S-
			ŚŚ
Bebeerine	1NNU:A	-10.0	
Selamectin	2NQ8:B	-10.0	

Amodiaquine	200S:B	-10.0		
Dihydroartemisinin	1ZW1:A	-9.9	ООО	
Quinine	2OL4:B	-9.9		
Cinchonine	2OL4:B	-9.9		
Rutilantinone	1V35:A	-9.8		
Coralyne	1VRW:A	-9.8		
Lycorine	1VRW:B	-9.8		
Methylbenzethonium	1VRW:B	-9.8		
Benzethonium	202Y:C	-9.8		
3,7-Dihydroxyflavone	200S:B	-9.8	HO O O HO	
Cabergoline	200S:B	-9.8		

Quinidine	1VRW:A	-9.7		
Cinchonidine	2OL4:B	-9.7		
Hydroquinidine	2OL4:B	-9.7	OH O O O	
Alexidine hydrochloride	20P0:B	-9.7	$\begin{array}{c c} & & & & \\ & & & & \\ & & & \\ & & & \\ & & & & \\ & & & & \\ & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & &$	
Gentian violet	1ZXB:B	-9.5		
Pergolide	1VRW:B	-9.4	S NH	
Rhodomyrtoxin B	1ZXB:A	-9.4	НО ОН ОН ОН ОН ОН ОН ОН ОН ОРИСКИИ СТАНИИ СТ	
Pararosaniline	1UH5:A	-9.3	NH ₂ +H ₂ N NH ₂	
Cycloheximide	1VRW:B	-9.3		
Mitomycin	1ZXB:A	-9.3		
Hydroxychloroquine	200S:B	-9.3		

Perhexiline	20P1:B	-9.1	
Quinacrine	1UH5:A	-9.0	
Chlorprothixene	1VRW:A	-9.0	
Cyclosporin A	20L4:A	-9.0	
Tilorone	2OL4:B	-9.0	
Propafenone	1UH5:B	-8.9	
Mitoxantrone	1VRW:B	-8.9	
Angloensin	202Y:C	-8.8	но
Chloroquine	200S:B	-8.8	
Pentamidine	200S:B	-8.8	H ₂ N H NH H ₂ N NH NH ₂

Acriflavinium	1ZW1:A	-8.7	H ₂ N ⁺ NH ₂	
Hycanthone	2OP0:B	-8.7		
Pyrimethamine	1NNU:B	-8.4	$H_2 N \xrightarrow{N}_{N} \xrightarrow{NH_2}_{N} Cl$	
Dactinomycin	2OL4:A	-8.4	$\begin{array}{ } \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ 0 \\ $	
Ciclopirox	1NNU:B	-8.3	O OH	
Anisomycin	200S:B	-8.2		
Sulcotidil	1VRW:B	-7.6	→ s → H → H → → → → → → → → → → → → → →	
Benzalkonium	2OL4:B	-7.3		
Amphotericin B	20P1:A	-7.1	$H_2N \xrightarrow{OH} H_2N OH$	
Cetylpyridinium	202Y:C	-6.9		

Acivicin	1ZW1:B	-6.5	
Cetrimonium	200S:B	-6.3	

Table S1. The list of small molecules from the MicroSource Spectrum and Killer Collection library [1] that were docked into *Pf*ENR. The scores were calculated by AutoDock Vina. The PDB ID is given as ID:Chain.

	NADH	crotonyl-CoA	Reference
K _m (mM)	0.24 <u>+</u> 0.04	0.17 <u>+</u> 0.06	current study
k _{cat} (sec⁻¹)	0.9 <u>+</u> 0.1	1.0 <u>+</u> 0.2	current study
k _{cat} /K _m (mM ⁻¹ sec ⁻¹)	3.8 <u>+</u> 1.0	6.0 <u>+</u> 1.1	current study
K _m (mM)	0.03 <u>+</u> 0.004	0.17 <u>+</u> 0.015	Ref 1
k _{cat} (sec ⁻¹)	NR	1.62 <u>+</u> 0.06	Ref 1
k _{cat} /K _m (mM ⁻¹ sec ⁻¹)	NR	9.8 <u>+</u> 0.96	Ref 1

Table S2. Kinetic parameters calculated for *Pf*ENR. The 100 μ L reaction volume contained a final concentration of 0.25 μ M ENR, 20 mM Tris/HCl buffer (pH 7.4), with 150 mM NaCl 100 μ M crotonyl-CoA, and 100 μ M NADH (Sigma). The enzyme was preincubated in 20 mM Tris/HCl buffer (pH 7.4), 150 mM NaCl and crotonyl-CoA, and initiated with NADH to reach its final concentration. The K_{m, crotonyl-CoA} was determined by varying the concentration of crotonyl-CoA while keeping the NADH concentration fixed at 100 μ M. Conversely, the K_{m, NADH} was determined by varying the concentration of NADH while keeping the crotonyl-CoA concentration fixed at 100 μ M.



Figure S1. 12% SDS-PAGE showing expression and purification of *P. falciparum* ENR in pET28a plasmid encoding N-terminus 6xHis-tag in *E.coli* BL21 cells. Lane (1) Expression of *Pf*ENR insoluble fraction, (2) soluble fraction, (3) flow-through of Ni-NTA, and (4) buffer wash of Ni-NTA. Lanes (5-11) step gradient of imidazole elution of *Pf*ENR.



Figure S2. The Michaelis-Menten plots for *P. falciparum* ENR measuring the consumption of NADH at 340 nm $(\varepsilon_{NADH} = 6.22 \text{ mM}^{-1} \text{ cm}^{-1})$ for varying concentrations of crotonyl-CoA (a) and NADH (b) at 27 °C. The 100 µL reaction volume contained a final concentration of 0.25 µM ENR, 20 mM Tris/HCl buffer (pH 7.4) with 150 mM NaCl, 100 µM crotonyl-CoA, and 100 µM NADH (Sigma). The enzyme was preincubated in 20 mM Tris/HCl buffer (pH 7.4), 150 mM NaCl, and crotonyl-CoA, and initiated with NADH to reach its final concentration. a) The K_{m, crotonyl-CoA} was determined by varying the concentration of crotonyl-CoA (10-300 µM) while keeping the NADH concentration fixed at 100 µM. b) The K_{m, NADH} value was revalidated by titrating NADH (20-500 µM) and keeping the crotonyl-CoA concentration constant at 100 µM. Data was collected in triplicate, and the individual values were within 10% error.





Figure S3. IC₅₀ binding curves for triclosan (TCL). The final 100 μ L reaction volume contained 50 μ M NAD⁺, 20 mM Tris/HCl buffer (pH 7.4), 150 mM NaCl, 200 μ M crotonyl-CoA, 100 μ M NADH, and 0.05 μ M ENR. The ENR cocktail was preincubated with a final concentration of 50 μ M NAD⁺ (Sigma), 20 mM Tris/HCl buffer (pH 7.4), 150 mM NaCl, and varying amounts of triclosan (300 nM to 0.3 nM) (5% v/v DMSO) at 25 °C for 45 minutes. IC₅₀ for triclosan (•) without NAD⁺ preincubation (•) with 50 μ M NAD⁺ preincubation. Data was collected in triplicate, and the individual values were within 10% error.







Figure S5. LC-MS Trace of compound 1 (NCI).



Figure S6. LC-MS Trace of compound 1 (Sigma-Aldrich).

Figure S7.



Figure S7. a) $K_{m, app}$ plotted as a function of varying celastrol concentration. $K_{m, app}$ is independent of varying celastrol concentration. b) Scatter plot of $V_{max, app}$ as a function of varying celastrol concentration. The $K_{i, app}$ (5.3 μ M) was calculated from the slope of the line ($r^2 = 0.89$). These experiments were repeated in triplicate.

Figure S8.



Figure S8. IC₅₀ binding curve for celastrol (compound 1) in the presence of 0.01% Triton-X. The final 100 μ L reaction volume contained 50 μ M NAD⁺, 20 mM Tris/HCl buffer (pH 7.4), 150 mM NaCl, 0.01% Triton-X, 200 μ M crotonyl-CoA, 100 μ M NADH, and 0.05 μ M ENR. ENR was preincubated at 25 °C for 45 minutes with a final concentration of 50 μ M NAD⁺, 20 mM Tris/HCl buffer (pH 7.4), 150 mM NaCl, and varying concentrations (0.9-50 μ M) of **1**. Data was collected in duplicate, and the individual values were within 10% error.





Figure S9. A comparison between the activity of *Pf*ENR a) with and without b) rapid dilution. a) The final 100 μ L reaction volume contained 50 μ M NAD⁺, 20 mM Tris/HCl buffer (pH 7.4), 150 mM NaCl, 0.01% Triton-X, 200 μ M crotonyl-CoA, 100 μ M NADH, 15 μ M of compound **1**, and 0.05 μ M ENR. Data was collected in duplicate with r² values greater than 0.97. b) 3 μ M *Pf*ENR was incubated with 15 μ M of compound **1** and 5 mM NAD⁺ for 45 minutes at 25 °C. The protein was diluted 100-fold with 200 μ M crotonyl-CoA and 100 μ M NADH to initiate the reaction. The two lines represent the activity of *Pf*ENR with celastrol (solid circle) and DMSO (open square). This experiment was conducted in triplicate with r² values greater than 0.97.

Figure S10.



Figure S10. Comparison of *Pf*ENR inhibition without iodoacetamide treatment (solid black) and with iodoacetamide treatment (checkered box). The final 100 μ L reaction volume contained 50 μ M NAD⁺, 20 mM Tris/HCl buffer (pH 7.4), 150 mM NaCl, 0.01% Triton-X, 200 μ M crotonyl-CoA, 100 μ M NADH, 50 μ M of compound **1**, and 0.05 μ M ENR. These experiments were conducted in triplicate.



Figure S11. The best-predicted celastrol docking pose, obtained when the molecule was docked into the 2OL4 structure. Part of the protein has been removed to facilitate visualization.