

Supplementary Information

Supplementary Figure 1 | Ridge detection algorithm overview. (a) PCA is used to identify the major direction in the RBJH. This direction is used, along with the coordinates of the global maximum, to create the major vector (v_0). Two additional vectors (v_{-1} , v_1) are created, at $\pm 15^\circ$ of the primary directions, to ensure complete coverage of the colorspace. (b) Local maxima tangent to each vector are identified to create basis for the RBJH ridge set. (c) The local maxima identified in each direction are combined to create a map of all local maxima in the RBJH. (d) Morphological dilation and thinning is applied to the density map to create the binarized ridge set.

Supplementary Figure 2 | Ridge set registration. (a) Binarized ridge set is further registered to identify tissue components. Tissue type are determined based on the distance (lower values correspond to closer components) from each of the four pure colors (red, blue, white, and black). Nuclei are defined as being closest to blue and black, ECM and cytoplasm are closest to white, and red blood cells are closest to red. (b) Overview of ridge set registration cases for H&E and IHC images. In cases where only one component is identified for H&E images, further processing is applied. Gaussian separation creates two components by cutting according to the distance from the global maximum according to a Gaussian fit.

Supplementary Figure 3 | Distance transformation algorithm overview. (a) Watershed segmentation is applied to the binarized ridge set. (b) Three transformation functions are created: i. Tissue region function (f_{region}), ii. Absorption function ($f_{\text{absorption}}$), and iii. Ridge function (f_{ridge}). (c) The three transformation functions are combined to create an overall transformation function that can be used as a basis for image transformation.

Supplementary Figure 4 | Overview of NLTD immunohistochemistry scoring technique. . Each immunohistochemically (DAB) stained image was separated into two images using the NLTD method, a nuclei-rich image and an antigen-rich image. Pre-processing steps were performed in order to only analyze nuclei rich regions where antigen staining was present, and avoid background areas where no staining should occur. Briefly, the nuclei-rich image was segmented using Otsu's thresholding technique. Small objects were removed from the image, followed by morphological opening and closing operations and another removal of small objects. After pre-processing, a transformation score was derived based on the ratio of antigen intensity to nuclei intensity.

Supplementary Figure 5 | Qualitative comparison of NLTD and color deconvolution. Comparison of the NLTD method with a freely available color deconvolution plugin (ImageJ "Color deconvolution" plugin; <http://www.mecourse.com/landinig/software/cdeconv/cdeconv.html>) with several built-in color deconvolution vectors. (a) Typical H&E image from breast cancer set. (b) Separation of input H&E image into nuclei (top) and non-nuclei (bottom) components. Shown are results from NLTD (left), two preset ImageJ/Fiji Color Deconvolution settings (middle-left, middle-right), and custom settings from user-input ROI using ImageJ/Fiji Color Deconvolution (right). (c) Typical immunohistochemistry image. (d)

Separation of input IHC image into nuclei (top) and antigen (bottom) components. Shown are results from NLTD (left) and preset ImageJ/Fiji Color Deconvolution settings (right). Qualitatively, the NLTD images show a smoother, better separated image for each component compared to the color deconvolution images.

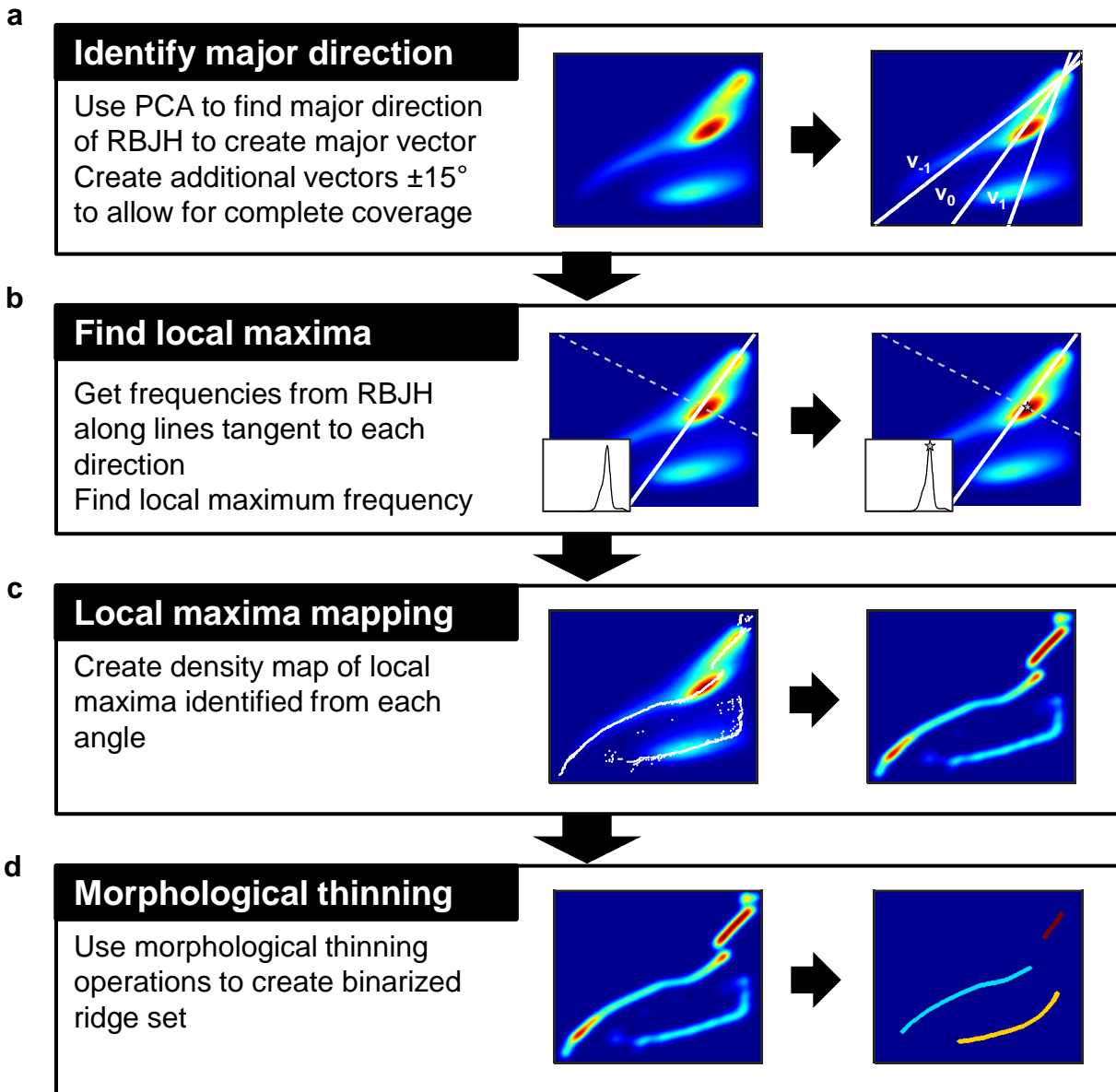
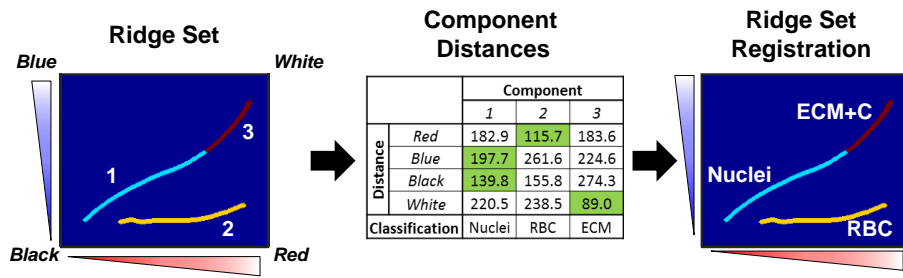
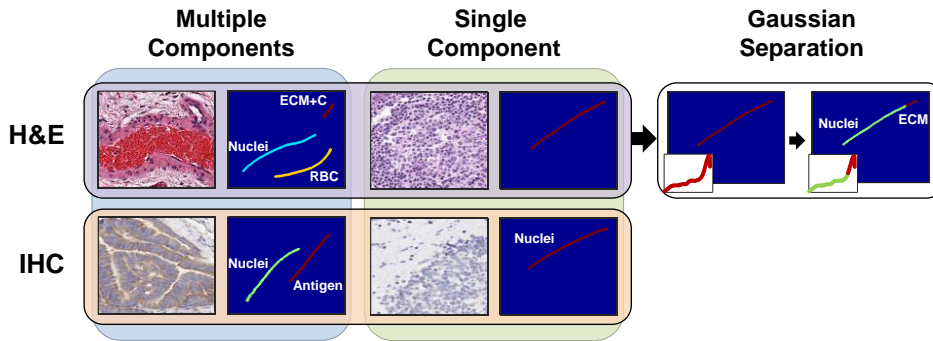


Figure S1

a**b****Figure S2**

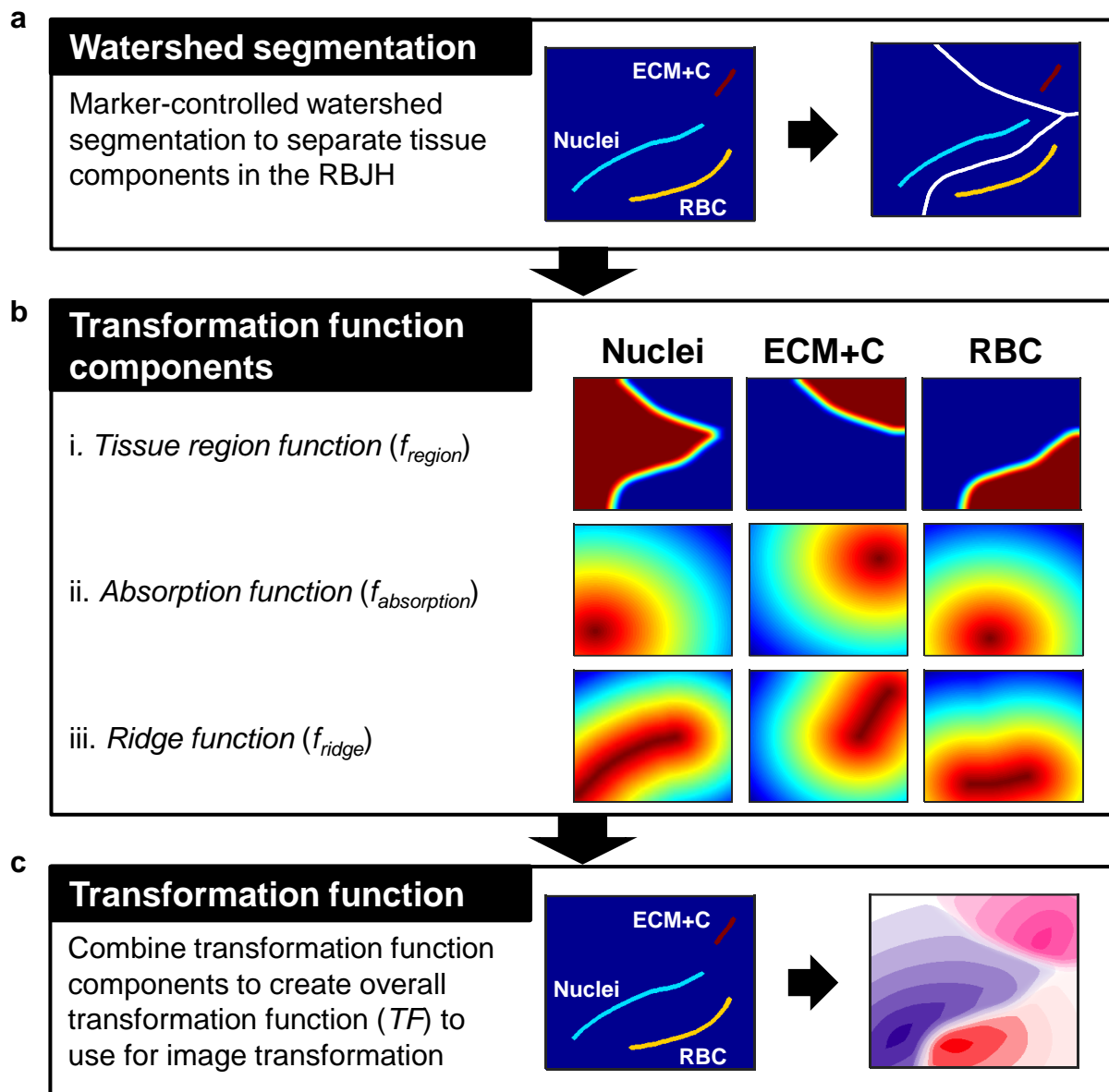


Figure S3

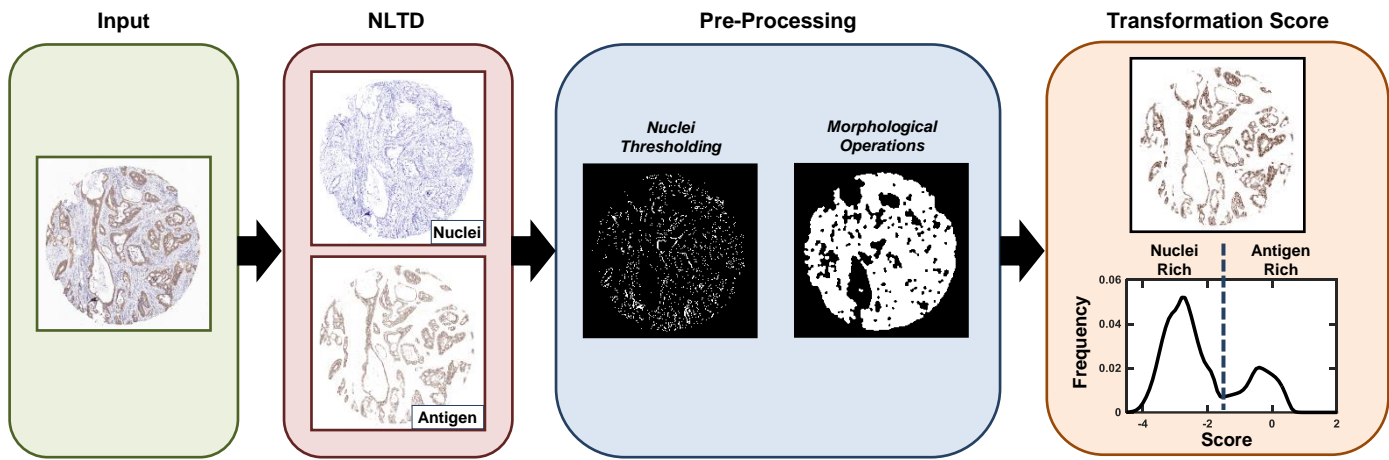


Figure S4

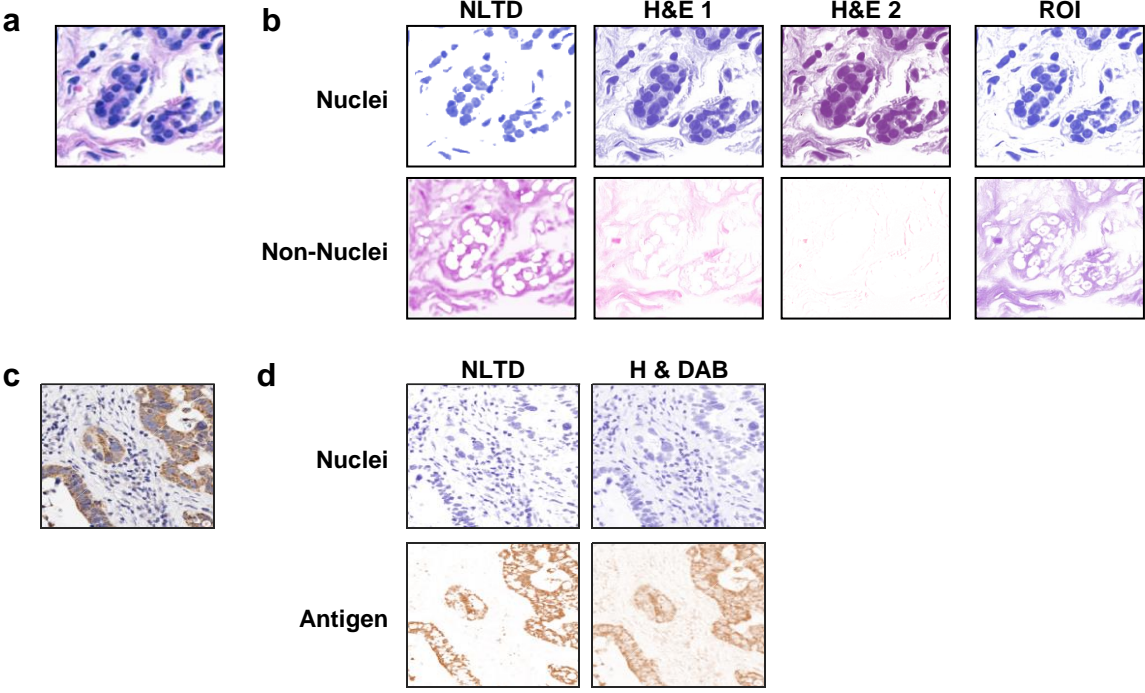


Figure S5