**Title:** Dual mechanisms regulating *glutamate decarboxylases* and accumulation of gamma-aminobutyric acid in tea (*Camellia sinensis*) leaves exposed to multiple stresses

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## **Supplementary Information**

Amino acid μg/g (fresh weight)	0 h	4 h		6 h	
		Aerobic	Anoxic	Aerobic	Anoxic
P-Ser	101.5±17.1b	66.5±4.5a	83.3±10.1ab	85.2±10.5ab	82.3±25.1ab
PEA	5.7±0.2a	8.3±2.2a	-	6.3±1.9a	-
Asp	122.0±14.7b	168.9±7.8c	13.5±2.3a	182.5±2.2c	12.3±0.9a
Thr	15.3±2.4a	23.5±0.9b	21.8±2.1ab	21.9±2.2ab	21.0±0.4b
Ser	38.3±6.9ab	40.8±2.7b	34.8±3.7ab	41.2±6.2ab	33.2±0.7a
Asn	5.8±0.4a	14.5±1.4c	9.1±1.2b	16.6±3.4c	9.7±0.8b
Glu	222.7±77.7c	335.7±20.1cd	101.9±17.6b	333.1±6.7d	45.8±10.5a
Thea	1075.9±122.8a	1276.6±112.5a	1281.9±187.2a	1202.3±385.2a	1136.5±216a
α-ΑΑΑ	7.6±1.6a	8.9±1.1a	8.2±0.4a	9.2±2.9a	6.4±1.2a
Gly	8.6±0.2a	11.9±0.4b	18.3±2.8c	10.9±1.6ab	17.1±0.2c
Ala	8.6±1.2a	12.7±1.2b	169.6±14.9c	14.8±3.3ab	137.7±11.6c
Cit	4.5±0.3a	4.5±0.1a	-	4.1±0.5a	-
α-ABA	2.4±0.3a	2.6±0.1a	2.5±0.4a	2.7±0.4a	2.6±0.2a
Val	22.8±1.6b	3.5±0.8a	34.5±4.3c	14.3±18.9abc	2.0±0.1a
Cys	3.4±0.4b	3.1±0.3ab	2.8±0.4ab	2.2±0.7ab	2.0±0.4a
Ile	8.6±2.5a	15.8±0.9c	11.5±1.9ab	16.9±2.4c	12.5±0.6ab
Leu	5.2±1.3a	15.4±1.0b	13.8±1.6b	15.1±1.6b	13.9±0.5b
Tyr	18.0±2.9a	41.9±1.1b	56.0±6.2c	39.8±3.4b	57.2±2.9c
Phe	18.5±0.9b	36.5±1.3c	10.0±1.1a	42.6±6.1c	9.0±0.5a
β-Ala	2.2±0.2a	3.8±0.6b	7.7±0.9c	3.3±0.5ab	7.9±0.4c
β-ΑΒΑ	0.6±0.1a	0.7±0.1a	0.6±0.2a	0.8±0.2a	0.7±0.3a
GABA	2.6±0.4a	2.0±0.5a	161.2±10.5b	4.3±1.3a	171.3±8.0b
Trp	16.7±4.2ab	22.9±1.5b	14.8±0.1a	21.5±5.9ab	16.9±3.9ab
EOHNH <sub>2</sub>	22.1±1.8a	23.4±1.1a	22.4±4.1a	20.0±1.9a	22.5±0.7a
Lys	5.4±1.3a	11.1±1.1b	11.1±1.5b	10.0±1.1b	11.1±0.8b
1Mehis	6.2±0.3b	10.3±0.2c	3.8±0.3a	9.7±2.1bc	3.7±0.9a
His	3.3±1a	5.8±1.5ab	14.4±2.4c	5.8±0.6b	16.1±2.5c
Arg	9.8±6.7a	14.3±4.1a	16.2±5.7a	16.1±3.2a	11.6±2.6a

 Table S1 Contents of free amino acids in picked (mechanically damaged) tea (*C. sinensis* cv. Jinxuan)

 leaves under aerobic and anoxic treatments.

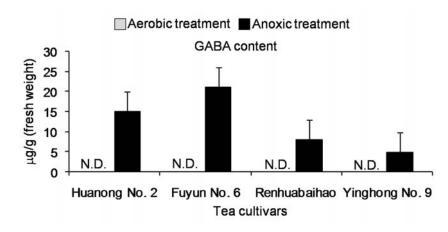
Data are expressed as mean  $\pm$  S.D. (n=3). -, not detectable. P-Ser, Phosphoserine. PEA, Phosphorylethanolamine. Asp, Aspartic acid. Thr, Threonine. Ser, Serine. Asn, Asparagine. Glu, Glutamate. Thea, Theanine.  $\alpha$ -AAA,  $\alpha$ -amino acetic acid. Gly, Glycine. Ala, Alanine. Cit, Citrulline.  $\alpha$ -ABA,  $\alpha$ -Aminobutyric acid. Val, Valine. Cys, Cystine. Ile, Isoleucine. Leu, Leucine. Tyr, Tyrosine. Phe, Phenylalanine.  $\beta$ -Ala,  $\beta$ -Alanine.  $\beta$ -ABA,  $\beta$ -Aminobutyric acid. GABA,  $\gamma$ -Aminobutyric acid. Trp, Tryptophan. EOHNH<sub>2</sub>, Ethanolamine. Lys, Lysine. 1Mehis, 1-Methyl histidine. His, Histidine. Arg, Arginine. The test tea samples were prepared in June, 2013. Different means with different letters in the same row are significantly different from each other ( $p \le 0.05$ ).

Table S2 Primers used in this study

For CsGAD CDS				
CsGAD1F	5'-GCGGCCGCATGGTTCTGTCAAAGACAACATCG-3'			
CsGAD1R	5'-GTCGACTTAGCAAACACCATTAGTCTTCTT-3'			
CsGAD2F	5'-CCCGGATCCATGGTTCTCTCAAAGACTGCTTC-3'			
CsGAD2R	5'-CCCAAGCTTCTAGCAAATCACTTGTGTTTTCC-3'			
CsGAD3F	5'-ACGCGTCGACCCATGGTTATCTCAAATGTTCAAACC-3'			
CsGAD3R	5'-ATAAGAATGCGGCCGCCTAACAAACTCCCTTTGTCTTACC-3'			
For mutants				
CsGAD1∆CR	5'-CCCAAGCTTTGACTCTTGGACTACAGC-3'			
CsGAD2∆CR	5'-CCCAAGCTTAGTCCCATTCTTCCCATTC-3'			
CsGAD3∆CR	5'-ATAAGAATGCGGCCGCCCTCTCTTCATCGCTT-3'			
For CsGAD tra	nsient expression			
CsGAD1F	5'-CTAGTCTAGAATGCACCATCACCATCACCATCGCGTTCTGTCAAAGACAACA-3'			
CsGAD1R	5'-CGCGGATCCCGTTAGCAAACACCATTAGTCT-3'			
CsGAD2F	5'-CTAGTCTAGAATGCACCATCACCATCACCATCGCGTTCTCTCAAAGACTGC-3'			
CsGAD2R	5'-CGCGGATCCCGCTAGCAAATCACTTGTGTT-3'			
CsGAD3F	5'-CTAGTCTAGAATGCACCATCACCATCACCATCGCGTTATCTCAAATGTTCAAAC-3'			
CsGAD3R	5'-CGCGTCGACCGCTAACAAACTCCCTTTGTCT-3'			
For qPCR				
Csβ-ActinF	5'-GCCATATTTGATTGGAATGG-3'			
Csβ-ActinR	5'-GGTGCCACAACCTTGATCTT-3'			
CsGABA-T1F	5'-TTCACAGATAACAAGTCACCTAAT-3'			
CsGABA-T1R	5'-TCTCACACTCTGCTCCAA-3'			
CsGABA-T2F	5'-TAGTATGTTGGCACCATTCAC-3'			
CsGABA-T2R	5'-ACCATAGACCAGCGAGAG-3'			
CsGABA-T3F	5'-CGCAGTAGAAGTAGCAGTTG-3'			
CsGABA-T3R	5'-GTCGGTTGTAAGAGATGTGAAT-3'			
CsSSADH1F	5'-CCACCAAGTTCCAGAGATAC-3'			
CsSSADH1R	5'-GCAAGTCCACAGGTAAGG-3'			
CsSSADH2F	5'-ACATTCGCTATAACTTCACCAT-3'			
CsSSADH2R	5'-TCCTCTCGGCAGATTAG-3'			
CsGDH1F	5'-AGGTGGAGTTACGGTTAGTTA-3'			
CsGDH1R	5'-GCACGAGCAACACGATTA-3'			
CsGDH2F	5'-TTGTCCTCCTATTACCTCCAT-3'			
CsGDH2R	5'-GAATCCAAGCCGAGAATGT-3'			
CsSSR1F	5'-AACATCAGCAGAAGGACAT-3'			
CsSSR1R	5'-ATCACCAAACACATCATATCAAG-3'			
CsSSR2F	5'-GTGCCATTAGTTCGCCAAT-3'			
CsSSR2R	5'-TGCTGATGCTTGAGAGGAA-3'			
CsGAD1F	5'-AGTGACATCCAGAAAGTCTTGCT-3'			
CsGAD1R	5'-CACCATTAGTCTTCTTCCTACTGAG-3'			
CsGAD2F	5'-TTCGACATCTGCAAGGTGCTCCA-3'			

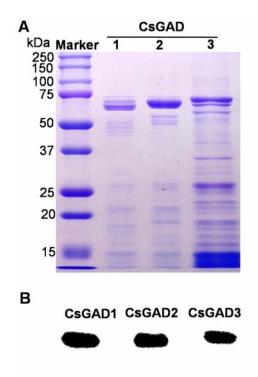
CsGAD2R	5'-ACTTGTGTTTTCCTAGCCAAGAC-3'
CsGAD3F	5'-TTTCACATAACAAATGCAACGTC-3'
CsGAD3R	5'-CTCCCTTTGTCTTACCACCCATA-3'
AtACT2F	5'-TATCGCTGACCGTATGAGCA-3'
AtACT2R	5'-TTACCTGCTGGAATGTGCTG-3'
AtGAD1F	5'-CGCTGAGAGACTTGTGATCG-3'
AtGAD1R	5'-TTCTTCACCGTGACCATCAA-3'
AtGAD2F	5'-ACACTCGCGGAGAGACTTGT-3'
AtGAD2R	5'-CCATCAGAATCTCCTTCTCCA-3'
AtGAD3F	5'-GCCAAGATGGCTAGTGGAAA-3'
AtGAD3R	5'-GGTAATACTTGCTACTAACGGAACG-3'
AtGAD4F	5'-GTTCACGCCAAGATGGCTAA-3'
AtGAD4R	5'-GCAAATTGTGTTCTTGTTGGTC-3'
AtGAD5F	5'-CGAGGCCTTGCAGATAGACT-3'
AtGAD5R	5'-GGACATCTTGGCAGTCTTCAC-3'

GABA-T, GABA transaminase. GAD, glutamate decarboxylase. GDH, glutamate dehydrogenase. SSADH, succinic semialdehyde dehydrogenase. SSR, succinic semialdehyde reductase. The Accession Numbers of Csβ-Actin, CsGABA-T1, CsGABA-T2, CsSSADH1, CsSSADH2, CsGDH1, CsGDH2, CsSSR1, CsSSR2, and AtACT2 are HQ420251, GBBZ01005465, KA282884, HP727388, HP728333, GBBZ01002276, GBBZ01009549, GAAC01043686, GARM01012969, and NM\_112764, respectively.

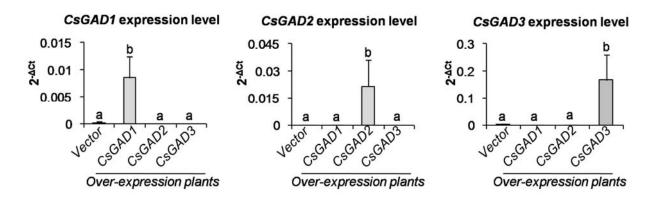


**Figure S1** Contents of GABA in different cultivars of picked tea leaves (with mechanical damage) in air and anoxia.

GABA, gamma-aminobutyric acid. The tea leaves were treated under anoxic condition for 6 h. N.D., not detectable. Data are expressed as mean  $\pm$  S.D. (n=3).

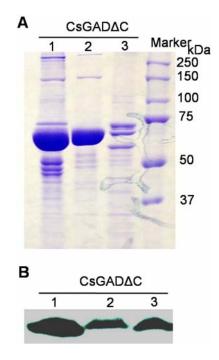


**Figure S2** SDS-PAGE (A) and Western Blot (B) analyses of CsGADs expressed in *E. coli*. GAD, glutamate decarboxylase. The CsGADs were the partially purified expressed S-tagged recombinant proteins.

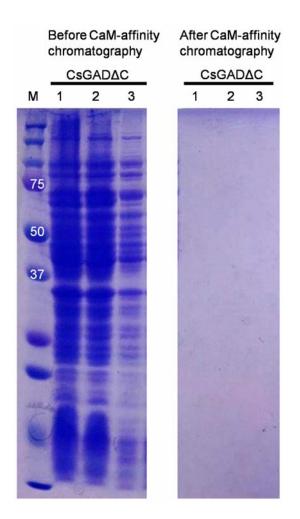


**Figure S3** *Agrobacterium*-mediated transient expression of *CsGADs* mRNAs in *Nicotiana benthamiana*.

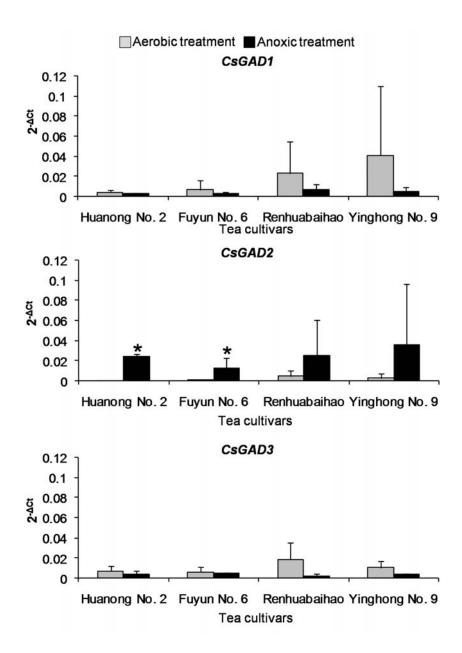
GAD, glutamate decarboxylase.  $2^{-\Delta^{Ct}}$  value was calculated based on comparison with beta-actin. Means distinguished with different letters are significantly different from each other ( $p \le 0.05$ ).



**Figure S4** SDS-PAGE (A) and Western Blot (B) analyses of CsGAD $\Delta$ C s expressed in *E. coli*. GAD $\Delta$ C, C-terminal truncated glutamate decarboxylase. The loading amounts of CsGAD1 $\Delta$ C, 2 $\Delta$ C and 3 $\Delta$ C were 0.3 µg, 0.1 µg, and 2 µg, respectively. The expressed CsGAD $\Delta$ Cs S-tagged recombinant proteins, were detected using S-tag antibody.



**Figure S5** Determination of CaM-binding ability of CsGADΔCs expressed in *E. coli*. GADΔC, C-terminal truncated glutamate decarboxylase. The CaM-affinity chromatography only detects proteins with CaM-binding ability. Any protein eluted from the CaM-affinity chromatography column is detected by SDS-PAGE, and is determined as having CaM-binding ability. No CsGADΔCs proteins were visible by SDS-PAGE, suggesting that the CsGADΔCs had no CaM-binding ability. M, molecular weight markers. A positive control was shown in the Figure 4A, suggesting that only the full-length CsGAD1 had CaM-binding ability.



**Figure S6** Expression levels of *CsGADs* in picked (mechanically damaged) tea leaves of different cultivars under aerobic and anoxic treatments

GAD, glutamate decarboxylase. The tea leaves were treated under anoxic condition for 6 h. Data are expressed as mean  $\pm$  S.D. (n=3).  $2^{-\Delta^{Ct}}$  value was calculated based on comparison with beta-actin. \*, *p* < 0.05.

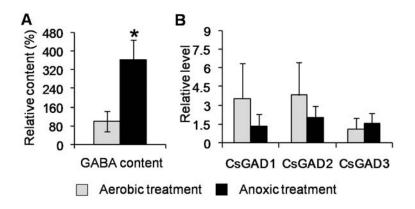


Figure S7 Effect of anoxia treatment on the GABA content and *CsGADs* mRNA levels in intact tea leaves

GABA, gamma-aminobutyric acid. GAD, glutamate decarboxylase. Tea leaves (*C. sinensis* cv. Jinxuan) were treated under anoxic conditions for 6 h. (A) The relative content (%) of GABA was calculated based on the GABA content of aerobic treatment (set as 100%). "\*" indicates that anoxic treatment significantly elevated GABA content (p < 0.05). (B) Transcript abundance was calculated based on the difference in cycle threshold (Ct) values between *CsGADs* and beta-actin transcripts by the normalized relative quantification  $2^{-\Delta\Delta}^{-Ct}$  method. The expression levels of the *CsGADs* in the tea leaves picked at 0 h were defined as 1.

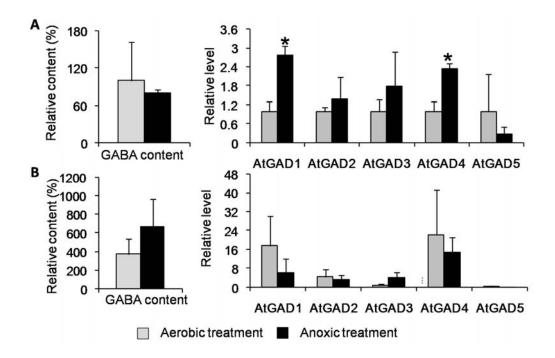


Figure S8 Effect of anoxia treatment on GABA content and levels of *AtGADs* mRNAs in the intact (A) and picked leaves (B) of *Arabidopsis thaliana* 

GABA, gamma-aminobutyric acid. GAD, glutamate decarboxylase. Leaves were treated under conditions of anoxia for 6 h. The relative content (%) of GABA was calculated based on the GABA content of intact leaves under aerobic treatment (set as 100%). "\*" indicates that anoxia significantly elevated *AtGAD1* and *AtGAD4* expression levels in the intact leaves (p < 0.05). Transcript abundance was calculated based on the difference in cycle threshold (Ct) values between *AtGADs* and beta-actin transcripts by the normalized relative quantification  $2^{-\Delta\Delta}^{Ct}$  method. The expression levels of the *AtGADs* of intact leaves under aerobic treatment were defined as 1.

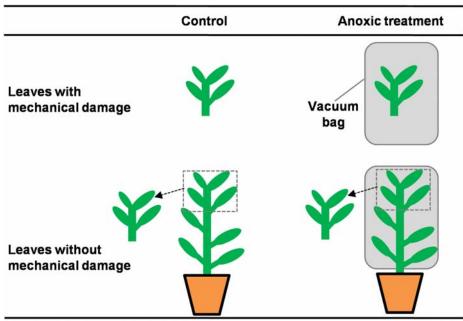


Figure S9 Anoxic treatments and mechanical damage applied to tea leaves.

Tea leaves (*C. sinensis* cv. "Jinxuan") were obtained from the tea seedlings grown under a controlled condition of 22-25°C and 12 h light/ 12 h dark. The anoxic treatments were carried out using a Vacuum Sealer (Deni, TVS-2013) at 25°C, 70% humidity in the dark. The air in the vacuum bag was drawn out by the Vacuum Sealer. The mechanical damage was induced by picking.