

**Microbial oil-degradation under mild hydrostatic pressure (10MPa): which pathways are impacted in piezosensitive hydrocarbonoclastic bacteria?**

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## Supplementary Information

**Figure S1** C12 bioavailability in *A. jadensis* KS\_339 (purple) and *A. dieselolei* KS\_293 (green) under atmospheric (0.1 MPa) and mild pressure (5 and 10MPa). Initial C12 concentration provided was equal to  $7.5 \text{ g L}^{-1}$ . Bars indicate 95% confidence intervals.

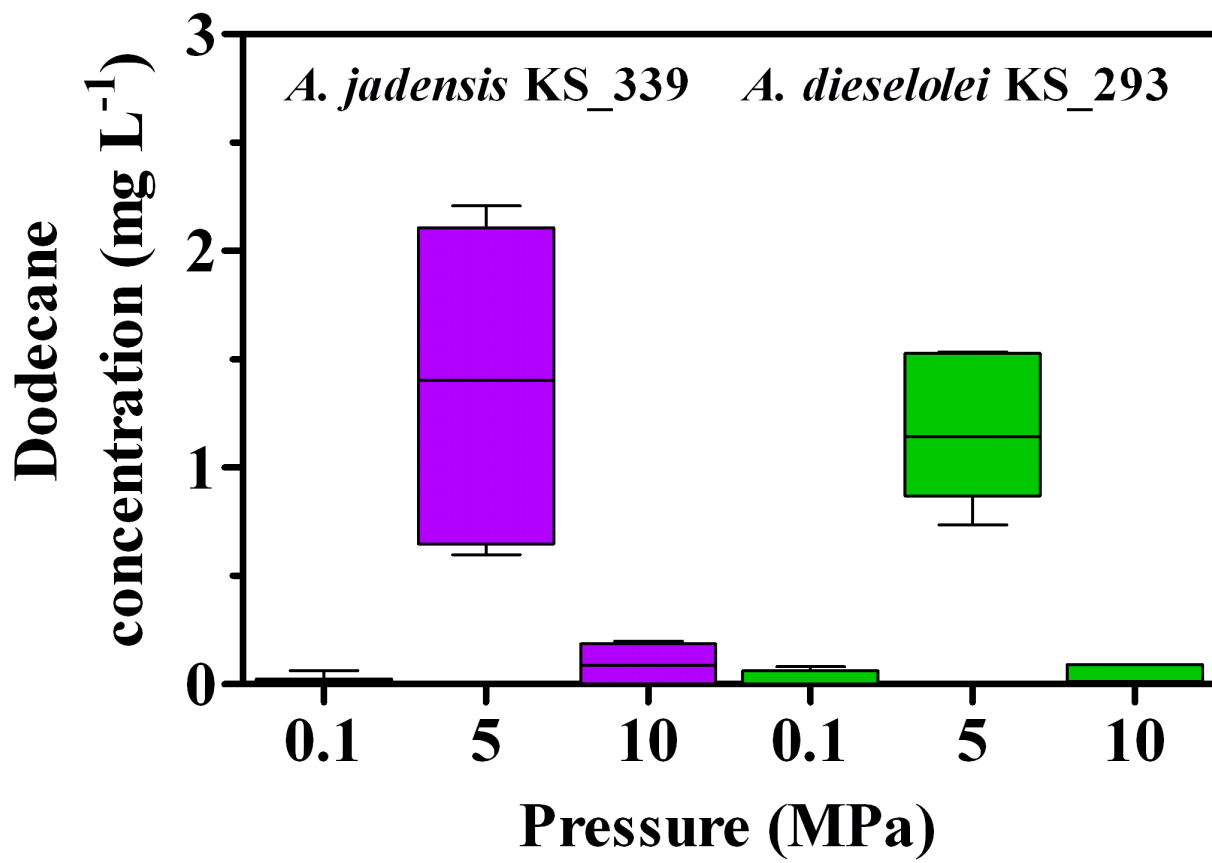
**Figure S2** Growth of *A. jadensis* KS\_339 (purple) and *A. dieselolei* KS\_293 (green) under atmospheric pressure (0.1 MPa) using air or pure O<sub>2</sub> as gas phase. Bars indicate 95% confidence intervals.

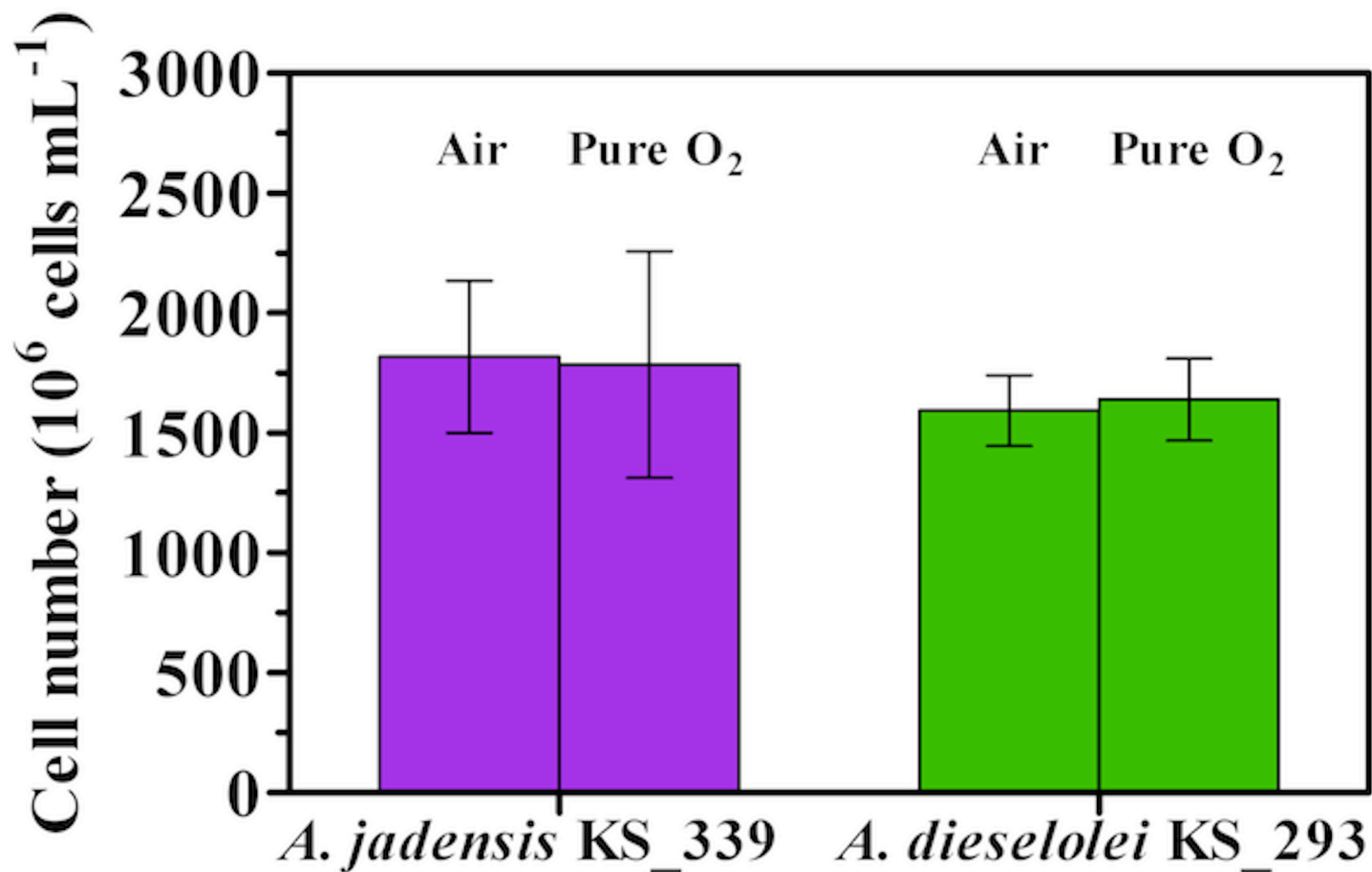
**Figure S3** Total CO<sub>2</sub> production in *A. jadensis* KS\_339 (purple) and *A. dieselolei* KS\_293 (green) under atmospheric (0.1 MPa) and mild pressure (5 and 10MPa). Bars indicate 95% confidence intervals.

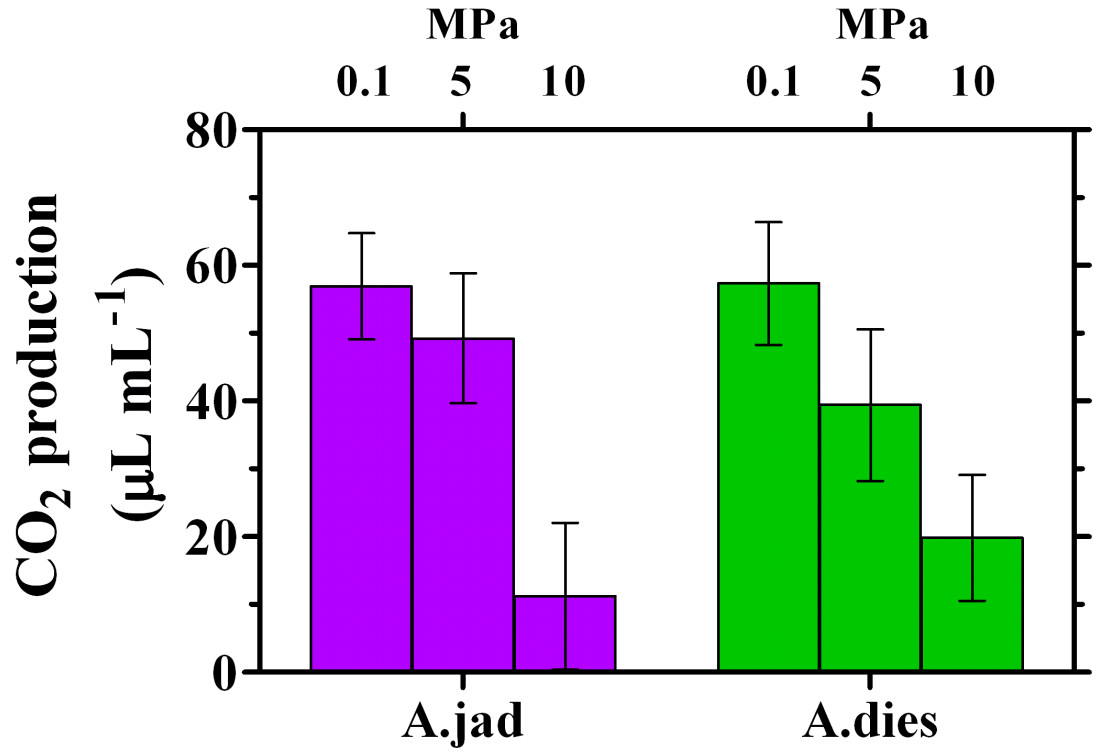
**Table S1** Total amount of O<sub>2</sub> transferred to the liquid phase by increased hydrostatic pressure to 10MPa. Measurements were conducted with sterile controls at 10MPa. Provided that diffusion is linear<sup>53</sup>, at 5MPa O<sub>2</sub> delivered to the culture was equal to 3.09mM.

**Table S2** Respiration capacity of *Alcanivorax* cells under 5 and 10MPa. Cell number and respiration rate values are derived from Fig. 1B and 3B

**Table S3** Expression of some typical pressure-resistance pool of genes in *A. dieselolei* KS\_293 cells under 10 and 0.1MPa.







**Table S1: Expression of some typical pressure-resistance pool of genes in *A. dieselolei* KS\_293 cells under 10 and 0.1MPa**

Function	Regulation	log2 FC	10MPa	0.1MPa	Cluster ID	Locus Tag	Description
<i>Cell division</i>							
-	<b>-0.71</b>	64.5	105.3	1439	B5T_03495	Cell division protein FtsL	
-	<b>-1.59</b>	1.6	4.7	1370	B5T_01893	Cell division topological specificity factor	
-	<b>-1.84</b>	10.0	35.8	646	B5T_01894	Cell division inhibitor, membrane ATPase	
-	<b>-1.87</b>	6.0	21.8	449	B5T_03484	Cell division protein FtsA	
-	<b>-1.96</b>	8.5	33.0	2138	B5T_03483	Cell division protein FtsZ	
-	<b>-3.5</b>	1.0	11.8	539	B5T_03489	Cell division protein FtsW	
<i>Outer Membrane Protein</i>							
-	<b>-0.68</b>	25.1	40.2	1785	B5T_02341	OmpA family protein	
-	<b>-0.77</b>	3.9	6.6	1985	B5T_02487	OmpA family protein	
-	<b>-0.99</b>	548.4	1088.7	1217	B5T_01469	Thrombospondin type 3 repeat family, OmpA-OmpF porin, OOP family	
-	<b>-1.05</b>	9.3	19.3	927	B5T_01929	Outer membrane protein assembly complex, YaeT protein	
-	<b>-1.38</b>	18.5	48.1	965	B5T_03555	Outer membrane protein transport protein (OMPP1/FadL/TodX)	
-	<b>-1.74</b>	2.2	7.2	980	B5T_01647	Efflux transporter, outer membrane factor lipoprotein, NodT family	
-	<b>-1.85</b>	6.8	24.7	1169	B5T_01197	Outer membrane assembly lipoprotein YfgL	
-	<b>-2.65</b>	27.9	175.6	588	B5T_03706	Outer membrane assembly lipoprotein YfiO	
-	<b>-3.19</b>	0.2	1.8	1031	B5T_03240	Outer membrane protein, OMP85 family, putative	
-	<b>-3.24</b>	23.9	225.7	613	B5T_01116	OmpW family	
-	<b>-3.59</b>	0.9	10.3	531	B5T_01930	Outer membrane protein (OmpH-like)	
-	<b>-5.03</b>	0.1	2.7	1457	B5T_00661	Type I secretion outer membrane protein, TolC family	
-	<b>-5.2</b>	0.5	17.9	1416	B5T_03628	Outer membrane lipoprotein LolB	
<i>Sigma Factor</i>							
-	<b>-0.78</b>	60.4	103.6	1579	B5T_00961	RNA polymerase sigma factor	
-	<b>-1.11</b>	288.6	622.9	1243	B5T_00231	RNA polymerase sigma factor	
-	<b>-1.33</b>	6.4	16.0	1698	B5T_00495	Regulator of RNA polymerase sigma(70)	
-	<b>-1.73</b>	203.6	674.8	798	B5T_01530	RNA polymerase sigma factor	
-	<b>-1.79</b>	6.9	23.8	1798	B5T_03577	RNA polymerase sigma-54 factor	
-	<b>-2.95</b>	4.3	33.5	363	B5T_00812	RNA polymerase sigma factor, sigma-70 family	
<i>Heat/Cold Shock Proteins</i>							
=	<b>0.44</b>	85.7	63.0	412	B5T_03857	Ribosome-binding factor A; <i>Csp</i> ;	
=	<b>0.03</b>	55.2	54.0	1277	B5T_03011	Hsp20/alpha crystallin family	

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-	<b>-1.88</b>	104.2	383.3	943	B5T_01982	Cold shock-like protein; <i>CspD</i>
-	<b>-1.89</b>	1.3	4.7	2071	B5T_03275	ATPase, histidine kinase-, DNA gyrase B-, and HSP90-like domain protein
-	<b>-1.98</b>	2.4	9.4	2179	B5T_00995	ATPase, histidine kinase-, DNA gyrase B-, and HSP90-like domain protein
-	<b>-2.14</b>	13.3	58.8	142	B5T_03709	ATPase, histidine kinase-, DNA gyrase B-, and HSP90-like domain protein
-	<b>-3.08</b>	2.1	17.3	1693	B5T_01542	ATPase, histidine kinase-, DNA gyrase B-, and HSP90-like domain protein
-	<b>-4.03</b>	0.4	6.9	382	B5T_01201	ATPase, histidine kinase-, DNA gyrase B-, and HSP90-like domain protein
-	<b>-4.06</b>	1.7	27.5	647	B5T_01484	ATPase, histidine kinase-, DNA gyrase B-, and HSP90-like domain protein
-	<b>-5.28</b>	0.4	13.8	640	B5T_00504	ATPase, histidine kinase-, DNA gyrase B-, and HSP90-like domain protein

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**Table S2: Total amount of O<sub>2</sub> transferred to the liquid phase by increased hydrostatic pressure to 10MPa. Measurements were conducted with sterile controls at 10MPa. Provided that diffusion is linear<sup>1</sup>, at 5MPa O<sub>2</sub> delivered to the culture was equal to 3.09mM.**

Hydrostatic pressure	O <sub>2</sub> in the headspace	Total head-space	Total O <sub>2</sub> moving to the water-phase	Total O <sub>2</sub> moving to the water-phase
MPa	%	mL	mL	mM
0.1	19.37 ±0.09	6.5	-	-
10	11.34 ±1.93	6.5	0.52 ±0.09	6.18 ±1.13



# Sheet1

**Table S3. Respiration capacity of *Alcanivorax* cells under 5 and 10MPa. Cell number and respiration rate values are derived from Fig. 1B and 3B**

Pressure	<i>A. jadensis</i>	<i>A. dieselolei</i>	<i>A. jadensis</i>	<i>A. dieselolei</i>	<i>A. jadensis</i>	<i>A. dieselolei</i>
MPa	Cell number (10 <sup>9</sup> cells/mL)		Respiration rate ( $\mu$ mol O <sub>2</sub> /10 <sup>9</sup> cells)		mM respiration	
5	0.86	0.79	4.1	5.7	3.6	4.5
10	0.54	0.44	10.5	8.6	5.7	3.8

## References

1. Enns, T., Scholander, P.F. & Bradstreet, E.D. The effect of hydrostatic pressure on gases dissolved in water. *J Phys Chem* **69**:389-391 (1965)