

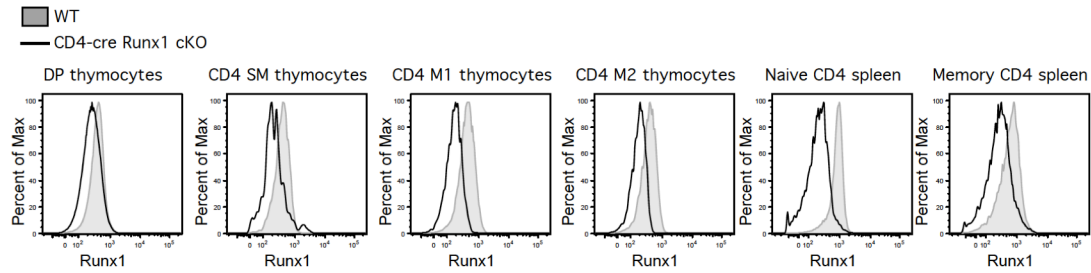
An Essential Role of Transcription Factor Runx1 for T Cell Maturation

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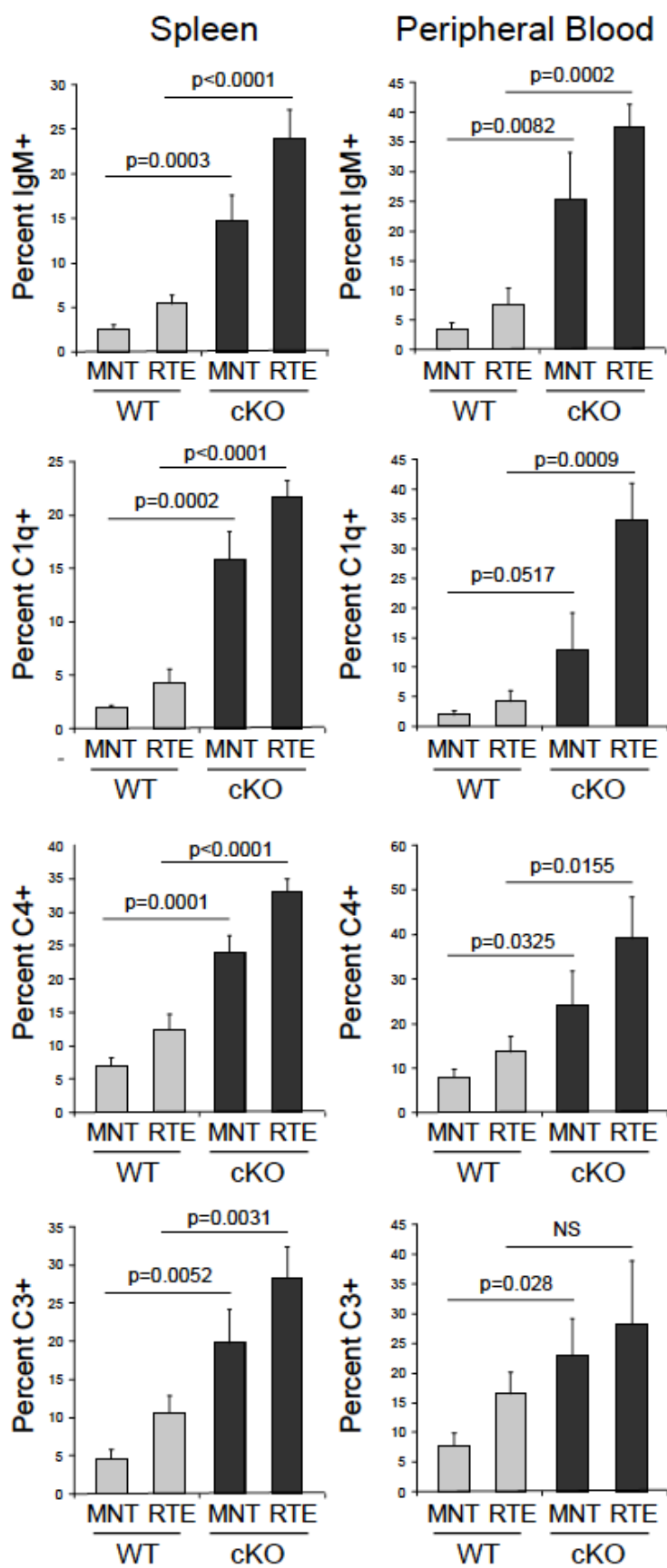
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Supplemental Figure 1

Runx1 protein expression throughout T cell development in CD4-cre Runx1 cKO mice and WT mice

Intranuclear Runx1 staining was performed on thymocytes and splenocytes from WT and CD4-cre Runx1 cKO mice. The protein levels of Runx1 in DP ($CD4^+CD8^+$), CD4 SM ($CD4^+CD8^-CD24^{hi}CCR7^{lo}$), CD4 M1 SP ($CD4^+CD8^-CD24^{hi}CCR7^{hi}$) thymocytes, CD4 M2 SP ($CD4^+CD8^-CD24^{lo}CCR7^{hi}$) thymocytes, naive ($CD4^+CD62L^+CD44^-$) and memory ($CD4^+CD62L^-CD44^+$) T cells from both WT and CD4-cre Runx1 cKO mice were examined. As intranuclear staining destroys GFP fluorescence, expression of Runx1 in RTEs was not examined. Data shown are representative of five WT and five Rag1-GFP CD4-cre Runx1 cKO mice from five separate experiments.

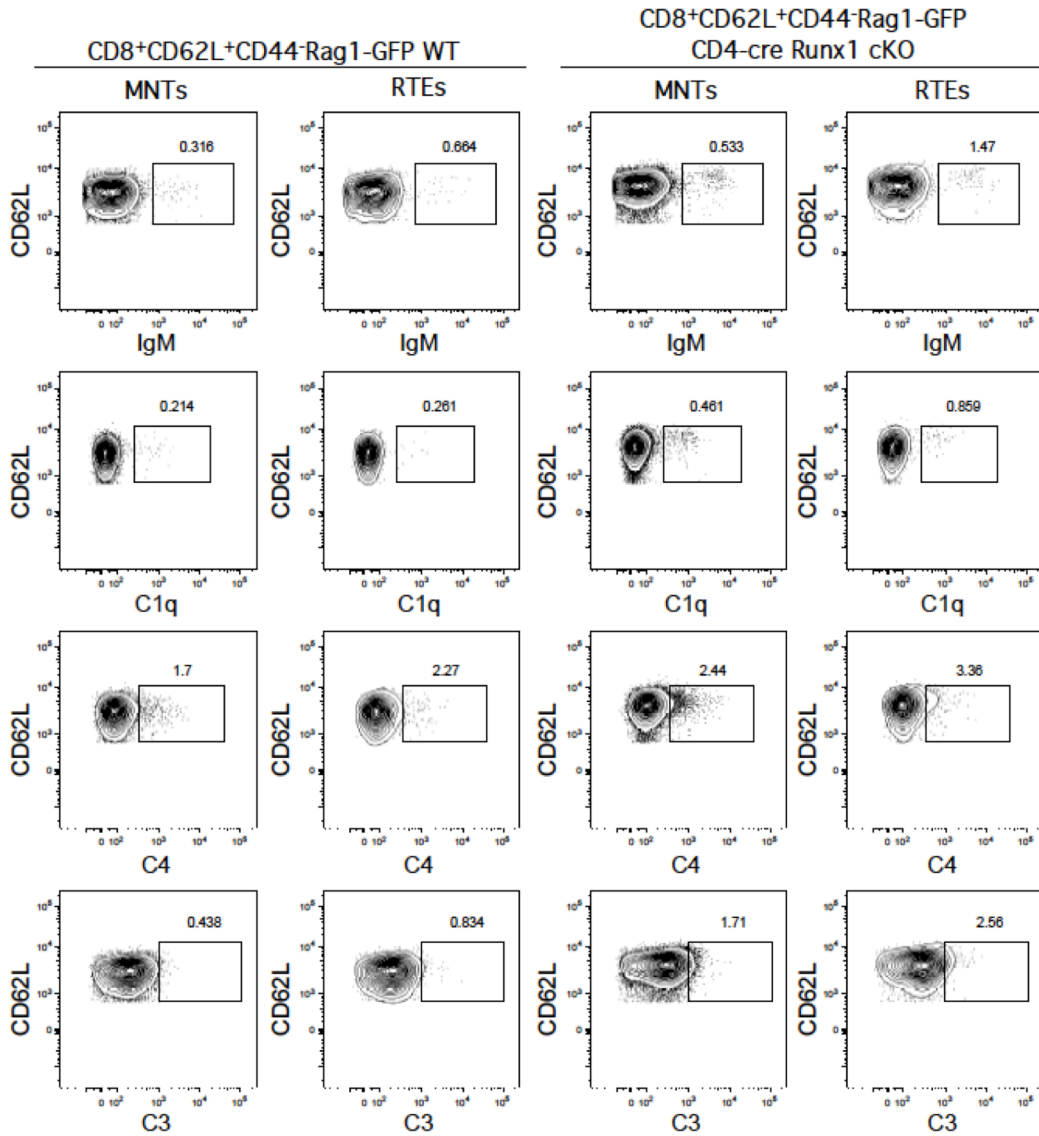


Supplemental Figure 2

Quantitation of IgM and complement protein deposition on splenic and peripheral blood Runx1-deficient CD4⁺ RTEs and MNTs as compared to WT

Deposition of IgM, C1q, C4 and C3 in splenic (left column) and peripheral blood (right column) CD4⁺ RTEs and MNTs from Rag1-GFP WT and Rag1-GFP CD4-cre Runx1 cKO mice as shown in Figure 3 was quantified across several experiments. For analysis of complement deposition in splenic CD4⁺ T cells, the results shown are quantified from at least 6 Rag-GFP WT and 5 Rag-GFP CD4-cre Runx1 cKO mice from 5 independent experiments. For the analysis of complement deposition in peripheral blood CD4⁺ T cells, the results shown are quantified from at least 6 Rag1-GFP WT and 3 Rag1-GFP CD4-cre Runx1 cKO mice. Error bars represent SEM, and significance was determined by unpaired Student's *t* test using GraphPad Prism.

Spleen

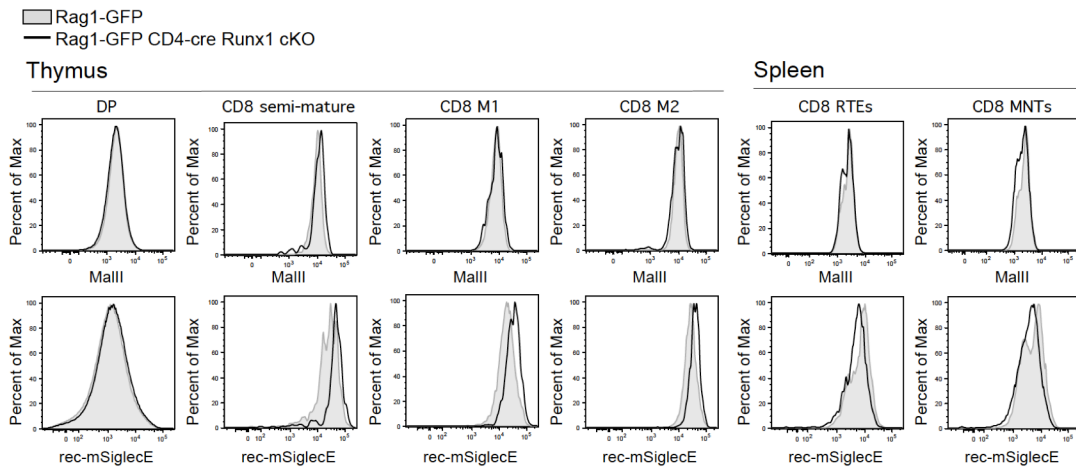


Supplemental Figure 3

IgM, C1q, C4 and C3 are not deposited on CD8⁺ T cells from CD4-cre Runx1 cKO mice

Splenic CD8⁺ RTEs and MNTs from Rag1-GFP WT and Rag1-GFP CD4-cre Runx1 cKO mice were examined for the deposition of IgM, C1q, C4 and C3 as denoted in the figure. Cells were incubated in GVB⁺⁺ buffer prior to staining with indicated Abs. CD8⁺ RTEs were

gated on CD62L⁺CD44⁻Rag1-GFP⁺ while CD8⁺ MNTs were gated on CD62L⁺CD44⁻Rag1-GFP⁻. Data shown are representative of six Rag1-GFP WT and six Rag1-GFP CD4-cre Runx1 cKO mice from five separate experiments.



Supplemental Figure 4

α 2,3- and α 2,8-sialylation on CD8 SP thymocytes and CD8⁺ T cells is not altered from CD4-cre Runx1 cKO mice

DP (CD4⁺CD8⁺), SM CD8 SP (CD8⁺CD24^{hi}CCR7^{lo}), M1 CD8 SP (CD8⁺CD24^{hi}CCR7^{hi}), M2 CD8 SP (CD8⁺CD24^{lo}CCR7^{hi}) thymocytes, splenic CD8⁺ RTEs (CD8⁺CD62L⁺CD44⁺Rag1-GFP⁺), CD8⁺ MNTs (CD8⁺CD62L⁺CD44⁻Rag1-GFP⁻) from Rag1-GFP WT (grey histogram) and Rag1-GFP CD4-cre Runx1 cKO mice (solid line) were examined for α 2,8-sialylation by using recombinant mSiglec-E Fc chimera (rec mSiglec-E) and α 2,3-sialylation using MaIII. Thymocytes were first gated on expression of Rag1-GFP to exclude recirculating mature T cells. Representative FACS analysis from at least three mice in each group from three separate experiments is shown.