

Supplementary information

Design, synthesis, crystallization and biological evaluation of new symmetrical biscationic compounds as selective inhibitors of human Choline Kinase $\alpha 1$ (ChoK $\alpha 1$)

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Table S1: Conditions reaction to obtain compound **7** and **8**

Synthesis of 7			
	Quantity	Time and Temperature	Yields
Refluxed	5 g of 6	8 h - 70 °C	21%
MW	1 g of 6	28' - 130 °C	20%
		14' - 140 °C	31%
		28' - 140 °C	35%
		56' - 140 °C	27%
		14' - 150 °C	25%
Synthesis of 8			
Refluxed	1 g of 7	5 h - 70 °C	39%
MW	100 mg of 7	21' - 120 °C	65%
	200 mg of 7	21' - 120 °C	52%
	300 mg of 7	21' - 120 °C	55%
		40' - 120 °C	50%

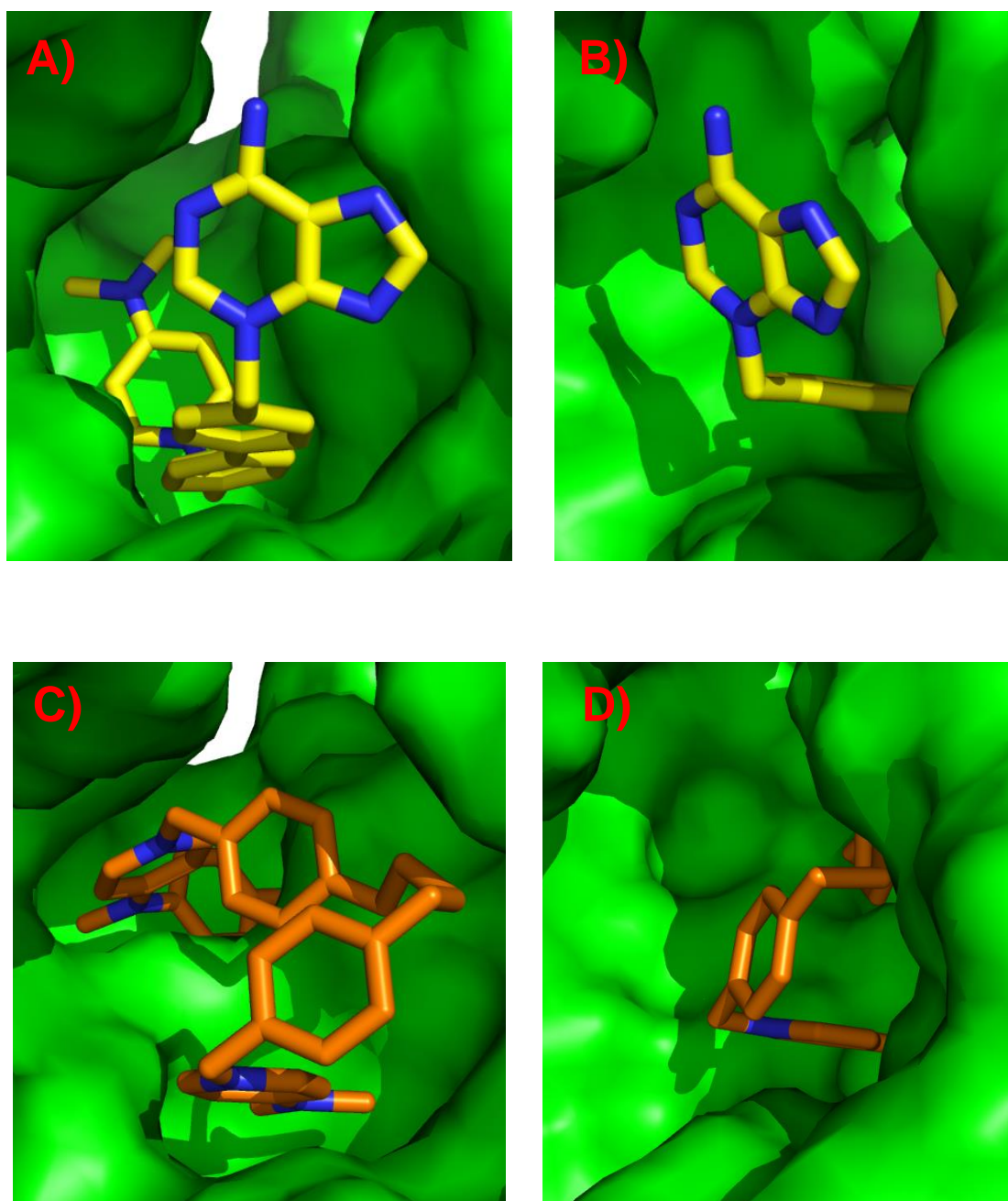


Figure S1. Detail view of the compounds **2** (carbon atoms in yellow color) and **4** (carbon atoms in cyan color) inserted into the Cho binding site represented by the protein surface. **(A)** The 1-(biphenyl-4-ylmethyl)-4-(dimethylamino)pyridinium fragment of compound **2** is inserted into the Cho binding site. **(B)** The adenine moiety is situated outside of the Cho binding site, and this fact justified the absence of interactions between the adenine moiety and the protein. **(C)** The 4-chloro-*N*-methylaniline fragment is inserted into the additional binding site, and the rest of the molecule is inserted into the ChoK binding site. **(D)** The second cationic head is also situated inside the protein.

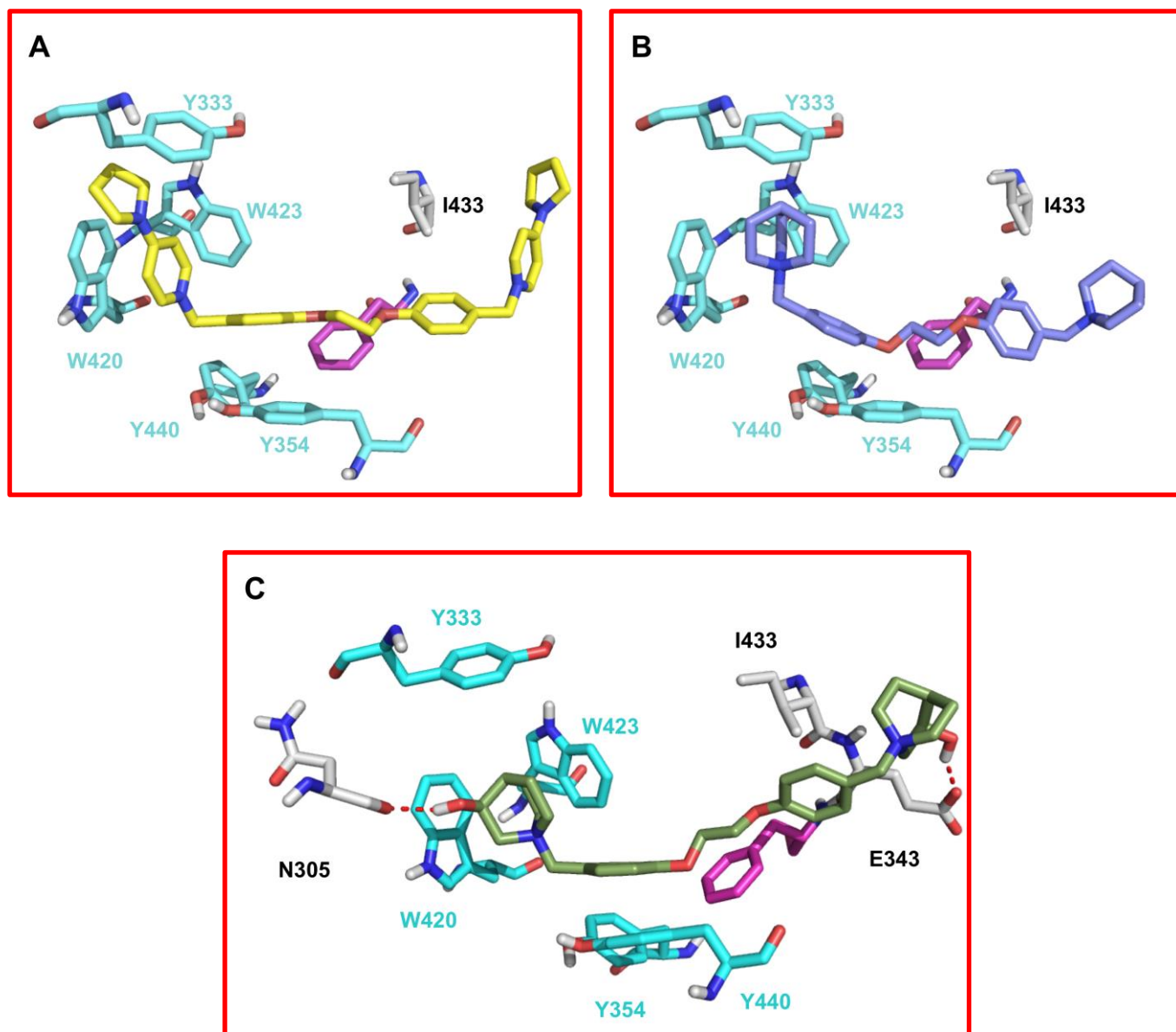


Figure S2. Poses resulting in the docking studies of compounds **10b** (A, carbon atoms in yellow color), **10d** (B, carbon atoms in purple color) and **10e** (C, carbon atoms in green color) inside the Cho binding site of ChoK α 1/2 complex (PDB ID: 4BR3)

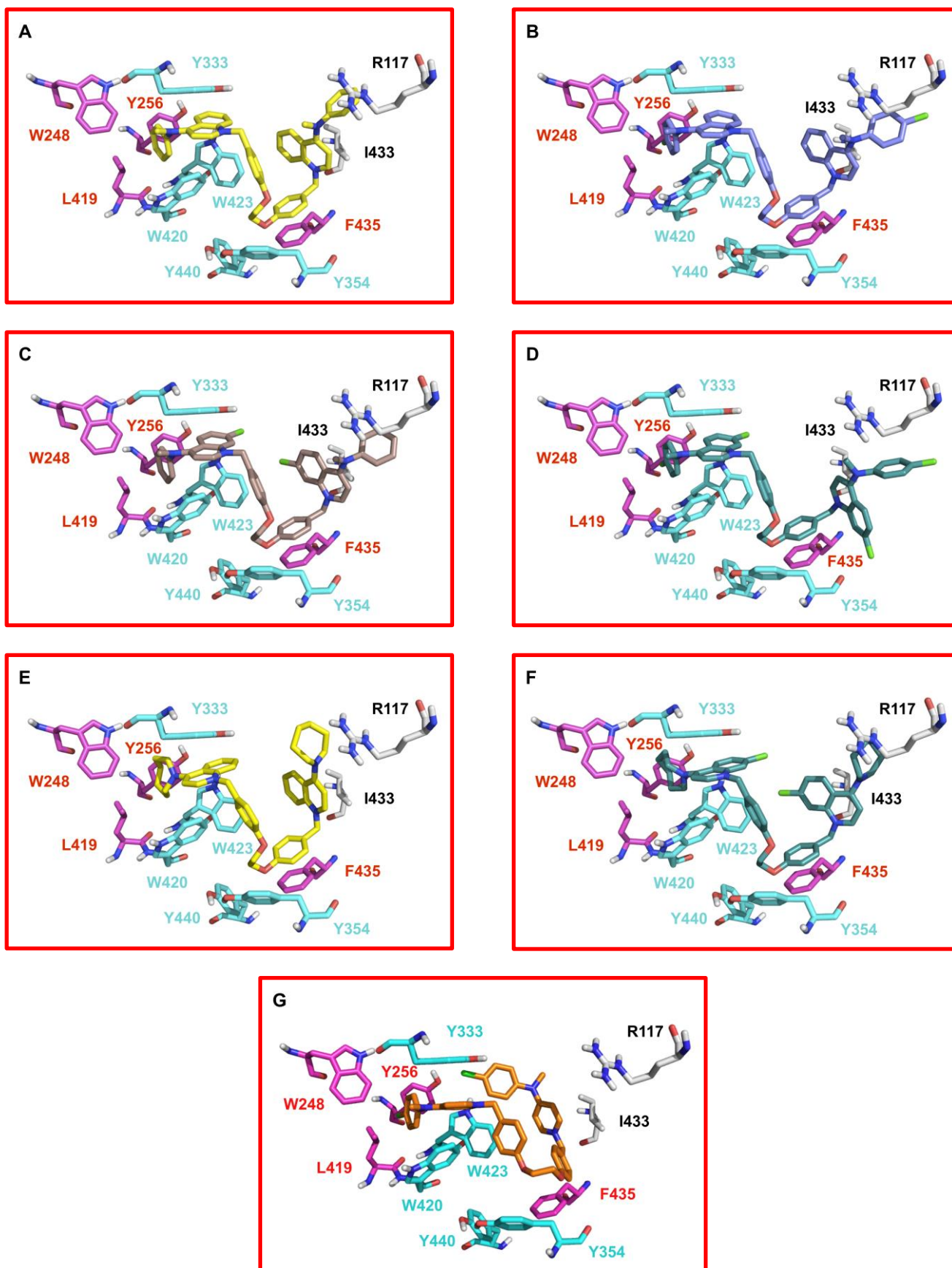


Figure S3. Poses resulting in the docking studies of compounds **10f** (A, carbon atoms in yellow color), **10g** (B, carbon atoms in blue-slate color), **10h** (C, carbon atoms in reddish-salmon color), **10i** (D, carbon atoms in blue color), **10j** (E, carbon atoms in yellow color), **10k** (F, carbon atoms in blue color) and **10c** (carbon atoms in orange color) inside the Cho binding site of ChoKa1/4 complex (PDB ID: 4CG8).

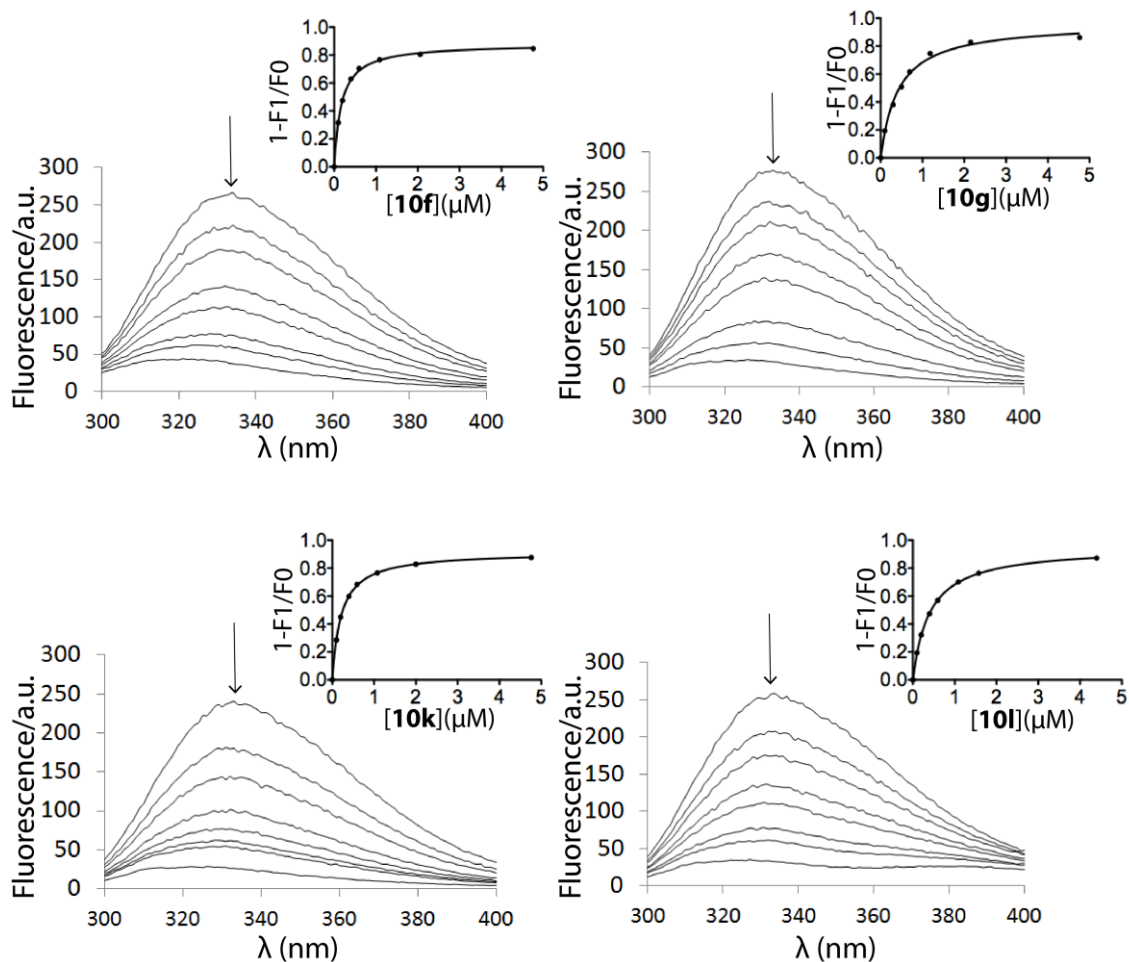


Figure S4: Quenching of intrinsic ChoK α 1 tryptophan fluorescence measured at increasing concentrations of compounds **10f**, **10g**, **10k** and **10l**. All data points represent the means \pm S.D. for three measurements. The K_d for the four compounds was determined by fitting fluorescence intensity data against its concentration. A black arrow indicates the decrease of the maximal fluorescence signal with increasing concentrations of each compound.

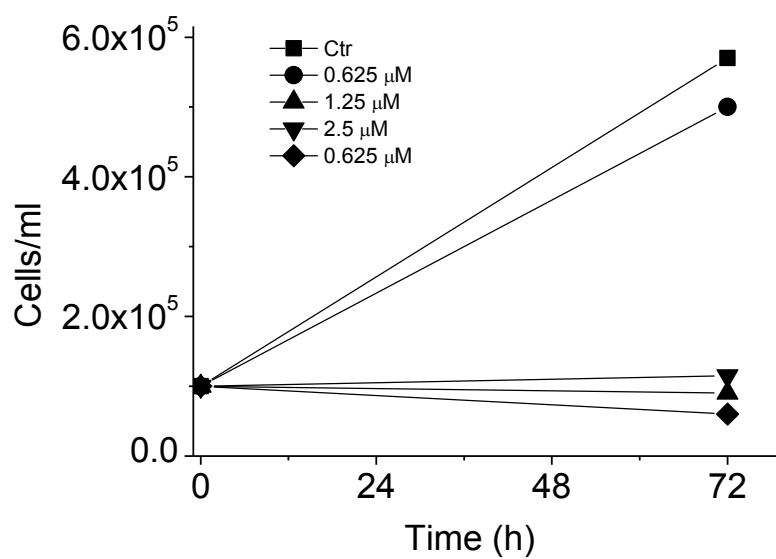


Figure S5

Jurkat cells were treated with the indicated concentration of **10a** for 72 h and then cells were harvested, washed, counted, and incubated in a drug-free medium. After a further 72 h of incubation, cell viability was analyzed by trypan blue exclusion assay as described in the experimental section.

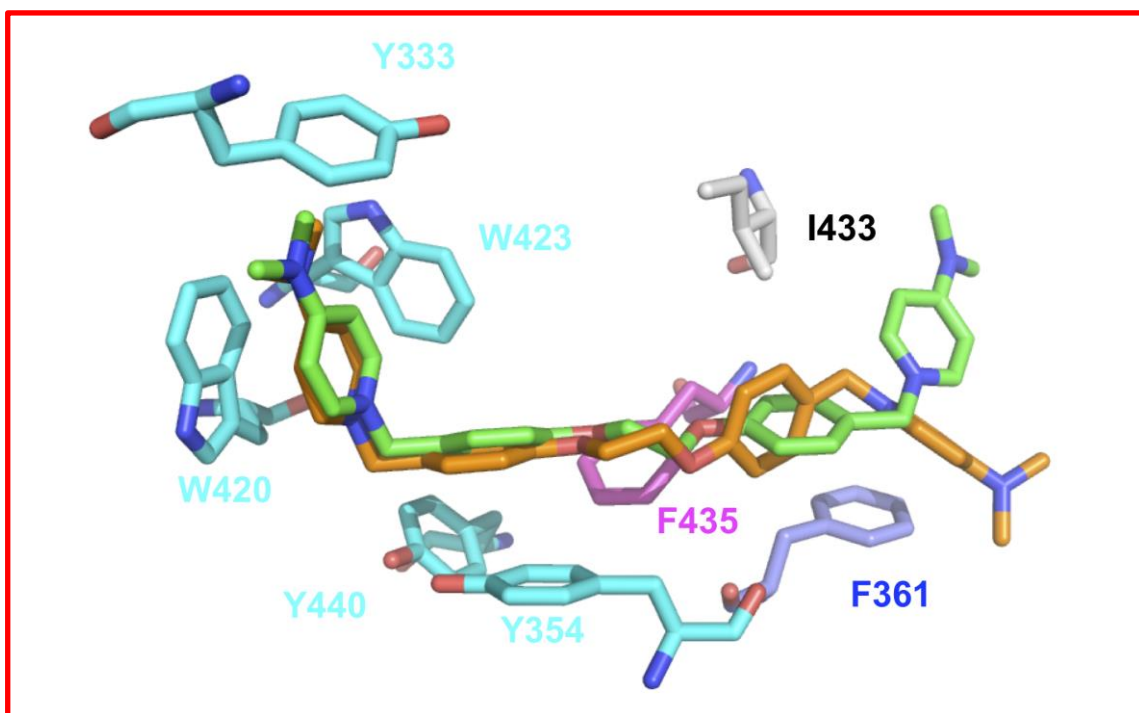


Figure S6. Superposition of the resulting pose of compound **10a** (carbon atoms in green color) with compound **10a** (carbon atoms in orange color) inserted into Cho binding site into the crystal structure (PDB ID: 4TST)

Table S2. Data collection and refinement statistics. Values in parentheses refer to the highest resolution shell. Ramachandran plot statistics were determined with PROCHECK.

	ChoKα1-10a
Space group	P4 ₃ 2 ₁ 2
Wavelength (Å)	1.0507
Resolution (Å)	58-1.45 (1.53-1.45)
Cell dimensions (Å)	<i>a</i> = 60.65 <i>b</i> = 60.65 <i>c</i> = 220.40
Unique reflections	74.167
Completeness	99.9 (99.4)
<i>R</i> _{sym}	0.058 (0.672)
<i>I</i> / <i>s</i> (<i>I</i>)	34.8 (5.0)
Redundancy	23.2 (14.0)
<i>R</i> _{work} / <i>R</i> _{free}	0.197/0.218
RMSD from ideal geometry, bonds (Å)	0.0048
RMSD from ideal geometry, angles (°)	1.079
< <i>B</i> > protein (Å ²)	18.11
< <i>B</i> > EB3D (Å ²)	30.50
< <i>B</i> > solvent (Å ²)	33.42
< <i>B</i> > Ethylenglycol (Å ²)	29.97
Ramachandran plot:	
Most favored (%)	97.61
Additionally allowed (%)	1.882.09
Generously allowed (%)	0.000.30
PDB ID	4TST

Experimental Procedures

General Experimental details

Chemistry. Melting points were taken in open capillaries on a Stuart Scientific SMP3 electrothermal melting-point apparatus and were uncorrected. Elemental analyses were performed on the Thermo Scientific Flash 2000 analyzer only on the final compounds tested as *HsChoK α 1* inhibitors. The measured values for C, H, and N agreed to within $\pm 0.40\%$ of the theoretical values. Analytical thin-layer chromatography (TLC) was performed using Merck Kieselgel 60 F254 aluminum plates and visualized by UV light or iodine. All evaporation occurred *in vacuo* with a Büchi rotary evaporator and the pressure controlled by a Vacuubrand CVCII apparatus. For flash chromatography, Merck silicagel 60 with a particle size of 0.040 – 0.063 mm (230 – 400 mesh ASTM) was used: 600 MHz ^1H and 151 MHz ^{13}C NMR Varian Direct Drive; a 400 MHz ^1H and 100 MHz ^{13}C NMR Varian Direct Drive; or 300 MHz ^1H and 75 MHz ^{13}C NMR Varian Inova Unity spectrometers at room temperature. Chemical shifts (δ) are quoted in parts per million (ppm) and are referenced to the residual solvent peak. Spin multiplicities are given as s (singlet), bs (broad singlet), d (doublet), t (triplet), and m (multiplet). High-resolution Nano-Assisted Laser Desorption/Ionization (NALDI-TOF) or Electrospray Ionization (ESI-TOF) mass spectra were carried out on a Bruker Autoflex or a Waters LCT Premier Mass.

General procedure A for the synthesis of the linker

1,2-bis(*p*-tolxy)ethane (7) [1]: To a solution of 0.4 g of NaOH in 5 mL of EtOH 1 g of 4-methoxyphenol (**6**) was added. The mixture was stirred for 30 min at room temperature and afterwards 0.4 mL of 1,2-dibromoethane was added before microwave irradiation at 140 °C for 28 min. The reaction was quenched by cooling (ice/water bath) and by addition of 25 mL distilled water. The solid was filtered

and washed twice with water H₂O (2 x 2 mL) and EtOH (1 x 2 mL), dried to vacuum to furnished a white solid identified as 1,2-bis(*p*-methylphenoxy)ethane (**7**). Yield 35%.

¹H NMR (300 MHz, CDCl₃) δ: 7.10-7.08 (d, *J* = 8.61 Hz, 4H), 6.86-6.84 (d, *J* = 8.44 Hz, 4H), 4.28 (s, 4H), 2.29 (s, 6H).

1,2-bis(4-(bromomethyl)phenoxy)ethane (8) [2]: To a solution of **7** (0.1, 25 mmol) in tetrachloromethane (3 mL), a mixture of dibenzoylperoxide (0.5 mg, 2 mmol) and NBS (9.08 mg, 51 mmol) was added. The mixture was purged with argon before microwave irradiation at 120 °C, 21 min. The precipitate was filtered and washed twice with diethylether (2 x 2 mL), and dried under vacuum to give **7** as a yellow solid. Yield: 65%,

¹H NMR (300 MHz, CDCl₃) δ: 7.35-7.32 (d, *J* = 8.73 Hz, 4H), 6.92-6.88 (d, *J* = 8.73 Hz, 4H), 4.50 (s, 4H), 4.32 (s, 4H).

General procedure B for the synthesis of the cationic heads

The cationic heads (**9a-d**) are commercially available while that the cationic heads (**9f-l**) were obtained following the procedure previously described [3-9], starting of 4-chloroquinoline and 7,4-dichloroquinoline which were treated with *N*-methylaniline, 4-chloro-*N*-methylaniline, perhydroazepine or pyrrolidine to obtain **9e** (4-*N*-methylanilinoquinoline), **9f** (4-(4-chloro-*N*-methylanilino)quinoline), **9g** (7-chloro-4-(*N*-methylanilino)quinoline), **9h** (7-chloro-4-(4-chloro-*N*-methylanilino)quinoline), **9i** (4-perhydroazepinoquinoline), **9j** (7-chloro-4-perhydroazepinoquinoline) and/or **9k** (7-chloro-4-(pyrrolidino)quinoline).

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