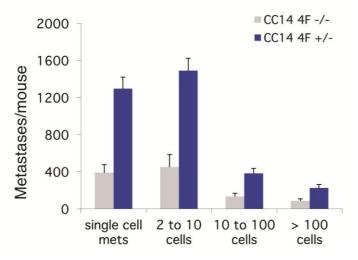
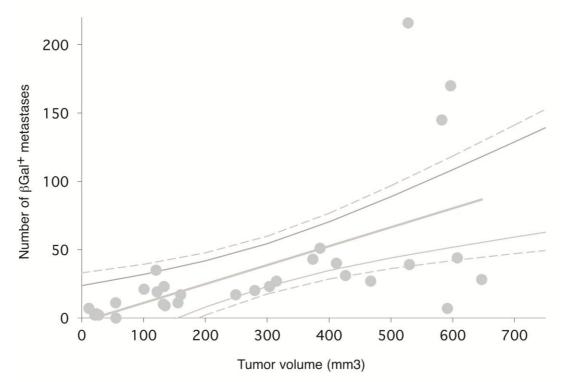
Supplementary Figures



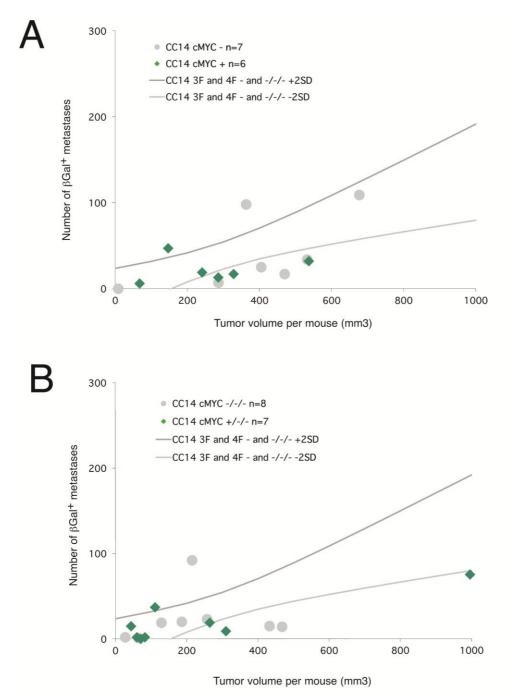
Supplementary Figure S1. Quantification of 4F-induced lung metastases by size in CC14 cells.

Lung metastases were scored after injection of human tumor cells into the circulation. In all cases p<0.05. See main text for details.



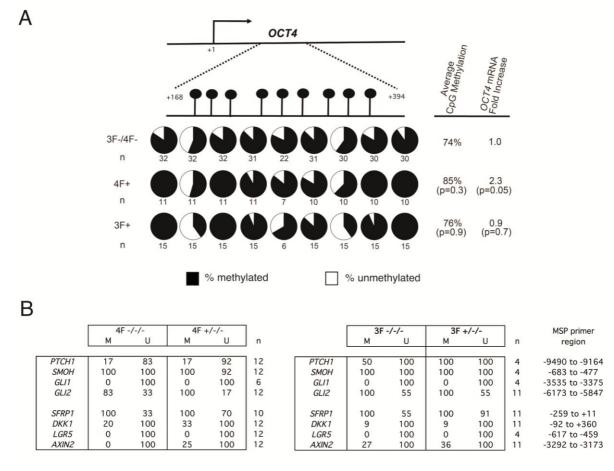
Supplementary Figure S2. Plot of the number of β Gal⁺ lung metastases versus total tumor volume per mouse for all CC14 controls.

The individual tumor values and metastases for 4F and 3F uninduced (-, -/-/-) samples is denoted with round markers. The trend line is shown bisecting the control space, flanked on either side by the average +/- 2 standard deviation (SD) space limits (with p<0.05; thin solid lines) or the +/- 3SD space limits (with p<0.01; broken thin lines). Note the presence of rare outliers.



Supplementary Figure S3. Plots of the number of β Gal⁺ lung metastases versus total tumor volume per mouse for cMYC tumors.

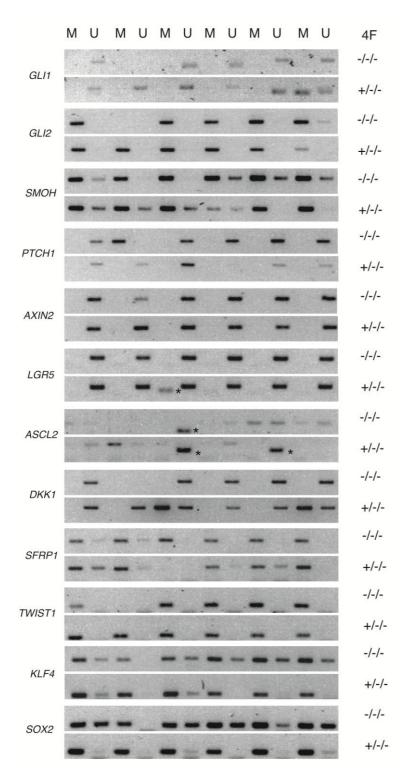
Plots for cMYC+ vs cMYC- (**A**) and for cMYC+/-/ and cMYC-/-/ (**B**) tumors, all compared with the +/- 2SD space of controls (see Fig S2). P values were determined by the unpaired two-tailed Student's t test comparing all samples within a given class (e.g. cMYC+) versus the entire control (all uninduced samples) pool and were as follows: Top) Tumor volume cMYC+ vs controls p=0.3. Number of metastases cMYC+ vs. controls p=0.3. Bottom) Tumor volume cMYC+/-/- vs controls p=0.9, and 3F+/-/- vs. controls p=0.8. Number of metastases cMYC+/-/- vs. controls p=0.8.



Supplementary Figure S4. Reprogramming-driven DNA methylation changes.

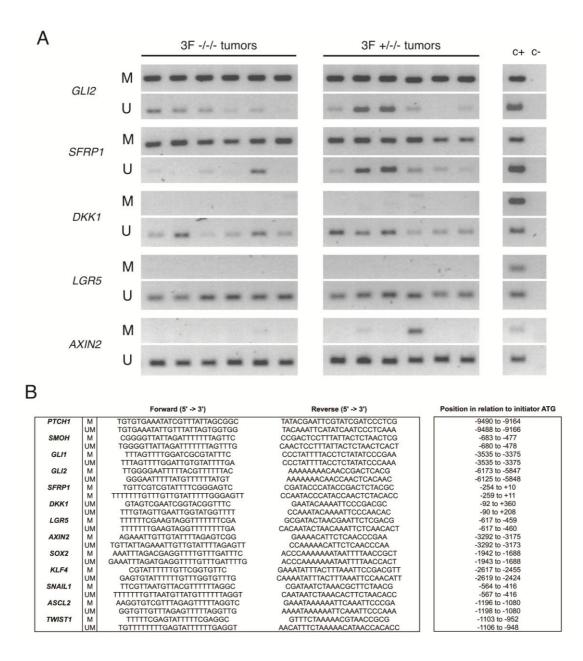
A) Reprogramming does not change the methylation of OCT4. Analysis was performed as for NANOG in Fig. 5; The upper diagram shows the position of analyzed CpG dinucleotides in the 5' promoter region of *OCT4*. The methylation status (represented in pie charts in the lower part of the diagram) was determined by bisulfite sequencing. Uninduced controls were pooled as there were no differences between 4F- vs. 3F- samples.

B) Quantification of methylation specific PCR (MSP) results for 3F+/-/- vs 3F-/-/- tumors (Figs. S4, S5) testing for both methylated and unmethylated status of selected CpG islands for different genes as indicated. The numbers of tumors analyzed (n) and the position of the islands (MSP primer region Fig. S5B) in relation to the initiator ATG are indicated.



Supplementary Figure S5. MSP analyses of the methylation status of CpG islands of selected genes.

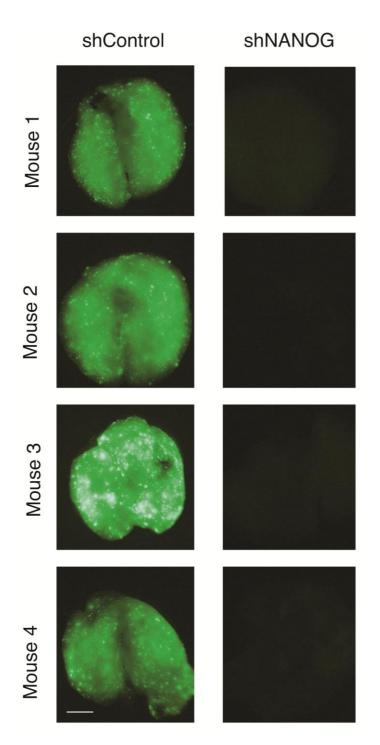
PCR reaction products for methylated (M) and unmethylated (U) states were loaded on agarose gels and photographed. The images show representative examples for the genes indicated in 4F+/-/- and 4F-/-/- tumors. Bands with asterisks denote primer dimers. (see Fig. 5)



Supplementary Figure S6. Gene regions analyzed by MSP.

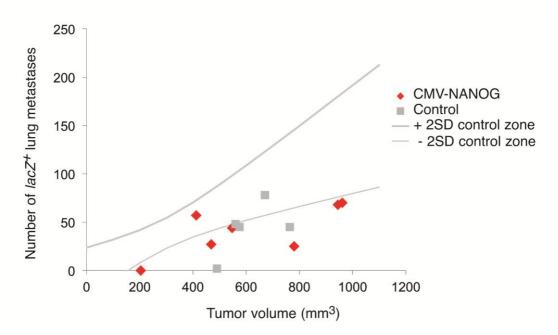
A) Examples of MSP reactions, amplified from 3F+/-/- and 3F-/-/- tumor DNA and run in agarose gels with positive fully methylated (c+) and negative fully unmethylated (c-) controls for individual genes as noted (see text). See Fig. S4 for of 4F+/-/- and 4F-/-/- MSP examples.

B) List of the position and sequences of MSP primers for selected genes as indicated.



Supplementary Figure S7. Loss of lung metastases driven by NANOG knockdown.

Panels show fluorescent images of dissected lungs from mice injected into the tail vein with control (*shControl*) and *shNANOG/P8* conditions. The left lung lobes were dissected and stained with XGAL to reveal β Gal⁺ metastases in great detail. Here the images show the overall GFP fluorescence from the *GFP*⁺/*LacZ*⁺ human colon cancer cells forming metastases. Note that cells with compromised NANOG fail to form GFP⁺ metastases. Scale bar = 3.2 mm for all panels.



Supplementary Figure S8. Overexpression of NANOG does not increase the number of metastases in the lungs from subcutaneous xenografts.

The graph shows the number of metastases and total tumor weight per animal for control CC14 and CC14-CMV-NANOG cells. Expression of NANOG over 100 fold was verified in the experimental xenografts at the end of the experiments by RT-qPCR (not shown).

Supplementary Table S1. RT-qPCR primers used.

Gene name	Direction	Sequence
NANOG	FW RV	AAATTGGTGATGAAGATGTATTCG GCAAAACAGAGCCAAAAACG
NANOGP8	FW RV	GCTGCCTTCAAGCATCTGTT TTTAAGAGGTGGACTGGAAAAA
OCT4 (5'UTR)	FW RV	GGGTTGAGTAGTCCCTTCGC TAGCCAGGTCCGAGGATCAA
SOX2 (5'UTR)	FW RV	GAGAGTGTTTGCAAAAGGGGG CGCCGCCGATGATTGTTATT
KLF4 (5'UTR)	FW RV	ATCTCGGCCAATTTGGGGTT TTGACGCAGTGTCTTCTCCC
GLI1	FW RV	AGCGTGAGCCTGAATCTGTG CAGCATGTACTGGGCTTTGAA
GLI2	FW RV	AGCAGCAGCAACTGTCTGAG CACATGAGCCGTGTCCAG
PTCH1	FW RV	GGCAGCGGTAGTAGTGGTGTTC TGTAGCGGGTATTGTCGTGTGTG
AXIN2	FW RV	AGTGTGAGGTCCACGGAA ACTGCCCACACGATAAGGAG
LGR5	FW RV	GGAGCATTCACTGGCCTTTA CTGGACGGGGATTTCTGTTA
сМҮС	FW RV	TGGTCTTCCCCTACCCTCTCAAC GATCCAGACTCTGACCTTTTGCC
DKK1	FW RV	TCCGAGGAGAAATTGAGGAA CCTGAGGCACAGTCTGATGA
SNAIL1	FW RV	AAGAGGCCTTCCCATGGCCATT GGGAGCTTCCCAGTGAGTCT
SNAIL2	FW RV	TTAGAACTCACACGGGGGAGAA GCTACACAGCAGCCAGATTC
ZEB2	FW RV	TCCTAATATTCCGCCTGTCG GGCATGAAAATGGAGTGGAT
FOXC2	FW RV	AGTTCATCATGGACCGCTTC GCTCCTCCTTCTCCTTGGAC
TWIST1	FW RV	GCCGGAGACCTAGATGTCATT CCCACGCCCTGTTTCTTTGA
HPRT	FW RV	AAGATGGTCAAGGTCGCAAG CTCCAGATGTTTCCAAACTCAAC
βΑCΤΙΝ	FW RV	ACAGAGCCTCGCCTTTGC GGAATCCTTCTGACCCATGC

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