Molecular characterization and silk gland expression of *Bombyx* engrailed and invected genes

(homeobox genes/compartments)

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ABSTRACT Genetic analysis in Drosophila has shown that engrailed (en) plays an important role in segmentation and neurogenesis. A closely related gene, invected (in), is coexpressed with en in the posterior developmental compartments where en is known to specify cell state. We report here the isolation of two en-like cDNAs from the middle silk glands of Bombyx mori larvae. Sequence analysis revealed that they are the counterparts of Drosophila en and in. Four highly conserved domains, including the homeodomain, were identified in these En and In proteins from Bombyx and Drosophila. In addition, two en-specific and one in-specific domains could also be found. These structurally homologous genes might share a similar role in Bombyx development. They were found to be coexpressed in the middle silk gland but not in the posterior silk gland during the fourth molt/fifth intermolt period. We speculate that these Bombyx en-like genes might be involved in the compartmentalization of the silk gland.

The Drosophila gene engrailed (en) was originally identified as a segmentation gene affecting cells in the posterior compartments (1-3). It was later found to be required also for the development of the central nervous system (4). Like many other developmental control genes in Drosophila (5), en is a homeobox-containing gene (6, 7). It encodes a nuclear protein that binds DNA specifically through its homeodomain (8) and can function as a transcription factor *in vitro* (9-11). A closely related gene, invected (*in*), was also identified in Drosophila that shares extensive homology with en and lies within 20 kilobases (kb) of en (12). Both *in* and en are expressed in the cells of the posterior compartments early in development and later in the central nervous system (12). The roles of *in*, if any, remain obscure.

Based on sequence similarity, en homologs have been isolated from many invertebrate and vertebrate species (13-21). Expression studies revealed that *en* probably plays a role in neurogenesis (14, 18, 21-28), as has been shown by genetic analysis in Drosophila (4). It might play roles both in compartmentalization of the developing neural tube and specification of particular neuronal populations. Recently, it was found that mice homozygous for a targeted mutation of one of the en-like genes, En-2, show defects in the development of the folia in the cerebellum providing further support for the neurogenetic function of en (29). During the process of segmentation, en is expressed in the posterior portion of each metamere in all arthropod species examined so far suggesting that the segmentation function of Drosophila en might be conserved in other arthropods (14, 30-32). In this respect, en expression has been used as a molecular marker to directly compare mechanisms of segmentation (32).

We have been analyzing the regulation of silk protein gene expression in *Bombyx mori* (for review, see ref. 33). Recently, we reported that the promoters of the silk protein genes contain clustered homeodomain binding sites (34, 35) and that several putative homeodomain-containing proteins that interact with these sites can be found in the silk gland extracts (36). As part of our efforts to characterize the homeobox genes that are possibly involved in the regulation of the silk protein genes, several homeobox-containing cDNAs have been isolated from silk gland cDNA libraries. In this report, we describe the molecular characterization of two Bombyx en-like cDNAs. Sequence analysis reveals that they are the counterparts of Drosophila en and in. We have named the corresponding genes Bombyx engrailed (Bm en) and Bombyx invected (Bm in), respectively. Throughout the fourth molt/fifth intermolt period, they are coexpressed in the middle silk gland but not in the posterior silk gland. A hypothesis that these en-like genes are involved in specifying compartments in the silk gland is discussed.

MATERIALS AND METHODS

Animals. B. mori eggs, from a Japanese strain (Kin-Shu), a Chinese strain (Sho-Wa), and a hybrid between them (Kin-Shu \times Sho-Wa) were purchased from Kanebo Silk (Kasugai City, Japan). Larvae were maintained aseptically at 25°C on an artificial diet and staged as described (37).

Cloning and Sequencing of cDNAs for *Bombyx en* and *in*. λ gt11 cDNA libraries were constructed from poly(A)⁺ RNA of the middle silk gland of 2-day-old fifth-instar B. mori larvae (Kin-Shu \times Sho-Wa) by standard procedures (38). Five Bm en and one Bm in clones were isolated by screening a random-primed cDNA library under conditions of low stringency at 30°C with an oligonucleotide probe, 5'-GAGGCG-CAGATCAAGATCTGTTCCAGAACAGGCGGGCC-AA-3', that spans the putative recognition helix of the en homeodomain (5). The hybridization buffer consisted of 50% (vol/vol) formamide, 5× SSC (1× SSC is 0.15 M NaCl/15 mM sodium citrate), 5× Denhardt's solution (1× = 0.02%bovine serum albumin/0.02% Ficoll/0.02% polyvinylpyrrolidone), 0.1% SDS, 5 mM sodium phosphate (pH 6.5), and denatured salmon testes DNA (250 μ g/ml). By rescreening libraries with the 5' portion of these isolated cDNA fragments, four additional Bm en and three additional Bm in clones were chosen for further characterization. The cDNA inserts were subcloned into pBluescript II KS(-) vector (Stratagene) and sequenced by the dideoxynucleotide method using the Sequenase protocol (United States Biochemical). Genomic fragments that contain the cDNA se-

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[‡]The sequences reported in this paper have been deposited in the GenBank data base (accession nos. M64335 and M64336).

quences corresponding to $Bm \ en$ and $Bm \ in$ were isolated from genomic λ and Cosmid libraries (C.-c.H. and K.U., unpublished) and sequenced with specific primers derived from the cDNA sequences.

RNA Extraction and Blot-Hybridization (Northern) Analysis. Total RNA was isolated by using an acid guanidinium thiocyanate/phenol/chloroform method (39) and poly(A)⁺ RNA was enriched by oligo(dT)-cellulose chromatography. The RNA was electrophoresed on 1% agarose/1.1 M formaldehyde gels (38) and transferred onto nylon membranes (Biodyne Electronics, Santa Monica, CA; Pall). Blots were hybridized with probes specific for *Bm en* and *Bm in* at 45°C, in 50% formamide/5× SSC/5× Denhardt's solution/0.1% SDS/5 mM sodium phosphate (pH 6.5)/denatured salmon testes DNA (250 μ g/ml). A control cDNA probe that hybridizes with an abundantly expressed transcript of unknown identity (C.-c.H. and Ping-Xian Xu, unpublished data) was also used to check the integrity of RNA.

RESULTS AND DISCUSSION

Cloning of cDNAs for Bombyx en and in. Low-stringency hybridization with oligonucleotide probes that code for the putative recognition helix of the Antennapedia and en homeodomains (5) yielded several positive clones in a screen of a middle silk gland cDNA library from 2-day-old fifth-instar larvae. Among them, two types of en-positive clones were found. These cDNA fragments were used to rescreen cDNA libraries and several overlapping clones were characterized by nucleotide sequence analysis. These analyses revealed that they correspond to the Bombyx homologs of Drosophila en and in (Bm en and Bm in, respectively).

The nucleotide and amino acid sequences of a composite cDNA molecule of Bm en are shown in Fig. 1. The 3363base-pair sequence reveals an open reading frame that yields a protein of 372 amino acids with a deduced molecular mass of 42 kDa. The methionine residue at nucleotide 379 is designated as the initiator because the N-terminal region of the predicted protein shows a strong similarity with that of the Drosophila En protein (see below). As shown in Fig. 2, the sequence of Bm in reveals an open reading frame that yields a protein of 476 amino acids with a deduced molecular mass of 54 kDa. We have chosen the methionine residue at nucleotide 251 to be the initiator because it is the first methionine residue after many termination codons and the sequence around it (AGAACGAAAATGGCC) is very similar to the sequence near the Bm en initiator (AacAC-GAgAATGGCC, where lowercase letters indicate the difference between the two sequences). No polyadenylylation signal or poly(A) tail can be found at the 3' ends of these cDNA clones. The transcripts of cDNAs for Bm en and Bm in are 5.1 and 6.2 kb long, respectively, indicating that these cDNA clones are only partial (see Fig. 5A).

Conserved Domains in the En and In Proteins. As summarized in Fig. 3, sequence comparison unambiguously revealed a distinction between the two En proteins and the two In proteins from *Bombyx* and *Drosophila*. In addition to the homeodomain, three conserved domains (I, II, and III) can be found in these four insect En-like proteins. The two En proteins share two additional conserved domains, one located at the N terminus and the other in the central part of the proteins. Near the N-terminal region of the two In proteins, one In-specific domain can be recognized. Furthermore, an Arg-Ser dipeptide sequence just after region II is only found in the two In proteins (Fig. 4A). As will be discussed below, this might serve as a hallmark of In proteins.

Among the four conserved domains found in these four insect En-like proteins, the homeodomain and regions II and III have been shown (16) to be present in *Drosophila* En and In and in mouse En-1 and En-2 proteins. Region I, which is

1 64 127 190 253 316	GCGC TCAC GCGC ACTC TTA/ ACAC	CAGTO CGCGO GCAA/ CGCGTO ATC/ CCGTO	GCCAC CCCGC AGGA1 ICAAA ACGCC CACGC	GTCC CGTC CCA1 CCA1 CCA1 CCA1 CCA1 CCA1	GCGT TACA GCTA CGAC CACGG	CAAA CGAC ATGA CGCC AACC	GAAA GCTC GCCTC GGGGC GCGTC	AAGT AAAA GAGCC GAA1 GACCC GCGC	GACO AAAA TCCO TCGA GCA1 TGGO	TATC AAAA TATCG TGTG CTCGC	GATT GATT GTCG CTCT TGCG	TGAC GTGT AATA CGGA CCGT	AGCI CGTC TTAA CATI CCCCA	CCGG CGTC TGAC AGTG ACGA	GTGT GTGT ATTC GAGG TGAA	CTG CGA GAA CAT CAA AGA
379	ATG	GCC	TTC	GAG	GAC	CGC	TGC	AGC	CCT	AGC	CAG	GCC	AAC	AGT	CCG	GGA
1	Met	Ala	Phe	Glu	Asp	Arg	Cys	Ser	Pro	Ser	Gln	Ala	Asn	Ser	Pro	Gly
427	CCG	GTG	ACG	GGG	CGA	GTC	CCG	GCG	CCG	CAC	GCC	GAG	ACC	CTC	GCA	TAC
17	Pro	Val	Thr	Gly	Arg	Val	Pro	Ala	Pro	His	Ala	Glu	Thr	Leu	Ala	Tyr
475	AGC	CCG	CAG	AGC	CAG	TAC	ACT	TGC	ACC	ACA	ATA	GAA	TCG	AAG	TAC	GAA
33	Ser	Pro	Gln	Ser	Gln	Tyr	Thr	Cys	Thr	Thr	Ile	Glu	Ser	Lys	Tyr	Glu
523	CGA	GGT	TCT	CCG	AAC	ATG	ACA	ATT	GTG	AAG	GTG	CAG	CCG	GAC	TCC	CCG
49	Arg	Gly	Ser	Pro	Asn	Met	Thr	Ile	Val	Lys	Val	Gln	Pro	Asp	Ser	Pro
571	CCT	CCC	AGC	CCG	GGA	CGC	GGT	CAG	AAC	GAG	ATG	GAA	TAC	CAG	GAC	TAC
65	Pro	Pro	Ser	Pro	Gly	Arg	Gly	Gln	Asn	Glu	Met	Glu	Tyr	Gln	Asp	Tyr
619	TAC	CGC	CCT	GAA	ACG	CCC	GAC	GTA	AAG	CCT	CAT	TTT	AGC	CGG	GAG	GAG
81	Tyr	Arg	Pro	Glu	Thr	Pro	Asp	Val	Lys	Pro	His	Phe	Ser	Arg	Glu	Glu
667	CAG	AGG	TTT	GAA	CTG	GAC	AGA	TCG	AGG	GGG	CAG	CGG	CTG	CAA	CCC	ACC
97	Gln	Arg	Phe	Glu	Leu	Asp	Arg	Ser	Arg	Gly	Gln	Arg	Leu	Gln	Pro	Thr
715	ACG	CCG	GTC	GCC	TTC	TCC	ATA	AAC	AAC	ATC	CTG	CAC	CCT	GAG	TTC	GGC
113	Thr	Pro	Val	Ala	Phe	Ser	Ile	Asn	Asn	11e	Leu	His	Pro	Glu	Phe	Gly
763	TTG	AAC	GCC	ATC	AGG	AAA	ACG	AGC	AAA	ATC	GAA	GGT	CCC	AAG	CCC	ATT
129	Leu	Asn	Ala	Ile	Arg	Lys	Thr	Ser	Lys	Ile	Glu	Gly	Pro	Lys	Pro	Ile
811	GGA	CCG	AAC	CAC	AGT	ATC	CTC	TAT	AAA	CCT	TAC	GAT	TTA	TCC	AAA	CCG
145	Gly	Pro	Asn	His	Ser	Ile	Leu	Tyr	Lys	Pro	Tyr	Asp	Leu	Ser	Lys	Pro
859	GAC	TTA	TCG	AAA	TAC	GGC	TTT	GAT	TAT	TTG	AAG	AGT	AAG	GAA	ACG	AGT
161	Asp	Leu	Ser	Lys	Tyr	Gly	Phe	Asp	Tyr	Leu	Lys	Ser	Lys	Glu	Thr	Ser
907	GAT	TGC	AAC	GCT	TTG	CCG	CCT	TTA	GGA	GGG	TTG	AGG	GAG	ACG	GTA	TCG
177	Asp	Cys	Asn	Ala	Leu	Pro	Pro	Leu	Gly	Gly	Leu	Arg	Glu	Thr	Val	Ser
955	CAG	ATC	GGC	GAA	CGC	TTG	TCC	AGA	GAC	AGG	GAG	CCT	CCG	AAG	AGT	CTG
193	Gin	Ile	Gly	Glu	Arg	Leu	Ser	Arg	Asp	Arg	Glu	Pro	Pro	Lys	Ser	Leu
1003	GAG	CAG	CAG	AAG	AGG	CCC	GAT	TCC	GCG	AGT	TCC	ATC	GTC	TCG	TCG	ACG
	Glu	Gln	Gln	Lys	Arg	Pro	Asp	Ser	Ala	Ser	Ser	Ile	Val	Ser	Ser	Thr
1051	TCG	AGC	GGC	GCG	GTT	TCC	ACC	TGC	GGA	AGC	TCG	GAC	GCG	AGC	TCC	ATC
225	Ser	Ser	Gly	Ala	Val	Ser	Thr	Cys	Gly	Ser	Ser	Asp	Ala	Ser	Ser	Ile
1099	CAG	TCC	CAG	AGC	AAC	CCT	GGC	CAG	CTG	TGG	CCA	GCC	TGG	GTA	TAC	TGC
241	Gln	Ser	Gln	Ser	Asn	Pro	Gly	Gln	Leu	Trp	Pro	Ala	Trp	Val	Tyr	Cys
1147	ACC	AGA	TAC	AGC	GAC	CGA	CCT	AGT	TCC	GGT	CCC	AGA	AGC	AGA	AGG	GTC
257	Thr	Arg	Tyr	Ser	Asd	Arg	Pro	Ser	Ser	Gly	Pro	Arg	Ser	Arg	Arg	Val
1195	AAA	AAG	AAA	GCA	GCG	CCC	GAA	GAG	AAG	AGA	CCA	AGG	ACC	GCT	TTC	AGC
273	Lys	Lys	Lys	Ala	Ala	Pro	Glu	Glu	Lys	Arg	Pro	Arg	Thr	Ala	Phe	Ser
1243	GGG	GCA	CAA	CTC	GCG	AGA	TTG	AAG	CAC	GAG	TTC	GCC	GAG	AAC	CGG	TAT
289	Gly	Ala	Gln	Leu	Ala	Arg	Leu	Lys	His	Glu	Phe	Ala	Glu	Asn	Arg	Tyr
1291	CTG	ACT	GAG	CGG	CGG	CGG	CAG	AGC	CTC	GCG	GCG	GAG	CTG	GGC	CTC	GCC
305	Leu	Thr	Glu	Arg	Arg	Arg	Gln	Ser	Leu	Ala	Ala	Glu	Leu	Gly	Leu	Ala
1339	GAG	GCG	CAG	ATC	AAG	ATT	TGG	TTC	CAG	AAC	AAA	CGG	GCC	AAG	ATC	AAG
321	Glu	Ala	Gln	11e	Lys	Ile	Trp	Phe	Gln	Asn	Lys		Ala	Lys	Ile	Lys
1387	AAG	GCG	TCG	GGC	CAG	AGG	AAC	CCG	CTG	GCG	TTG	CAG	CTC	ATG	GCC	CAG
337	Lys	Ala	Ser	Gly	Gln	Arg	Asn	Pro	Leu	Ala	Leu	Gln	Leu	Met	Ala	Gln
1435	GGT	CTC	TAC	AAC	CAC	AGC	ACC	GTC	ACG	GAG	AGC	GAC	GAC	GAA	GAG	GAG
353	Gly	Leu	Tyr	Asn	His	Ser	Thr	Val	Thr	Glu	Ser	Asp	Asp	Glu	Glu	Glu
1483 369	ATC Ile	AA1 Asi	GTT Val	ACG Thr	TAA	AGG	AGAT	GGTG	CGAT	TGTT	TTGG	TGAC	GTCA	CGAT	AACG	GTTT
1541 1604 1667	TCG GAA	GGAA	ATTI	TTAG	TTTG GACG	GAAC	ACCT	TCGT	GGTA	ATTG	TGTT	TACT GACC	TTCT GTAA	CGAA TGCA	AGGG	AAAC
1730 1793	AAG	TTTC	TTTT	CGGG	TTTA	GCTA	CTTC	CAAA	TGTT	GCCA	TATG	AAAA TGTA	TACA	TAAT	TTTT	CGGT
1856	TCT CTA	TACI	GATI	AATA	TATT	TATT	TGTA	ATAG	ATTA	ATGG	CAAT	AGTC	AAGT	ACAA	GTTA	GTTC AAAA
2045	CAC	TTG	TGTA	TTGG	CAAA		TACG	GACT	CATT	TATA	TTTG	ACAC	AGGA	CAAA	ATTG	ACAG
2171		AATA	GAAT	CGTT	TGGA	ATCA	CTCA	AGTC	TATG	GAGT	AGTT	TCGT	GCTG	TCCC	ACGG	AGAC
2297	ATC	AGAG	TTTT	TGTA	CTGC	TGAC	TACA	TCCA		TAGT	TTTT	TAAA	AATA	CGAC	TCTG	ACGT
2423	TCI	TCAC	CAGI	TGAC	AATG	CTTC	ATCA	TATT	CCTC	TCAG	TTTC	CTCG	GTAA	AAGT	ACTC	GAAA
2549		AAT	CATI	AAAI	GAAG	TAAA	CAAA	TAAT	GTTA	TTGG	AAAT	TTAA	AAAA		TCAT	CGAT
2675	TC/	TAA	TTA	TTAA	TATT	TCTT	TTGG	AATC	TTAC	GAAA	AAAA	CAAC	AGAA	CCGA	TTTA	AATT
2738	AA1	TGA/	TCA1	TGCG	ACGT	AAGG	TTCA	TACT	GTCG	TACT	TTTT	AAGA TAA1	TAAG	ATAA TGCT	GCGA	GGTG TATC
2864 2927	AT/	ACC/		GATA	AACT	ATTA	GTTT	AGCT	AAAC	GTAA	CGTT	TTAG TTAA	TAAA	AGTA	GCAA	TGAA TAAA
2990 3053) TAA	AAAI	TAAA1	ATTG	TTTT CTCA	ATAA	CGAA	AAAG	CACG	AGCA	TCTT	AATG	CCGA	ACTA	ATCT	GTTT
3116	ČC1	CGAT	ACGO	GTGC	TAAT	TTTT	AAGT	TAAT	CGGA	CGTT	TTTT	TTTG	TGAA	ATCA	TATA	CTTT
3242	AAA	GCG/		ACTI	TATG	ACGI	ATCO	TCGT	TGTO		CAAA	ATCI	GCTA	TGTGA	CAAA	AACT
-	-															

FIG. 1. Nucleotide and deduced amino acid sequences of *Bombyx en*. The nucleotide sequence of a composite cDNA for *Bm en* is shown with the deduced amino acid sequence of the Bm En protein. The homeodomain is boxed and the three homologous domains (regions I, II, and III in Fig. 3) are marked with heavy underlines. The two En-specific domains (striped boxes in Fig. 3) are underlined and the positions of the introns are indicated by arrowheads.

located in the N-terminal half of these proteins, is also conserved in the vertebrate En-like proteins (refs. 14 and 21;

62 125 188	AG TTCG GATA ATAC	ATC/ ATAC AGC/	CGCG TCGA CGGT CGCAC	GACA GACA GATO	ACA1 TTAC GTG1 AAG1	TCA/	AGA/	TTTG TAAC	AAAAC STTTI CATTO SGTAA	TCG1	GTT	TCT/	GGCC GTGC1 GTGAC	ACCO TTTT GGAG	TAAC TTTT GTTC	GCA TTGT GAGG GAAA
251	ATG	GCC	GCT	GTA	TCC	GCC	CAT	ATG	CAG	GAC	ATT	AAG	ATC	CAA	GAC	CAG
1	Met	Ala	Ala	Val	Ser	Ala	His	Met	Gln	Asp	Ile	Lys	Ile	Gln	Asp	Gln
299	AGC	GAT	GAC	GAT	CCA	ТАС	TCT	CCG	AAC	ACG	AGA	GAC	ACG	ACA	AGT	CCA
17	Ser	Asp	Asp	Asp	Pro	Туг	Ser	Pro	Asd	Thr	Arg	Asp	Thr	Thr	Ser	Pro
347	GAG	TGC	CAC	GAC	GAT	GAG	AAA	TCG	GAA	GAC	ATA	AGC	ATC	CGT	TCA	TCC
33	Glu	Cys	His	Asp	Asp	Glu	Lys	Ser	Glu	Asp	Ile	Ser	Ile	Arg	Ser	Ser
395	TCT	TTC	TCC	ATC	CAC	AAC	GTG	CTT	AGA	AGG	AGC	GGG	ACA	ACA	GCA	GCC
49	Ser	Phe	Ser	Ile	His	Asn	Val	Leu	Arg	Arg	Ser	Gly	Thr	Thr	Ala	Ala
443	CTG	ACA	ATG	TCT	TTT	CGA	CGG	AAA	AGC	TCT	TGG	AGA	ATC	CCG	AAT	TTC
65	Leu	Thr	Met	Ser	Phe	Arg	Arg	Lys	Ser	Ser	Trp	Arg	Ile	Pro	Asn	Phe
491	GAT	GAC	AGA	AAC	ACA	GAG	AGT	GTA	AGT	CCC	GTT	GTT	GAA	GTG	AAT	GAA
81	Asp	Asp	Arg	Asn	Thr	Glu	Ser	Val	Ser	Pro	Val	Val	Glu	Val	Asd	Glu
539	AGA	GAA	ATA	AGC	GTG	GAC	GAT	GGT	AAT	TCT	TGC	TGT	AGC	GAC	GAC	ACC
97	Arg	Glu	Ile	Ser	Val	Asp	Asp	Gly	Asn	Ser	Cys	Cys	Ser	Asp	Asp	
587	GTG	TTG	TCA	GTT	GGA	AAC	GAG	GCA	CCC	GTA	TCC	AAC	TAC	GAA	GAG	AAA
113	Val	Leu	Ser	Val	Gly	Asp	Glu	Ala	Pro	Val	Ser	Asn	Tyr	Glu	Glu	Lys
635	GCC	AGC	CAG	AAT	ACC	CAC	CAA	GAA	CTG	ACC	TCC	TTC	AAA	CAC	ATA	CAA
129	Ala	Ser	Gln	Asn	Thr	His	Glu	Glu	Leu	Thr	Ser	Phe	Lys	His	Ile	Gln
683	ACA	CAC	TTG	AGC	GCC	ATA	TCG	CAG	CTG	AGC	CAA	AAC	ATG	AAT	GTG	GCC
145	Thr	His	Leu	Ser	Ala	Ile	Ser	Gln	Leu	Ser	Gln	Asn	Met	Asn	Val	Ala
731	CAA	CCG	CTG	CTA	TTA	CGG	CCG	AGT	CCA	ATT	AAC	CCA	AAC	CCA	ATA	ATG
161	Gln	Pro	Leu	Leu	Leu	Arg	Pro	Ser	Pro	Ile	Asn	Pro	Asn	Pro	Ile	Met
779	TTC	CTA	AAC	CAA	CCG	CTT	CTG	TTC	CAA	AGT	CCG	ATC	TTG	AGC	CAA	GAC
177	Phe	Leu	Asn	Gln	Pro	Leu	Leu	Phe	Gln	Ser	Pro	Ile	Leu	Ser	Gln	Asp
827	TTA	AAA	GGT	ATG	CCC	AAC	AGA	CAA	ACA	GCC	AAC	GTG	ATC	AGT	CCA	ACG
193	Leu	Lys	Gly	Met	Pro	Asn	Arg	Gln	Thr	Ala	Asn	Val	Ile	Ser	Pro	Thr
875	TTT	GGC	TTA	AAT	TTC	GGT	ATG	AGA	TTG	AAG	GCC	AAT	CAT	GAA	ACA	CGA
209	Phe	Gly	Leu	Asn	Phe	Gly	Met	Arg	Leu	Lys	Ala	Asd	His	Glu	Thr	Arg
923	ACG	AGG	TCT	GAT	GAG	AAT	CGG	TAT	TCG	AAG	CCG	GAA	GAA	TCT	AGA	GAT
225	Thr	Arg	Ser	Asp	Glu	Asn	Arg	Tyr	Ser	Lys	Pro	Glu	Glu	Ser	Arg	Asp
971	TAC	ATC	AAT	CAG	AAC	TGC	CTT	AAG	TTT	AGC	ATA	GAT	AAT	ATT	TTA	AAA
241	Tyr	Ile	Asd	Gln	Asb	Cys	Leu	Lys	Phe	Ser	Ile	Asp	Asn	Ile	Leu	Lys
1019	GCG	GAC	TTC	GGA	AGG	AGG	ATC	ACC	GAT	CCT	TTG	CAC	AAA	AGG	AAA	GTG
257	Ala	Asp	Phe	Gly	Arg	Arg	Ile	Thr	Asp	Pro	Leu	His	Lys	Arg	Lys	Val
1067	AAG	ACG	AGA	TAC	GAG	GCT	AAA	CCT	GCT	CCA	GCA	AAA	GAC	АСТ	GCG	GCT
273	Lys	Thr	Arg	Tyr	Glu	Ala	Lys	Pro	Ala	Pro	Ala	Lys	Asp	Тћг	Ala	Ala
1115	TTT	GCT	CCG	AAG	CTG	GAC	GAA	GCG	AGG	GTA	CCT	GAC	ATC	AAA	ACA	CCA
289	Phe	Ala	Pro	Lys	Leu	Asp	Glu	Ala	Arg	Val	Pro	Asp	Ile	Lys	Thr	Pro
1163	GAC	AAA	GCT	GGA	GCC	ATC	GAC	CTT	TCT	Lys	GAC	GAT	AGC	GGA	AGC	AAT
305	Asp	Lys	Ala	Gly	Ala	Ile	Asp	Leu	Ser		Asp	Asp	Ser	Gly	Ser	Asd
1211	TCT	GGA	TCA	ACC	TCC	GGT	GCA	ACT	TCA	GGC	GAC	AGT	CCG	ATG	GTG	TGG
321	Ser	Gly	Ser	Thr	Ser	Gly	Ala	Thr	Ser	Gly	Asp	Ser	Pro	Met	Val	<u>Trp</u>
1259 337	CCC Pro	GCG Ala	TGG Trp	GTG Val	TAC Tyr	TGT Cys	ACG Thr	AGG Arg	TAC Tyr	AGC	GAT Asp	CGA	CCC Pro	AGT Ser	TCC Ser	GGA Gly
1307	AGA	AGT	CCT	CGC	ACC	AGA	CGA	CCG	AAG	AAG	CCG	CCC	GGA	GAC	ACC	GCC
353	Arg	Ser	Pro	Arg	Thr	Arg	Arg	Pro	Lys	Lys	Pro	Pro	Gly	Asp	Thr	Ala
1355	AGC	AAT	GAC	GAG	AAG	AGA	CCA	AGG	ACC	GCA	TTC	TCC	GGA	CCA	CAG	CTC
369	Ser	Asn	Asp	Glu	Lys	Arg	Pro	Arg	Thr	Ala	Phe	Ser	Gly	Pro	Gln	Leu
1403	GCG	AGG	CTA	AAG	CAC	GAG	TTC	GCG	GAG	AAC	CGG	TAT	CTG	ACA	GAG	CGG
385	Ala	Arg	Leu	Lys	His	Glu	Phe	Ala	Glu	Asn	Arg	Tyr	Leu	Thr	Glu	Arg
1451	CGG	CGG	CAG	AGC	CTC	GCG	GCG	GAG	CTG	GGC	CTC	GCC	GAG	GCG	CAG	ATC
401	Arg	Arg	Gln	Ser	Leu	Ala	Ala	Glu	Leu	Gly	Leu	Ala	Glu	Ala	Gln	Ile
1499	AAG	ATC	TGG	TTC	CAG	AAC	AAA	CGG	GCC	AAG	ATC	AAG	AAG	GCG	TCG	GGC
417	Lys	Ile	Trp	Phe	Gln	Asn	Lys		Ala	Lys	Ile	Lys	Lys	Ala	Ser	Gly
1547	CAG	AGG	AAC	CCA	CTG	GCA	CTG	CAG	CTC	ATG	GCC	CAG	GGC	CTC	TAC	AAC
433	Gln	Arg	Asn	Pro	Leu	Ala	Leu	Gln	Leu	Met	Ala	Gln	Gly	Leu	Tyr	Asn
1595	CAC	AGC	ACC	GTG	CCG	CTG	ACA	AAG	GAA	GAG	GAG	GAA	TTA	GAG	ATG	AAG
449	<u>His</u>	Ser	Thr	Val	Pro	Leu	Thr	Lys	Glu	Glu	Glu	Glu	Leu	Glu	Met	Lys
1643 465	GCG Ala	AGA Arg	GAA Glu	AGA Arg	GAG Glu	AGA Arg	GAG Glu	CTG Leu	AAG Lys	AAT	AGA Arg	TGT Cys	TAA	AAC	GGCT	TTCA
1693	GTA	AGAG	TGGT	AGTG	TGTT	CTTG	GTAA	TACT	AGTC	AAAC	TTAC	TTAC	CTTA	CAAC	CCAA	AACT
1756	TAC	TCTT	TTAC	TGGT	GGTA	GGAC	CACT	TGTG		CGCG	CGGA	TAGG	TACC	ACCA	CCCT	GCTT
1819	ATT	TCTG	CCGT	AAAG	CAGT	AATG	CGTT	TCGG		AAGG	GTGG	GGCA	GTCG	TTGT	AACT	ATAC
1882	TGA	даат	TTAG	AACT	TGTA	TCTC	AAGG	TGGG	GCGC	GTTT	ACGT	TGTA	GATG	TCTA	TGGG	CTCC
1945	AGT	Аасс	ACTT	AACA	CCAG	GTGG	GCTG	TGAG	CTCG	TCCA	CCCA	TCAA	ATCA	ATTT	CTAA	ATTT
2008	CAT	Тата	AGCG	AAGT	TATT	TTCT	AATG	TTTG	CCAA	ACCG	AAAA	TAAT	TCTA	ATAA	CACG	TTGA
2071 2134 2197	CTG TCG		TCAC ATTT	CGGT CCTA	GCCC ATTG	TACG TTTC	CGAC TTTA	GCAC AGCT	TGAA GCAA	GTAA GAAG	TTAT CTCT	AATA	GCAG AAGC	TTTT	CATA	ACAG ACAG
2260 2323	TTT	CTTT	TATT	GCTT	GTAA	CCCT	GGAC	GAGC	TCAC	corr	CACT	TGAT	CTTT	ACTO	CTT.	0000
	AGC	CCAT	AGAC	ATCT	ACAA	CGTA	AATG	CGGC	TACC	CACC	TTGA	GACA	CAAG	TTGT	AAGG	TCTC
2386	AGC ACT GGG	CCAT TTTA TGGT	AGAC ACAG GGTA	ATCT TACA GCTA	ACAA ACTG CCCG	CGTA CTGC TGCG	AATG AATT GACT	CGGC CAAA CACA	TACC CCGA AGAC	CACC	TTGA CATT TACC	GACA ACTG ACCA	CAAG CTTC GTAA	TTGT ACGG TTTT	AAGG CAGG AGAT	TCTC CATA TGAA
2386 2449 2512 2575 2638	AGC ACT GGG TTC. TGA ATT	CCAT TTTA TGGT AATT AAGC TTAT	AGAC ACAG GGTA TTAC GCCA TAAT	ATCT TACA GCTA AAAG TTGC CAAA	ACAA ACTG CCCG GCTA GACC TCGA	CGTA CTGC TGCG TAAT CATC AAAT	AATG AATT GACT ATTG AGAT TTAA	CGGC CAAA CACA TTAT GTCG CTTC	TACC CCGA AGAC CGTT GTGG TGCC	CACC AACG ATCC TTCG ACAA	TTGA CATT TACC AGAA CACT	GACA ACTG ACCA CTAT ATAC	CAAG CTTC GTAA GCGT CGCC CGCC	TTGT ACGG TTTT GAAC CAGT	AAGG CAGG AGAT ACTC ATTG	TCTC CATA TGAA TTAC AATT AGCA

2827 CTTAAACATGCGTTATAGATTTATCAACACCCTACCTCGTGCCGAATTC

FIG. 2. Nucleotide and deduced amino acid sequence of *Bombyx* in. The nucleotide sequence of the longest cDNA clone for Bm in is shown with the deduced amino acid sequence of the Bm In protein. An In-specific domain (solid box in Fig. 3) is underlined. Other symbols are as described in Fig. 1.

A. L. Joyner, personal communication). This region spans a conserved core of 12 amino acids with additional similarities in the flanking sequences between the two En proteins and between the two In proteins (Fig. 4A). In particular, region I of the Bombyx and Drosophila In proteins is highly conserved (15 out of 16 residues are identical). Region II spans a region of 17 amino acids and is the most conserved region (it is identical in all En-like proteins examined so far). The homeodomain of the two Bombyx proteins shows very high sequence similarity (59 out of 60 amino acids are identical) as compared with the two Drosophila proteins (52 out of 60 amino acids are identical). The two honeybee en-like genes, E30 and E60, also have only one amino acid difference in their homeodomain (13). Apparently, Drosophila en and in homeodomains are more divergent than other En-like proteins present within the same species (13, 14, 16). Region III spans a region of 18 amino acids and is also highly conserved in various organisms. Region III is identical in the two Bombyx En-like proteins as well as the two honeybee en-like genes (13)

The N terminus is probably an important feature of the insect En proteins because it is highly conserved between the Bombyx and Drosophila En proteins (11 of 15 amino acids are identical; Fig. 4B) and is identical in the Drosophila melanogaster and Drosophila virilis En proteins (40). Though the two Drosophila En proteins are very similar, an intervening stretch of 31 amino acids can be found in the D. virilis En protein to demarcate this N-terminal region and the next conserved region in the Drosophila proteins. This conservation of the N-terminal region is apparently confined to these insect En proteins because no significant similarity in this region can be found in the two insect In proteins and other vertebrate En-like proteins (ref. 21; A. L. Joyner, personal communication). Another conserved region in the insect En proteins is near the center of the proteins (8 out of 12 amino acids are identical between Bombyx and D. melanogaster, and 9 out of 12 amino acids are identical between Bombyx and D. virilis; Fig. 4B). Besides these conserved regions, the Bombyx and Drosophila En proteins also show an indication of sequence similarity in a region between the central Enspecific domain and region II. Though their sizes are very different, both regions possess a high serine content (16 out of 54 amino acids in Bombyx and 38 out of 112 amino acids in Drosophila). This serine-rich region of the Drosophila En protein has been suggested to be the site for posttranslational modification by a serine-specific protein kinase (41). Between the two insect In proteins, only an additional conserved region of 14 amino acids (8 out of 14 amino acids are identical) can be identified near the N terminus (Fig. 4B).

Despite extensive homologies between the two En proteins, the Bm En protein lacks some prominent features of the Drosophila En protein, like the polyglutamine and polyalanine stretches (6). It has been suggested that en might play a similar role in body segmentation in all insects (14, 30-32). Consistent with this hypothesis, Bm en and Bm in are expressed during the process of segmentation in the embryo (C.-c.H., unpublished data). If we assume that the insect En proteins perform a similar function in transcriptional regulation, the poly(amino acid) stretches that have been proposed as transcriptional regulatory domains in the Drosophila En protein might be dispensable in Bombyx. In this respect, it is worthwhile to mention that the two Xenopus En-2 proteins also lack these polyglutamine and polyalanine stretches (21). In contrast, the conserved regions reported here are likely to be important structural and/or functional domains in these En-like proteins.

Phylogeny of en and in. Southern blot hybridization revealed that *Bm* en and *Bm* in are the only en-like genes in the *Bombyx* genome (data not shown). Genomic clones harboring these cDNA sequences were isolated and partially charac-



FIG. 3. Schematic representation of the *Bombyx* and *Drosophila* En-like proteins. The *B. mori* En (Bm en) and In (Bm in) and the *D. melanogaster* En (Dm en) and In (Dm in) proteins are portrayed with various conserved domains indicated. The positions of the four conserved domains found in all these En-like proteins, the homeodomain (HD), and regions I-III are represented by stippled boxes aligned with dotted lines. The striped boxes indicate the positions of the En-specific domains and the solid boxes indicate the positions of the In-specific domains (see text for more details).

terized by nucleotide sequence analysis using specific primers derived from the cDNAs. These analyses revealed the presence of two introns in the protein coding region of $Bm \ en$ at exactly the same positions as in *Drosophila en* (Fig. 1). Similar to *Drosophila in*, an additional intron is found between the region II and the homeobox region of $Bm \ in$ (Fig. 2). This indicates that $Bm \ in$ also contains a 6-nucleotide miniexon as found in *Drosophila in* (12). This exon is not found in *Drosophila en* (40) and, apparently, is also absent in $Bm \ en$ (Fig. 1). The Arg-Ser sequence encoded by this exon thus appears to be a hallmark of In proteins. In this respect, it is interesting to find that the only En-like protein found in the grasshopper also contains this sequence (14) and the mouse En-1 and En-2 proteins do not possess this sequence (16).

Whether the two *en*-like genes found in the honeybee represent the counterparts of *en* and *in* is still unclear. It is interesting to find that the two honeybee *en* genes do not possess an intron in the homeobox region while their *Bombyx* and *Drosophila* counterparts do (13). This intron is also absent in the mouse (16), zebrafish (19), and sea urchin (20). Though we do not know at present whether there are additional introns in the untranslated regions of *Bm en* and *Bm in*, the *Bombyx* and *Drosophila* genes are apparently similar in their exon-intron organization. These observations strongly



FIG. 4. Homologous domains between the *Bombyx* and *Drosophila* En-like proteins. (A) Homologous domains in En-like proteins. The homeodomain is boxed and the three homologous domains are marked. Identical amino acids are indicated by a dash and an asterisk indicates a gap in the sequence. Other symbols are the same as in Fig. 3. (B) En- and In-specific domains. Dv en represents the sequence of D. virilis En.

suggest that en and in were present in the last ancestor common to both Bombyx and Drosophila. Based on the observations that only one *en*-like gene could be found in the grasshopper, Patel et al. (14) suggested that en and in may have arisen by duplication some time in the insect lineage after the last ancestor common to both Drosophila and the grasshopper. This hypothesis suggests that the two en genes found in vertebrates also arose as an independent duplication of an ancestral en gene early in the chordate lineage. The same conclusion has been suggested by Dolecki and Humphreys (20) after finding a single en gene in the sea urchin. Moths and flies are believed to have diverged about 240 million years ago, subsequent to the separation of their ancestors from those of the grasshopper and honeybee (42). Our data are consistent with the above hypothesis on the phylogeny of en.

Coexpression of *Bombyx en* and *in* in the Middle Silk Gland. Northern blot hybridization analysis revealed that two transcripts are encoded by *Bm en* and by *Bm in* in the middle silk gland of Kin-Shu \times Sho-Wa larvae (Fig. 5 *B* and *C*). By ribonuclease protection and Northern blot hybridization analyses using RNA samples taken from the Kin-Shu and the Sho-Wa strains, we found that the cDNA sequences reported here are derived from the Kin-Shu strain (data not shown). Fig. 5*A* shows that a single *Bm en* transcript of 5.1 kb and a single *Bm in* transcript of 6.2 kb were detected in the Kin-Shu middle silk gland RNA sample (lanes 2 and 4). Both of them are shorter than their Sho-Wa counterparts, which are 5.5 kb (*Bm en*) and 7.4 kb (*Bm in*) long. Though the molecular basis for this length polymorphism is unknown, it might be due to a strain-specific variation in the long untranslated regions.

In Fig. 5B, we investigated the developmental profile of the Bm en and Bm in transcripts in the posterior and the middle silk gland during the transition from the fourth molt to the fifth intermolt. Both transcripts were barely detectable in the middle silk gland during the molting stage (15 h after the fourth apolysis; lane 5) and the fourth ecdysis (lane 6). Their levels increase gradually during the fifth intermolt (Fig. 5B, lanes 7 and 8, and also Fig. 5C) and peak between 48 h and 72 h after ecdysis (Fig. 5C). A trace amount of the Bm en and Bm in transcripts could also be detected in the middle silk gland during the fourth intermolt (Fig. 5C, lane 1). However, we could not detect any of these transcripts in the posterior silk gland at any stage examined so far (Fig. 5B, lanes 1-4).

The expression of $Bm \ en$ and $Bm \ in$ in the middle but not posterior silk gland is intriguing because *Drosophila en* is known to be expressed in the posterior developmental compartments derived from the ectoderm and to specify cell state (ref. 43 and references therein). The silk gland is an ectodermal derivative (44) and forms three morphologically distinct regions, the anterior, middle, and posterior silk glands (see ref. 33). The middle silk gland specifically expresses a number of glue protein genes, such as the sericin-1 gene, and the posterior silk gland specifically expresses the silk fiber genes, such as the fibroin gene. $Bm \ en$ and $Bm \ in$ might





FIG. 5. Coexpression of Bombyx en and in in the middle silk gland during the fifth larval instar. (A) A Northern blot of poly(A)⁺ RNA (5 µg) isolated from the posterior (lanes 1 and 3) and middle silk gland (lanes 2 and 4) of 2-day-old fifth-instar larvae (Kin-Shu strain) was hybridized with an en-specific probe (lanes 1 and 2) and an in-specific probe (lanes 3 and 4). (B) Northern blot analysis of poly(A)⁺ RNA (5 µg) isolated from the posterior silk gland (lanes 1-4) and the middle silk gland (lanes 5-8) of Kin-Shu × Sho-Wa larvae. Lanes: 1 and 5, 15 h after the fourth apolysis; 2 and 6, the fourth ecdysis; 3 and 7, 24 h after the fourth ecdysis; 4 and 8; 48 h after the fourth ecdysis. The blot was hybridized with an *en*-specific probe (en), an *in*-specific probe (in), and a control probe (C). (C) Northern blot analysis of $poly(A)^+$ RNA (5 μg) isolated from the middle silk gland of Kin-Shu × Sho-Wa larvae. Lanes: 1, 72 h after the third ecdysis; 2, the fourth ecdysis; 3-6, 24, 48, 72 and 144 h, respectively, after the fourth ecdysis. Blots were hybridized as described in B.

similarly be involved in specifying compartments in the silk gland. A possible role for them might be the transcriptional regulation of genes specifically expressed in the silk gland. In this respect, the silk protein genes that possess homeodomain binding sites in their promoters are candidate genes (34-36). Further studies of Bm en and Bm in should provide information about development of the silk gland and about body segmentation in this intermediate germ-band insect.

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