Supplemental Information

Heparin binding preference and structures in the fibroblast growth factor family parallel their evolutionary diversification

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Table of contents

Figure S1 Stabilization effect of heparin on FGF10	3
Figure S2 Stabilization effect of heparin on FGF4	4
Figure S3 Stabilization effect of heparin on HT-FGF6	5
Figure S4 Stabilization effect of heparin on FGF17	6
Figure S5 Stabilization effect of heparin on FGF20	7
Figure S6 The average molecular mass per volume unit volume and the differential	8
refractive index of FGF20	
Table S1 Summary of peptides of FGFs identified by lysine targeted Protect and	9
Label structural proteomics	
Figure S7 Peptides corresponding to trypsin digested FGF3	10
Figure S8 Peptides corresponding to thermolysin digested FGF3	11
Figure S9 Peptides corresponding to chymotrypsin digested FGF3	12
Figure S10 Peptides corresponding to trypsin digested FGF10	13
Figure S11 Peptides corresponding to chymotrypsin digested FGF10	14
Figure S12 Peptides corresponding to chymotrypsin digested FGF4	15
Figure S13 Peptides corresponding to chymotrypsin digested FGF4 (2)	16
Figure S14 Peptides corresponding to thermolysin digested FGF4	17
Figure S15 Peptides corresponding to trypsin digested HT-FGF6	18
Figure S16 Peptides corresponding to chymotrypsin digested HT-FGF6	19
Figure S17 Peptides corresponding to Glu-C digested FGF17	20
Figure S18 Peptides corresponding to chymotrypsin digested FGF17	21
Figure S19 Peptides corresponding to thermolysin digested FGF17	22
Figure S20 Peptides corresponding to trypsin and Glu-C digested FGF17	23
Figure S21 Peptides corresponding to trypsin digested FGF17	24
Figure S22 Peptides corresponding to chymotrypsin digested FGF20	25
Figure S23 Peptides corresponding to thermolysin digested FGF20	26

Figure S1 Stabilization effect of heparin on FGF10. Differential scanning fluorimetry of 5 μ M FGF10 in the presence of various concentrations of heparin. (*a*) melting curve profiles of FGF10 (5 μ M) with a range of heparin concentrations (0-100 μ M). (*b*) first derivative of the melting curves of FGF10 in (*a*). (*c*) peak of the first derivative of the melting curves from (*b*), which is the melting temperature, Tm (mean of triplicate ± S.E.).



Figure S2 Stabilization effect of heparin on FGF4. Differential scanning fluorimetry of 5 μ M FGF4 in the presence of various concentrations of heparin. (*a*) melting curve profiles of FGF4 (5 μ M) with a range of heparin concentrations (0-100 μ M). (*b*) first derivative of the melting curves of FGF4 in (*a*). (*c*) peak of the first derivative of the melting curves from (*b*), which is the melting temperature, Tm (mean of triplicate ± S.E.).



Figure S3 Stabilization effect of heparin on HT-FGF6. Differential scanning fluorimetry of 5 μ M HT-FGF6 in the presence of various concentrations of heparin. (*a*) melting curve profiles of HT-FGF6 (5 μ M) with a range of heparin concentrations (0-100 μ M). (*b*) first derivative of the melting curves of HT-FGF6 in (*a*). (*c*) the melting temperature of Halo protein with and without heparin (2.5 μ M and 10 μ M). (*d*) peak of the first derivative of the melting curves from (*b*), which is the melting temperature, Tm (mean of triplicate ± S.E.).



Figure S4 Stabilization effect of heparin on FGF17. Differential scanning fluorimetry of 5 μ M FGF17 in the presence of various concentrations of heparin. (*a*) melting curve profiles of FGF17 (5 μ M) with a range of heparin concentrations (0-100 μ M). (*b*) first derivative of the melting curves of FGF17 in (*a*). (*c*) peak of the first derivative of the melting curves from (*b*), which is the melting temperature, Tm (mean of triplicate ± S.E.).



Figure S5 Stabilization effect of heparin on FGF20. Differential scanning fluorimetry of 5 μ M FGF20 in the presence of various concentrations of heparin. (*a*) melting curve profiles of FGF20 (5 μ M) with a range of heparin concentrations (0-100 μ M). (*b*) first derivative of the melting curves of FGF20 in (*a*). (*c*) peak of the first derivative of the melting curves from (*b*), which is the melting temperature, Tm (mean of triplicate ± S.E.).



Figure S6 The average molecular mass per volume unit volume and the differential refractive index of FGF20. The red line shows the different reflective index (dRI). The blue line indicates the molecular mass of FGFs.



Table S1

Summary of peptides of FGFs identified by lysine targeted Protect and Label structural proteomics

Labelled peptides were identified by MALDI-Q-TOF and analysed by MS-digest from the package ProteinProspector v 5.12.3. A full list of identified peptides is provided in this table. Four proteases used for protein digestion were trypsin (TRY), thermolysin (THE), chymotrypsin (CHY), Glu-c (GLU) and mixture of trypsin and Glu-C (TG).

Р	eptide	Sequences	Proteases	Residues	HBS	Spectrum
FGF3	1	YC(Carbamidomethyl)ATK(biotin)YHLQ	(THE)	49-57	1	S8
	2	RTQK(biotin)SSLFLPR	(TRY)	171-181	1	S7
FGF10	1	FSFTK(biotin)Y	(CHY)	83-88	1	S11
FGF17	1	AK(biotin)LIVETDTF	(CHY)	84-93	1	S18
	2	AHFIK(biotin)R	(TRY)	172-177	2	S21
	3	EAHFIK(biotin)R	(TG)	171-177	2	S20
	4	LYQGQLPFPNHAEK(biotin)QK(biotin)QF	E (THE)	178-196	2	S19
	5	K(biotin)QK(biotin)QFE	(GLU)	191-196	2	S17
FGF4	1	VAMSSK(biotin)GK(biotin)	(CHY)	137-145	1	S12
	2	IALSKNGKTKKGNRVSPT(3biotin/1acety	I) (THE)	179-196	1	S13
	3	AGDYLLGIK(biotin)R	(THE)	73-82	3	S13
FGF6	1	QGTYIALSK(biotin)Y	(CHY)	177-186	1	S16
	2	ATPSFQEEC (carbamidomethyl) K (biotin)	F (CHY)	149-156	3	S16







Figure S8 Peptides corresponding to thermolysin digested FGF3, (Data summary in Table 2 and S1).



Figure S9 Peptides corresponding to chymotrypsin digested FGF3, (Data summary in Table 2).



Figure S10 Peptides corresponding to trypsin digested FGF10, (Data summary in Table 2).

Figure S11 Peptides corresponding to chymotrypsin digested FGF10, (Data summary in Table 2 and S1).





Figure S12 Peptides corresponding to chymotrypsin digested FGF4, (Data summary in Table 3 and S1). Due to variability in the digestion of FGF4, two spectra are shown (Figures S12 and S13).

Figure S13 Peptides corresponding to chymotrypsin digested FGF4, (Data summary in Table 3 and S1). Due to variability in the digestion of FGF4, two spectra are shown (Figures S12 and S13).





Figure S14 Peptides corresponding to thermolysin digested FGF4 (2), (Data summary in Table 3).



Figure S15 Peptides corresponding to trypsin digested HT-FGF6, (Data summary in Table 3).

Figure S16 Peptides corresponding to chymotrypsin digested HT-FGF6, (Data summary in Table 3 and S1).





Figure S17 Peptides corresponding to Glu-C digested FGF17, (Data summary in Table 4 and S1).

Figure S18 Peptides corresponding to chymotrypsin digested FGF17, (Data summary in Table 4 and S1).



Figure S19 Peptides corresponding to thermolysin digested FGF17, (Data summary in Table 4 and S1).



Figure S20 Peptides corresponding to trypsin and Glu-C digested FGF17, (Data summary in Table S1).









Figure S22 Peptides corresponding to chymotrypsin digested FGF20, (Data summary in Table 5).



