SUPPLEMENTAL MATERIAL

Lim et al., http://www.jem.org/cgi/content/full/jem.20151750/DC1

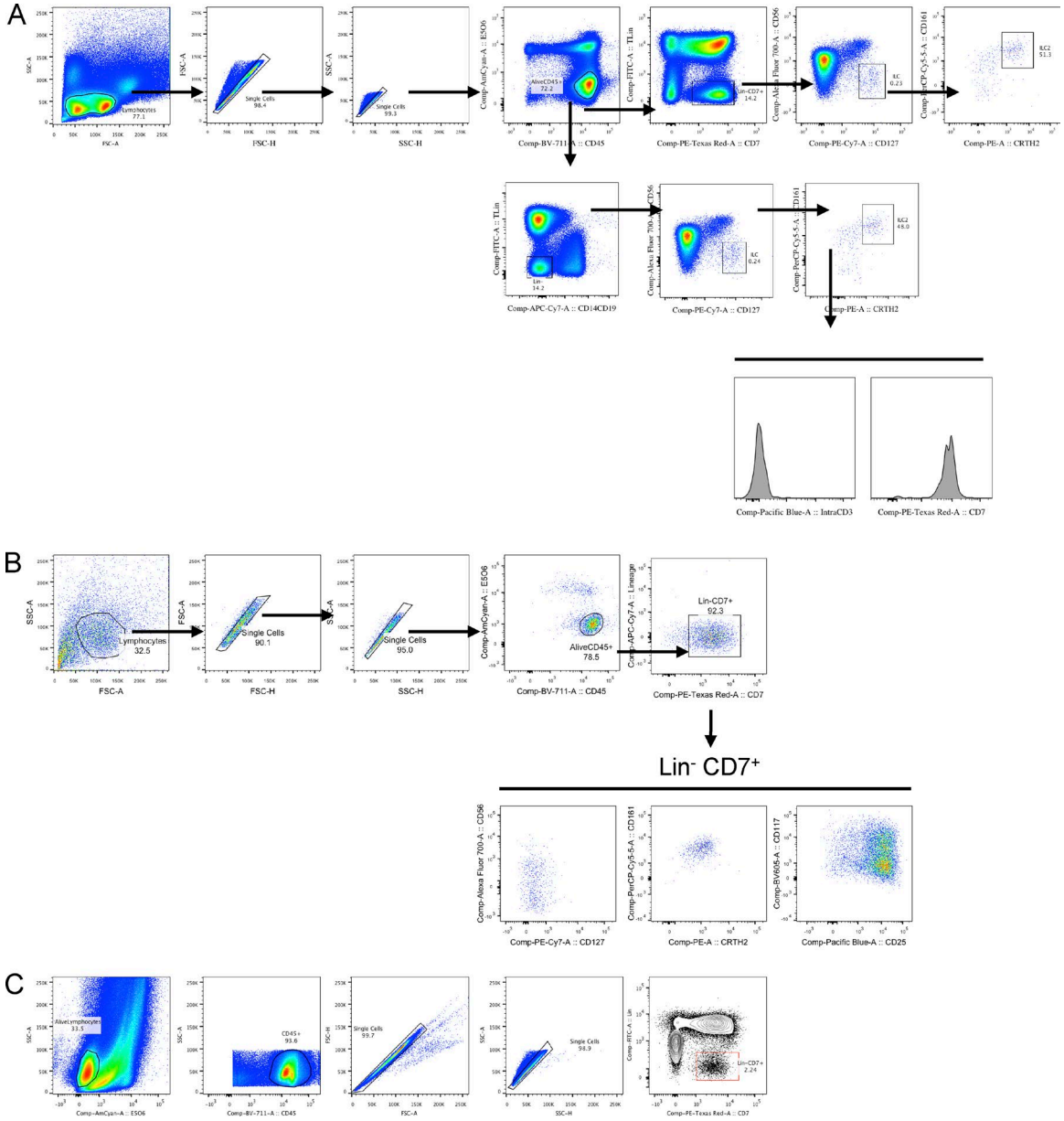

Figure S1. **ILC2 gating strategy.** FACS analysis of ILCs from human peripheral blood (A), culture-expanded ILC2 (B), or intestinal ILCs (C). (A) Cells were first gated on lymphoid size and side scatter; singlet and viable $CD45^{+}Lin^{-}CD7^{+}CD127^{Hi}$ cells that expressed high levels of CRTh2 and CD161 were considered to be ILC2. (B) Bulk ILC2 cultured with irradiated PBMCs in IL-2, -7, -25, and -33 for 7 d. Culture-expanded cells were gated on lymphoid size and side scatter; singlet, viable $CD45^{+}Lin^{-}CD7^{+}$ cells were then analyzed for ILC2 markers as shown. (C) Intestinal ILC2 were gated on lymphoid size and side scatter; singlet, viable $CD45^{+}Lin^{-}$ $CD7^{+}$ cells were then analyzed.

Table S1. **Primers used for single cell Biomark analysis**

Gene name	Forward primer (5′→3′)	Reverse primer (5′→3′)
IL7R	GGAGAAAGTGGCTATGCTCAA	CTGCGATCCATTCACTTCCA
ACTB	CCAACCGCGAGAAGATGAC	TAGCACAGCCTGGATAGCAA
B2M	TCCGTGGCCTTAGCTGTG	CCCAGACACATAGCAATTCAGG
GATA3	CACGGTGCAGAGGTACCC	AGGGTAGGGATCCATGAAGCA
GAPDH	ACACCATGGGGAAGGTGAAG	GTGACCAGGCGCCCAATA
RORA	CAGCAGATAACGTGGCAGAC	GGCACACAATTGCCACATCA
ID2	CTCAACACGGATATCAGCATCC	CACACAGTGCTTTGCTGTCA
AHR	TAGGCTCAGCGTCAGTTACC	TGGCCTCCGTTTCTTTCAGTA
TNFRSF25	CAGGGCGGCACTCGTA	AGCCTCTGCAACAAAACAGAC
MAF	TCGACGACCGCTTCTCC	ATCACCTCCTCCTTGCTGAC
CCR6	AGGCAGCGATGTCTGTGAA	AGCTCAAGCCCCAACATCA
PTGDR2	TCCAGGGCTGGAATCCTGT	GGCAGAGTGGCTTCAGTGT
CD40L	GAGGCCAGCAGTAAAACAAC	AGTTGTTGCTCATGGTGTAGTA
IL17RB	TGGTGGCAGGGATCTATCTA	GCAGTAGTGTGGTGGTAGAA
ICOS	AGTCTGCATTTTGGGATGCA	GTCGTGCACACTGGATGAA
IL13	TGCAGTGCCATCGAGAAGAC	TCGGACATGCAAGCTGGAAA
IL1RL1	GGCGACCAGGTCCTTCAC	AGGGGCTCCGATTACTGGAA
CSF2	TGATGGCCAGCCACTACAA	CAAAGGGGATGACAAGCAGAAA
ZEB2	AGGCCAATGGGCAAGAAGAA	AGGTCAGCAGTTGGGCAAA
EOMES	CTGTGGCAAAGCCGACAATA	CTCATCCAGTGGGAACCAGTA
IFNG	ACTGCCAGGACCCATATGTAA	GTTCCATTATCCGCTACATCTGAA
TBX21	GGGCGTCCAACAATGTGAC	CCGTCGTTCACCTCAACGATA
IRF4	CACCATGACAACGCCTTACC	CGAGGGGTGGCATCATGTA
IL5	ACTCTGAGGATTCCTGTTCCTGTA	CCAGTGTGCCTATTCCCTGAAA
NFIL3	CCGCCCCTTTCTTTCTCC	GGATAAATCCGTCAGGCTCCTTA
PPARG	TAGATGACAGCGACTTGGCAATA	TGGGCTTCACATTCAGCAAAC
AREG	GGTGGTGCTGTCGCTCTT	GCTTCCCAGAGTAGGTGTCATT
IL4	CAGCTGATCCGATTCCTGAAA	GTTGGCTTCCTTCACAGGAC
RORC	CAAGACTCATCGCCAAAGCA	TTTCCACATGCTGGCTACAC
CCR7	GTGGTGGCTCTCCTTGTCA	TGTGGTGTTGTCTCCGATGTA
KIT	GGATTCCCAGAGCCCACAA	ACATCCACTGGCAGTACAGAA
CCR2	GCTGAGAAGCCTGACATACCA	GGGAAATGCGTCCTTGTTCAA

Table S2. **Characteristics of Crohn's disease patients**

Patient ID	Age	Gender	Active medications	Disease duration	Indication for surgery
1	62	Female	Vedolizumab	31 yr	Intestinal stricture
2	65	Male	None	42 yr	Intestinal stricture
3	19	Male	Budesonide and antibiotics	8 mo	Intestinal fistula
4	38	Male	Azathioprine and Infliximab	16 mo	Intestinal stricture

Table S3. **Antibodies used for flow cytometric analysis**

Antibody	Clone	Manufacturer
CD3	OKT3	eBioscience
CD5	L17F12	eBioscience
TCRαβ	IP16	eBioscience
TCRγδ	B1.1	eBioscience
CD14	TÜK4	Miltenyi Biotec
CD19	LT19	Miltenyi Biotec
CD7	M-T701	BD
CD56	B159	BD
CD127	eBioRDR5	eBioscience
CD161	DX12	BD
CRTH2	BM16	Miltenyi Biotec
CD117	104D2	BioLegend
CD25	BC96	eBioscience
KLRG1	2F1/KLRG1	BioLegend
CCR6	11A9	BD
CD11a	HI111	BioLegend
CD2	RPA-2.10	BioLegend
CD90	5E10	BD
IL1R1	NA[a]	R&D Systems
IL17RB	NA[a]	R&D Systems
ST2	B4E6	MD Biosciences
GATA-3	TWAJ	eBioscience
T-BET	eBio4B10	eBioscience
RORγt	AFKLS-9	eBioscience
IL-13	JES10-5A2	BD
IL-5	JES1-39D10	BioLegend
IL-17A	CZ8-23G1	Miltenyi Biotec

[a]NA, not available.