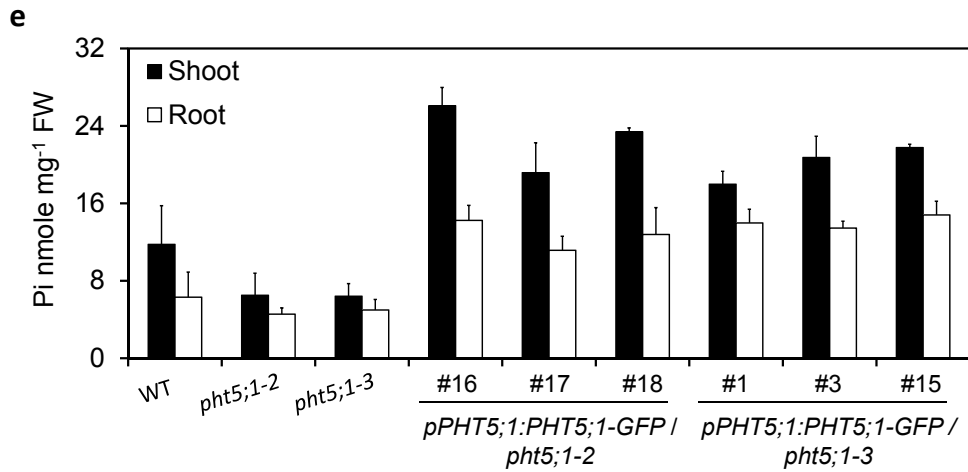
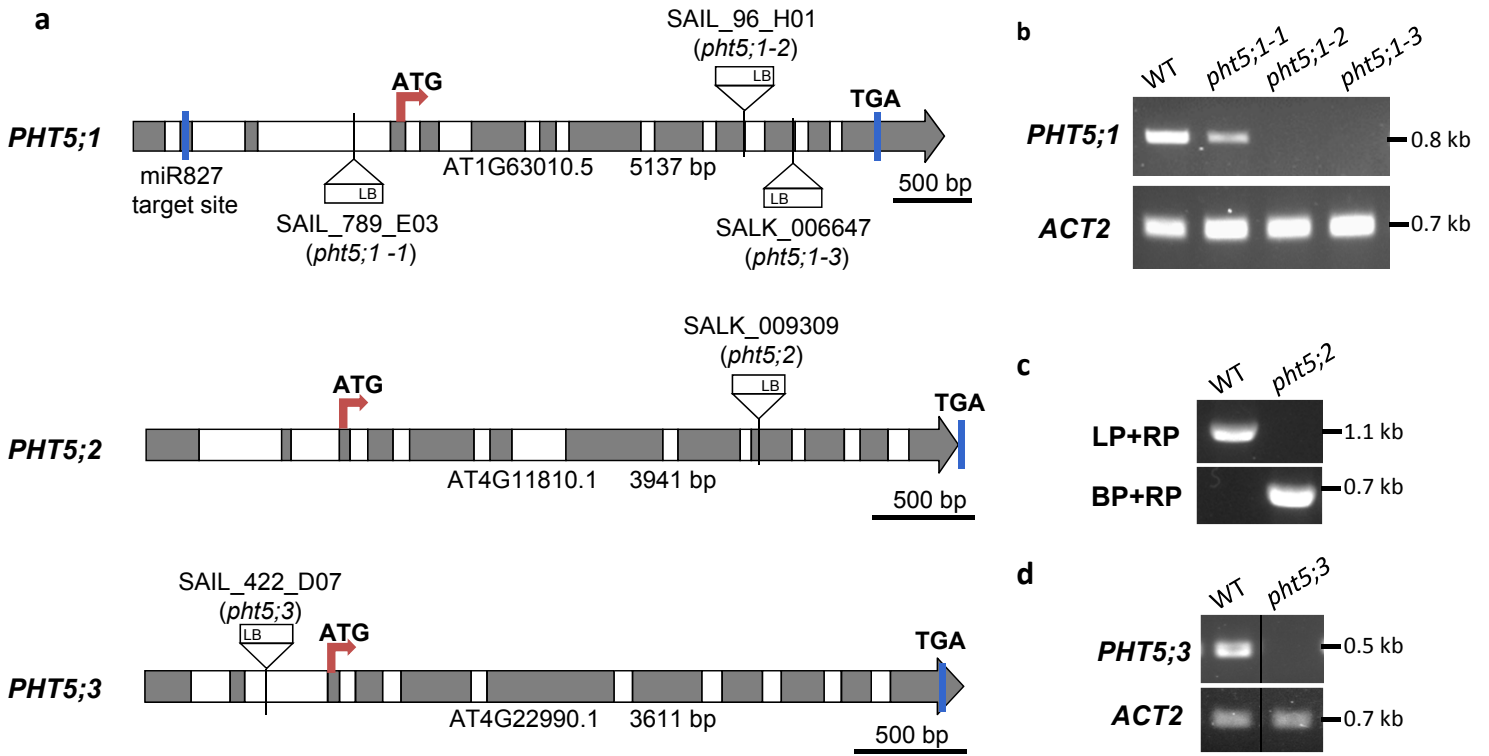


**Supplementary Figure 1 | Subcellular localization of *Arabidopsis* PHT5;2 and PHT5;3**

Expression of *p35S:AtPHT5;2-GFP* and *p35S:AtPHT5;3-GFP* in *Arabidopsis* mesophyll protoplasts (**a,b**) and tobacco (*Nicotiana benthamiana*) leaves (**c,d**). Scale bar, 10  $\mu$ m.

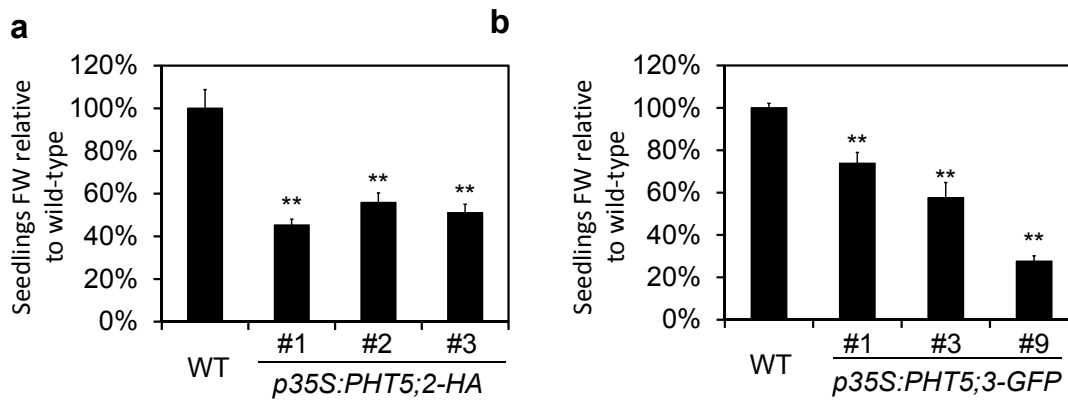


**Supplementary Figure 2 | Characterization and complementation of the mutants of the *PHT5s***

(a) Schematic representation of T-DNA insertion in the mutants of *PHT5s*.

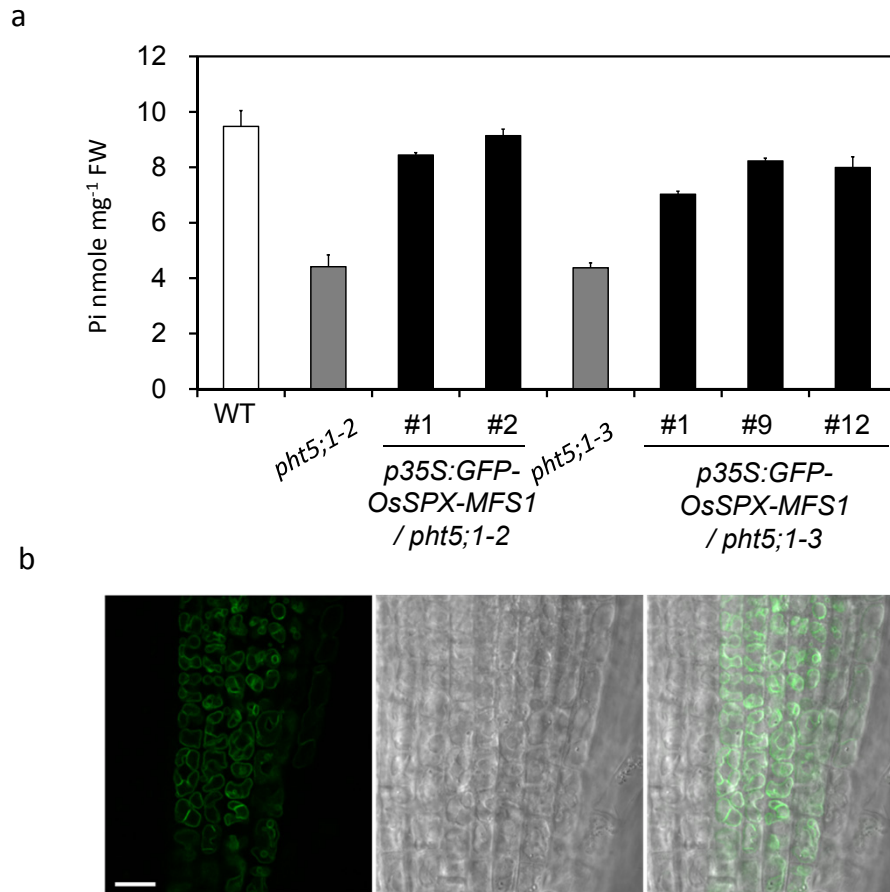
(b,d) RT-PCR analyses of *PHT5;1* and *PHT5;3* transcripts in the respective mutants of *PHT5s*. (c) Because of extremely low expression of *PHT5;2*, PCR analysis of genomic DNA was employed to confirm homozygous T-DNA inserted mutation at *PHT5;2* locus (e)

Complementation of the *pht5;1-2* or *pht5;1-3* mutant by the expression of *PHT5;1-GFP* driven by its native promoter. Three independent transgenic lines in each mutant background are shown. Error bar, s.d. ( $n=3$ ).



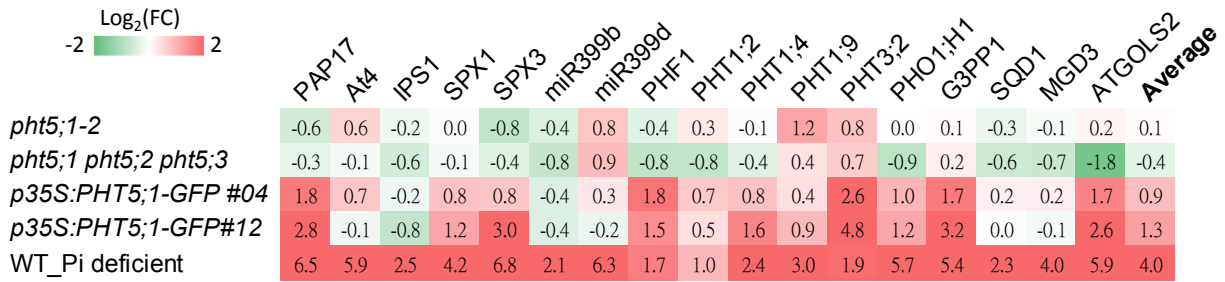
**Supplementary Figure 3 | Overexpression of *Arabidopsis* *PHT5;2* and *PHT5;3* leads to retarded growth**

The relative fresh weight of seedlings of *PHT5;2-HA*-overexpressing lines (11-day-old, **a**) and *PHT5;3-GFP*-overexpressing lines (12-day-old, **b**) to the corresponding wild-type plants (WT). Three independent transgenic lines are shown for each construct. Error bar, s.d. ( $n=3$ ), \* $P<0.05$ , \*\* $P<0.01$ , Student's *t*-test. Results were reproducible in at least three independent experiments.



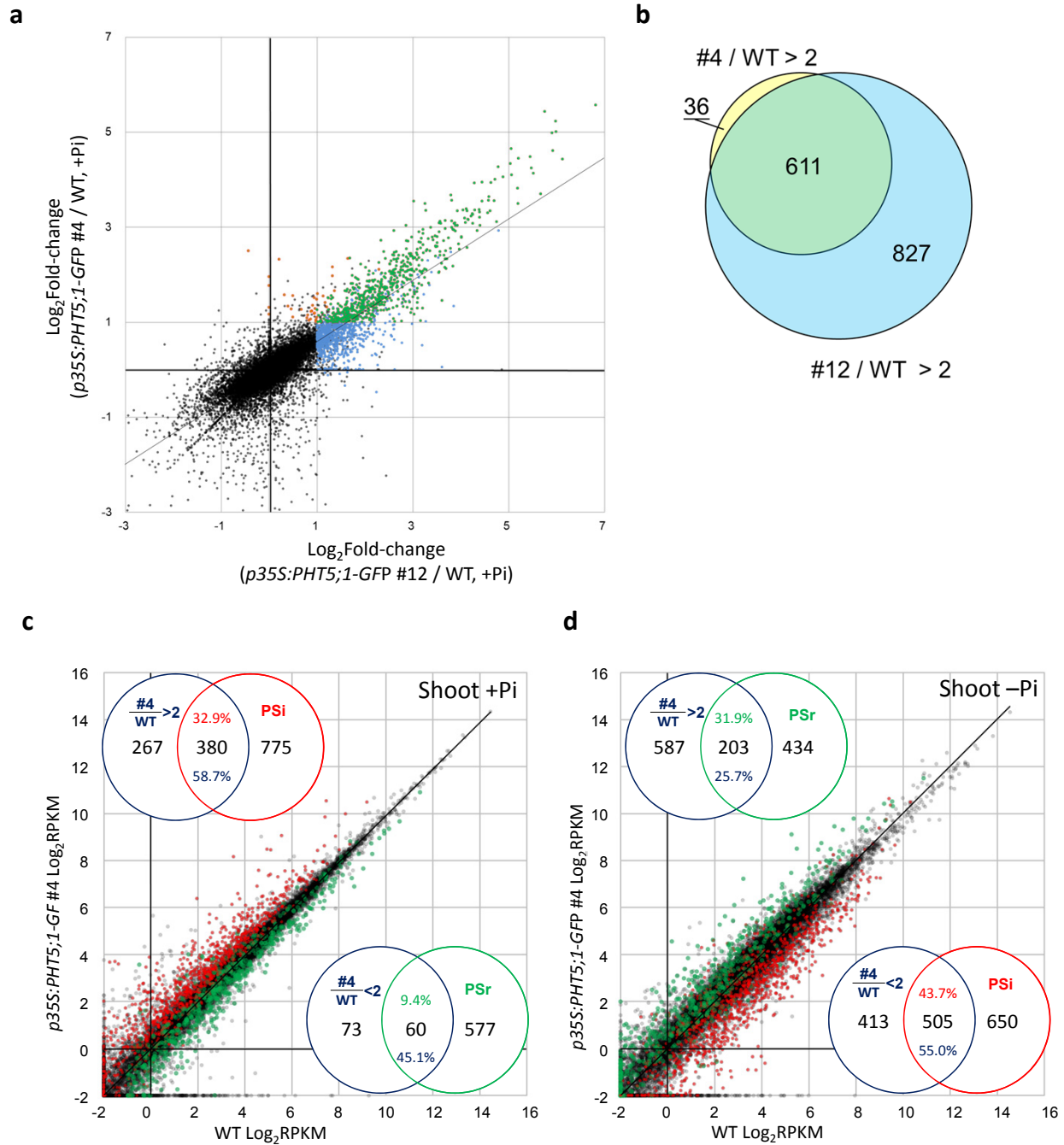
**Supplementary Figure 4 | Characterization of function and sub-cellular localization of OsSPX-MFS1 in *Arabidopsis***

**(a)** Pi concentrations of the first leaf of 14-day-old *Arabidopsis* wild type (WT), *pht5;1-2* and *pht5;1-3* plants and *p35S:GFP-OsSPX-MFS1*-overexpressing lines grown under +Pi conditions. Two-three independent transgenic lines in each mutant background are shown. Error bar, s.d., technique replicates  $n=2$ . **(b)** Fluorescence signal (left), bright field (middle) and merged image (right) of *pht5;1-3* mutant transformed with a *p35S:GFP-OsSPX-MFS1* construct. Scale bar, 10  $\mu\text{m}$



**Supplementary Figure 5 | Overexpression of *PHT5;1* leads to the upregulation of Pi starvation-induced genes under Pi-sufficient conditions.**

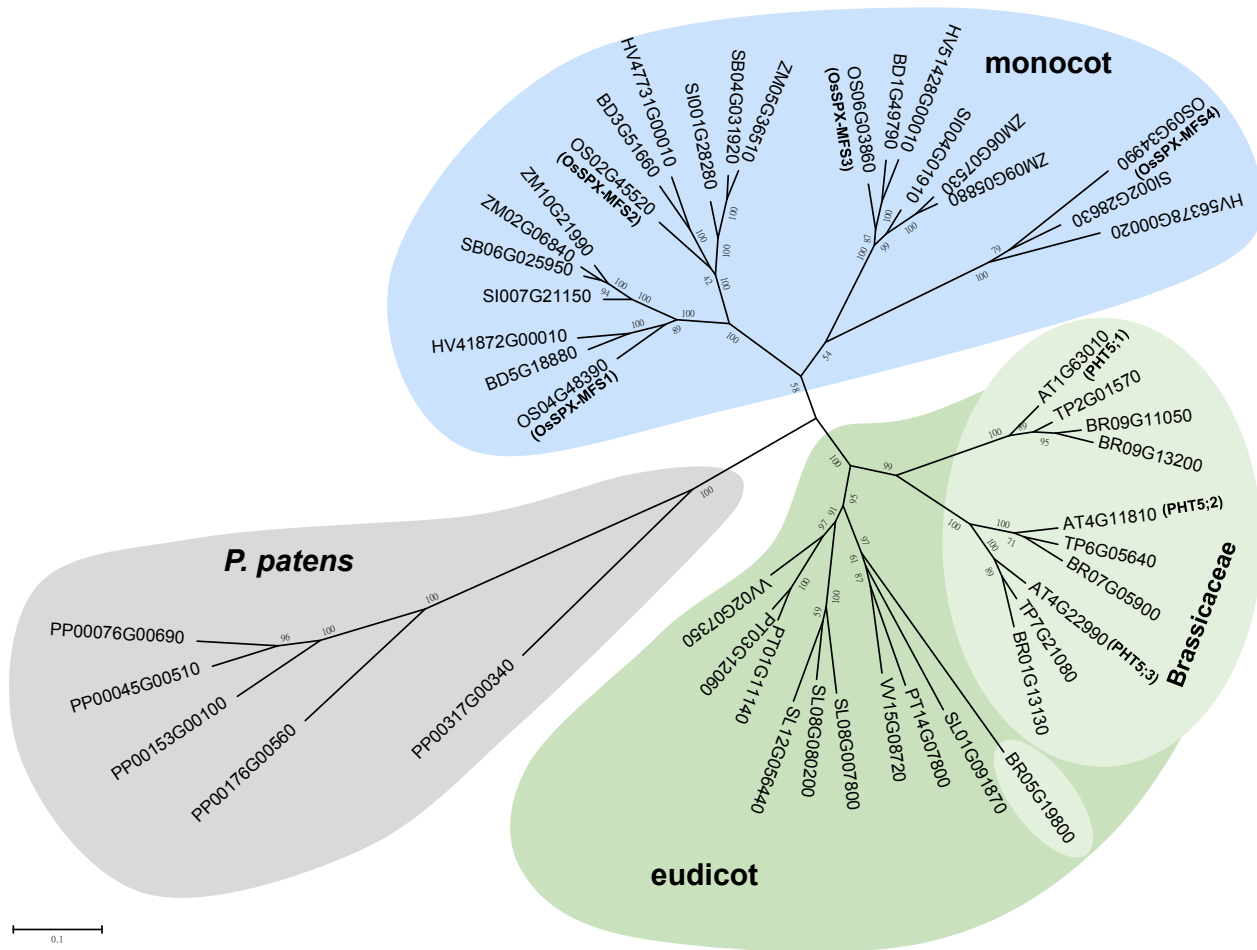
Heat map of qRT-PCR results reveals the upregulation of many Pi starvation-induced (PSi) genes in the shoot of 10-day-old *PHT5;1*-overexpressing lines, implying that a lower cyt-Pi level of the *PHT5;1*-overexpressing lines likely induced Pi-starvation responses. In contrast, these PSi genes were downregulated in the *pht5* triple mutants. The numbers indicate the log<sub>2</sub> values of fold changes of gene expression relative to WT plants grown under Pi sufficiency. Results were reproducible in three independent experiments.



**Supplementary Figure 6 | Overexpression of *PHT5;1* leads to mis-regulation of PSR genes.**

(a) The positive correlation of the gene expression pattern between two *PHT5;1*-overexpression lines, #4 and #12, grown under Pi-sufficient conditions (Pearson coefficient of correlation  $r = 0.73$ ). (b) Venn diagram showing the overlap of differentially expressed genes in the *PHT5;1*-overexpressing lines #4 and #12.

Scatter plots showing the relationship of the expression of an individual gene (a single dot) in the shoots between WT and *PHT5;1* overexpressing plants (line #4) grown under Pi sufficient medium (c) or starvation for 3 days (d). The Pi-starvation-upregulated and -downregulated genes are marked in red and green, respectively. The Venn diagrams in each patterns indicate the proportion of upregulated or downregulated genes in *PHT5;1* overexpressing lines belonging to PSR genes of WT.



**Supplementary Figure 7 | Phylogenetic analysis of SPX-MFS homologs.**

The coding DNA sequences of 47 *SPX-MFS* homologs in 13 different species obtained from PLAZA 3.0 (<http://bioinformatics.psb.ugent.be/plaza/>) were aligned using ClustalW. The phylogenetic tree was built based on Maximum Likelihood with the Tamura-Nei model. The scale bar indicates genetic distance of branch length measured in the number of nucleotide substitutions per site. *Oryza sativa japonica* (OS), *Sorghum bicolor* (SB), *Zea mays* (ZM), *Setaria italic* (SI), *Brachypodium distachyon* (BD), *Hordeum vulgare* (HV), *Arabidopsis thaliana* (AT), *Brassica rapa* (BR), *Thellungiella parvula* (TP), *Populus trichocarpa* (PT), *Vitis vinifera* (VV), *Solanum lycopersicum* (SL) and *Physcomitrella patens* (PP).

**Supplementary Table 1 | List of primers**

<b>Gene</b>		<b>Sequences (5'→3')</b>
<b>Q-RT-PCR</b>		
AT1G63010	Forward	TAACCAATCTACTTTGCCGGGAT
<i>PHT5;1</i>	Reverse	CTCTGAATGAAATGCATAGCCATAC
AT4G22990	Forward	GGATCGGCGAGAGCAGTGAA
<i>PHT5;3</i>	Reverse	CTTTAACGGTACACAGTCGCTTATAT
AT4G05320	Forward	GCCCTTGATAATCCCTGATGAATAAG
<i>UBQ10</i>	Reverse	AAAGAGATAACAGGAACGGAAACATAGT
AT3G17790	Forward	TTGTTGACACGACTCCTTTTCGT
<i>PAP17</i>	Reverse	ATCACGGAGAAGAGCTTTGACAT
AT5G03545	Forward	AACCCTAACACATCCATTGTTGAA
<i>AT4</i>	Reverse	GAAACTTGAACCTCTCAAACCCT
AT3G09922	Forward	TTTGGAGAATAGTCAGACCAGTGC
<i>IPS1</i>	Reverse	TCACTATAAAGAGAATCGGAAGCA
AT5G20150	Forward	GATTCCATTGTTGGAGCAAGA
<i>SPX1</i>	Reverse	AATCTGTTAGCTTCTTCTATTGTA
AT2G45130	Forward	CCGATCTCTTCATCTATCTT
<i>SPX3</i>	Reverse	TTACGATAATGTCATATTGCGT
AT1G63005	Forward	AGGTCCTTTACTTCCAAATATACACATACA
<i>MIR399B</i>	Reverse	TCGATCATCGGAAATTTTCGA
AT2G34202	Forward	TTACTGGGCGAATACTCCTATGG
<i>MIR399D</i>	Reverse	ATTTTACTTGCATATCTAGCCAATGC
AT3G52190	Forward	GCCTTCTGAGGATCATAGTAG
<i>PHF1</i>	Reverse	CGCTGCAACAAGATAAGGAAG
AT5G43370	Forward	AGCCATCATTGGAGCCTTC
<i>PHT1;2</i>	Reverse	ACCTTAGCCTTGTCTTGATT
AT2G38940	Forward	GGTCCCAATAGTTTAGGTGAT
<i>PHT1;4</i>	Reverse	AGTTGCTAGAGACAAGGAGAA
AT1G76430	Forward	GAAGATCGTTAGAAGAGAACGAA
<i>PHT1;9</i>	Reverse	GTATTGTCTCCGAAGTAACTCA
AT3G48850	Forward	GCTCCATCTGTAAGTGCATA
<i>PHT3;2</i>	Reverse	CTGGTAGTTCTGTTTCCCTT
AT1G68740	Forward	GATGAAGAAGACTAATCTGATC
<i>PHO1;H1</i>	Reverse	GTTCTTTCAAATCTGACGAAGC
AT3G47420	Forward	AGGCGAATATTTATCGGACGAA
<i>G3PP1</i>	Reverse	ACACCTCCTACATCGAACATTGTC



Gene		Sequences (5'→3')
AT4G33030	Forward	CATTGACTCCTATTGCCTCCATT
<i>SQD1</i>	Reverse	TCCCTGTCAAAGCCTTCCAT
AT2G11810	Forward	TGCGGCCGGAACAAAG
<i>MGD3</i>	Reverse	CCTTGACCGGAATCTTCCATT
AT1G56600	Forward	AATATAATCATCGAGCTCGT
<i>GOLS2</i>	Reverse	TTATAGTCATGAAGAGGCG
<b>RT-PCR</b>		
AT3G18780	Forward	GTAGTCAACAGCAACAAAGGAGAGC
<i>ACTIN2</i>	Reverse	ATGGCTGAGGCTGATGATATTCA
AT1G63010	Forward	TTGTGAGTGCAAGTGCTCTTG
<i>PHT5;1</i>	Reverse	TACCCGCGATAACAACGATAG
AT4G22990	Forward	TCGTGTGATCCCCTTAGTGTC
<i>PHT5;3</i>	Reverse	TTTCCTTTCTCACCGTATCCC
<b>CONFIRM T-DNA</b>		
SAIL_BP	BP	TAGCATCTGAATTTTCATAACCAATCTCGATACAC
SALK_BP	BP	TGGTTCACGTAGTGGGCCATCG
<i>pht5;1-1</i>	LP	TTTTCACTTGAATGCACGAG
	RP	GAAATCTTTGAGAACATGGCG
<i>pht5;1-2</i>	LP	TTGTGAGTGCAAGTGCTCTTG
	RP	TACCCGCGATAACAACGATAG
<i>pht5;1-3</i>	LP	TACCCGCGATAACAACGATAG
	RP	TTGTGAGTGCAAGTGCTCTTG
<i>pht5;2</i>	LP	GGTTTGTGGTGTGTTATCGG
	RP	GACGCTAACAGAATTTGCCTG
<i>pht5;3</i>	LP	TCGTGTGATCCCCTTAGTGTC
	RP	TTTCCTTTCTCACCGTATCCC
<b>CONSTRUCTS</b>		
<i>pPHT5;1:GUS</i>	Forward	CTTTTAATCGCAGAAAGCAGAGAGC
	Reverse	CTGTTCTTTACCTTGTTTATATCCGTCT
<i>pPHT5;2:GUS</i>	Forward	CTTCTACACTGAAAGAAAGTCACAGAG
	Reverse	GGAGATGGATATGAGCAGATAGCTAATC
<i>pPHT5;3:GUS</i>	Forward	AGAAACGAATCACGTGGCTTTGTTTTT
	Reverse	TTTCTACAACAAAACAAGAATCAGAGAC
<i>p35S:PHT5;1-GFP</i>	Forward	ATGGTGGCTTTTGGGAAATACTTGCAGC
	Reverse	ATAGAGTGAGTTATAAGTACAACAAGTAGC
<i>p35S:PHT5;2-GFP</i>	Forward	ATGGTCGCCTTTGGAAAAAAGCTCAAGG
	Reverse	ATACAAGGAGTTATAAGTATAACAAGTAGC

Gene		Sequences (5'→3')
<i>p35S:PHT5;2-HA</i>	Forward	ATGGTCGCCTTTGGAAAAAAGCTCAAGG
	Reverse	TTAAGCGTAATCTGGAACATCGTATGGGTAATACAA GGAGTTATAAG
<i>p35S:PHT5;3-GFP</i>	Forward	ATGGTAGCCTTCGGGAAAAAACTCAAGG
	Reverse	ATACAAGGAGTTATAAGTAAAACAAG
<i>pPHT5;1:PHT5;1-GFP</i>	Forward	CTTTTAATCGCAGAAAGCAGAGAGC
	Reverse	TCAATAGAGTGAGTTATAAGTACAAC
<i>p35S:GFP-OsSPX-MFS1</i>	Forward	ATGGTTAATTTTGGAAAGAAGTTG
	Reverse	TTAGTACAGGGTGTGTATGTG
<i>OsSPX-MFS1-ECFP</i>	Forward	ATGGTTAATTTTGGAAAGAAGTTG
	Reverse	GTACAGGGTGTGTATGTGCAG