	AMPPNP	ADP	ADP-A1F3-ssDNA	ano
Wavelength	0.97914	1 00001	0.97776	0.9753986
Resolution range	33.62 - 1.38	57 21 - 2 269	75 12 - 3 19	48 56 - 1 695
Resolution range	$(1\ 43\ -1\ 38)$	$(2\ 35\ -\ 2\ 269)$	$(3\ 304\ -\ 3\ 19)$	(1.756 - 1.695)
Space group	P 21 21 21	(2.33 2.20)) P 21 21 21	(5.504 5.17) P 31 21 1	P 21 21 21
Unit cell	67 238 74 93 107 729	67 173 74 032 109 153	86 743 86 743 149 222	66 783 70 747 103 985
	90 90 90	90 90 90	90 90 120	90 90 90
Total reflections	804634 (79664)	327618 (31138)	110624 (11373)	295055 (8181)
Unique	110840 (10818)	25850 (2511)	11303 (1117)	48257 (2538)
reflections	110040 (10010)	25050 (2511)	11505 (1117)	40237 (2330)
Multiplicity	73(74)	127(124)	98(102)	61(32)
Completeness	0.99 (0.98)	1 00 (1 00)	1.00 (1.00)	0.87(0.46)
(%)	0.55 (0.50)	1.00 (1.00)	1.00 (1.00)	0.07 (0.10)
Mean I/sigma(I)	18 60 (2 39)	27 56 (2.82)	15 62 (2,59)	24 99 (2.22)
Wilson B-factor	15.34	50.21	107.94	21.69
R-merge	0.06131 (0.9668)	0.05905 (1.057)	0 1117 (1 061)	0.04385(0.4887)
R-meas	0.06619 (1.039)	0.06164 (1.102)	0 118 (1 118)	0.0477 (0.5862)
CC1/2	0 999 (0 739)	1 (0 906)	0.998 (0.932)	0.999(0.812)
CC*	1 (0.922)	1 (0.900)	1 (0.982)	1 (0 947)
Reflections used	110825 (10818)	25839 (2510)	11299 (1117)	48248 (2537)
in refinement	110020 (10010)	20009 (2010)	112)) (1117)	10210 (2007)
Reflections used	5444 (546)	1343 (134)	568 (46)	2355 (128)
for R-free	5111 (510)	10 10 (10 1)		2500 (120)
R-work	0 1676 (0 2464)	0 1945 (0 2832)	0 2212 (0 3554)	0 1693 (0 2241)
R-free	0 1828 (0 2699)	0 2490 (0 3519)	0.2212 (0.5551)	0.1099(0.2211) 0.1940(0.2538)
CC(work)	0.961 (0.846)	0.955 (0.878)	0.919 (0.555)	0.964 (0.885)
CC(free)	0.958 (0.820)	0.938 (0.783)	0.926 (0.554)	0.952 (0.853)
Number of non-	4218	3585	3616	3908
hydrogen atoms				•,•••
macromolecules	3459	3481	3584	3461
ligands	41	29	32	5
Protein residues	429	431	429	429
RMS(bonds)	0.006	0.008	0.003	0.012
RMS(angles)	1.05	0.85	0.74	1.28
Ramachandran	99	96	93	97
favored (%)				
Ramachandran	0.94	3.3	6.1	2.3
allowed (%)				
Ramachandran	0.23	0.7	0.47	0.7
outliers (%)				
Rotamer outliers	1.8	3.4	1.6	3.9
(%)				
Clashscore	5.01	11.40	6.27	5.48
Average B-factor	24.24	67.57	118.06	30.71
macromolecules	21.61	67.82	118.13	29.67
ligands	24.35	65.05	110.98	29.24
solvent	36.90	56.57		38.88
Number of TLS	7	5	6	
groups				

Supplementary Table S1. Data collection and refinement statistics.

Statistics for the highest-resolution shell are shown in parentheses.

Data collection (Se-Met)	
Wavelength (Å)	0.9792
Resolution range	49.64 - 2.438 (2.525 - 2.438)
Space group	P 21 21 21
Unit cell	66.948 73.969 107.31 90 90 90
Total reflections	300540 (29886)
Unique reflections	20489 (2007)
Multiplicity	14.7 (14.9)
Completeness (%)	1.00 (1.00)
Mean I/sigma(I)	39.76 (7.59)
Wilson B-factor	39.80
R-merge	0.07222 (0.4397)
R-meas	0.0748 (0.4551)
CC1/2	1 (0.968)
CC*	1 (0.992)
Anomalous signal and phasing	
Anomalous completeness	99.8 (91)
Anomalous multiplicity	7.8 (7.6)
CC ano	0.847 (0.178)
R-ano	0.059 (0.428)
Mean I/sigma(I) ano	2.149 (1.026)
FOM	0.43
FOM after density modification	0.69
Refinement	
Reflections used in refinement	20479 (2007)
Reflections used for R-free	1006 (102)
R-work	0.2139 (0.2723)
R-free	0.2364 (0.2549)
CC(work)	0.944 (0.843)
CC(free)	0.927 (0.813)
Number of non-hydrogen atoms	3609
macromolecules	3447
ligands	32
Protein residues	427
RMS(bonds)	0.020
RMS(angles)	1.88
Ramachandran favored (%)	92
Ramachandran allowed (%)	6.4
Ramachandran outliers (%)	1.2
Rotamer outliers (%)	9.4
Clashscore	22.42
Average B-factor	53.27
macromolecules	53.34
ligands	83.42
solvent	44.20
Number of TLS groups	0

Supplementary Table S2. SAD data collection and phasing statistics

Statistics for the highest-resolution shell are shown in parentheses.

Supplementary Table S3. SAXS data collection and analysis

Data-collection parameters (BsPif1)				
Instrument	BL19U2			
Beam geometry (mm)	0.40×0.15(H×V)			
Wavelength (Å)	1.033 Å			
q range (Å ⁻¹)	0.01-0.5			
Exposure time (s) / nb frames	1 / 10			
Concentration range (mg.ml ⁻¹)	1-7			
Temperature	288			
$I(0) (cm^{-1}) [from P(r)]$	4350			
Rg(Å) [from P(r)]	28.0			
I(0) (cm ⁻¹) [from Guinier]	4350			
Rg (Å) [from Guinier]	28.0			
$D_{max}(Å)$	95.6			
Porod estimate (Å ³)	95744			
Molecular-mass determination				
Partial specific volume (cm ³ .g ⁻¹)	0.738			
Contrast (\times 10 ¹⁰ cm ⁻²)	2.872			
Molecular mass M _r [from I(0)]	59917			
Calculated monomeric Mr from	46673			
sequence				
Data processing				
Primary data reduction	XX			
Data processing	PRIMUS			
Ab initio analysis	DAMMIF			
Number of models	50			
Model ²	1.200 ± 0.031			
Validation and averaging	DAMAVER			
NSD	0.559 ± 0.084			
Rigid-body modeling	DADIMODO			
Computation of	CRYSOL			
model intensities				
Model ²	2.2			

SUPPLEMENTARY FIGURES LEGENDS

Supplementary Fig. S1. Schematic diagram of BsPif1 DNA helicase. (a) Coloring denotes the 1A (blue), 1B (orange), 2A (red), 2B (cyan), CTD (pink) domain. (b) Structural alignment of BsPif1 with human Pif1, ScPif1, RecD2 and Dda. Secondary structure alignment is for BsPif1. Conserved helicase motifs (I to VI) and specific Pif1 motifs (A, B, C and PFSS) are indicated.

Supplementary Fig. S2. Representative regions of the electron density. Electron density results from a map calculated using $2F_0$ -Fc coefficients contoured at 2 sigmas in the nucleotide binding site. (a) AMPPNP; (b) ADP•AlF3; (c) ADP; (d) apo structure.

Supplementary Fig. S3. The putative water molecule could act as the nucleophile for ATP hydrolysis. The yellow broken line connects the presumptive attacking water and the γ -phosphate atom.

Supplementary Fig. S4. Localization of mutated residues at BsPif1-ssDNA-ADP•AlF3 ternary complex (a) and in BsPif1-apo (b).

Supplementary Fig. S5. Comparison of BsPif1 structures. Structures are superimposed on 1A domain. (a) Comparison of AMPPNP structure (blue) on ADP-AIF3-ssDNA structure (red). C α r.m.s.d is 3.0 Å. (b). Comparison of ADP structure (green) on ADP-AIF3-ssDNA structure (red). C α r.m.s.d is 3.29Å. Structural changes are limited in BsPif1 belonging to space group P212121 C α r.m.s.d between AMPPNP and ADP is 1.23Å.

Supplementary Fig. S6. Disulphide bridge in the L95C/E339C mutant. (a) BsPif1-ssDNA-ADP•AlF3 ternary complex structure shows that the domain 2B is far away from domain 1B. (b) introducing of cysteine residues at positions of I339 and L95 establish a disulphide bridge between domains 1B and 2B. (c) The $2F_0$ -Fc electron density map contoured at 2σ level shows clearly the disulphide bridge between the two mutated residues.

Supplementary Fig. S7. (a) Presentation of the spatial configurations of the Motif A-C and PFSS motif. (b) Interactions of PFSS with the pin-loop. The residues in PFSS helix (green) and in pin-loop (orange) are shown.

Supplementary Fig. S8. Solution conformations of BsPif1. Fit of the experimental data obtained with BsPif1 protein alone to the calculated intensities from the model by CRYSOL with BsPif1 "apo" P212121 crystal form (a) and BsPif1 "complex" in P3121 crystal form (b). Initial χ^2 is 5.5 and 7.5, respectively. The residual (I_{exp}(q)-I_{calc}(q))/s_{exp} is indicated below. (c) The final χ^2 after flexible fitting is 2.2. (d) Superposition of SAXS envelope with "apo" structure (cyan) and model after flexible fitting (green) The 1B domain is in blue (X-ray structure) and orange (model).

Supplementary Fig. S9. (a-c) Comparison of the conformation of 1B domain in BsPif1, RecD2 and Dda. The structures were superimposed on 1A domain. (a) BsPif1 structure. Colors are the same as in Figure 1. (b). Comparison with RecD2 (3GPL, magenta). (c) Comparison with Dda (3UPU, yellow). (d-f) Comparison of Pin/wedge organization SF1B and SF1A helicases. (d) Structure of BsPif (this work), (e) PcrA (PDB code 3PJR) and (f) UvrD (PDB code 3LFU) have been superimposed on 1A domain and are shown in the same orientation. The wedge and the helix which may have the same function as PFSS are colored in orange and green respectively.



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В





Supplementary Figure S4





Supplementary Figure S6



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