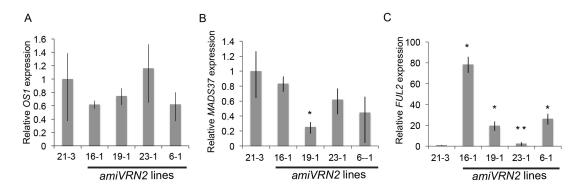
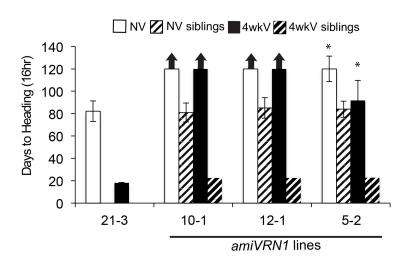


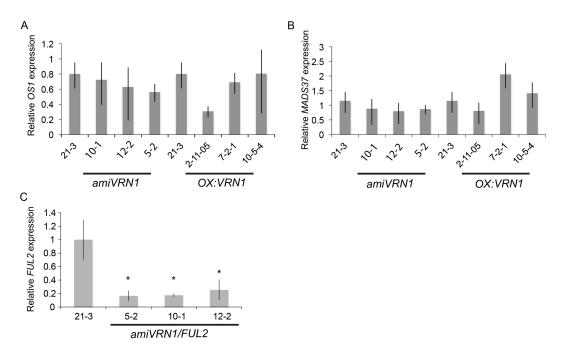
**Supplemental Figure 1.** Days to heading of Bd21-3 and *OX:VRN1* lines grown in 20,16, 15,14, 12, 10, and 8 hours of lights. Plants were were vernalized (V) as an imbibed seed for 4 weeks before outgrowth in the indicated photoperiods. Non-vernalized (NV) plants and V plants were both moved into the indicated photoperiod at the same time. Arrows indicates that no plants flowered during the course of the experiment.



Supplemental Figure 2. *BdOS1*, *BdMADS37* and *BdFUL2* gene expression in leaves of Bd21-3 and *amiVRN2* plants grown in 16h long days without vernalization. qRT-PCR expression data sampled from the youngest leaf at the third leaf stage of Bd21-3 wild-type (WT) and *amiVRN2* plants for *BdOS1* (A), *BdOS2* (B), and *BdFUL2*. Bars represent the average of three biological replicates +/- SD. Expression analysis was repeated with similar results. Single asterisks indicate P-values < 0.01 and two asterisks indicate P values < 0.05.



**Supplemental Figure 3.** Days to heading of Bd21-3, and *amiVRN1* plants non-vernalized (NV) and vernalized for 4 weeks (4wkV) at 5°C at the 3rd leaf stage in a 16-h photoperiod before outgrowth for 120 days in a 16-h photoperiod. Arrows indicates that no plants flowered during the course of the experiment. Asterisks (\*) indicates that some plants in this treatment did not flower during the course of the 120 day experiment. Siblings are plants segregating from the same primary transformant mother plant that does not carry the transgene.



**Supplemental Figure 4.** *BdOS1* (A) and *BdMADS37* (B) gene expression in the youngest leaves of Bd21-3 wild type (WT), *amiVRN1*, and *OX:VRN1* plants at the three leaf stage grown without vernalization under a 20h photoperiod. Bars represent the average of three biological replicates +/- SD. Expression analysis was repeated with similar results. (C) *BdFUL2* gene expression in the third leaf of Bd21-3 and *amiVRN1/FUL2* lines which knockdown mRNA expression of *VRN1* and *FUL2* simultaneously. See Figure 6 for *VRN1* expression in the *amiVRN1/FUL2* lines. Asterisk indicates P value <.05.

## **Supplemental Table 1**

Genotyping overexpression

pANIC 6 series AcV5 R
VRN2 cDNA F no cacc
VRN2 cDNA R no stop
amiRNA

aga cca gcc gct cgc atc ttt cca ag
ATGTCGACATGTGGCATGTGCG
TCGGAGCCATCCGAGGTCGAG

G-11491-GW-attB1-F G-11494-GW-attB2 R BdVRN2miR-sI-C BdVRN2miR-all-C BdVRN2miR\*sIII-C BdVRN2miR\*alV-C

BdVRN2miR\*aIV-C
BdVRN1miR-s1-C
BdVRN1miR-aII-C
BdVRN1miR-a\*III-C
BdVRN1miR-a\*IV-C
BdVRN1L1/2miR-sI-271
BdVRN1L1/2miR-aII-271
BdVRN1L1/2miR-\*sIII-271
BdVRN1L1/2miR-\*sIII-271

**Genotyping amiRNAs** Universal F amiRNA

Phylogenetics CO-like\_994f CO-like\_1175r for qPCR

NassPul\_qVRN2\_f NassPul\_qVRN2\_r NassPub\_qVRN2\_f NassPub\_qVRN2\_alt.r Mnutans\_5'\_\_166R A\_sativa\_VRN2\_58F A\_sativa\_VRN2\_132R Grass\_UBQ5\_F Grass\_UBQ5\_R Lolium\_EF1alphaF Lolium\_EF1alphaR BdFUL2 qPCR F1

BdFUL2 qPCR R1

GGGGACAAGTTTGTACAAAAAAGCAGGCTCATCGGATCCCAGCAGCAGCACACAAAAAAGCAGGACCACTTTGTACAAGAAAGCTGGGTCTCGGTACCGCTGCTGATGCTGATGCCAT

GGGGACCACTTTGTACAAGAAAGCTGGGTCTCGGTA
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tgCAGGTACCTATGATCCAAGTActgctgctgctacagcc
ctCAGGTTCCTTTGATCCAAGTAttcctgctgctaggctg
aaTACTTGGATCAAAGGAACCTGagagaggcaaaagtgaa
agTTCAGAAATCGACTCGCGCATcaggagattcagtttga
tgATGCGCGAGTCGATTTCTGAActgctgctgctacagcc
ctATGCGGGAGACGATTTCTGAAttcctgctgctaggctg
aaTTCAGAAATCGTCCCGCATagagaggccaaaagtgaa
agTTTCCTATATTCGTGACGCCAcaggagattcagtttga
tgTGGCGTCACGAATATAGGAAActgctgctgctacagcc
ctTGGCGACACCAATATAGGAAAAttcctgctgctacagcc
aaTTTCCTATATTCGTGCCCAagagaggcaaaagtgaa

ctgatgcatatacatgatggc

GAGAAGCARATCCGSTAYGMGTC CGGAACCAYCCGAGGTSRAG

AGGAAAGCTTACGCCGAGAT
GGTGGAGTCTGCTTGGATGT
GGTACGAGTCCAGGAAAGCA
GAGGTCGAGTCTCCTTGGATGT
CTGCAGTGGTCTCATTTCCTC
GTTTGTTTGCTCTGGTGGTGT
CAAATCCGGTATGAGTCAAGAAAA
ATGTACCTTGGCGAAGCGGCCT
CGCCGACTACAACATCCAG
TCACCTTCTTGTGCTCTTGC
CCTTGCTTGAGGCTCTTGAC
GTTCCAATGCCACCAATCTT
CAAGGGCAAGCTCTACGAGT
AAGAGCCTTTTTCAGCATACGA

**Supplemental Figure 5. Alignment of** *VRN2/GhD7/CO9* **genes and outgroups used for phylogenetic analysis**. Apparent frameshifts in the alignment are due to the removal of unalignable regions, which resulted in higher support values at several nodes, presumably due to reduction of random noise in the data versus true phylogenetic signal. See nexus file.