

Supplementary Information

Title: Integrated Platform for Three-dimensional Quantitative Analysis of Spatially Heterogeneous Metastasis Landscape

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Supplementary Legends

Supplementary Figure S1 | Tissue optical clearing based imaging pipeline for molecular

phenotyping of brain metastasis. (a) Schematic for the timeline of the experimental pipeline.

(b) Brain tissue slice before (top) and after (bottom) tissue clearing and refractive index

matching. **(c)** 3D multiphoton raw image (top) and surface generated image (bottom)

demonstrating multiplexed staining for metastases (K8, green), EdU-tagged nuclei (orange), and

astrocytes (GFAP, red). **(d)** 3D surface generated image of highly branched vasculature in mouse

brain (red).

Supplementary Figure S2 | Metastasis segmentation based on tumor specific staining and

nuclear of tumor cells. (a) Image of anti-cytokeratin K8 staining (K8, red) detected in

leptomeninges of normal brain. Left DAPI stain (blue), middle K8 stain, right merge. **(b)** 2D

overlay of K8 and DAPI (colored images; K8, green; DAPI, blue) and corresponding DAPI/K8

segmented images (white) in two metastasis models. **(c)** 3D DAPI staining and overlay of K8

and DAPI (colored images; K8, green; DAPI, blue) and corresponding DAPI/K8 segmented

images (white) in two metastasis models. **(d)** 3D multiphoton image of DAPI (blue) and K8

(green) stained PNA.Met1 metastases (left), corresponding segmented 3D image (middle), and

overlap of original image and segmented data demonstrating nearness of segmentation to the

ground truth (right).

Supplementary Figure S3 | 3D segmentation of astrocytes. (a) Schematic representing the

incomplete view of astrocytes obtained by 2D imaging (left) and a 2D multiphoton image of

astrocytes (GFAP, red). **(b)** 3D multiphoton images captures gross astrocyte morphology, including fine protrusions. **(c)** 2D segmentation of astrocytes.

Supplementary Figure S4 | Heterogeneous Astrocyte Response to MDA.MB.231br Brain

Metastases. (a) 3D surface generated image of global MDA-MB-231.Br brain metastases (green) and associated astrocytes (red) (left), demonstrating high gliosis levels (top right) and low gliosis levels (bottom right). **(b)** Plot of relationship between the proliferative index and the square root (Sqrt) of the gliosis index for small (left), medium (middle), and large (right) MDA-MB-231.Br brain metastases.

Supplementary Figure S5 | 3D reconstructed view of metastasis associated vasculature. (a)

Representative surface generated images global (left), surface (middle), and intratumoral (right) metastasis-associate vasculature in tumor B and tumor D. **(b)** Schematic of concentric zone analysis of intratumoral and peritumoral vascularization index. **(c)** Plots of vascularization index per intratumoral zones (negative numbers) and peritumoral zones (positive numbers) for metastases A, B, C, and D. $\text{Vascularization index} = \text{Total vasculature voxel volume} / \text{Total zone voxel volume}$.

Supplementary Video S1 | Global View of MDA-MB-231.Br Brain Metastasis Landscape.

3D view of MDA-MB-231.Br brain metastases identified by DAPI (blue) clustering and astrocytes (GFAP, orange) demonstrating both the global view achieved with the simultaneous ability to reach single cell resolution.

Supplementary Video S2 | Multiplexed Staining of MDA-MB-231.Br Brain Metastasis

Landscape. 3D view of MDA-MB-231.Br brain metastasis landscape with multiplexed staining for brain metastases (K8, green), nuclei (DAPI, blue), EdU-tagged nuclei (EdU, purple), and astrocytes (GFAP, orange). Brain metastasis staining reveals highly irregular shape of MDA-MB-231.Br brain metastases and proliferative heterogeneity.

Supplementary Video S3 | Multiplexed Staining of PNA.Met1 Brain Metastasis Landscape.

3D view of PNA.Met1 brain metastasis landscape with multiplexed staining for brain metastases (K8, green), EdU-tagged nuclei (EdU, orange), and astrocytes (GFAP, red). Brain metastasis staining reveals highly irregular shape of PNA.Met1 brain metastases and proliferative heterogeneity.

Supplementary Video S4 | 3D View of PNA.Met1 Metastasis-Associated Vasculature. 3D

surface generation of metastasis-associated vasculature (red) shows thin, highly branched vessels on the metastasis surface and few, but large blood vessels infiltrating the tumor.

Supplementary Table 1 Performance evaluation of the imaging segmentation classifier

| | Tumor | Astrocytes | Proliferation | Vessels |
|-----------|--------|------------|---------------|---------|
| Precision | 0.9698 | 0.9352 | 0.9200 | 0.9333 |
| Recall | 0.9373 | 0.9349 | 0.9983 | 0.9507 |
| Accuracy | 0.9963 | 0.9758 | 0.9971 | 0.9980 |
| F1 Score | 0.9532 | 0.9350 | 0.9575 | 0.9417 |

Note: The F1 score is the harmonic mean of precision and recall and is a widely used measure to evaluate the performance of the classifier. The error of the volume quantification can be calculated by the following equation:

$$Error_{volume} = \frac{Calculated\ Volume}{True\ Volume} - 1 = \frac{TP + FP}{TP + FN} - 1 = \frac{Recall}{Precision} - 1$$

In this equation, TP denotes true positive, FP denotes false positive, and FN denotes false negative. According to the evaluation, the error of the tumor volume quantification is around -3.35%, the error of the astrocytes volume quantification is around -0.03%, the error of proliferating cells volume quantification is around +8.51%, and the error of the blood vessel volume quantification is around +1.86%.

The error of the ratio quantification can be calculated by the following equation:

$$\begin{aligned} Error_{ratio} &= \frac{Calculated\ Ratio}{True\ Ratio} - 1 \\ &= \frac{Calculated\ volume_{other\ component} / Calculated\ volume_{tumor}}{True\ volume_{other\ component} / True\ volume_{tumor}} - 1 \\ &= \frac{1 + Error_{volume\ other\ component}}{1 + Error_{volume\ tumor}} - 1 \end{aligned}$$

The error of the tumor-astrocyte ratio quantification is around +3.44%, the error of the tumor-proliferating cells ratio quantification is around +12.27%, and the error of the tumor-vessel ratio quantification is around +5.39%.

Supplementary Table 2 Dyes, Stains, and Antibodies Compatible with Various Tissue Clearing

Methods

| Dye/antibody | Source | Application | SMART 3D compatibility | Reference |
|--|-----------------------------|----------------------------|------------------------|----------------------|
| DAPI | Life Technologies (D1306) | nuclear staining | Yes | SMART3D |
| Hoechst | Life Technologies (H3570) | nuclear staining | Yes | SMART3D |
| Dil/DiO/DiR | Life Technologies (V-22889) | membrane staining | Yes (with Scale S) | SMART3D |
| EdU | Life Technologies (C10339) | proliferation marker | Yes | SMART3D |
| mouse anti-GFAP | CST (3670S) | astrocyte marker | Yes | SMART3D |
| rabbit anti-GFAP | CST (12389) | astrocyte marker | Yes | SMART3D |
| rabbit anti-cytokeratin 8 | Abcam (ab53280) | epithelial cell marker | Yes | SMART3D |
| dextran, fixable, Alexa-fluor conjugated | Life Technology (D22914) | pseudo-blood vessel marker | Yes | SMART3D |
| Parvalbumin | Abcam | neuron marker | Not Tested | CLARITY ¹ |
| Tyrosine Hydroxylase | Abcam | neuron marker | Not Tested | CLARITY ¹ |
| Propidium Iodide | Life Technology | nuclear staining | Yes | CLARITY ¹ |
| anti-Neurofilament NF-H | Aves | neuron marker | Not Tested | CLARITY ¹ |
| anti-GFP | Invitrogen | GFP | Not Tested | CLARITY ¹ |
| mouse anti- α -smooth muscle actin | Sigma (C6198) | vascular structure marker | Not Tested | CUBIC ² |
| mouse anti-Pan cytokeratin antibody | Abcam (ab78478) | epithelial cell marker | Not Tested | CUBIC ² |
| chicken anti-tyrosine hydroxylase | Aves | neuron marker | Not Tested | PACT ³ |
| rabbit anti-integrin b4 | Santa Cruz | general cellular staining | Not Tested | PACT ³ |
| rabbit anti-b5 IgG | Santa Cruz | general cellular staining | Not Tested | PACT ³ |
| rabbit anti-beta tubulin IgG | Santa Cruz | beta tubulin marker | Not Tested | PACT ³ |
| NeuroTrace 530 / 615 Red Fluorescent Nissl Stain | Life Technology (N-21482) | neuron marker | Yes | PACT ³ |
| mouse anti-amyloid- β | Covance (SIG-39347) | amyloid plaque makerer | Not Tested | ScaleS ⁴ |

| | | | | |
|-------------------------------------|---------------------------|-------------------------------------|------------|---------------------|
| rabbit anti-NeuN | Millipore (ABN78C3) | neuron marker | | ScaleS ⁴ |
| rabbit anti-Iba1 | Wako (019-19741) | macrophage/microglia marker | Yes | ScaleS ⁴ |
| goat anti-TrkC | R & D Systems (AF1404) | DRG neuron marker | Not Tested | iDISCO ⁵ |
| goat anti-mouse TrkB | R & D Systems (AF1494) | DRG neuron marker | Not Tested | iDISCO ⁵ |
| goat anti-human ROBO3 | R & D Systems (AF3076) | roundabout homolog 3 | Not Tested | iDISCO ⁵ |
| goat anti-mouse Ret | R & D Systems (AF482) | DRG neuron marker | Not Tested | iDISCO ⁵ |
| goat anti-choline acetyltransferase | Millipore (AB114P) | cholinergic neuron marker | Not Tested | iDISCO ⁵ |
| anti-tyrosine hydroxylase | Millipore (AB152) | dopaminergic neuron marker | Not Tested | iDISCO ⁵ |
| guinea pig anti-VGLUT2 | Millipore (AB2251) | vesicular glutamate transporter 2 | Not Tested | iDISCO ⁵ |
| mouse anti-neuN | Millipore (MAB377) | neuronal nuclei | Not Tested | iDISCO ⁵ |
| rabbit anti-c-Fos | Santa Cruz (sc-52) | neuron activity marker | Not Tested | iDISCO ⁵ |
| goat anti-AQP2 | Santa Cruz (sc-9882) | kidney collection duct marker | Not Tested | iDISCO ⁵ |
| rabbit anti-E-cadherin | CST (3195) | intestinal microvillousities marker | Not Tested | iDISCO ⁵ |
| anti-cleaved caspase 9 | CST (9509) | apoptotic cell marker | Not Tested | iDISCO ⁵ |
| anti-cleaved caspase 3 | CST (9661) | apoptotic cell marker | Not Tested | iDISCO ⁵ |
| rabbit anti-N cadherin | Abcam (ab12221) | cadherin-2 | Not Tested | iDISCO ⁵ |
| rabbit anti-FOXP2 | Abcam (ab16046) | developing neuron marker | Not Tested | iDISCO ⁵ |
| rabbit anti-histone H3 | Abcam (ab5176) | mitotic cell maker | Not Tested | iDISCO ⁵ |
| rabbit anti-laminin | Sigma-Aldrich (L9393) | kidney tubule maker | Not Tested | iDISCO ⁵ |
| rat anti-mouse CD31 | BD Bio (550274) | vasculature marker | Not Tested | iDISCO ⁵ |
| rabbit anti-GFP | Life Technology (CAB4211) | GFP | Yes | iDISCO ⁵ |

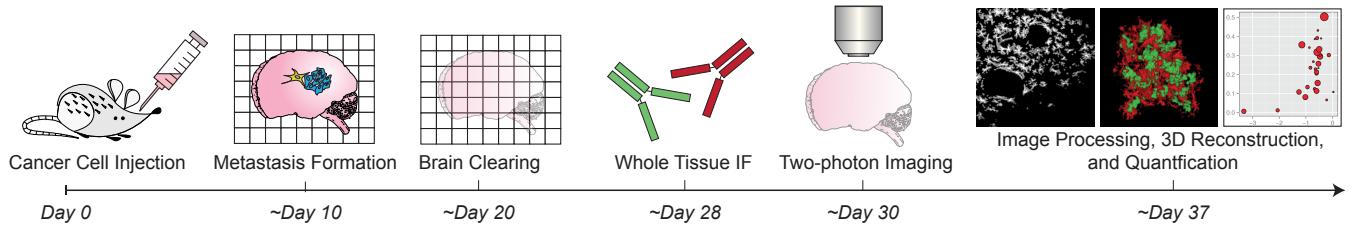
Reference cited:

1. Chung, K. *et al.* Structural and molecular interrogation of intact biological systems. *Nature* **497**, 332–337 (2013).
2. Tainaka, K. *et al.* Whole-body imaging with single-cell resolution by tissue decolorization. *Cell* **159**, 911–924 (2014).
3. Yang, B. *et al.* Single-cell phenotyping within transparent intact tissue through whole-body clearing. *Cell* **158**, 945–958 (2014).
4. Hama, H. *et al.* ScaleS: an optical clearing palette for biological imaging. *Nat. Neurosci.* **18**, 1518–1529 (2015).
5. Renier, N. *et al.* iDISCO: A Simple, Rapid Method to Immunolabel Large Tissue Samples for Volume Imaging. *Cell* **159**, 896–910 (2014).

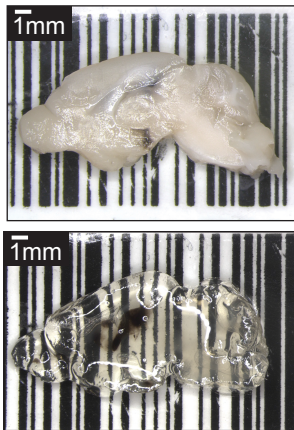
Supplementary Figure 1

a

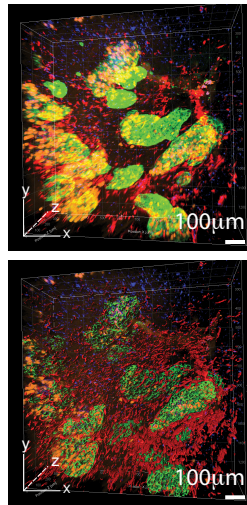
Timeline for Applying Integrated Platform



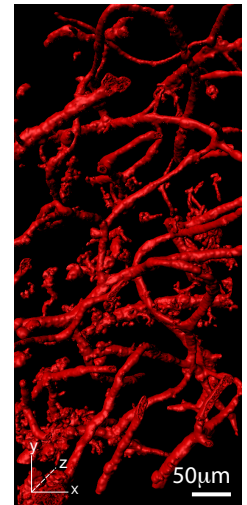
b



c

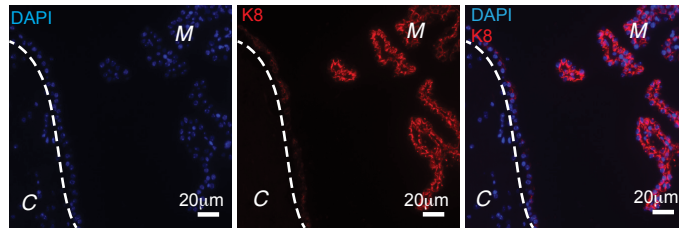


d



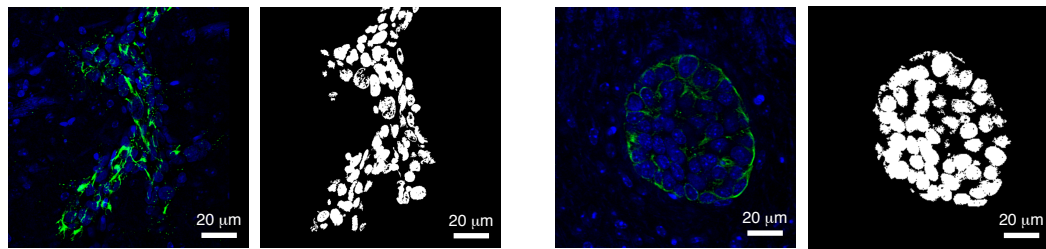
Supplementary Figure 2

a



C: cerebral cortex M: meningeal structures

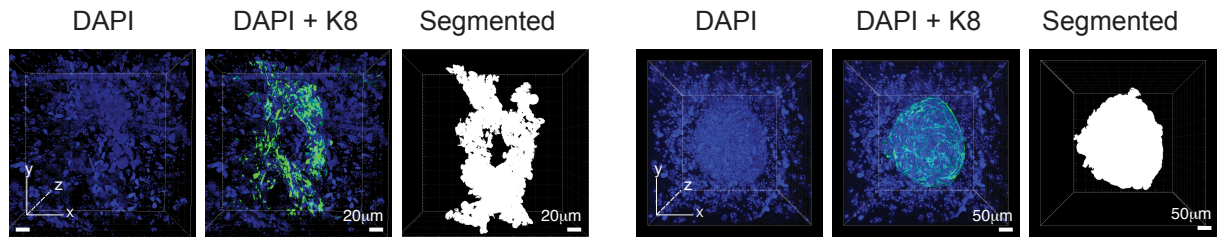
b



MDA-MB-231.Br

PNA.Met1

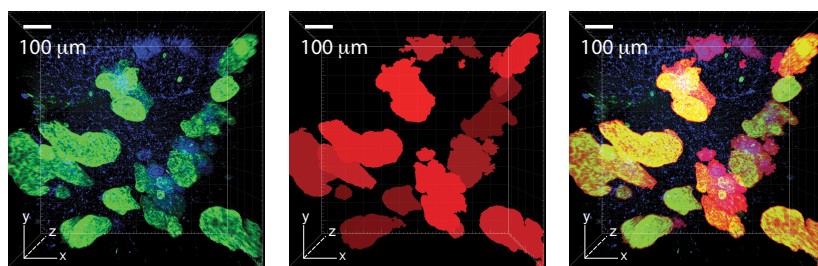
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MDA-MB-231.Br

PNA.Met1

d



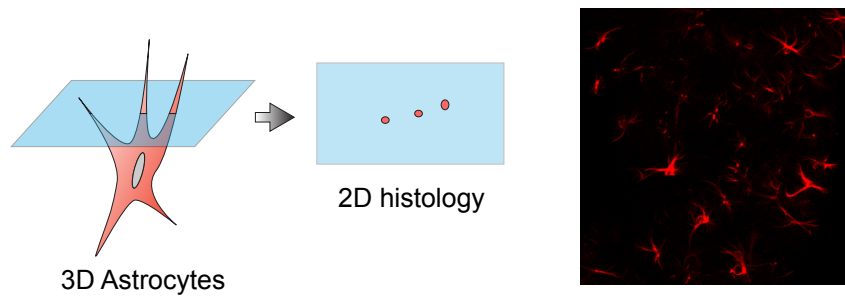
Original

Segmented

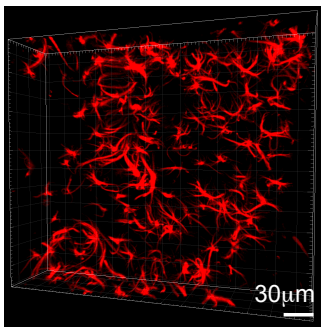
Overlay

Supplementary Figure 3

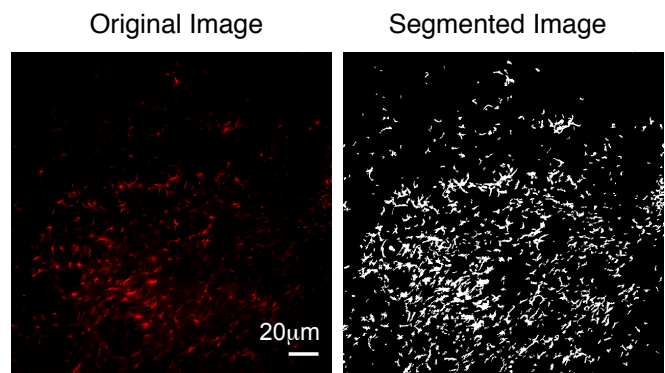
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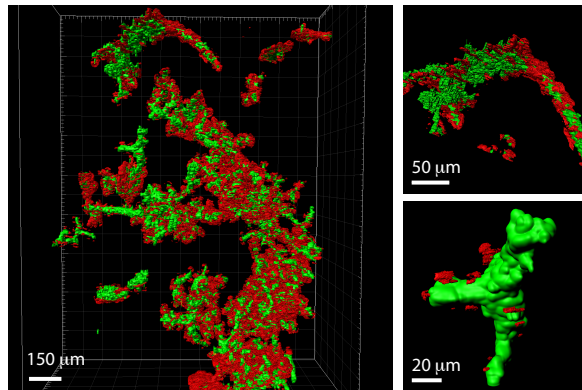


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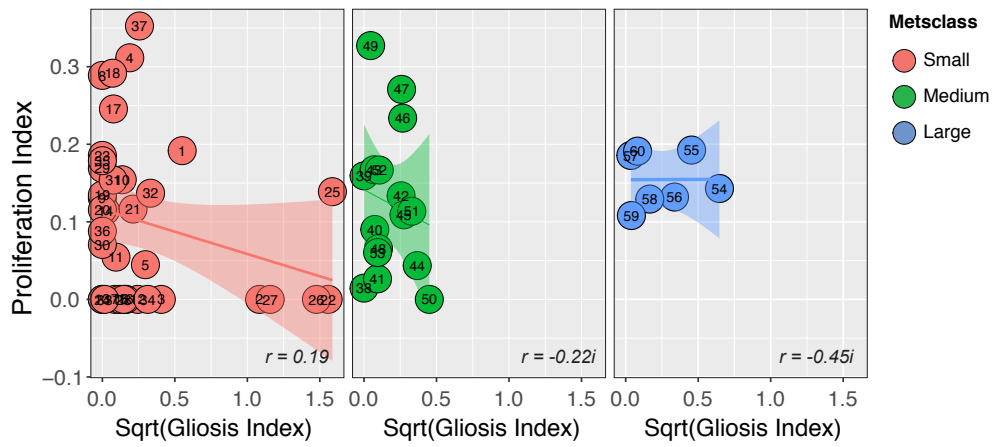


Supplementary Figure 4

a

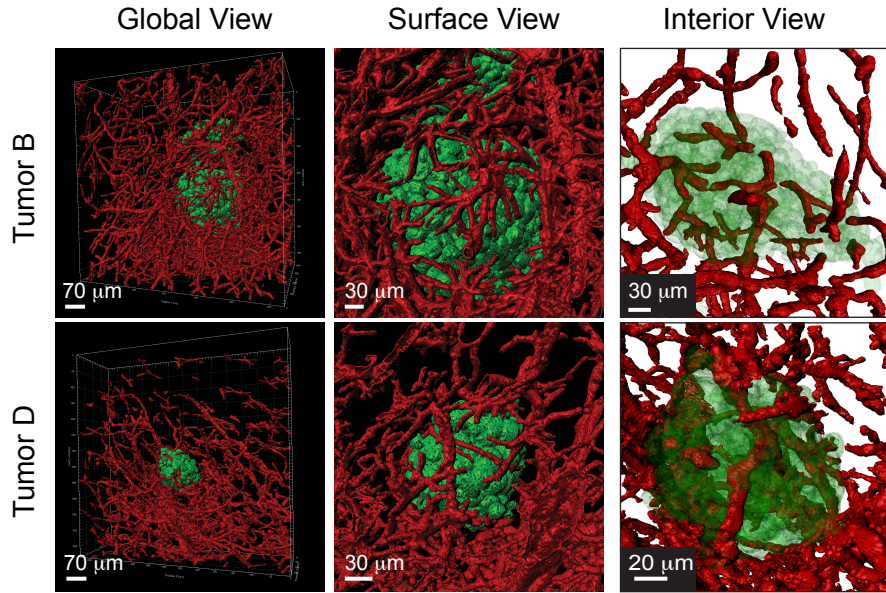


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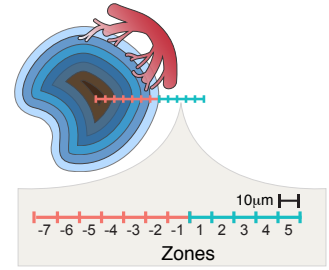


Supplementary Figure 5

a



b



c

