

Within the *blood vessel* network, nine of IL-1 $\beta$ 's top 20 neighbors are not top neighbors in the tissue-naïve network. Eight of these genes are transcriptionally regulated in vascular cells based on IL-1 $\beta$  treatment (Fig. 2b). For example, MCL-1 is an anti-apoptotic gene involved in survival of endothelial cells<sup>1,2</sup>. It is rapidly induced by growth and inflammatory stimuli in vascular cells<sup>3,4</sup> to control vascular apoptosis<sup>5</sup>. PAI-1 (also called SERPINE1) is produced and secreted by vascular cells<sup>6</sup> in response to various agonists such as hormones, growth factors, endotoxin, cytokines and phorbol esters<sup>7</sup>. Specifically, IL-1 $\beta$  increases PAI-1 mRNA and protein expression in a dose- and time-dependent manner in endothelial cells<sup>8</sup>. PAI-1 plays an important role in several vascular pathologies, including atherosclerosis and vascular remodeling<sup>6,9</sup>. TNFAIP2 (also B94) is known as a primary inflammatory response gene induced in endothelial cells by inflammatory stimuli, including IL-1 $\beta$ <sup>10</sup>; it is also induced in endothelial progenitor cells involved in vascular repair<sup>11</sup>. Chemokines CXCL2 (GRO-beta) and CXCL3 (GRO-gamma) are known to function in the endothelial inflammatory response<sup>12-16</sup>. A20 (also known as TNFAIP3) is induced in response to injury and to inflammatory stimuli in vascular cells<sup>17</sup>. A20 is involved in neovascularization that is strongly induced by and plays a role downstream of IL-1 $\beta$  in vascular endothelial and smooth muscle cells<sup>17-19</sup>. Inflammation-induced IER3/IEX-1<sup>20</sup> plays a role in vascular endothelial and smooth muscle physiology and cardiovascular disease<sup>21-24</sup>. IL-8 is involved in the establishment and preservation of the inflammatory microenvironment of the blood vessel, and is implicated in cardiovascular diseases including atherosclerosis, hypertension and coronary artery disease (CAD)<sup>25</sup>.

The ninth gene, GLIPR1, does not show a transcriptional response but is among the top genes linked to IL-1 $\beta$  in the *blood vessel* network. GLIPR1 belongs to the CAP superfamily, several members of which are involved in innate immune response<sup>26,27</sup>, and has links to endothelial inflammatory modulation<sup>28-30</sup> and angiogenesis<sup>31</sup>. GLIPR1 has been shown to be regulated post-transcriptionally, potentially via subcellular localization<sup>32</sup>, and thus lack of differential expression in steady state levels of its mRNA might not be unexpected. This is similar to what we observe for IL-1R, the receptor for IL-1 $\beta$ , which is internalized upon treatment and precedes IL-1 $\beta$  induced gene-expression<sup>33</sup>. Altogether, this demonstrates that tissue-specific functional networks discover both tissue-specific transcriptional and post-transcriptional regulatory responses.

## References

1. Erwert, R. D. *et al.* Shiga toxin induces decreased expression of the anti-apoptotic protein Mcl-1 concomitant with the onset of endothelial apoptosis. *Microb Pathog* **35**, 87–93 (2003).
2. Ikezoe, T. *et al.* Thrombomodulin protects endothelial cells from a calcineurin inhibitor-induced cytotoxicity by upregulation of extracellular signal-regulated kinase/myeloid leukemia cell-1 signaling. *Arter. Thromb Vasc Biol* **32**, 2259–2270 (2012).
3. Zhang, D., Li, F., Weidner, D., Mnjoyan, Z. H. & Fujise, K. Physical and functional interaction between myeloid cell leukemia 1 protein (MCL1) and Fortilin. The potential role of MCL1 as a fortilin chaperone. *J Biol Chem* **277**, 37430–37438 (2002).
4. Bannerman, D. D. *et al.* A constitutive cytoprotective pathway protects endothelial cells from lipopolysaccharide-induced apoptosis. *J. Biol. Chem.* **276**, 14924–32 (2001).
5. Yang, Z. *et al.* Cardiovascular inflammation and lesion cell apoptosis: a novel connection via the interferon-inducible immunoproteasome. *Arter. Thromb Vasc Biol* **29**, 1213–1219 (2009).
6. Diebold, I., Kraicun, D., Bonello, S. & Görlach, A. The “PAI-1 paradox” in vascular remodeling. *Thromb. Haemost.* **100**, 984–91 (2008).
7. Lijnen, H. R. & Collen, D. Mechanisms of physiological fibrinolysis. *Baillieres. Clin. Haematol.* **8**, 277–90 (1995).
8. Schleef, R. R., Bevilacqua, M. P., Sawdey, M., Gimbrone Jr., M. A. & Loskutoff, D. J. Cytokine activation of vascular endothelium. Effects on tissue-type plasminogen activator and type 1 plasminogen activator inhibitor. *J Biol Chem* **263**, 5797–5803 (1988).
9. Lijnen, H. R. Pleiotropic functions of plasminogen activator inhibitor-1. *J. Thromb. Haemost.* **3**, 35–45 (2005).
10. Sarma, V., Wolf, F. W., Marks, R. M., Shows, T. B. & Dixit, V. M. Cloning of a novel tumor necrosis factor-alpha-inducible primary response gene that is differentially expressed in development and capillary tube-like formation in vitro. *J Immunol* **148**, 3302–3312 (1992).
11. Navarro-Sobrinho, M. *et al.* The angiogenic gene profile of circulating endothelial progenitor cells from ischemic stroke patients. *Vasc Cell* **5**, 3 (2013).
12. Okada, M. *et al.* Detection of up-regulated genes in thrombin-stimulated human umbilical vein endothelial cells. *Thromb Res* **118**, 715–721 (2006).
13. Mattaliano, M. D. *et al.* LOX-1-dependent transcriptional regulation in response to oxidized LDL treatment of human aortic endothelial cells. *Am J Physiol Cell Physiol* **296**, C1329–37 (2009).
14. Hamid, C. *et al.* Anti-beta2GPI-antibody-induced endothelial cell gene expression profiling reveals induction of novel pro-inflammatory genes potentially involved in primary antiphospholipid syndrome. *Ann Rheum Dis* **66**, 1000–1007 (2007).

15. Krogmann, A. *et al.* Inflammatory response of human coronary artery endothelial cells to saturated long-chain fatty acids. *Microvasc Res* **81**, 52–59 (2011).
16. Gargalovic, P. S. *et al.* The unfolded protein response is an important regulator of inflammatory genes in endothelial cells. *Arter. Thromb Vasc Biol* **26**, 2490–2496 (2006).
17. Verstrepen, L. *et al.* Expression, biological activities and mechanisms of action of A20 (TNFAIP3). *Biochem Pharmacol* **80**, 2009–2020 (2010).
18. Dixit, V. M. *et al.* Tumor necrosis factor- $\alpha$  induction of novel gene products in human endothelial cells including a macrophage-specific chemotaxin. *J Biol Chem* **265**, 2973–2978 (1990).
19. Heyninck, K. & Beyaert, R. The cytokine-inducible zinc finger protein A20 inhibits IL-1-induced NF- $\kappa$ B activation at the level of TRAF6. *FEBS Lett* **442**, 147–150 (1999).
20. Arlt, A. & Schafer, H. Role of the immediate early response 3 (IER3) gene in cellular stress response, inflammation and tumorigenesis. *Eur J Cell Biol* **90**, 545–552 (2011).
21. Sommer, S. L. *et al.* Elevated blood pressure and cardiac hypertrophy after ablation of the gly96/IEX-1 gene. *J Appl Physiol* **100**, 707–716 (2006).
22. Shahid, M. *et al.* Impaired 3',5'-cyclic adenosine monophosphate-mediated signaling in immediate early responsive gene X-1-deficient vascular smooth muscle cells. *Hypertension* **56**, 705–712 (2010).
23. De Keulenaer, G. W. *et al.* Identification of IEX-1 as a biomechanically controlled nuclear factor- $\kappa$ B target gene that inhibits cardiomyocyte hypertrophy. *Circ Res* **90**, 690–696 (2002).
24. Schulze, P. C. *et al.* Biomechanically induced gene iex-1 inhibits vascular smooth muscle cell proliferation and neointima formation. *Circ Res* **93**, 1210–1217 (2003).
25. Apostolakis, S., Vogiatzi, K., Amanatidou, V. & Spandidos, D. A. Interleukin 8 and cardiovascular disease. *Cardiovasc Res* **84**, 353–360 (2009).
26. Gibbs, G. M., Roelants, K. & O'Bryan, M. K. The CAP superfamily: cysteine-rich secretory proteins, antigen 5, and pathogenesis-related 1 proteins--roles in reproduction, cancer, and immune defense. *Endocr Rev* **29**, 865–897 (2008).
27. Ren, C., Ren, C. H., Li, L., Goltsov, A. A. & Thompson, T. C. Identification and characterization of RTVP1/GLIPR1-like genes, a novel p53 target gene cluster. *Genomics* **88**, 163–172 (2006).
28. Wang, Y. L. *et al.* Cobra CRISP functions as an inflammatory modulator via a novel Zn<sup>2+</sup>- and heparan sulfate-dependent transcriptional regulation of endothelial cell adhesion molecules. *J Biol Chem* **285**, 37872–37883 (2010).

29. Bonura, A. *et al.* Cloning and expression of a novel component of the CAP superfamily enhanced in the inflammatory response to LPS of the ascidian *Ciona intestinalis*. *Cell Tissue Res* **342**, 411–421 (2010).
30. Szyperski, T., Fernandez, C., Mumenthaler, C. & Wuthrich, K. Structure comparison of human glioma pathogenesis-related protein GliPR and the plant pathogenesis-related protein P14a indicates a functional link between the human immune system and a plant defense system. *Proc Natl Acad Sci U S A* **95**, 2262–2266 (1998).
31. Satoh, T. *et al.* Adenoviral vector-mediated mRTVP-1 gene therapy for prostate cancer. *Hum Gene Ther* **14**, 91–101 (2003).
32. Asojo, O. A., Koski, R. A. & Bonafe, N. Structural studies of human glioma pathogenesis-related protein 1. *Acta Crystallogr D Biol Crystallogr* **67**, 847–855 (2011).
33. Hansen, B., Dittrich-Breiholz, O., Kracht, M. & Windheim, M. Regulation of NF-kappaB-dependent gene expression by ligand-induced endocytosis of the interleukin-1 receptor. *Cell Signal* **25**, 214–228 (2013).