

Increased N6-methyladenosine in Human Sperm RNA as a Risk Factor for Asthenozoospermia

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Fig. S1. Melt curves and standard curves of real-time PCR for *METTL3* and *METTL14*. (A-B), melt curve and standard curve for *METTL3*. (C-D), melt curve and standard curve for *METTL14*.

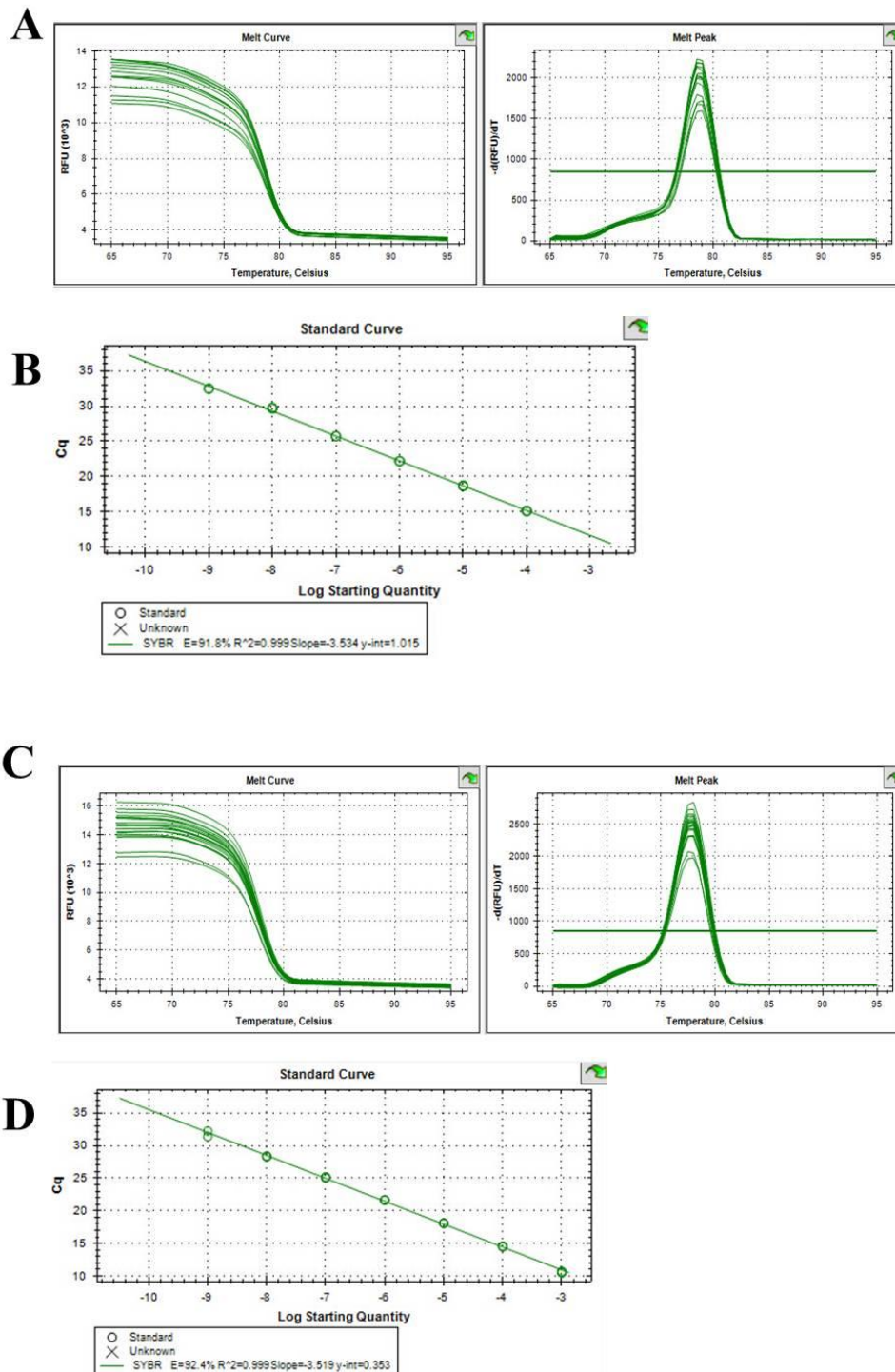


Fig. S2. Correlation of m⁶A contents with gene expressions in sperm RNA. (A-E), Correlation analysis between m⁶A contents and mRNA expression in all subjects.

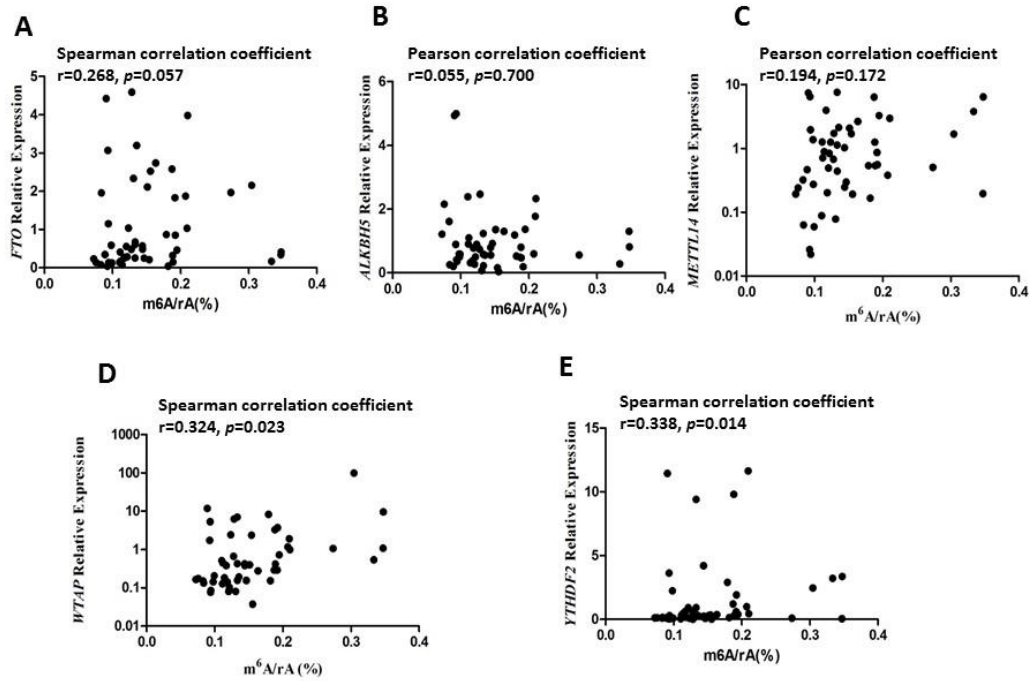


Fig. S3. Microscopic examination of the sperms before and after purification (400×).

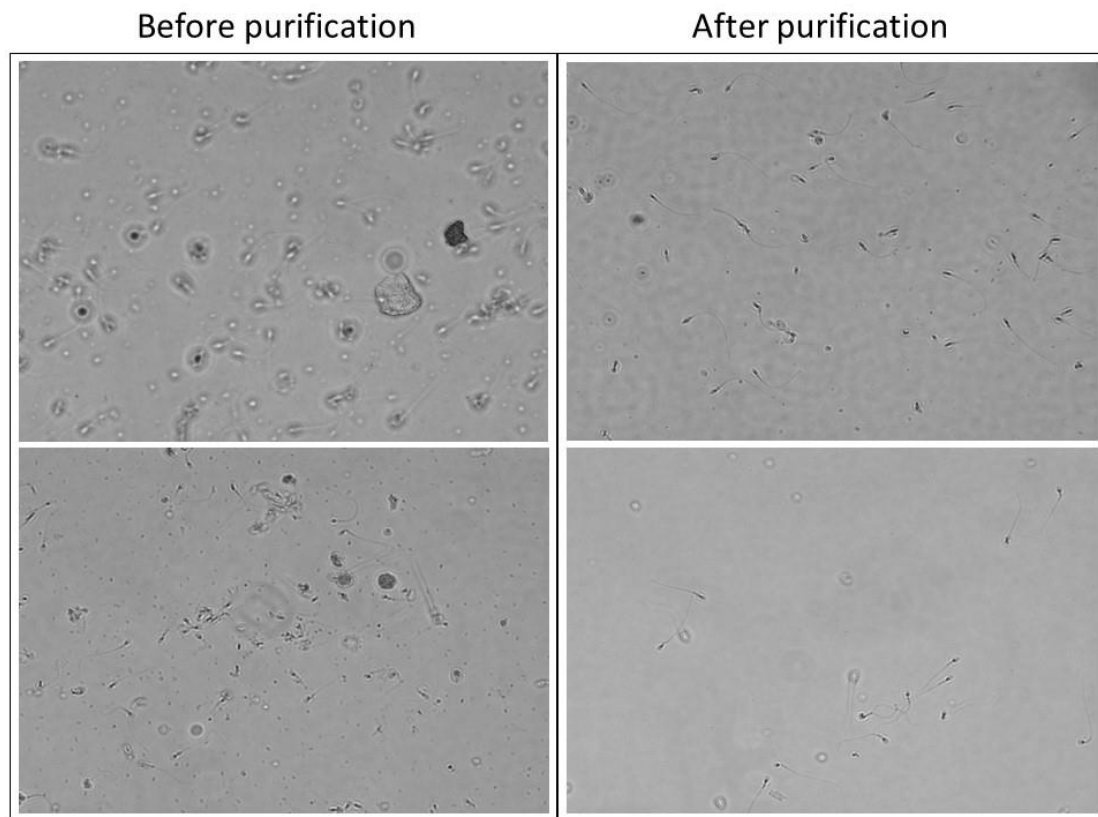


Table S1. Linearity, LOD and LOQ of m⁶A by LC-ESI-MS/MS.

Analyte	Linear range (m ⁶ A/rA,%)	Regression line		R ²	LOD (fmol)	LOQ (fmol)
		Slope	Intercept			
m ⁶ A	0.02-2	0.0396 ± 0.0003	0.0012 ± 0.0001	0.997 7	1.2	4.0

Table S2. Preparation of the quality control (QC) samples with m⁶A-containing oligonucleotides.

Methylation level	Molar ratio (%)	
	m ⁶ A/A (%)	RNA standard 1
0.050	0.16	99.84
0.100	0.30	99.70
0.300	0.91	99.09
RNA standard 1 (15-mer RNA)	5'- UAAUAC(m ⁶ A)GAGAAAUC-3'	
RNA standard 2 (10-mer RNA)	5'-AUCUAUAUGC-3'	

Table S3. Validation of the method for the detection of m⁶A with QC samples.

Nominal [m⁶A]/[A],%		0.050	0.100	0.300
	Measured mean			
Day 1	[m ⁶ A]/[A],%	0.044	0.091	0.292
<i>n</i> =3	RSD ^a (%)	5.7	4.3	1.8
	RE ^b (%)	-12.1	-9.2	-2.7
	Measured mean			
Day 2	[m ⁶ A]/[A],%	0.055	0.093	0.321
<i>n</i> =3	RSD (%)	9.7	4.6	4.4
	RE (%)	10.1	-7.4	7.1
	Measured mean			
Day 3	[m ⁶ A]/[A],%	0.053	0.106	0.283
<i>n</i> =3	RSD (%)	10.6	5.1	4.8
	RE (%)	6.4	5.6	-5.7

^a Relative standard deviation. ^b Relative error.

Table S4. The results of m⁶A contents in semen RNA

Sample Name	m6A/A(%)	SD	RSD
C1	0.15	0.007	4.69
C2	0.12	0.012	10.42
C3	0.19	0.005	2.52
C4	0.11	0.021	18.62
C5	0.08	0.005	6.72
C6	0.10	0.012	11.95
C7	0.08	0.007	8.99
C8	0.09	0.003	2.80
C9	0.14	0.001	0.85
C10	0.11	0.011	9.45
C11	0.09	0.009	9.74
C12	0.12	0.007	6.10
C13	0.13	0.014	10.56
C14	0.12	0.004	3.85
C15	0.07	0.006	8.38
C16	0.09	0.000	0.16
C17	0.11	0.006	4.98
C18	0.08	0.007	8.72
C19	0.16	0.000	0.15
C20	0.13	0.005	3.81
C21	0.19	0.029	15.28
C22	0.13	0.006	4.71
C23	0.21	0.003	1.52
C24	0.21	0.006	3.00
C25	0.16	0.012	7.85
C26	0.19	0.033	17.02
C27	0.12	0.001	0.86
C28	0.09	0.001	1.21
C29	0.21	0.005	2.21
C30	0.18	0.010	5.80
C31	0.14	0.014	9.78
C32	0.15	0.013	8.68
A1	0.35	0.010	2.95
A2	0.11	0.003	2.61
A3	0.14	0.008	5.88
A4	0.18	0.014	7.65
A5	0.13	0.000	0.19
A6	0.10	0.012	12.70
A7	0.27	0.004	1.60
A8	0.19	0.002	0.98

A9	0.19	0.011	5.58
A10	0.15	0.011	7.12
A11	0.12	0.011	8.71
A12	0.09	0.005	5.12
A13	0.33	0.029	8.61
A14	0.35	0.037	10.60
A15	0.13	0.004	3.35
A16	0.09	0.004	4.02
A17	0.10	0.012	11.89
A18	0.19	0.021	10.97
A19	0.30	0.008	2.73
A20	0.13	0.003	2.05

Table S5. Primers used for real-time PCR

Genes	Primers
GAPDH	sense 5'-TCTATAAATTGAGCCCGCAGC-3'
	anti-sense 5'-CCAATACGACCAAATCCGTTG-3'
FTO	sense 5'-CTTCACCAAGGAGACTGCTATTTTC-3'
	anti-sense 5'-CAAGGTTCCCTGTTGAGCACTCTG-3'
ALKBH5	sense 5'-TCCAGTTCAAGCCTATTTCG-3'
	anti-sense 5'-CATCTAATCTTGTCTTCCTGAG-3'
METTL14	sense 5'-GAGTGTGTTTACGAAAATGGGGT-3'
	anti-sense 5'-CCGTCTGTGCTACGCTTCA-3'
METTL3	sense 5'-AGATGGGGTAGAAAGCCTCCT-3'
	anti-sense 5'-TGGTCAGCATAGGTTACAAGAGT-3'
WTAP	sense 5'-TTGTAATGCGACTAGCAACCAA-3'
	anti-sense 5'-GCTGGGTCTACCATTGTTGATCT-3'
YTHDF2	sense 5'-CCTTAGGTGGAGCCATGATTG-3'
	anti-sense 5'-TCTGTGCTACCCAACCTTCAGT-3'