Supplemental Materials Molecular Biology of the Cell

Wei et al.

Supplementary Material

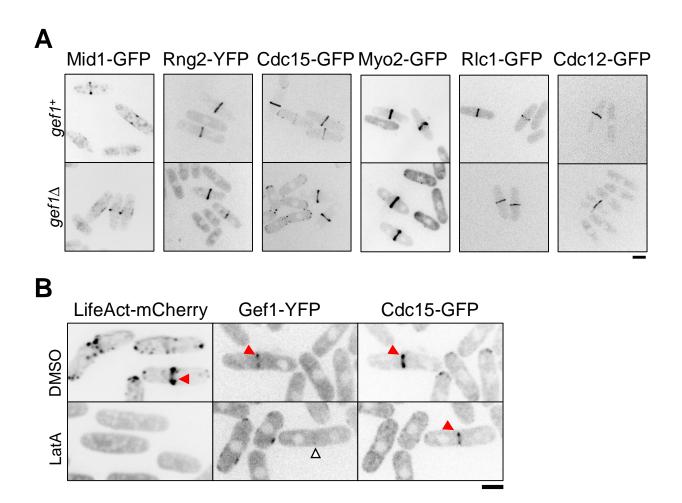
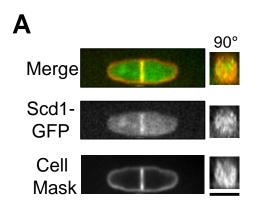


Figure S1: A. Gef1 is not required for recruitment of actomyosin ring assembly proteins. Localization of Mid1-GFP, Rng2-YFP, Cdc15-GFP, Myo2-GFP, Rlc1-GFP and Cdc12-GFP in *gef1*⁺ and *gef1* Δ cells were observed at the site of cell division. **B. Gef1 localization to the division site is actin dependent.** Cell expressing LifeAct-mCherry or Gef1-3xYFP and Cdc15-GFP treated with DMSO and 100 µM of Latrunculin A (LatA) for 30 mins. Red arrowheads indicate the presence and white arrowheads indicate absence of signals at cell division site. Bars, 5 µm.



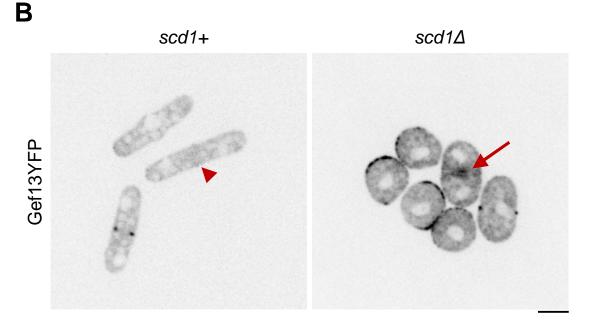


Figure S2 : (A) Scd1 localizes to the ingressing membrane during cytokinesis. Scd1-GFP expressing cells were stained with the membrane dye Cell Mask. 3D reconstructed cells division site is shown after 90° rotation. (B) Scd1 is required for proper Gef1 localization at the division site and cell cortex. Gef1-3xYFP expressing *scd1*+ and *scd1* Δ cells were imaged. Red arrowhead shows loss of Gef1-3xYFP signal at the site of cell division post ring constriction. Red arrow shows Gef1-3xYFP at the division site post ring constriction. Gef1-3xYFP signal in non-dividing cells is more depolarized and randomly distributed at the cell cortex.

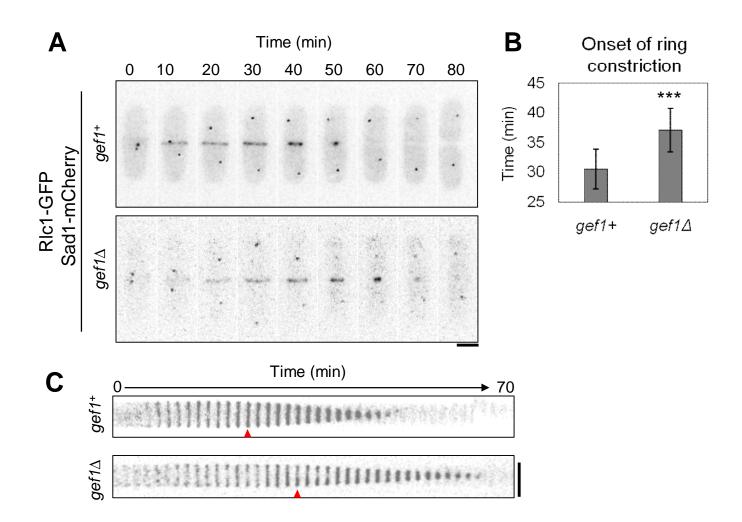


Figure S3: Gef1 is required for timely onset of actomyosin ring constriction. A. Actomyosin ring protein Rlc1-GFP and spindle pole body marker Sad1-mCherry observed over time in *gef1*⁺ and *gef1*^{Δ} cells at the site of cell division during cytokinesis. B. Quantification of onset of ring constriction in *gef1*⁺ and *gef1*^{Δ} cells. Student's t-test, ****p* < 0.001. C. Kymograph of Rlc1-GFP at the actomyosin ring overtime since onset of spindle pole body separation. Scale bar, 5 µm. m, minutes.

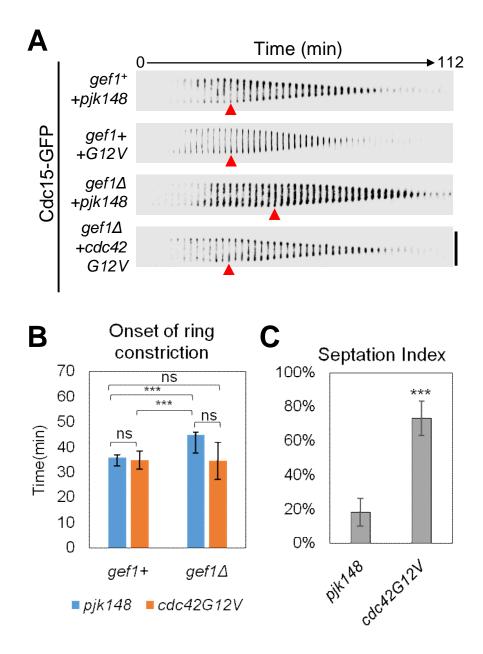


Figure S4: Constitutively active Cdc42 rescues delay in onset of ring constriction in *gef1* Δ cells. (A) Kymographs of cytokinetic ring represented by Cdc15-GFP in indicated cells. Red arrowheads- onset of cytokinetic ring constriction. Bars, 5 µm. (B) Quantification of onset of cytokinetic ring constriction as shown in A . n > 16 cells. (C) Septation index of control and Cdc41G12V overexpressing cells. n>16. ***, *p*<0.001. ns, not significant . Error bars, SD.

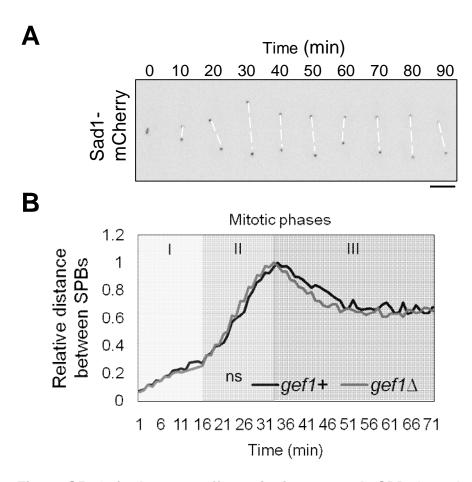
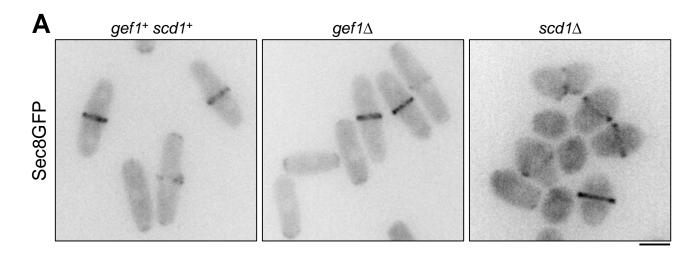


Figure S5: Gef1 does not affect mitotic events. A. SPB dynamics since onset of anaphase by Sad1-mCherry.White dashed lines indicate the distance between the two SPBs each time point. Bar, 5 μ m. B. Quantification of the distance between two spindle pole bodies over time in *gef1*⁺ and *gef1* Δ cells. n > 14 cells. Three stages are defined by SPBs' distance: I, Anaphase A; II, Anaphase B; III, Telophase. ns, not significant. m, minutes.



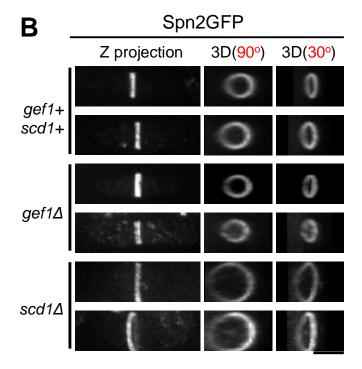


Figure S6. **Gef1 and Scd1 do not affect the localization of the Exocyst complex or the septin ring during cytokinesis.** (A) Cells as indicated expressing the exocyst protein Sec8-GFP were imaged. (B) Indicated cells expressing the septin ring protein Spn2-GFP were analyzed during cytokinesis. 3D reconstructed rings rotated at 90° and 30° are shown. Bars, 5µm

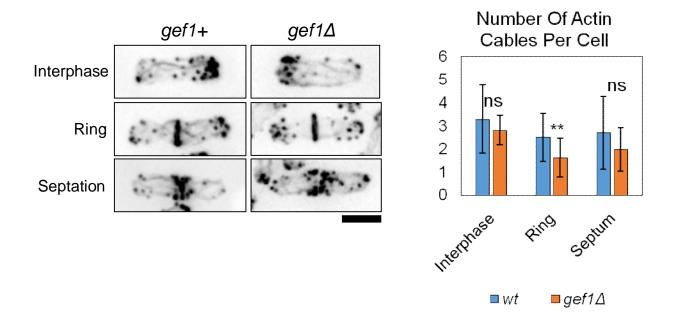


Figure S7: Gef1 is required for non-medial actin cables during cytokinesis.

Quantificantion of actin cables observed in Alexa-fluor phalloidin stained $gef1^+$ and $gef1\Delta$ cells during interphase, ring phase of cytokinesis and during the septum phase. n> 10. **, p=0.004. ns, not significant. Error bars, SD.

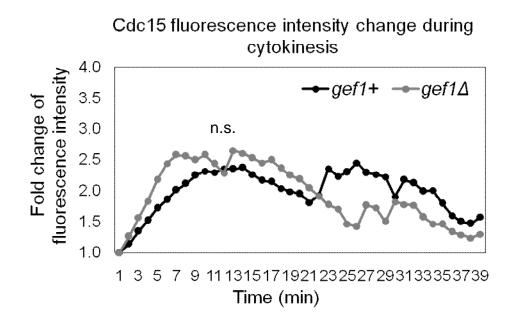


Figure S8: Gef1 des not affect the recruitment of the actomyosin ring protein Cdc15 during ring maturation. Quantification of fold increase in Cdc15-GFP levels at the cell division site in gef1+ and $gef1\Delta$ cells over time. Time 0 is onset of maturation phase as determined by the first appearance of a fully assembled Cdc15-GFP ring. Student's t-test; ns, not significant. min, minutes.

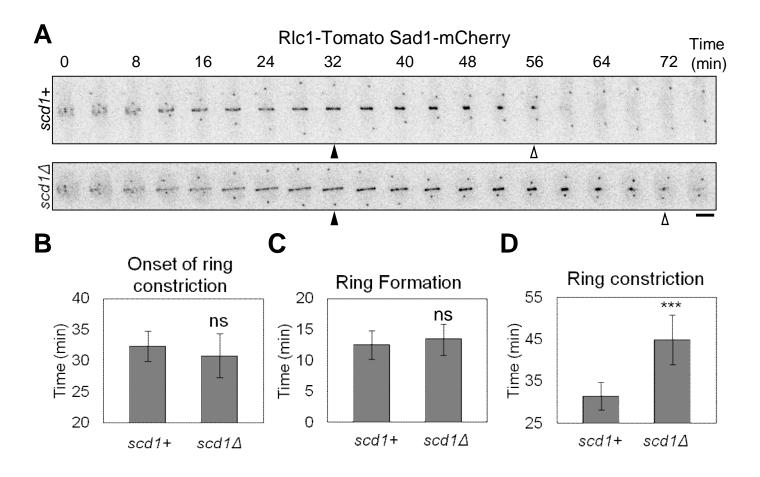


Figure S9: Scd1 is not required for timely onset of actomyosin ring constriction. (A) Actomyosin ring protein Rlc1-GFP and spindle pole body marker Sad1-mCherry observed over time in *scd1*⁺ and *scd1* Δ cells at the site of cell division during cytokinesis. Bar, 5 µm. (B) Quantification of onset of ring constriction in *scd1*⁺ and *scd1* Δ cells. (C) Quantification of actomyosin ring assembly in *scd1*⁺ and *scd1* Δ cells. ns, not significant. (D) Quantification of duration of actomyosin ring constriction in *scd1*⁺ and *scd1* Δ cells. ***, *p*<0.0001. min, minutes.

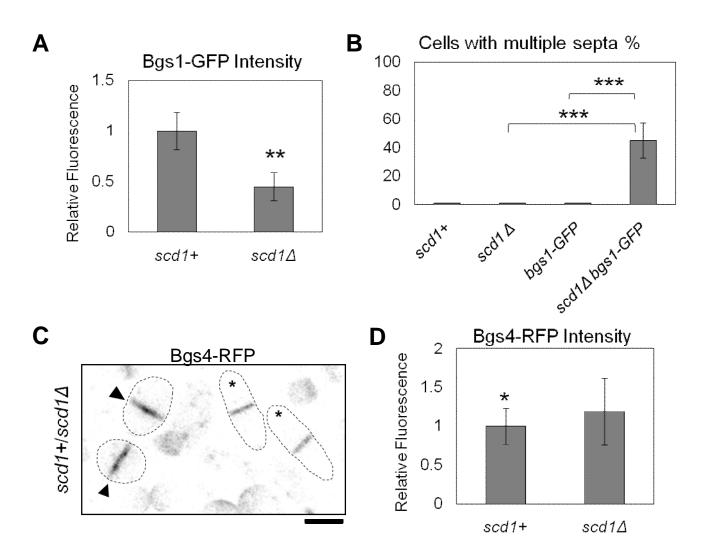


Figure S10. Scd1 is required for normal septum formation. (A) Quantification of Bgs1-GFP levels at the cell division site in *scd1*⁺ and *scd1*^{Δ} cells as shown in Figure 8A. n>23 **, *p*=0.0014. (B) Quantification of multi-septated cells over total septated cells in indicated cells as shown in Figure 8B. n>286. ***, *p*= 0.0001. (C) Cells expressing Bgs4-RFP in *scd1*⁺ and *scd1*^{Δ} cells analyzed in the same field. *scd1*⁺ cells are depicted by asterisks and *scd1*^{Δ} cells are depicted by arrowheads. Bars, 5 µ m (D) Quantification of Bgs4-RFP levels at the cell division site in the indicated cells as shown in C. n>21. **, *p*=0.04. Error bars, SD.

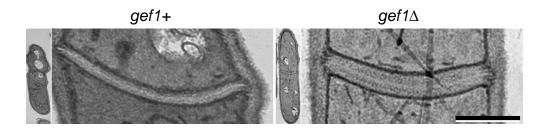


Figure S11. Loss of *gef1* does not show septum morphology defects but shows thicker septum. Electron microscopy images of *gef1*⁺ and *gef1* Δ cells. Bar, 1µm.

SUPPLEMENTARY MOVIE LEGENDS

Movie S1: **Spatiotemporal localization of CRIB-3XGFP during cytokinesis.** Left panel shows CRIB-3XGFP. Right panel shows cytokinetic ring marker Rlc1-Tomato and SPB marker Sad1-mCherry. Frame interval is 2 minute. Frame rate is 2 fps.

Movie S2: **Spatiotemporal localization of CRIB-3XGFP in** *gef1***∆ cells during cytokinesis.** Left panel shows CRIB-3XGFP. Right panel shows cytokinetic ring marker Rlc1-Tomato and SPB marker Sad1-mCherry.Frame interval is 2 minute. Frame rate is 2 fps.

Movie S3: **Spatiotemporal localization of CRIB-3XGFP in** *scd1***∆ cells during cytokinesis.** Left panel shows CRIB-3XGFP. Right panel shows cytokinetic ring marker Rlc1-Tomato and SPB marker Sad1-mCherry.Frame interval is 2 minute. Frame rate is 2 fps.

Movie S4: Loss of Gef1 delays onset of ring constriction. Cdc15-GFP shows the cytokinetic ring in the cell division site in upper panel. Sad1-mCherry as SPB marker is shown in the bottom panel. Left panel shows a $gef1^+$ cell, right panel shows a $gef1\Delta$ cell. Both $gef1^+$ and $gef1\Delta$ movies start from initial SPB separation. Frame interval is 2 minute. Frame rate is 2 fps.

Movie S5: **Bgs1 recruitment to the division site is delayed in** *gef1* Δ **cells.** Bgs1-GFP at the cell division site is shown in upper panel. Bottom panel shows cytokinetic ring marker Rlc1-Tomato and SPB marker Sad1-mCherry. Left panel shows a *gef1*⁺ cell, right panel is a *gef1* Δ cell. Both *gef1*⁺ and *gef1* Δ movies start playing from separation of SPBs. Sad1-mCherry as SPB marker was used. Frame interval is 2 minute. Frame rate is 2 fps.

Movie S6: Loss of Scd1 delays ring constriction but not onset of ring constriction. Rlc1-Tomato shows the cytokinetic ring in the cell division site. Sad1-mCherry represents SPBs separation. Left panels show bright field images. Right panels shows both Rlc1-Tomato and Sad1-mCherry. Upper panel shows a $scd1^+$ cell, bottom panel shows a $scd1\Delta$ cell. Both $scd1^+$ and $scd1\Delta$ movies start playing from separation of SPBs. Frame interval is 2 minute. Frame rate is 2 fps.

Strain	Genotype	Source
PN972	h	P. Nurse
PPG5660	h⁺ cdc42-1625-kanMX leu1-32 ura4-D18	(Martin et al., 2007)
YMD208	cdc15-Tomato-NAT ^r gef1-3xYFP-kanMX6 ade6-M216/704 his7⁺	This study
YMD373	cdc15-Tomato-NAT ^r scd1-3xGFP-kanMX ad6-m216 leu1- 32 ura4-d18 his7⁺	This study
MBY6843	h+ LifeAct-mCherry: leu+ ade6-M216 leu1-32 ura4-D18	(Huang et al., 2012)
YMD317	CRIB-GFP-ura4 ⁺ rlc1-Tomato-NAT ^r sad1-mCherry:kanMX ade6-M21X leu1-32 ura4-D18 his7 ⁺	This study
YMD488	Δgef1∷ura4⁺ CRIB-GFP-ura4⁺,rlc1-Tomato-NAT′ sad1- mCherry:kanMX ura4-D18 leu1-32 his7⁺	This study
YMD530	scd1::ura4 ⁺ CRIB-GFP-ura4 ⁺ rlc1-Tomato-NAT ^r sad1- mCherry:kanMX ade6-M21X leu1-32 ura4-D18 his7 ⁺	This study
YMD133	cdc15-GFP: kanMX6 sad1-mCherry:kanMX ade6-M21X leu1-32 ura4-D18	This study
YMD131	Δgef1∷ura4⁺ cdc15-GFP: kanMX6 sad1-mCherry:kanMX ade6-M21X leu1-32 ura4-D18	This study
YMD542	Δgef1::ura4 ⁺ bgs1D::ura4 Pbgs1::GFP-bgs1:leu1 ⁺ Rlc1- Tomato-NAT ^r Sad1-mCherry:kanMX leu1-32 ura4-D18	This study
YMD546	bgs1D::ura4 Pbgs1::GFP-bgs1:leu1 ⁺ Rlc1-Tomato-NAT ^r Sad1-mCherry:kanMX leu1-32 ura4-D18	This study
YMD498	cdc15-GFP: kanMX6 sad1-mCherry:kanMX ade6-M21X leu1-32 ura4-D18 pjk148-nmt41x-leu1⁺	This study
YMD473	∆gef1::ura4⁺ cdc15-GFP: kanMX6 sad1-mCherry:kanMX ade6-M21X leu1-32 ura4-D18 pjk148-nmt41x-leu1⁺	This study
YMD474	Δgef1::ura4 ⁺ cdc15-GFP: kanMX6 sad1-mCherry:kanMX ade6-M21X leu1-32 ura4-D18 pjk148-nmt41x-cdc42G12V- leu1 ⁺	This study
PPG2601	h⁺ Δgef1::ura4⁺ ura4-D18 leu1-32	(Coll et al., 2003)
KLG2955	h⁻ mid1∆∷ura4⁺	K. Gould
YMD377	Δgef1∷ura4⁺ mid1∆∷ura4⁺ ura4-D18 leu1-32	This study
KGY3019	h⁺ cdc15-GFP: kanMX6 ade6-M210 leu1-32 ura4-D18	(Carnahan and Gould, 2003)
YMD61	Δgef1∷ura4⁺ cdc15-GFP: kanMX6 ade6-M210 leu1-32 ura4-D18	This study

Table S1.	S.	pombe strains used in this study.	
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PN527	h ⁻ ade6-M210 leu1-32 his3-D1 ura4-D18	P. Nurse
FV242+	h ⁹⁰ leu1 ura4-d18 ade6 h210 scd1::ura4	(Li et al., 2000)
PPG3750	h ⁻ bgs1D::ura4 Pbgs1::GFP-bgs1:leu1 leu1-32 ura4-D18 hys3-D1	(Cortes et al., 2002)
YMD501	h ⁹⁰ scd1::ura4 bgs1D::ura4 Pbgs1::GFP-bgs1:leu1 ⁺ leu1- 32 ura4-D18 hys3-D1	This study
PPG4608	h ⁻ bgs4::ura4 Pbgs4::GFP-bgs4:leu1 leu1-32 ura4-D18 hys3-D1	(Cortes et al., 2005)
YMD504	h ⁹⁰ scd1::ura4⁺ bgs4::ura4 Pbgs4::GFP-bgs4:leu1 leu1-32 ura4-D18 hys3-D1	This study
YMD64	Δgef1::ura4⁺ mid1-GFP ura4-D18 leu1-32	This study
YDM403	h ⁻ mid1-GFP:kanMX6 ade6-M210 leu1-32 ura4-D18	(Bahler et al., 1998)
YMD76	∆gef1::ura4⁺ KanMX::YFP-rng2 ade6-M21X ura4-D18 leu1-32	This study
JW937	h ⁻ kanMX6-Prng2-mYFP-rng2 ade6-M210 leu1-32 ura4- D18	(Wu et al., 2003)
KGY3019	cdc15-GFP::kanMX, ade6-m210 ura4-d18 leu1-32	(Carnahan and Gould, 2003)
YMD183	Δgef1::ura4⁺ kanMX-GFP-myo2 ade6-M210 ura4-D18 leu1-32	This study
JW766	kanMX-GFP-myo2 ade6-M210 leu1-32 ura4-D18	(Wu et al., 2003)
YMD156	∆gef1::ura4⁺ rlc1-GFP:ura4⁺ ade6-M210 ura4-D18 leu1-32	This study
KGY1278	h ⁺ rlc1-GFP:ura4 ⁺ ade6-M210 leu1-32 ura4-D18	(Naqvi et al., 2000)
YMD204	∆gef1::ura4⁺ rlc1-GFP:ura4⁺ sad1-mCherry:kanMX ade6- M210 ura4-D18 leu1-32	This study
YMD165	rlc1-GFP:ura4+ sad1-mCherry:kanMX ade6-M210 ura4- D18 leu1-32	This study
YMD275	bgs4::ura4 Pbgs4::RFP-bgs4:leu1 cdc11-GFP:ura4⁺ ade6- M210 leu1-32 ura4-D18	This study
YMD288	∆gef1::ura4⁺ bgs4::ura4 Pbgs4::GFP-bgs4:leu1 rlc1- GFP:ura4+ ade6-M210 leu1-32 ura4-D18	This study
YMD 457	Δgef1::ura4⁺ bgs1D::ura4 Pbgs1::GFP-bgs1:leu1 leu1-32 ura4-D18 hys3-D1	This study
YMD590	myo52GFP sad1-mCherry:kanMX ade6-M210 ura4-D18 leu1-32	This study
YMD594	∆gef1::ura4⁺ myo52GFP sad1-mCherry:kanMX ade6-M210 ura4-D18 leu1-32	This study
YMD 600	bgs4::ura4 Pbgs4::GFP-bgs4:leu1 leu1-32 ura4-D18 hys3-	This study

D1 cdc11-GFP:kanR ade6-M21x

YMD617	Δgef1::ura4⁺ bgs4::ura4 Pbgs4::GFP-bgs4:leu1 leu1-32 ura4-D18 hys3-D1 cdc11-GFP:kanR ade6-M21x	This study
FV1405	scd1::ura4⁺ Gef13yfp	This study
YMD566	Pjk148cdc42g12v cdc15gfp sad1-mCherry:kanMX ade6- M210 ura4-D18 leu1-32	This study
YMD606	Pjk148 ade6-M210 leu1-32 his3-D1 ura4-D18	This study
YMD610	Pjk148-nmt41-cdc42G12V ade6-M210 leu1-32 his3-D1 ura4-D18	This study
YSM1720	sec8-GFP-ura4+	Sophie Martin
YMD84	Δgef1::ura4 ⁺ sec8-GFP-ura4+	This study
YMD508	scd1::ura4⁺ sec8-GFP-ura4+	This study
Spn2GFP	Spn2-GFP::KanR Leu 1-32, ura4-D18,ade 6-m210 h-	(An et al., 2004)
YMD613	Δgef1::ura4⁺ Spn2-GFP::KanR Leu 1-32, ura4-D18,ade6- m210	This study
YMD615	scd1::ura4 ⁺ Spn2-GFP::KanR Leu 1-32, ura4-D18,ade6- m210	This study
YMD527	rlc1-tomato-NATr ade6-M21X leu1-32 his7+ sad1- mCherry:kanMX ura4-D18	This study
YMD625	scd1::ura4 ⁺ rlc1-tomato-NATr ade6-M216 leu1-32 ura4- D18 his7+ sad1mcherry	This study

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