

Supplementary Information

Moonlighting of *Helicobacter pylori* catalase protects against complement-mediated killing by utilizing the host molecule vitronectin

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Supplementary Table 1: Correlation of Vn binding and KatA surface exposure

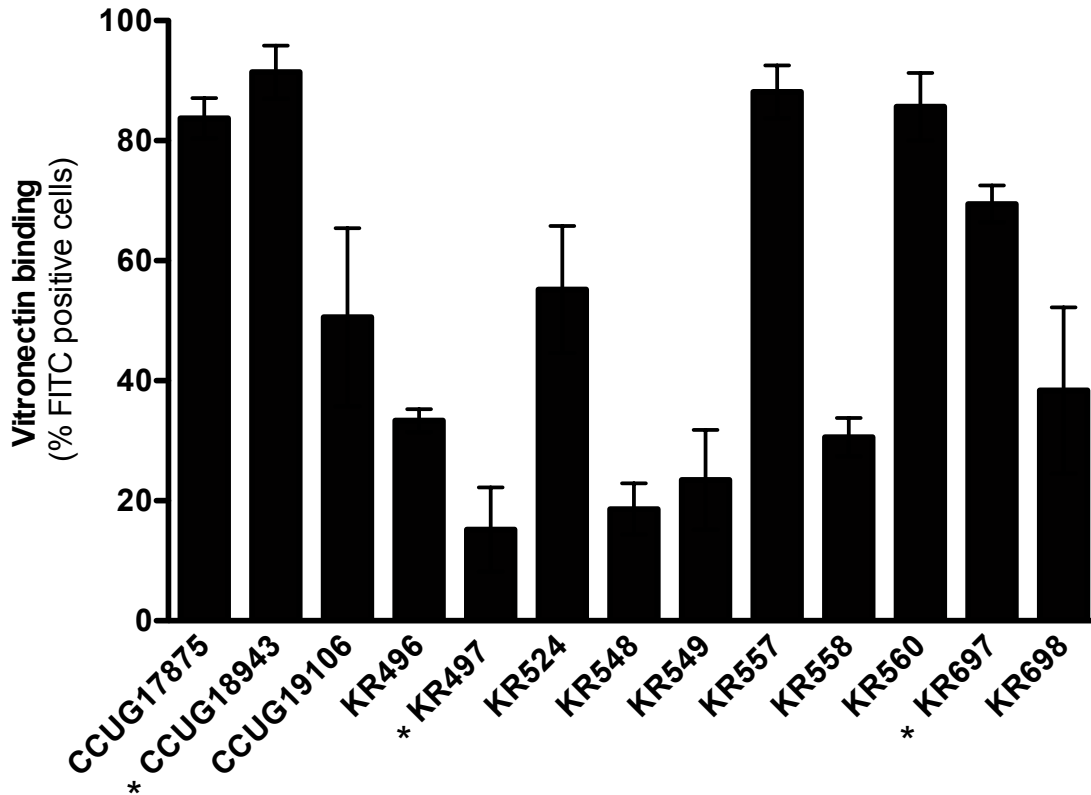
Strain	Vn binding (%)		anti-KatA binding (%)	
	Mean	SD	Mean	SD
CCUG18943	91.42	4.89	58.36	1.48
KR557	88.13	4.38	27.56	15.19
CCUG17875	83.73	3.35	56.19	13.94
KR560	75.06	5.60	59.96	6.59
KR697	69.47	3.07	47.34	26.76
KR524	55.20	10.59	21.75	2.81
CCUG19106	50.60	14.84	nd	nd
KR698	38.38	13	6.47	3.88
KR496	33.31	84	32.38	16.56
KR558	30.60	1.91	nd	nd
KR549	23.47	3.17	23.93	4.71
KR548	18.63	4.27	21.07	3.57
KR497	15.20	3.48	5.10	0.51
CCUG18943 Δ katA	na	na	28.6	6.39
KR697 Δ katA	na	na	9.31	4.12
KR497 Δ katA	na	na	4.96	2.83

Supplementary Table 2: Bacterial strains

Strain	Description	Reference/ Source
<i>E. coli</i>		
DH5 α	F- Φ 80 <i>lacZ</i> Δ M15 Δ (<i>lacZYA-argF</i>) U169 <i>recA1 endA1 hsdR17</i> (rK-, mK+) <i>phoA supE44</i> λ - <i>thi-1 gyrA96</i> <i>relA1</i>	Invitrogen
BL21 (DE3)	<i>fhuA2 [lon] ompT gal</i> (λ DE3) [<i>dcm</i>] Δ <i>hsdS</i> λ DE3 = λ <i>sBamHI</i> <i>o</i> Δ <i>EcoRI-B</i> <i>int::(lacI::PlacUV5::T7 gene1) i21</i> Δ <i>nin5</i>	Novagen
<i>H. pylori</i>		
CCUG18943	<i>cagA</i> ⁺	Culture Collection, University of Göteborg, Sweden (CCUG)
CCUG18943 Δ <i>katA</i>	<i>katA</i> deficient CCUG18943	This study
KR697	clinical isolate, <i>cagA</i> ⁺	This study
KR697 Δ <i>katA</i>	<i>katA</i> deficient KR697	This study
KR497	clinical isolate, <i>cagA</i> ⁺	This study
KR497 Δ <i>katA</i>	<i>katA</i> deficient KR497	This study
KR496	clinical isolate, <i>cagA</i> ⁺	This study
KR524	clinical isolate, <i>cagA</i> ⁺	This study
KR548	clinical isolate, <i>cagA</i> ⁺	This study
KR549	clinical isolate, <i>cagA</i> ⁺	This study
KR557	clinical isolate, <i>cagA</i> ⁺	This study
KR558	clinical isolate, <i>cagA</i> ⁺	This study
KR560	clinical isolate, <i>cagA</i> ⁺	This study
KR698	clinical isolate, <i>cagA</i> ⁺	This study
CCUG17875	<i>cagA</i> ⁺	CCUG
CCUG19106	<i>cagA</i> ⁻	CCUG

Supplementary Table 3: Primers and plasmids

Plasmids	
pET26	Protein expression vector, Km ^R
pET26 <i>katA</i>	Expression vector for <i>H. pylori</i> CCUG18943 KatA
pET26 <i>katA</i> 1-49	Expression vector for <i>H. pylori</i> CCUG18943 KatA aa 1-49 fragment
pET26 <i>katA</i> 51-505	Expression vector for <i>H. pylori</i> CCUG18943 KatA aa 51-505 fragment
pET26 <i>katA</i> 51-488	Expression vector for <i>H. pylori</i> CCUG18943 KatA aa 51-488 fragment
pET26 <i>katA</i> 51-400	Expression vector for <i>H. pylori</i> CCUG18943 KatA aa 51-400 fragment
pET26 <i>katA</i> 350-505	Expression vector for <i>H. pylori</i> CCUG18943 KatA aa 350-505 fragment
Oligonucleotides (5' – 3')	
<i>katA</i> _for2	ATGCCTGACATATGGTTAATAAAGATGTGAAACAAA CCACTGC
<i>katA</i> _for51	ATATCTGACATATGCCTGAAAGGGTAGTGCATGCTAA AGGAAGC
<i>katA</i> _for350	ATGCCTGACATATGAATTATCCTCAAATACCGGTAA TAAACCAAGA
<i>katA</i> _for401	ATGCCTGACATATGAAGTTCAACTTAGCTCATATTGA GAAAGAG
<i>katA</i> _rev49	ATCCTGACTCGAGCCTTTCTCTGTCAAACGCTGCTAA CTT
<i>katA</i> _rev488	ATTCTGACTCGAGCTTTTGGTGTTTTTCAAGAGCTTTT TAACTCCCTC
<i>katA</i> _rev505	ATATGACTCGAGCTTTTTCTTTTTTGTGTGGTGCATGT CTTTTCC
Kan_F	ATGAGCCATATTCAACGGGAAAC
Kan_R	TTAGAAAAACTCATCGAGCATCAA
UF_ <i>katA</i> _F1	GCGTTATCTCGTGTGCATGTTCCAGC
UF_ <i>katA</i> _R6_Kan	TCCCGTTGAATATGGCTCATCTTCTTTTCTTTATTGA TTTAAATTTT
UF_ <i>katA</i> _R4_Kan	TCCCGTTGAATATGGCTCATCTTTTTTCTTTATTGA CTTCAAATCT
DF_ <i>katA</i> _F2_Kan	TGCTCGATGAGTTTTTCTAACCCTTTTCTTTAAGCGTT CTTATTTTT
DF_ <i>katA</i> _R2	ATTCCTAGATTTTCATGCCTTTAGAC

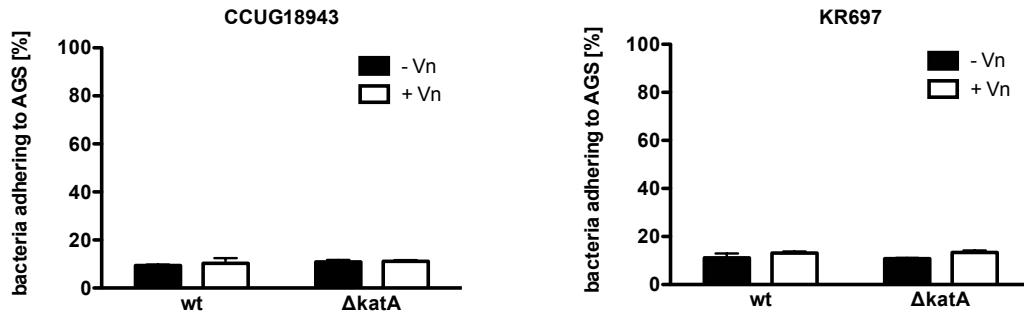


Supplementary Figure S1. Vitronectin binding differs between *H. pylori* isolates.

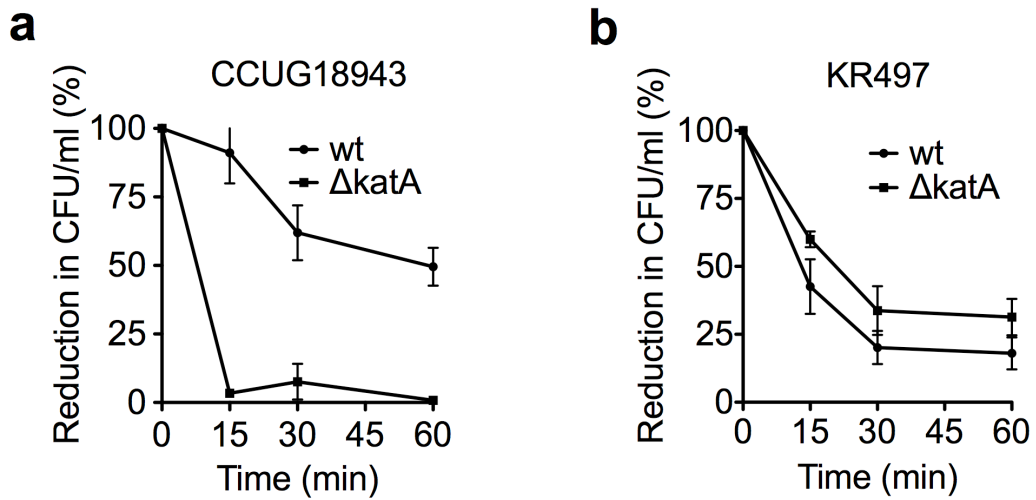
Clinical isolates of *H. pylori* were incubated with 3 μ g of vitronectin purified from human serum. The percentage of vitronectin-binding was determined by flow-cytometry using FITC-labeled α -Vn antibody. Data presented are the mean and SD of at least three independent experiments. Asterisks indicate the three strains chosen for further analyses.

18943	MVNKDVKQTTAFGAPVWDDNNVITAGPRGPVLLQSTWFLEKLAADFDRERIPERVVHAKGS	60
697	MVNKDVKQTTAFGAPVWDDNNVITAGPRGPVLLQSTWFLEKLAADFDRERIPERVVHAKGS	60
26695	MVNKDVKQTTAFGAPVWDDNNVITAGPRGPVLLQSTWFLEKLAADFDRERIPERVVHAKGS	60
497	MVNKDVKQTTAFGAPVWDDNNVITAGPRGPVLLQSTWFLEKLAADFDRERIPERVVHAKGS	60
18943	GAYGTFTVTKDITKYTKAKIFSKVGKKTECFFRFSTVAGERGSADAVRDPRGFAMKYYTE	120
697	GAYGTFTVTKDITKYTKAKIFSKVGKKTECFFRFSTVAGERGSADAVRDPRGFAMKYYTE	120
26695	GAYGTFTVTKDITKYTKAKIFSKVGKKTECFFRFSTVAGERGSADAVRDPRGFAMKYYTE	120
497	GAYGTFTVTKDITKYTKAKIFSKVGKKTECFFRFSTVAGERGSADAVRDPRGFAMKYYTE	120
8943	EGNWDLVGNNTPVFFIRDIAIKFPDFIHTQKRDPQTNLPNHDMVWDFWSNVPESLYQVTWV	180
697	EGNWDLVGNNTPVFFIRDIAIKFPDFIHTQKRDPQTNLPNHDMVWDFWSNVPESLYQVTWV	180
26695	EGNWDLVGNNTPVFFIRDIAIKFPDFIHTQKRDPQTNLPNHDMVWDFWSNVPESLYQVTWV	180
497	EGNWDLVGNNTPVFFIRDIAIKFPDFIHTQKRDPQTNLPNHDMVWDFWSNVPESLYQVTWV	180
18943	MSDRGIPKSFHRMDGFGSHTFSLINAKGERFVVKFHFETMQGVKHLTNEEAAEIRKYDPP	240
697	MSDRGIPKSFHRMDGFGSHTFSLINAKGERFVVKFHFETMQGVKHLTNEEAAEVRKYDPP	240
26695	MSDRGIPKSFHRMDGFGSHTFSLINAKGERFVVKFHFHTMQGVKHLTNEEAAEVRKYDPP	240
497	MSDRGIPKSFHRMDGFGSHTFSLINAKGERFVVKFHFHTMQGVKHLTNEEAAEIRKHDPP	240
18943	SNQRDLFNAIARGDFPKWKLSIQVMPEEDAKKYRFHPFDVTKIWLQDYPLMEVGIVELN	300
697	SNQRDLFNAIARGDFPKWKLSIQVMPEEDAKKYRFHPFDVTKIWLQDYPLMEVGIVELN	300
26695	SNQRDLFNAIARGDFPKWKLSIQVMPEEDAKKYRFHPFDVTKIWLQDYPLMEVGIVELN	300
497	SNQRDLFNAIARGDFPKWKLSIQVMPEEDAKKYRFHPFDVTKIWLQDYPLMEVGIVELN	300
18943	KNPENYFAEVEQAAFSPANVVPVIGYSPDRMLQGRVLSYGDTHRYRLGVNYPQIPVKNKPR	360
697	KNPENYFAEVEQAAFSPANVVPVIGYSPDRMLQGRVLSYGDTHRYRLGVNYPQIPVKNKPR	360
26695	KNPENYFAEVEQAAFSPANVVPVIGYSPDRMLQGRVLSYGDTHRYRLGVNYPQIPVKNKPR	360
497	KNPENYFAEVEQAAFSPANVVPVIGYSPDRMLQGRVLSYGDTHRYRLGVNYPQIPVKNKPR	360
18943	CPFHSSSRDGYMQNGYYGSLQNYTPSSSLPGYKEDKSARDPKFNLAHIEKEFEVWNWDYRA	420
697	CPFHSSSRDGYMQNGYYGSLQNYTPSSSLPGYKEDKSARDPKFNLAHIEKEFEVWNWDYRA	420
26695	CPFHSSSRDGYMQNGYYGSLQNYTPSSSLPGYKEDKSARDPKFNLAHIEKEFEVWNWDYRA	420
497	CPFHSSSRDGYMQNGYYGSLQNYTPSSSLPGYKEDKSARDPKFNLAHIEKEFEVWNWDYRA	420
18943	EDSDYYTQPGDYRSLPADEKERLHDTIGESLAHVTHKEIVDKQLEHFKKADPKYAEGVK	480
KR697	EDSDYYTQPGDYRSLPADEKERLHDTIGESLAHVTHKEIVDKQLEHFKKADPKYAEGVK	480
26695	DDSDYYTQPGDYRSLPADEKERLHDTIGESLAHVTHKEIVDKQLEHFKKADPKYAEGVK	480
497	DDSDYYTQPGDYRSLPADEKERLHDTIGESLAHVTHKEIVDKQLEHFKKADPKYAEGVK	480
18943	KALEKHQKMMKDMHGKDMHHTKTKK 505	
697	KALEKHQKMMKDMHGKDMHHTKTKK 505	
26695	KALEKHQKMMKDMHGKDMHHTKTKK 505	
497	KALEKHQKMMKDMHGKDMHHTKTKK 505	

Supplementary Figure S2. KatA sequences of strong, intermediate and weak vitronectin binding *H. pylori* strains show only small differences. KatA amino acid sequences of *H. pylori* CCUG18943, KR697, Hp22965, and KR497 were aligned using Clustal Omega (<http://www.ebi.ac.uk/Tools/msa/clustalo/>). The putative vitronectin-binding site is marked in grey and positions where changes in the amino acid sequence occur are marked in yellow.



Supplementary Figure S3. The presence of KatA and vitronectin does not affect adhesion of *H. pylori* to AGS cells. Gastric adenocarcinoma cells (AGS) were incubated with *H. pylori* at multiplicity of infection (MOI) 100 in the presence or absence of vitronectin. Cells and bacteria were incubated for a period of 3 h, cells were lysed and dilutions of bacteria plated on GC plates for CFU counts. The relative number of bacteria, which attached to the AGS cells is shown. Data presented are the mean and SD of three independent experiments performed in technical triplicate.



Supplementary Figure S4: KatA confers increased complement resistance in high-Vn binding strains. The resistance of CCUG18943 (a) and KR497 (b) wt and $\Delta katA$ to serum complement was tested in a series of assays using 5% normal human serum (NHS) of donors negative for anti-*H. pylori* IgG. Survival rates at 15, 30 and 60 min after addition of serum were determined by plate count. Depicted is the reduction in CFU/ml in percent. Results are the mean and SE of at least three independent experiments performed in technical duplicate.

Supplementary Methods

H. pylori adhesion to AGS cells

The human gastric adenocarcinoma cell line (AGS) was maintained in F-12 medium (GIBCO), 10% FBS at 37°C, 5% CO₂. Cells were routinely subcultured every 2–3 days. AGS cells were seeded into wells of cell culture dishes and grown till confluency and starved overnight prior to infection. Bacteria were grown on GC plates for 72 hours. To observe the effect of vitronectin on adherence of *H. pylori* to AGS cells, serum purified Vitronectin was added at a concentration of 10 µg/ml at 4°C for 1 h followed by two washes prior to the addition of *H. pylori* at an MOI of 1:100. Cells were incubated for a period of 3 hours after infection, washed thoroughly and cells were collected by treatment with citric acid buffer. Cells were then lysed with glass beads, vortexed, and serial dilutions plated on GC agar plates. Colony counts were performed after 72 h incubation.