

## T cell infiltration in the CNS of tamoxifen-treated DTA mice during the late-onset disease.

Staining for CD3 (green) revealed increased T cell infiltration in different areas of the CNS in tamoxifen-treated *PLP/CreER<sup>T</sup>*;*ROSA26-eGFP-DTA* (DTA) mice (lower panel) at 53 weeks post-injection (p.i.) compared with that in control (*ROSA26-eGFP-DTA*) mice (upper panel). Higher magnification of the CD3+ cells is shown in the inset (lower panel, corpus callosum). Images are representative of three mice/genotype. Scale bar: 100 µm. Sections were counterstained with DAPI (blue) to show cell nuclei.



### Tamoxifen-independent recombination in oligodendrocytes of the reporter mice.

(a) Expression of the reporter gene protein YFP (green) was detected in a subpopulation of oligodendrocytes stained for CC-1 (red) in all CNS areas examined of the untreated  $PLP/CreER^T$ ; RosaEYFP (reporter w/o 4-HT; lower panel) mice at the age of 52 weeks. CC1/YFP stainings in corresponding areas of control littermate mice (RosaEYFP) are shown in the upper panel. The CC1/YFP double positive cells (yellow) are marked by arrows. Images are representative of three mice/genotype. Scale bar: 100 µm. (b) Counts of the CC1/YFP double positive cells labeled in different CNS areas of the untreated reporter mice showed that tamoxifen-independent recombination occurs in ~ 6%-14% of oligodendrocytes at 26 weeks and it tends to be increased in most areas (6%-20%) at 52 weeks (brain stem p =0.0084). Graph in **b** shows means (+ SEM), n=4 mice per group, two-tailed unpaired Student's *t* test, \*\*p <0.01.



### Tamoxifen-independent demyelination in untreated DTA mice.

(a) Electron microscopy analysis demonstrated normal myelin levels in the cervical cord and milder demyelination in the cerebellum, corpus callosum, optic nerve and brain stem areas of the untreated  $PLP/CreER^T$ ; ROSA26-eGFP-DTA (DTA w/o 4-HT) mice as compared to tamoxifen-treated  $PLP/CreER^T$ ; ROSA26-eGFP-DTA (DTA) mice, which showed more severe myelin loss in the same areas. Images are representative of three mice/genotype. Scale bars: 2 µm. (b) Morphometric analysis of the corpus callosum area showed that numbers of unmyelinated axons were significantly lower (-30%) in the 52-week old untreated DTA mice as compared to tamoxifen-treated DTA mice at 53 weeks post-injection suggesting milder myelin loss in the former animals. Control versus tamoxifen-treated DTA mice: p=0.0001, control vs. untreated DTA mice: p=0.0001, tamoxifen-treated DTA mice, n=4 for untreated DTA mice. \*\*\*p <0.001, \*\*p <0.01, one way ANOVA with Tukey's post-hoc test. (c) Scatter plots of g-ratios versus axon diameters indicate significantly increased g-ratios (thinner myelin) in axons of the corpus callosum in both the tamoxifen-treated DTA (mean g ratio of 0.80 ± 0.004) and the untreated DTA mice (mean g ratio of 0.82 ± 0.005) as compared to controls (ROSA26-eGFP-DTA, mean g ratio of 0.77 ± 0.005). Control versus tamoxifen-treated DTA mice: p<000.1, one-way Anova followed by Tukey's *post-hoc* analysis test. Forty axons/mouse, n=3 mice per group.



### The untreated DTA mice do not develop a late-onset clinical phenotype.

(a-b) The untreated  $PLP/CreER^{T}$ ; ROSA26-eGFP-DTA (DTA w/o 4-HT) mice displayed normal coordination skills on the rotarod (a) and they did not show significant weight loss (b) as compared to control littermate (ROSA26-eGFP-DTA) mice during the time points that the tamoxifen-treated DTA mice were severely affected. Data are presented as the mean ( $\pm$  SEM), n= 5 mice/genotype for each data point shown in the graphs in **a** and **b**, p >0.05; two-way ANOVA with Bonferroni post-hoc analysis.



## Tamoxifen-independent demyelination in untreated DTA mice does not elicit an immune response against myelin.

(a) Staining for the pan T cell marker CD3 showed similar, low numbers of T cells in different CNS areas of the 52-week old untreated  $PLP/CreER^{T}$ ; ROSA26-eGFP-DTA (DTA w/o 4-HT) mice and in corresponding CNS areas of their littermate controls (ROSA26-eGFP-DTA mice). Images are representative of three mice/genotype. Scale bar: 100 µm. Spleen, cervical lymph nodes, and CNS samples were collected from age matched control littermate mice and untreated DTA mice. (b) Total splenoctyes ( $1x10^{6}$  cells per well) were cultured in the presence of medium alone, anti-CD3 ( $1\mu$ g/mI),

OVA<sub>323-339</sub>, PLP<sub>139-151</sub>, PLP<sub>178-191</sub>, MBP<sub>84-104</sub>, MOG<sub>35-55</sub>, and whole recombinant rat MOG protein (10µg/ml). The cultures were pulsed with tritiated thymidine (1µCi) at 24 hours and cultures were harvested at 72 hours post culture to determine the level of cellular proliferation. Cells from (**c**) spleen, (**d**) cervical lymph nodes, and (**e**) CNS were stained for the presence of total infiltrating lymphocytes (CD45hi), microglia (CD45lo/CD11b+), total CD3+, total CD4+ T cells (CD3+/CD4+), T<sub>reg</sub> cells (CD3+/CD4+/CD25+/FoxP3+), effector CD4+ T cells (CD3+/CD4+/CD44hi/FoxP3-), total monocytes (M0; CD3-/CD11b+/CD11c-), and total dendritic cells (DC; CD3-/CD11b-/CD11c+). No significant difference in the number of the various immune cells populations were found within the spleen, cervical lymph nodes, or the CNS (n=3) via two-way ANOVA with Bonferroni post-hoc analysis for differences between control mice and tamoxifen-treated DTA mice. The data are presented as the means (+ SEM). Sections in **a** were counterstained with DAPI (blue) to visualize cell nuclei. Abbreviations: CPM, counts per minute.



**Supplementary Figure 6** 

## Adoptive transfer of MOG-specific T cells derived from tamoxifen-treated DTA mice and 2D2 TCR transgenic mice into naive $Rag1^{-/-}$ mice versus wild-type C57BL/6 mice.

Splenic cells from tamoxifen-treated *PLP/CreER<sup>T</sup>*; *ROSA26-eGFP-DTA* (DTA) mice at 40-52 weeks post-injection or agematched control (*ROSA26-eGFP-DTA*) mice were cultured for 72 h at in the presence of MOG<sub>35-55</sub> peptide (20 µg/ml) plus IL-12 (10ng/ml) and IL-2 (100U/ml). Cultured cells ( $10x10^6$  blast cells) from tamoxifen-treated DTA mice were transferred into either naïve *Rag1<sup>-/-</sup>* recipient mice (n=7) (**a**) or naïve wildtype C57BL/6 recipient mice (n=7) (**b**). Naïve CD4+ T cells were isolated from 2D2 MOG<sub>35-55</sub>-specific TCR transgenic mice, and activated in the presence plate-bound anti-CD3 (1ug/ml) plus various Teff cell lineage promoting conditions as indicated. 2D2 T cells ( $3x10^6$  cells per mouse) activated in the presence of Th1 cell- [IL-12 (10ng/ml), IL-2 (100U/ml), and anti-IL-4 (1µg/ml)] or Th17 cell- [TGF- $\beta$  (10ng/ml), IL-6 (50ng/ml), IL-23 (10ng/ml), anti-IL-4 (1µg/ml), and anti-IFN- $\gamma$  (1µg/ml)] promoting conditions were transferred into *Rag1<sup>-/-</sup>* mice versus wildtype C57BL/6 mice (n=5), and the recipient mice were followed for disease (**c**). 2D2 T cells ( $3x10^6$  cells per mouse) activated in the presence of Th0- [IL-2 (100U/ml)], Th1 cell- or Th17 cell-promoting conditions were transferred into *Rag1<sup>-/-</sup>* mice (n=5), and the recipient mice were followed for disease (**d**). One representative experiment of three is presented. The data is presented as the mean Clinical Score (± SEM).



## Adoptive transfer of MOG-specific T cells derived from tamoxifen-treated DTA mice causes mild demyelination in naive $Rag1^{-/-}$ mice.

Splenic cells from tamoxifen-treated (*PLP/CreER<sup>T</sup>*;*ROSA26-eGFP-DTA*) DTA mice at 40-52 weeks post-injection or agematched control (*ROSA26-eGFP-DTA*) mice were cultured for 72 h at in the presence of MOG<sub>35-55</sub> peptide (20 µg/ml). Cultured cells from tamoxifen-treated DTA mice (2x10<sup>6</sup> blast cells) were transferred into naïve *Rag1<sup>-/-</sup>* recipient mice (**a**) Forty-two days after transfer, Electron microscopy analysis revealed foci of myelin loss in the lower lumbar spinal cord and cerebellar white matter in *Rag1<sup>-/-</sup>* mice inoculated with T cells from tamoxifen-treated DTA mice (DTA recipient) as compared with those inoculated with cells from control mice (littermate recipient). Inflammatory cells were also frequently detected in the cerebellum (asterisk). More-extensive demyelination was seen in the brain stem gray matter. Images are representative of three mice/genotype. Scale bars: 2 µm. (**b**) Demyelination in the brain stem of *Rag1<sup>-/-</sup>* mice inoculated with cells from tamoxifen-treated DTA mice was shown by significantly reduced mean (+ SEM) numbers of myelinated axons (~30% fewer, *p*=0.0363). N=3 mice/group; \**p* <0.05 with two-tailed unpaired Student's *t* test.



## $MOG_{35-55}$ -PLG-induced tolerance inhibits activated DTA mouse-derived $MOG_{35-55}$ -specific T cell entry into the CNS.

Representative flow plots for the data presented in **Fig. 7c** and **e** are presented. The data is presented as the (**a**) IFN- $\gamma$ +, (**b**) IL-17+, and (**c**) Ki67+ singlet live CD45hi/CD4+ T cells within the CNS from control (*ROSA26-eGFP-DTA*) or tamoxifen-treated *PLP/CreER<sup>T</sup>*; *ROSA26-eGFP-DTA* (DTA)-derived cell recipient mice that received OVA<sub>323-339</sub>-PLG (Red line) versus MOG<sub>35-55</sub>-PLG treatment (Blue line). Similar analyses were completed for splenocyte samples assessing (**d**) IFN- $\gamma$ +, (**e**) IL-17+, and (**f**) Ki67+ singlet live CD45hi/CD4+ T cells.



# $MOG_{35-55}$ tolerance inhibits CD4<sup>+</sup> T cell infiltration, proliferation and production of the proinflammatory cytokines in the CNS of the tamoxifen-treated DTA mice.

Representative flow plots of cells isolated from the CNS (**a-g**) and the spleens (**h-m**) of the tamoxifen-treated *PLP/CreER<sup>T</sup>*;*ROSA26-eGFP-DTA* (DTA) mice at 39 weeks post-injection after receiving a single treatment with the MOG<sub>35-55</sub>-PLG or the OVA<sub>323-339</sub>-PLG control peptide at 32 weeks post-injection. Lower numbers of the total infiltrating lymphocytes (CD45hi+ stained cells) were detected in the CNS of the MOG<sub>35-55</sub>-tolerized DTA mice, as compared to the

 $OVA_{323-339}$ -tolerized (control) DTA mice (n=6). The later mice also showed, lower numbers of the effector T cells (CD3+/CD4+ double positive cells) and proliferating interferon- $\gamma$ -positive effector T cells (Ki67+/INF- $\gamma$ + stained cells) and no changes for the proliferating IL-17-positive effector T cells (Ki67+/IL-17+ stained cells). No differences were detected in the numbers of the different immune cell populations in the spleens between the two groups of mice.