

Induction of Human iPSC-Derived Cardiomyocyte Proliferation Revealed by Combinatorial Screening in High Density Microbioreactor Arrays

Supplementary Information

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Supplementary Table 1 - HDMA Physical Parameters

Parameter	Unit	HDMA	96-well Plate	24-well Plate
Single Culture Chamber Dimensions				
Diameter	mm	0.513	6.35	15.5
	m	5.13E-04	6.35E-03	1.55E-02
Culture Area	mm ²	0.207	31.7	188
	cm ²	2.07E-03	3.17E-01	1.88E+00
	m ²	2.07E-07	3.17E-05	1.88E-04
	mm ²	0.207	31.7	200
Nominal Culture Area Specified	cm ²	2.07E-03	3.17E-01	2.00E+00
	m ²	2.07E-07	3.17E-05	2.00E-04
	mm ²	0.207	31.7	200
Height	mm	0.1	2.5	2.5
	m	1.00E-04	2.52E-03	2.50E-03
Nominal Volume	uL	0.021	80	500
	mL	2.07E-05	0.08	0.50
	m ³	2.07E-11	8.00E-08	5.00E-07
	mm ² /mm ³	10	0.40	0.40
Surface-Area-to-Volume (SAV) Ratio	mm ² /mm ³	10	0.40	0.40
Plate to Bioreactor Concentration Factor	-	25	1.0	1.0
Minimum Feature Size	mm	0.05		
Maximum Feature Aspect Ratio (height/width)	-	2		
Cell Seeding				
Nominal Cell Surface Density	/cm ²	4.00E+04	4.00E+04	4.00E+04
Nominal Cell Seeding Suspension Concentration	/mL	4.00E+06	1.59E+05	1.60E+05
HDMA Single Column Dimensions				
Interconnect Width	mm	0.08		
Rows		50		
Column Area (Wells only)	mm ²	10.3		
	cm ²	0.103		
Total Column Area incl. Interconnects	mm ²	10.6		
	cm ²	0.106		
Volume	uL	1.06		
HDMA Entire Culture Array Dimensions				
Columns		162		
Nominal Culture Array Area	mm ²	1709		
	cm ²	17.09		
Volume	uL	171		
HDMA Entire Culture Array Dimensions				
Nominal Flow Conditions				
Area per column	cm ²	0.106		
Flow	uL/h/cm ²	1		
Channel Flow	uL/h	0.148		
	m ³ /s	4.12E-14		
Total Flow	uL/h	24.000		
Number of Syringes	-	8		
Syringe Flow	uL/h	3.000		
^aReynolds Number, Re				
Fluid Density	kg/m ³	1000		
Average Velocity (chamber)	m/s	8.022E-07		
Average Velocity (interconnect)	m/s	4.115E-06		
Hydraulic Diameter (chamber)	m	1.67E-04		
Hydraulic Diameter (interconnect)	m	1.00E-04		

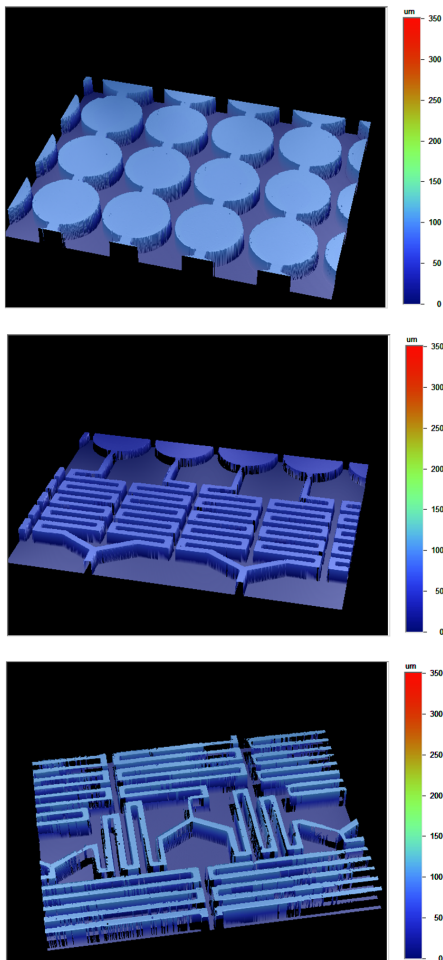
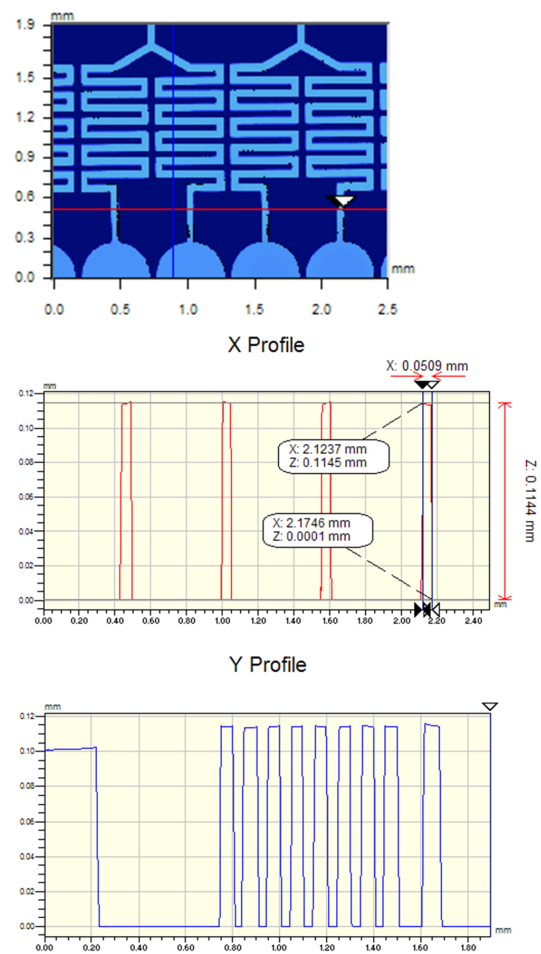
Fluid Viscosity	Pa.s	1.00E-03
Re (chamber)	-	1.34E-04
Re (interconnect)	-	4.12E-04
^bPeclet Number, P_e		
Diffusivity (O ₂)	m ² /s	3.30E-09
Diffusivity (Glucose)	m ² /s	6.00E-10
Diffusivity (Albumin)	m ² /s	7.00E-11
Diffusivity (40kDa GF)	m ² /s	8E-11
Pe (chamber) O ₂	-	4.07E-02
Pe (chamber) (Glucose)	-	2.24E-01
Pe (chamber) (Albumin)	-	1.92E+00
Pe (chamber) (GF)	-	1.68E+00
^cShear Stress, τ		
Shear stress (chamber)	Pa	4.81E-05
Shear stress (interconnect)	Pa	9.88E-05
Fluid Flow and Replacement		
Mean Residence Time (Chamber)	h	0.140
Mean Residence Time (Column)	h	7.12
Space Velocity (Chamber)	/h	7.17
Space Velocity (Column)	/h	0.140

Notes

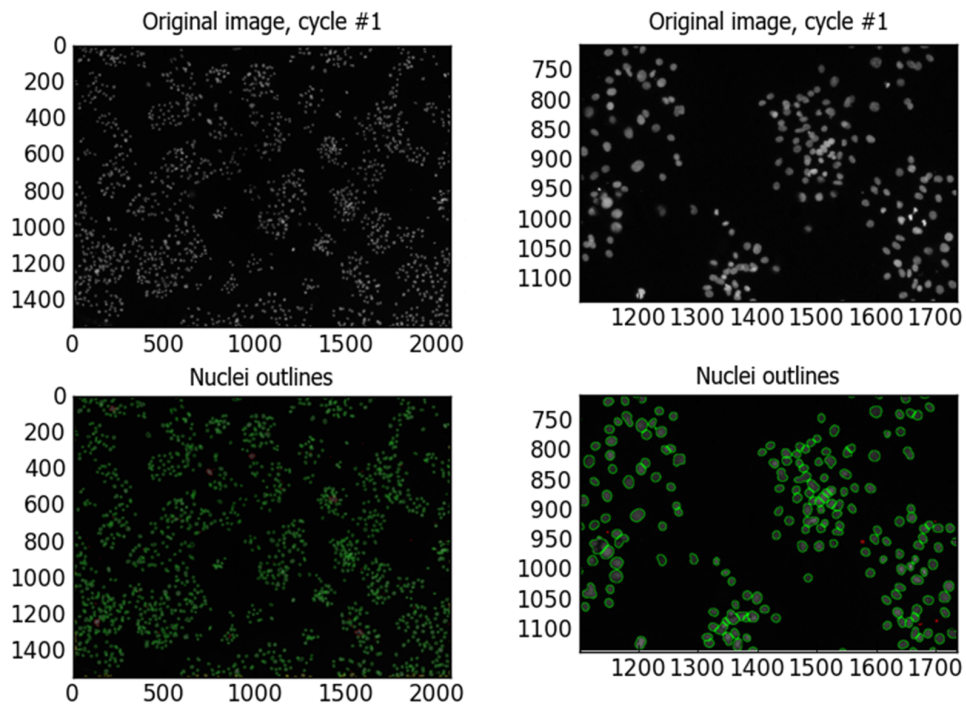
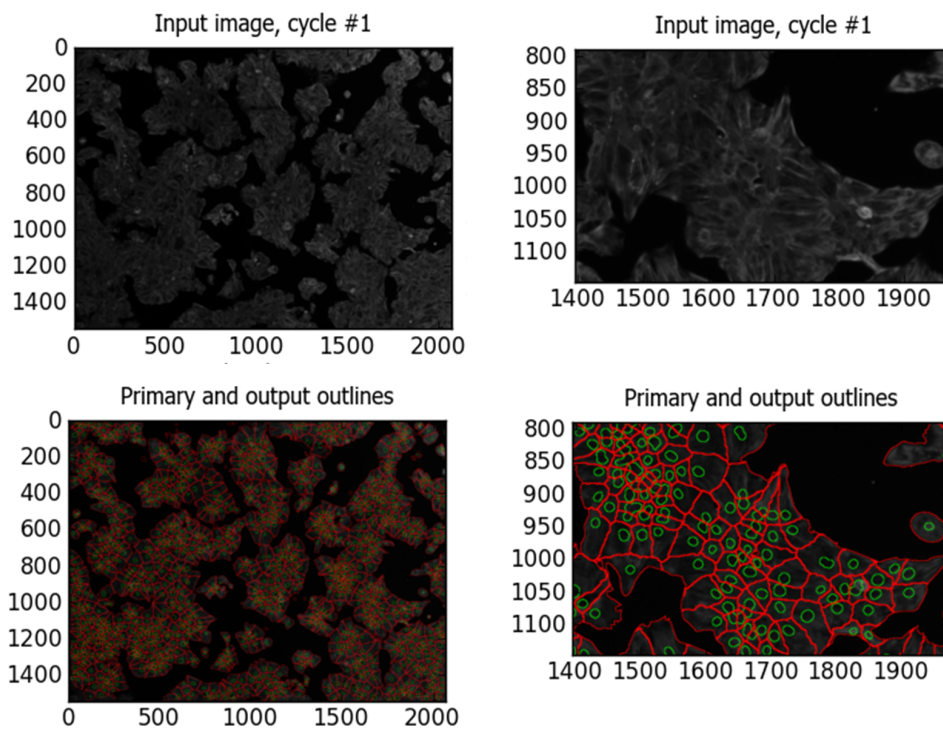
^a Calculated as $Re = \rho Q D_h / \mu A$, where density (ρ) = 1×10^3 kg/m³; Q represents flowrate (m³/s); D_h represents hydraulic diameter; viscosity (μ) = 10^{-3} Pa.s; and A represents cross-sectional area. Density and viscosity assumed as for water.

^b Calculated as $Pe = Q D_h / D_{AB} A$, where D_{AB} represents the diffusion coefficient.

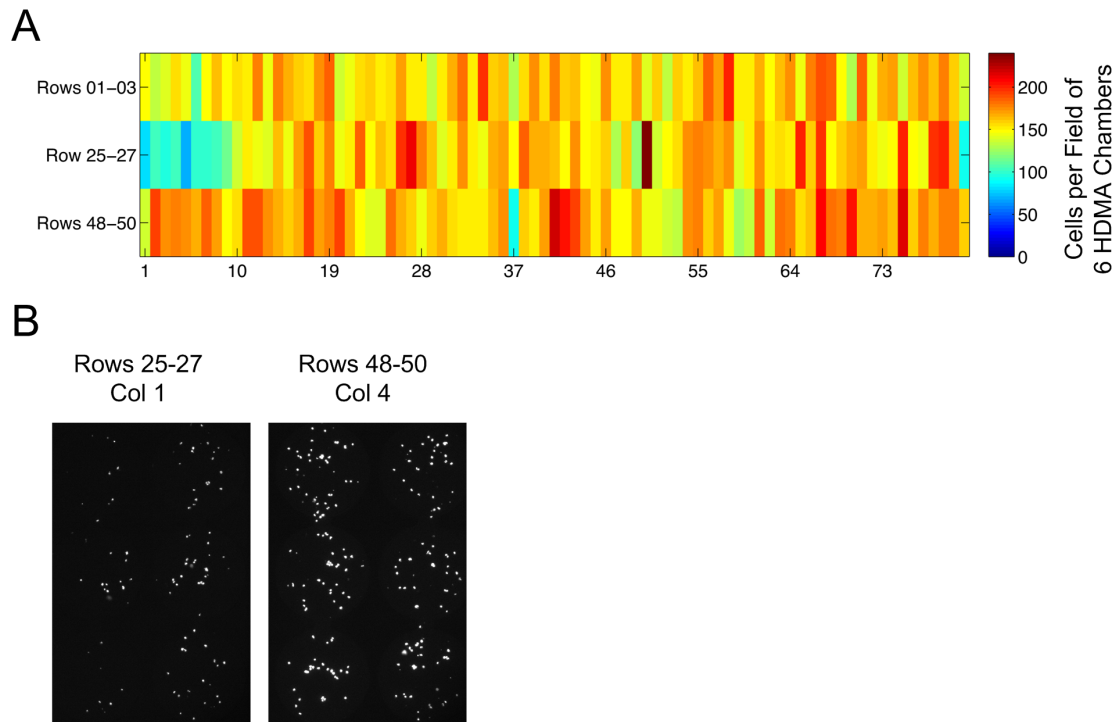
^c Shear stress estimated as $\tau = 6 \mu Q / h^2 w$, where height (h) = 2.5×10^{-4} m; and width (w) = 1.63×10^{-3} m for chamber at full width and 2.5×10^{-4} m for chamber interconnect.

A**B**

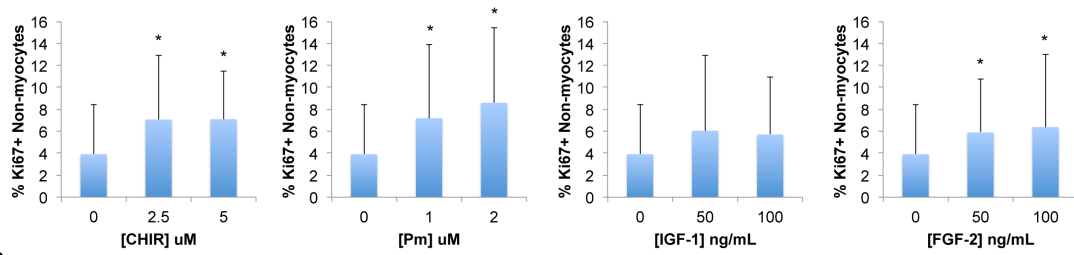
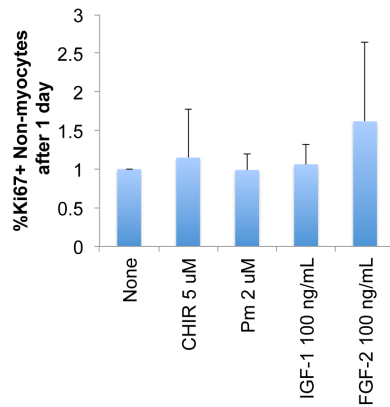
Supplementary Figure 1 - Optical Surface Profilometry. **A** 3D renderings of optical surface profiles of HDMA SU-8 masters. 2.5 mm x 1.9 mm sections of the cell culture array and fluidic channels are shown. **B** 2D representation and traced profiles showing height and width of microfabricated features (horizontal and vertical axes not in proportion).

A**Nuclei Detection (Hoechst)****B****Cell Detection (cTnT)**

Supplementary Figure 2 – Image Cytometry Processing by CellProfiler. **A** CellProfiler input and output images for primary object (nuclei) identification, whole image and close-up, from static control experiments. **B** CellProfiler input and output images for secondary object (cell) identification, whole image and close-up, from static control experiments.



Supplementary Figure 3 - Cell Seeding Distribution. **A** Heatmap showing distribution of fixed, Hoechst-labelled HES3 hESCs after injection into HDMA at 2×10^6 cells/mL. Image fields containing 6 HDMA chambers (three rows of one column pair) were acquired and nuclei counted by image cytometry. Sets of three rows were taken from the top (Rows 01-03), middle (Rows 25-27), and bottom (Rows 48-50) were measured. **B** Example images of a less dense (left), and normal density (right) condition.

A**B**

Supplementary Figure 4 - Non-myocyte Proliferative Response in HDMA Screening. **A** Data from individual column-pairs representing treatment with individual factors in HDMA. Bars represent mean and error bars represent S.D. of 50 rows from each column-pair where corresponding chambers in the 2 replicate columns are averaged. * indicates $p < 0.05$ versus control (no factors) by unpaired two-tailed t -test. **B** Quantification of percentage cTnT-Ki67⁺ non-myocytes after 24 h treatment in static cultures. Bars represent mean +/- S.D. of 3 independent experiments including separate cardiomyocyte inductions, normalised to control (None) conditions. No treatments were significantly different (paired two-tailed t -test) against control conditions.

Supplementary Videos

Supplementary Video 1 – Assay startpoint, video of spontaneously contracting C32 iPSC-derived cardiomyocytes in HDMA.

Supplementary Video 2 - Assay endpoint (3 days), video of spontaneously contracting C32 iPSC-derived cardiomyocytes in HDMA.