#### **Identification of Susceptibility Loci for Cutaneous Squamous Cell Cancer**

#### **Supplementary Materials**

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*Case verification.* Potentially eligible cases were identified from electronic pathology records of all pathology specimens from all KPNC medical facilities. All pathology specimens are assigned Systematized Nomenclature of Medicine (SNOMED) codes to classify diagnoses by organ (topography code) and morphological alteration (morphology code). The pathology records of all potential cases were queried for any report with a skin SNOMED topology code and cutaneous SCC morphology code. All pathology reports for potential SCC cases were reviewed and assigned case status (definite SCC, possible SCC, no SCC) by the study dermatologist (MA) based on the microscopic description of the pathology specimen in the electronic report. Only cases assigned definitive SCC status were included in the analysis.

*Subject Quality Control (QC)*. We used an ordered filtering process (**Table S8**) to exclude potentially eligible subjects with the following properties:

a) Ambiguous genetic sex (n=65), as determined by X-chromosome homozygosity rates between 0.2 and 0.8.

- b) Ambiguous SCC diagnosis (n = 1753).
- c) SNP call rate  $(CR)$  < 0.97 or SNP heterozygosity > 0.4 (n=59).
- d) Close genetic relationship to another eligible subject (n=3,696), defined by having expected number of alleles shared IBD greater than or equal to 0.25, as determined using 99,679 weakly correlated (pairwise  $R^2$  < 0.5) SNPs typed on all four arrays. We selected the case in discordant case-control pairs and the subject with greater CR in concordant pairs.

e) Absence of EUR principal components of ancestry (PCAs) (n = 329).

We implemented this filtering using PLINK v1.9 (https://www.cog-genomics.org/plink2/) (Chang et al., 2015) and custom programming in the R programming language. A total of 5,955 subjects failed the filter, leaving 67,867 eligible subjects, whom we classified in two array-based groups: the 61,457 subjects typed on the EUR array with the Axiom v1 reagent (screening phase), and the remaining 6,410 subjects typed on other arrays (replication phase). (**Table S9**).

**Genotype QC.** Genotype quality control has been described previously [Kvale 2015, http://www.ncbi.nlm.nih.gov/pubmed/26092718]. In addition, we excluded typed SNPs with CR<0.9.

*SNP Imputation.* Genotype imputation was performed by first pre-phasing the data with SHAPE-IT v2.r644 (Delaneau et al., 2012), using the family structure of first-degree cryptically related subjects. We then imputed a total of 31,081,353 variants from the 1000 Genomes Project (phase I integrated release, March 2012, with Aug 2012 chromosome X update, with singletons removed) as a cosmopolitan reference panel with IMPUTE2 v2.2.2 (Howie et al., 2009; Howie et al., 2011; Howie et al; 2012). We excluded 21,445,450 SNPs with minor allele frequency (MAF) <1% in either screening or replication phase and 25,435 SNPs with low imputation accuracy (defined as Information ≤ 0.3, where Information is the metric used in Impute2 to estimate the imputation accuracy (Marchini and Howie, 2010)). These exclusions left 9,610,468 variants for analysis.

*Population stratification.* We determined principal components of ancestry (PCAs) for the combined data from screening and replication phases using the smartpca program in the EIGENSOFT4.2 software package (Patterson et al., 2006), as has been described (Banda 2015, http://www.ncbi.nlm.nih.gov/pubmed/26092716). For each of the screening and replication phases, we then used PLINK to evaluate SCC-association with each SNP's allelic count using the Armitage-Cochran trend statistic, adjusted for gender, the first 10 principal components of ancestry, and for the replication phase, the genotyping array. We combined regression coefficients and P-values obtained from screening and replication phases using Cochran's method (Cochran 1954) implemented in R, and identified as significant those SNPs with combined P-values  $<$  5x10<sup>-8</sup> (Pe'er et al., 2008). We checked for residual population stratification by examining the inflation factor  $\,\lambda_{\rm GC}$  (Devlin and Roeder, 1999), and quantile-quantile plots of percentiles of the observed distribution of test statistics versus those expected under the global null hypothesis. We found little evidence for confounding by population stratification, as shown by the plot in **Figure S5,** and as indicated by an inflation factor of  $\lambda$  = 1.05.

*Data visualization and SNP annotation*. We visualized results using Manhattan plots created in R with QQman package (Turner, 2014) and plots generated by LocusZoom (Pruim et al., 2010) and HaploView (Barrett et al., 2005). To identify independent associations within a region, we used step-wise multiple logistic regressions, adjusted for the most strongly associated SNPs in the region. We also used the Annovar package (Wang et al., 2010) to perform RefSeq gene-based annotations of SCC-associated SNPs. We also used Annovar to map SNPs to the RegulomeDB (Boyle et al., 2012), GWAScatlog (Welter et al., 2014), and ClinVar (Landrum et al., 2014) databases to identify potential regulatory SNPs, and/or SNPs that have been previously associated with skin-pigmentation phenotypes or skin cancers, in addition to a PubMed search. We used the UCSC Coordinate Conversion (LiftOver, https://genome.ucsc.edu/cgi-bin/hgLiftOver) to convert genome coordinates in GWAScatlog from hg38 to hg19.

*Joint effects of pigment-related SCC genotypes and pigmentation phenotypes*. We used bivariate analyses to examine whether the pigment-related SNPs in Table 1A of the text are associated with SCC risk purely via their effects on skin pigmentation type (skin color, sun sensitivity and tanning ability), or through a mechanism independent of skin type (**Figure S6**). Since we lacked subjects' self-reported or clinically assessed skin phenotypes, we estimated them using their total counts of "risk alleles" for each skin phenotype, where the "risk allele" is the one associated positively with fair skin, sun sensitivity or inability to tan**.** Specifically, we compiled sets of SNPs associated (P<10<sup>-5</sup>) with each of three skin pigmentation phenotypes, using the GWASs cited in the GWAScatalog (http://www.ebi.ac.uk/gwas/). To exclude highly correlated SNPs (R<sup>2</sup>>0.85), we estimated pairwise R<sup>2</sup> based on the genotype data of the 1000 Genomes Project phase III subjects with European ancestry (CEU, TSI, FIN, GBR and IBS). The genotype data were extracted using the data slicer

[http://browser.1000genomes.org/Homo\_sapiens/UserData/SelectSlice] and R<sup>2</sup> was estimated using HaploView (Barrett et al., 2005). We used PHASE, version 2.1, (Stephens et al., 2001, 2003) to estimate the ASIP haplotypes based on genotype data. **Table S4** shows the SNPs used for each of the skin phenotypes. We also assigned each subject a polygenetic SCC risk score calculated as the total count of SCC risk alleles of the six pigment-related SNPs in text **Table 1**.

*Gene expression associations*. We examined whether the SCC-associated SNPs with univariate P-values meeting the genome-wide threshold  $5x10^{-8}$  are also associated with expression of nearby genes in both sun-exposed (lower leg) and non-sun-exposed (supra-pubic) skin tissue, in the version 6 analysis release (phs000424.v6.p1) of the Genotype-Tissue Expression project (GTEx Consortium, 2015). Gene expression association P-values were computed for SNP-gene pairs based on 302 samples of sun-exposed skin and 196 samples of non-sun-exposed skin using the interface available at http://www.gtexportal.org/home/testyourown

*Allelic effects of SNPs in Table 1*. For each of the SNPs at the 10 susceptibility loci in text Table 1, we used a likelihood ratio statistic (LRS) to evaluate how well allelic effects on SCC risk were

captured by a one-degree-of-freedom (DF) log-additive logistic model relative to a co-dominant (two DF) model. The LRS has a chi-square distribution with one DF under the null hypothesis that the log-additive allelic model fits well. We found significant improvement in fit for the co-dominant model relative to the log-additive model only for SNP rs1126809 in TYR at 11q24 (P = 3.6x10<sup>-3</sup>). SCC odds-ratios for heterozygote and homozygote carriers of the A-allele of this SNP were 1.13 (95% confidence interval: 1.07-1.19) and 1.51 (1.39-1.64), respectively. The co-dominant model also provided significant improvement in fit relative to a recessive model (P = 2.6x10<sup>-6</sup>).

*Potential interaction between pairs of significant SNPs*. We evaluated departures from an additive model for the dependence of SCC log odds on subjects' genotypes (g<sub>1</sub>,g<sub>2</sub>) at pairs of significant SNPs, for each of the 45 possible pairs of the 10 SNPs in Table 1 of the text. Specifically, we modeled the log odds of SCC as  $\alpha+\beta_1g_1+\beta_2g_2+\delta g_1g_2$  , and tested the null hypothesis  $\,\delta$  =  $0$  , using a likelihood ratio test. We found significant (P<0.05) evidence for interaction between the five SNP pairs shown in **Table S7.** Note that the interaction term is negative for four of the five pairs, which may relate to the roles of the genes in synthesizing and transporting melanin (**Figure S7).** 

#### **REFERENCES**

Banda Y, Kvale MN, Hoffmann TJ, Hesselson SE, Ranatunga D, Tang H, et al. Characterizing Race/Ethnicity and Genetic Ancestry for 100,000 Subjects in the Genetic Epidemiology Research on Adult Health and Aging (GERA) Cohort. Genetics. 2015;200(4):1285-95.

Barrett JC, Fry B, Maller J, Daly MJ. Haploview: analysis and visualization of LD and haplotype maps. Bioinformatics. 2005;21(2):263-5.

Bastiaens MT, ter Huurne JA, Kielich C, Gruis NA, Westendorp RG, Vermeer BJ, et al. Melanocortin-1 receptor gene variants determine the risk of nonmelanoma skin cancer independently of fair skin and red hair. Am J Hum Genet. 2001;68(4):884-94.

Branicki W, Brudnik U, Kupiec T, Wolanska-Nowak P, Szczerbinska A, Wojas-Pelc A. Association of polymorphic sites in the OCA2 gene with eye colour using the tree scanning method. Ann Hum Genet. 2008;72(Pt 2):184-92.

Branicki W, Brudnik U, Wojas-Pelc A. Interactions between HERC2, OCA2 and MC1R may influence human pigmentation phenotype. Ann Hum Genet. 73. England2009. p. 160-70. Box NF, Duffy DL, Irving RE, Russell A, Chen W, Griffyths LR, et al. Melanocortin-1 receptor genotype is a risk factor for basal and squamous cell carcinoma. J Invest Dermatol. 2001;116(2):224-9.

Boyle AP, Hong EL, Hariharan M, Cheng Y, Schaub MA, Kasowski M, et al. Annotation of functional variation in personal genomes using RegulomeDB. Genome Res. 2012;22(9):1790-7. Chang CC, Chow CC, Tellier LC, Vattikuti S, Purcell SM, Lee JJ. Second-generation PLINK: rising to the challenge of larger and richer datasets. Gigascience. 2015;4:7.

Cochran WG. The combination of estimates from different experiments. Biometrics. 1954;10:101- 29.

Delaneau O, Marchini J, Zagury JF. A linear complexity phasing method for thousands of genomes. Nat Methods. 2012;9(2):179-81.

Devlin B, Roeder K. Genomic control for association studies. Biometrics. 1999;55(4):997-1004. Duffy DL, Montgomery GW, Chen W, Zhao ZZ, Le L, James MR, et al. A three-single-nucleotide polymorphism haplotype in intron 1 of OCA2 explains most human eye-color variation. Am J Hum Genet. 2007;80(2):241-52.

Edwards M, Bigham A, Tan J, Li S, Gozdzik A, Ross K, et al. Association of the OCA2 polymorphism His615Arg with melanin content in east Asian populations: further evidence of convergent evolution of skin pigmentation. PLoS Genet. 2010;6(3):e1000867.

Eiberg H, Troelsen J, Nielsen M, Mikkelsen A, Mengel-From J, Kjaer KW, et al. Blue eye color in humans may be caused by a perfectly associated founder mutation in a regulatory element located within the HERC2 gene inhibiting OCA2 expression. Hum Genet. 2008;123(2):177-87.

GTEx Consortium. The Genotype-Tissue Expression (GTEx) pilot analysis: multitissue gene regulation in humans. Science. 2015;348(6235):648-60.

Han J, Kraft P, Nan H, Guo Q, Chen C, Qureshi A, et al. A genome-wide association study identifies novel alleles associated with hair color and skin pigmentation. PLoS Genet. 2008;4(5):e1000074.

Howie BN, Donnelly P, Marchini J. A flexible and accurate genotype imputation method for the next generation of genome-wide association studies. PLoS Genet. 2009;5(6):e1000529. Howie B, Fuchsberger C, Stephens M, Marchini J, Abecasis GR. Fast and accurate genotype imputation in genome-wide association studies through pre-phasing. Nat Genet. 2012;44(8):955-9. Howie B, Marchini J, Stephens M. Genotype imputation with thousands of genomes. G3 (Bethesda). 2011;1(6):457-70.

Kayser M, Liu F, Janssens AC, Rivadeneira F, Lao O, van Duijn K, et al. Three genome-wide association studies and a linkage analysis identify HERC2 as a human iris color gene. Am J Hum Genet. 2008;82(2):411-23.

Kennedy C, ter Huurne J, Berkhout M, Gruis N, Bastiaens M, Bergman W, et al. Melanocortin 1 receptor (MC1R) gene variants are associated with an increased risk for cutaneous melanoma which is largely independent of skin type and hair color. J Invest Dermatol. 2001;117(2):294-300. Kvale MN, Hesselson S, Hoffmann TJ, Cao Y, Chan D, Connell S, et al. Genotyping informatics and quality control for 100,000 subjects in the Genetic Epidemiology Research on Adult Health and Aging (GERA) Cohort. Genetics. 2015;200(4):1051-60.

Landrum MJ, Lee JM, Riley GR, Jang W, Rubinstein WS, Church DM, et al. ClinVar: public archive of relationships among sequence variation and human phenotype. Nucleic Acids Res.

2014;42(Database issue):D980-5.

Marchini J, Howie B. Genotype imputation for genome-wide association studies. Nat Rev Genet. 2010;11(7):499-511.

Mengel-From J, Borsting C, Sanchez JJ, Eiberg H, Morling N. Human eye colour and HERC2, OCA2 and MATP. Forensic Sci Int Genet. 2010;4(5):323-8.

Nan H, Kraft P, Qureshi AA, Guo Q, Chen C, Hankinson SE, et al. Genome-wide association study of tanning phenotype in a population of European ancestry. J Invest Dermatol. 2009;129(9):2250-

7.

Palmer JS, Duffy DL, Box NF, Aitken JF, O'Gorman LE, Green AC, et al. Melanocortin-1 receptor polymorphisms and risk of melanoma: is the association explained solely by pigmentation phenotype? Am J Hum Genet. 2000;66(1):176-86.

Patterson N, Price AL, Reich D. Population structure and eigenanalysis. PLoS Genet. 2006;2(12):e190.

Pe'er I, Yelensky R, Altshuler D, Daly MJ. Estimation of the multiple testing burden for genomewide association studies of nearly all common variants. Genet Epidemiol. 2008;32(4):381- 5.

Pruim RJ, Welch RP, Sanna S, Teslovich TM, Chines PS, Gliedt TP, et al. LocusZoom: regional visualization of genome-wide association scan results. Bioinformatics. 2010;26(18):2336-7. Sturm RA, Duffy DL, Zhao ZZ, Leite FP, Stark MS, Hayward NK, et al. A single SNP in an evolutionary conserved region within intron 86 of the HERC2 gene determines human blue-brown eye color. Am J Hum Genet. 2008;82(2):424-31.

Stephens M, Donnelly P. A comparison of bayesian methods for haplotype reconstruction from population genotype data. Am J Hum Genet. 2003;73(5):1162-9.

Stephens M, Smith NJ, Donnelly P. A new statistical method for haplotype reconstruction from population data. Am J Hum Genet. 2001;68(4):978-89.

Sulem P, Gudbjartsson DF, Stacey SN, Helgason A, Rafnar T, Magnusson KP, et al. Genetic determinants of hair, eye and skin pigmentation in Europeans. Nat Genet. 2007;39(12):1443-52. Turner SD. qqman: an R package for visualizing GWAS results using QQ and manhattan plots. 2014; Doi: http://dx.doi.org/10.1101/005165.

Wang K, Li M, Hakonarson H. ANNOVAR: functional annotation of genetic variants from highthroughput sequencing data. Nucleic Acids Res. 2010;38(16):e164.

Welter D, MacArthur J, Morales J, Burdett T, Hall P, Junkins H, et al. The NHGRI GWAS Catalog, a curated resource of SNP-trait associations. Nucleic Acids Res. 2014;42(Database issue):D1001- 6.

### **Supplementary Tables**

### **Table S1. SNPs in HERC2 and OCA2**

#### **A. Pigment-related SNPs**



### **B. SNPs achieving genome-wide significance after adjusting for ten SNPs in Table 1**



\* Impute-2 measure of imputation accuracy;<br> $\frac{1}{1}$  Odds ratio per minor allele associated with SCC risk and 95% confidence interval;<br> $\frac{1}{1}$  mOR = odds-ratio per minor allele, adjusted for gender, ancestry, array/reag

genotypes at 10 SNPs in Table 1 of text.

## **Table S2. Nonsynonymous SNPs in MC1R previously associated with pigmentation and skin cancer**



\* Impute-2 measure of imputation accuracy;

† The allele that was associated with skin cancer-predisposing pigmentation traits;

‡ Odds ratios (and 95% confidence intervals) associated with the risk of SCC in the present study.

## **Table S3. Odds-ratios (ORs) for chromosome 16 SNPs, after adjustment for the total dosage of the MC1R nonsynonymous SNPs\***



\* SNPs listed in Table S.4 with MAF≥0.01 and with an rs number.



## **Table S4. SNPs determining polygenic scores for: A. skin color, B. sun sensitivity and C. tanning ability\***

 $^*$  all SNPs obtained from GWAS catalogue (with p<5x10<sup>-6</sup>), excluding redundant ones either due to high LD with other SNPs in the table or deemed non-independent signal from other SNPs;<br><sup>†</sup> allele associated with high-risk pigment phenotype;<br><sup>‡</sup> OR and 95% CI associated with SCC in the present study;<br><sup>§</sup> Replacing rs15

¶ Not in GWAS catalog, but a significant predictor of skin sensitivity to sun in Sulem et al., 2007.

#### **Table S5. SCC risk in relation to joint tertiles of SCC risk allele count and score for: A. skin color; B. sun sensitivity and C. tanning ability**



risk alleles of top SNPs at six pigment-related loci;

† OR=odds-ratio, adjusted for gender, PCAs and genotyping array/reagent. Marginal OR's for each score are also adjusted for tertiles of the other score;

‡ CI=95% confidence interval;

 $\frac{8}{3}$  polygenic skin color score: higher allele counts associated with lighter skin color;

<sup>¶</sup> number of SNP alleles associated with burning;<br><sup>\\</sup> number of SNP alleles associated with tanning.



### **Table S6. Effects of SNP rs4455710 at 6p21, by history of smoking and immunosuppression**

 $*$  Interaction P-value = 0.3833

 $†$  Interaction P-value = 0.4483

‡ 42 with organ transplant, 52 with chronic lymphocytic leukemia (CLL), and 17 with HIV infection.

§ 49 with organ transplant, 99 with CLL and 207 with HIV infection.

# **Table S7. Significant (P<0.05) evidence for epistasis between pairs of SNPs in text Table 1**



\* log odds-ratio;

 $<sup>†</sup>$  SD = standard deviation;</sup>

‡ on log-additive scale;

§ interaction based on chi-squared likelihood ratio test.

## **Table S8. Subject Quality Control**



 $\check{C}$  CR = call rate;

 $^\dagger$  het = heterozygosity (observed heterozygotes/total number of SNPs with no missing genotypes);<br> $^\ddag$  expected number of alleles shared IBD >= 0.25.

# **Table S9. Distribution of Eligible Subjects by Study Phase, Case-control Status and Gender**



### **Supplementary Figures**



**Figure S1.** Manhattan plot enlargement showing significance levels for selected SNPs in 0.35 Mb region at Chr15q13. SNPs listed include top SNP (red), those previously associated with pigmentation *traits* (italics) and those that achieved significance after adjusting for all ten SNPs in Table 1 (bold).



**Figure S2.** Squared correlation coefficients  $(R^2)$  between pairs of statistically significant and previously reported SNPs in the HERC2/OCA2 region on chromosome 15q13, based on Caucasian 1000 Genomes data. Black squares indicate perfect correlation.



**Figure S3.** Manhattan plot enlargement showing 3.5 Mb region at the 20q11 locus. The upper panel shows the name and location of genes in the region, with an arrow indicating the transcribed strand of a gene and ticks indicating exons. The genes *RALY* (containing the top SNP rs6059655) and *ASIP* (associated with skin pigmentation and skin cancer) are shown in boxes. The lower panel shows the significance levels of the SNPs in this region.



**Figure S4.** Manhattan plot enlargement showing 0.55 Mb region at the 6p21 locus. The upper panel shows the name and location of genes in the region, with an arrow indicating the transcribed strand of a gene and ticks indicating exons. The box encloses the gene *HLA-DQA1* containing the top SNP rs4455710. The lower panel shows the significance levels of the SNPs in this region.



**Figure S5.** Plot of P-values > 10<sup>-8</sup> corresponding to percentiles of the empirical distribution of 9,610,468 squared Cochran-Armitage trend statistics (vertical axis) versus P-values of percentiles of a central chisquared variable with one degree of freedom (horizontal axis).



**Figure S6.** Possible mechanisms for association of SCC risk with SNPs at pigment-related loci



**Figure S7.** MC1R, located on the plasma membrane of melanocytes, activates the cAMP signaling pathway that increases MITF expression, while ASIP and other MC1R-antagonists block this pathway. MITF up-regulates the transcription of major melanogenic enzymes such as tyrosine kinase encoded by TYR, and can also activate expression of IRF4, which cooperates with MITF to activate TYR expression. The membrane-associated transporters SLC45A2 and OCA2 are involved in post-translation processing and transporting of melanosomal proteins into the melanosomes and thereby supporting melanin production. Melanin in the melanocytes is then transported to keratinocytes, where it forms a coat around their nuclei and protects them from UVR-induced DNA damage.