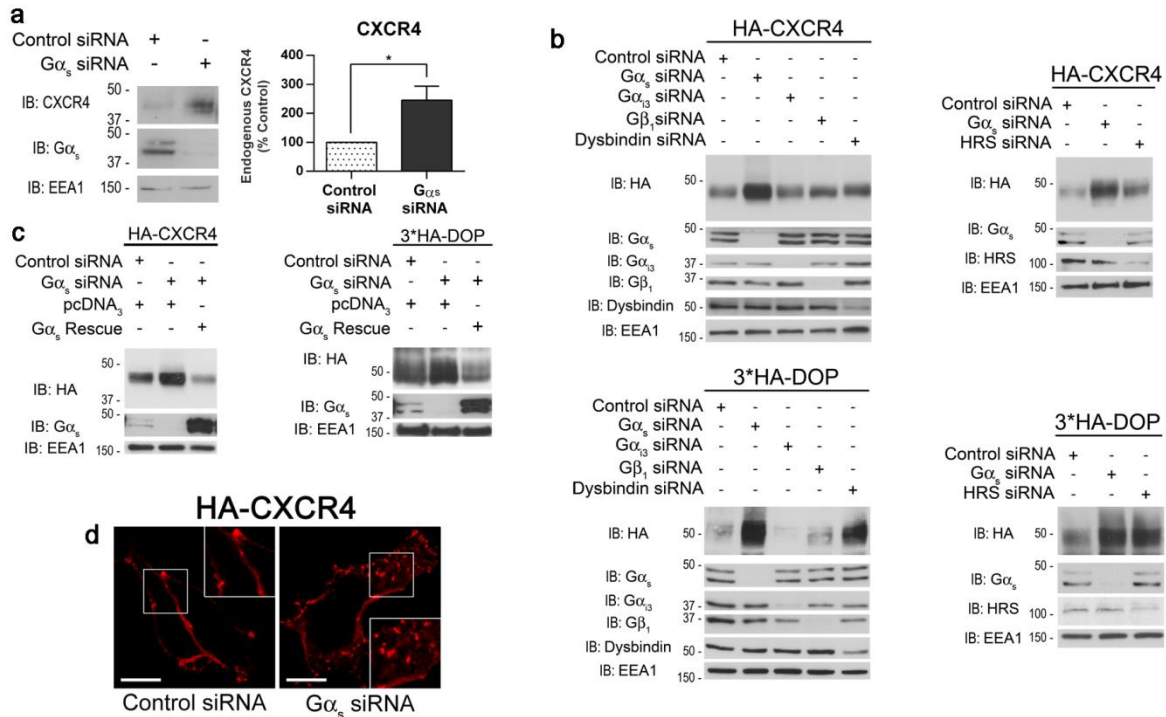
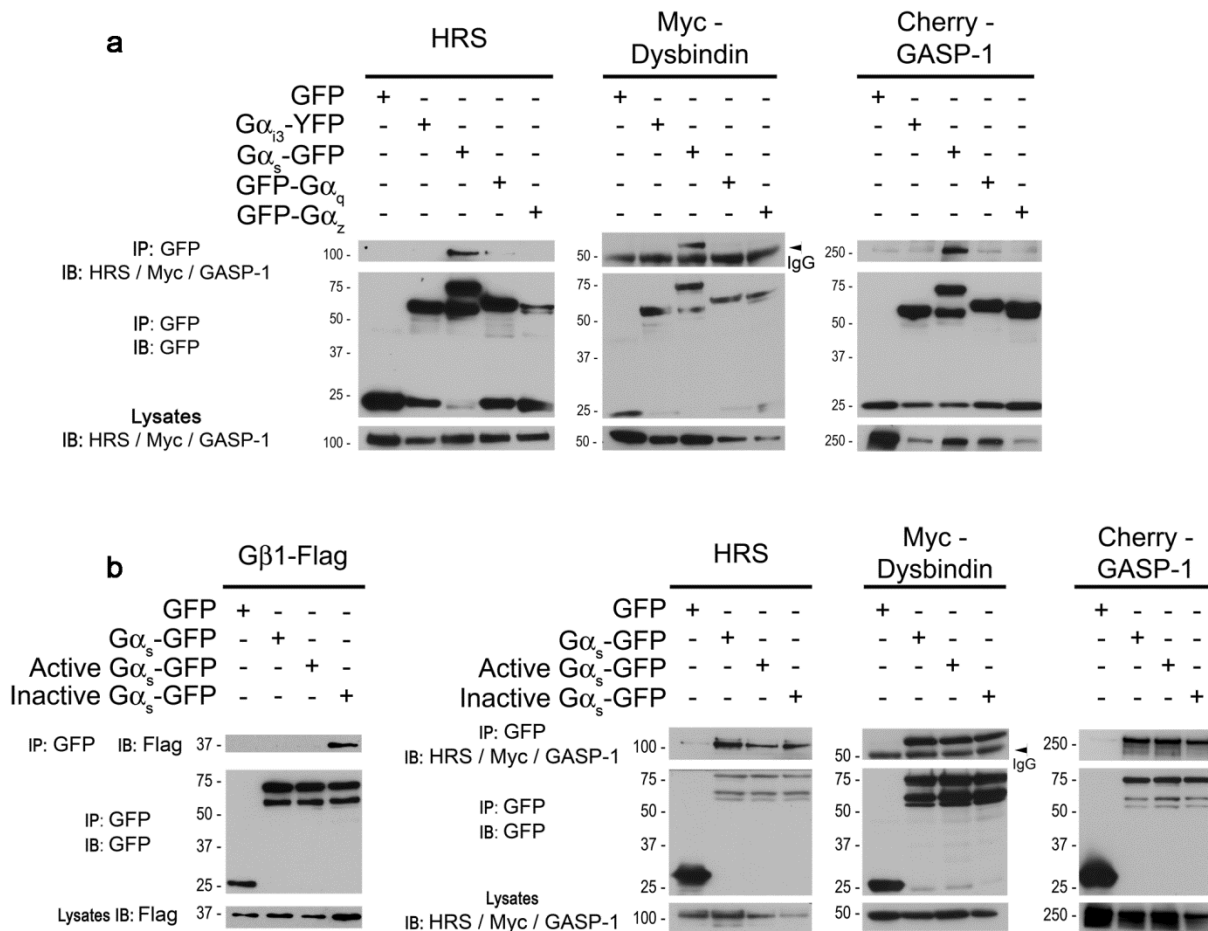


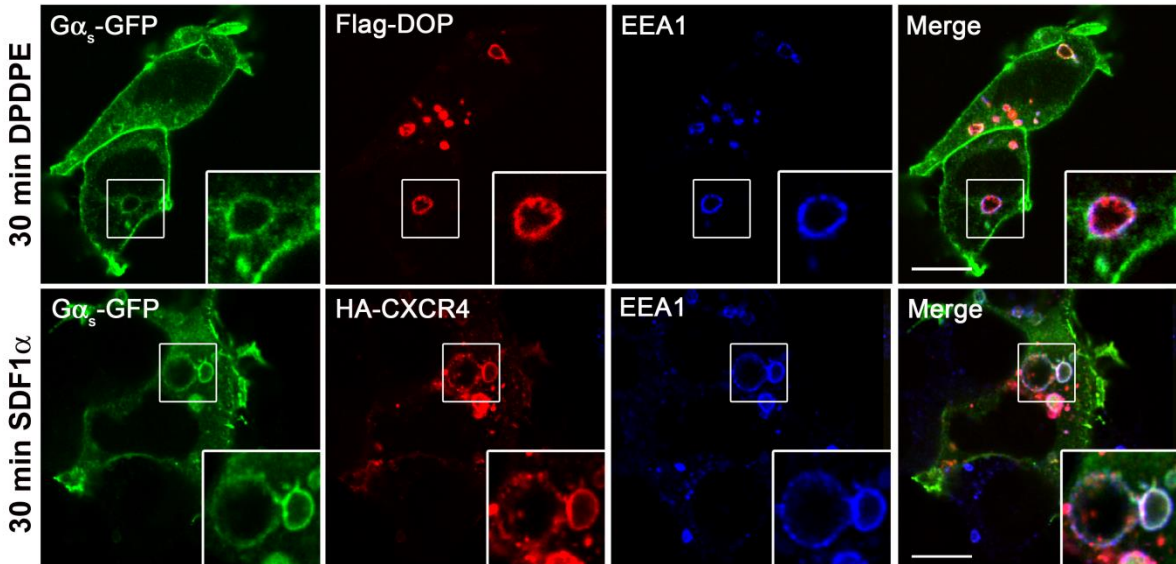
Supplementary Information



Supplementary Figure 1. Specificity of the effect of G α_s knockdown on the basal levels of CXCR4 and DOP. (a) HeLa cells endogenously expressing CXCR4 receptor were transfected with control or G α_s siRNA for 72 h. Steady-state levels of these tagged-GPCRs were analyzed by immunoblotting using anti-CXCR4 antibody. EEA1, loading controls. The histogram depicts the quantification of CXCR4 endogenous level of expression presented as percentage of levels quantified in control cells. Values shown are means \pm SEM of ≥ 3 separate experiments. * $p < 0.05$. (b) Effect of depletion of other heterotrimeric G proteins and components of the endosomal sorting machinery on the basal levels of HA-CXCR4 and Flag-DOP. HEK293 cells were transfected with control, G α_s , G α_{13} , G β_1 , dysbindin or HRS siRNA. Twenty-four hours later, cells were transfected with HA-CXCR4 and HA-DOP and incubated for another 48 hours. Cells were lysed and analyzed by immunoblotting (IB) using the indicated antibody. EEA1 was used as a loading controls. (c) Expression of G α_s rescues the basal levels of HA-CXCR4 and HA-DOP. Control or G α_s siRNA-treated HEK293 cells expressing HA-CXCR4 or HA-DOP were transfected with pcDNA3 (control vector) or siRNA-resistant versions of the short and long forms of G α_s (G α_s Rescue) 10 h after the initial human G α_s siRNA transfection. Cells were lysed and analyzed by immunoblotting (IB) using the indicated antibody. EEA1 was used as loading controls. (d) Immunofluorescence analysis of the steady state distribution of HA-CXCR4 in control or G α_s siRNA-treated HEK293 cells. Cells were fixed, permeabilized, immunostained with anti-HA antibodies and processed for confocal microscopy analysis. In controls cells, HA-CXCR4 localized at cell surface whereas in G α_s siRNA treated cells, GPCR localized at cell surface and in intracellular compartments. Scale bars, 10 μ m.



Supplementary Figure 2. Dysbindin and GASP-1 interact specifically with G α_s -GFP independently of its activity state. (a) Dysbindin and GASP-1 do not interact with other G α proteins. Lysate from HEK293 cells transiently transfected with GFP, G α_s -GFP, G α_{13} -GFP, G α_q -GFP or G α_z -GFP together with HRS, Myc-dysbindin or Cherry-GASP1 were immunoprecipitated (IP) with anti-GFP and immunoblotted (IB) using the indicated antibody. Only G α_s -GFP coimmunoprecipitated with all these proteins. (b) Constitutively active and inactive forms of G α_s similarly co-immunoprecipitated with dysbindin, HRS and GASP-1. HEK293 cells were transiently transfected with GFP, wild-type G α_s -GFP, constitutively active G α_s -GFP (Q227L) or inactive G α_s -GFP (G226A; R280T; T284D; I285T and A366S) together with HRS, Myc-dysbindin or Cherry-GASP1. Cells were lysed, immunoprecipitated (IP) with anti-GFP and immunoblotted (IB) using the indicated antibody. Coimmunoprecipitation of endogenous G β 1 was used as a control to confirm the activation status of G α_s mutants since G β 1 only binds inactive G α_s .



Supplementary Figure 3. $G\alpha_s$ colocalized with internalized GPCR on early endosomes. $G\alpha_s$ -GFP and FLAG-DOP or HA-CXCR4 are together on early endosomes. HEK293 cells stably expressing Flag-DOP or transiently expressing HA-CXCR4 were transfected with $G\alpha_s$ -GFP and Rab5Q79L, which created enlarged early endosomes. Cell-surface Flag-DOP or HA-CXCR4 was labeled with anti-Flag or HA at 4°C before incubation at 37°C in the presence of agonist for 30 min. Cells were fixed, permeabilized, stained with anti-GFP and anti-EEA1 (endogenous marker of early endosomes) and examined by confocal microscopy. The merged image indicates colocalization between $G\alpha_s$ and internalized Flag-DOP or HA-CXCR4 on EEA1-labeled enlarged endosomes. Scale bars, 10 μ m.