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**Supplemental Information**

**Regulation of PI3K by PKC and MARCKS: Single-Molecule Analysis of a Reconstituted Signaling Pathway**

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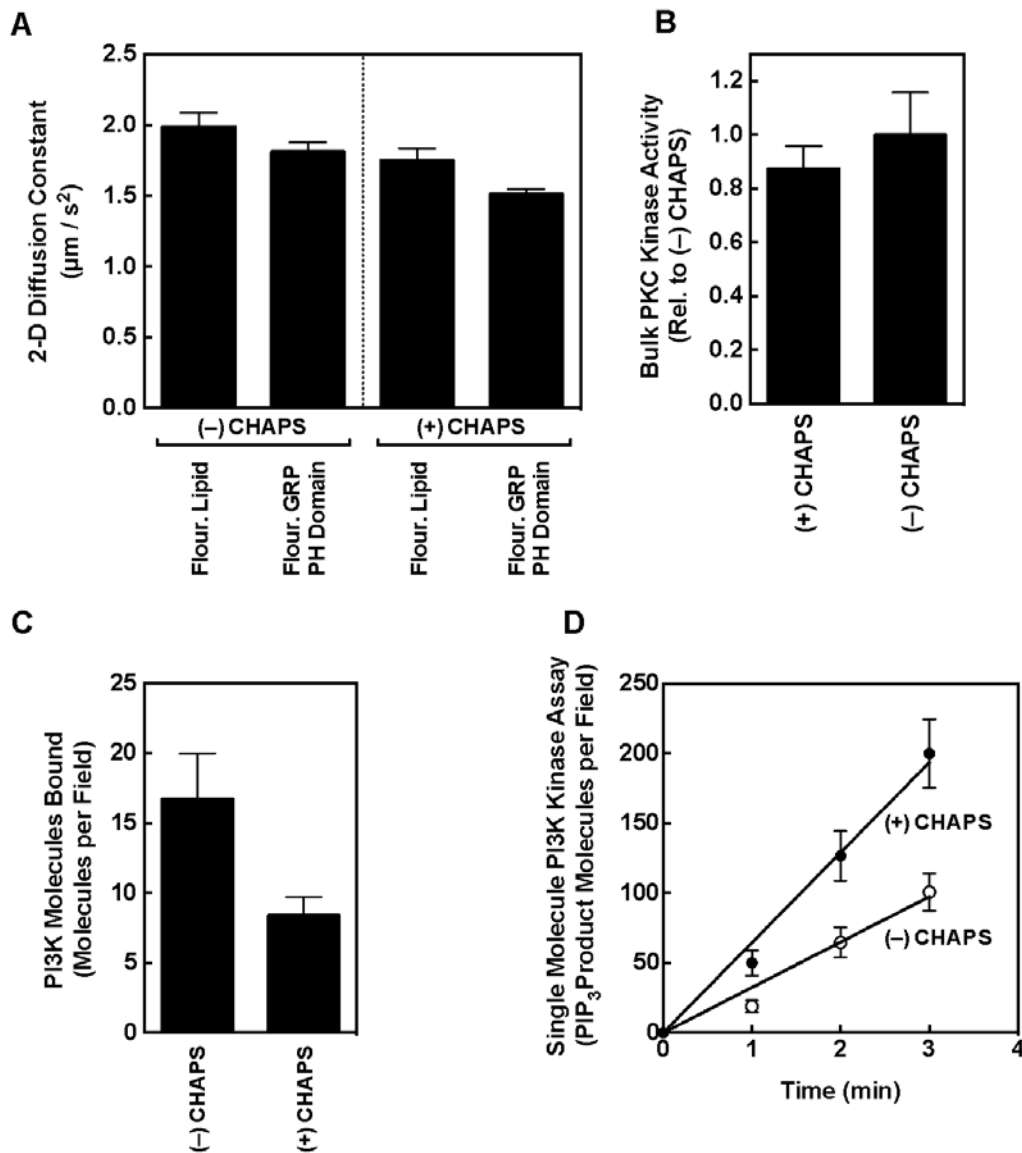
**Supporting Material**

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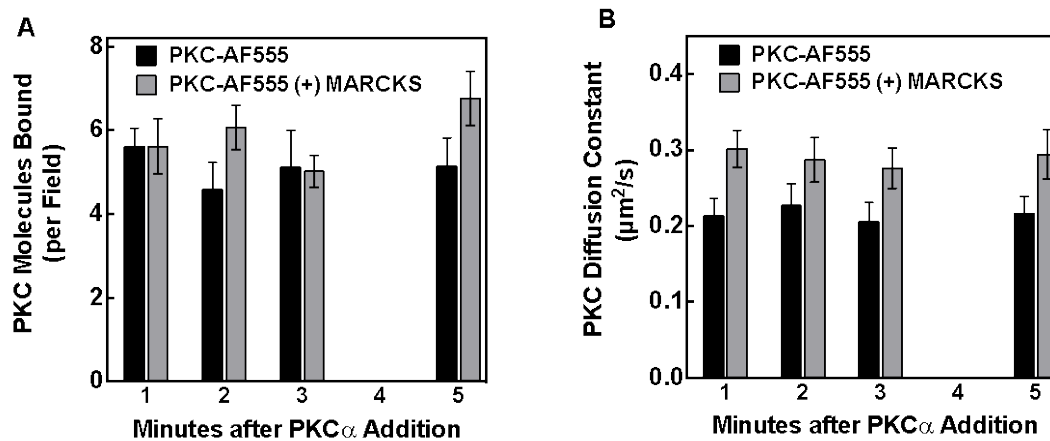
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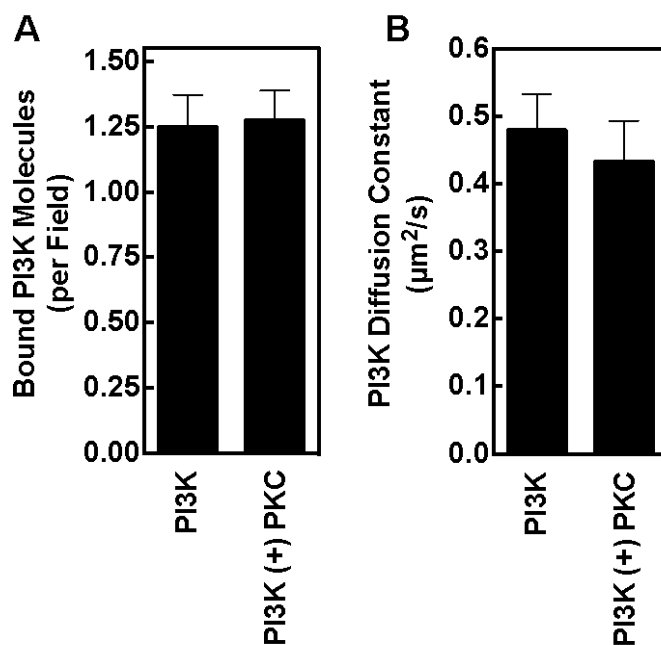
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**(Supporting Material, Figure S1)** Control experiments to test effect of CHAPS on fluorescent lipid and protein diffusion and kinase activity. **(A)** CHAPS has no significant effect on the diffusion of the fluorescent lipid LRB-PE, but causes a small (<15%), reproducible ( $p < 0.05$ ) diffusional slowing of fluorescent GRP1 PH domain bound to PIP<sub>3</sub>. **(B)** CHAPS detergent has no significant effect on PKC $\alpha$  kinase activity. **(C)** CHAPS significantly decreases binding of PI3K $\alpha$  to the target membrane ( $p < 0.05$ ). **(D)** CHAPS significantly increases PI3K $\alpha$  lipid kinase activity ( $p < 0.01$ ). Single molecule TIRF was carried out at  $21.5 \pm 0.5$  °C on PE/PS/DAG/PIP<sub>3</sub>  $\pm$  200 ppb LRB-PE **(A)** or standard PE/PS/DAG/PIP<sub>2</sub> **(B-D)** supported bilayers as detailed in Fig. 4.



(Supporting Material, Figure S2) Control experiments testing the effect of MARCKSp on fluorescent Ca<sup>2+</sup>-PKC $\alpha$  binding and diffusion. (A) MARCKSp does not significantly affect Ca<sup>2+</sup>-PKC $\alpha$  binding to bilayers consistent with its high affinity for PS even in the absence of PIP<sub>2</sub> (1). (B) MARCKSp generates a small (<25%), reproducible ( $p < 0.005$ ) increase in the 2-D diffusion speed of membrane-bound Ca<sup>2+</sup>-PKC $\alpha$ , likely due to PIP<sub>2</sub> ligand sequestration which prevents C2 domain binding to PIP<sub>2</sub> as well as preventing the resulting diffusional slowing (2). Single molecule TIRF assay at  $21.5 \pm 0.5$  °C on standard PE/PS/DAG/PIP<sub>2</sub> supported bilayers as detailed in Fig. 4. In these experiments PKC $\alpha$  was employed at the standard total concentration (Table 1) used in other experiments, but at this concentration the density of membrane bound PKC $\alpha$  is too high to resolve single particles for counting and tracking. Thus, a mixture of fluorescent (0.005%) and dark, unlabeled PKC $\alpha$  (99.995%) was employed yielding the density of bound fluorescent PKC $\alpha$  indicated in (A).



(Supporting Material, Figure S3) Control experiment testing for direct interaction between PKC and PI3K on the membrane surface. PKC $\alpha$  is observed to have no significant effect on (A) the membrane binding or (B) the 2-D diffusion constant of fluorescent PI3K on standard PE/PS/DAG/PIP<sub>2</sub> supported bilayers as detailed in Fig. 4.

## REFERENCES

1. Manna, D., N. Bhardwaj, M. S. Vora, R. V. Stahelin, H. Lu, and W. Cho. 2008. Differential roles of phosphatidylserine, PtdIns(4,5)P<sub>2</sub>, and PtdIns(3,4,5)P<sub>3</sub> in plasma membrane targeting of C2 domains. Molecular dynamics simulation, membrane binding, and cell translocation studies of the PKC $\alpha$  C2 domain. *J Biol Chem* 283:26047-26058.
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