

Figure S1

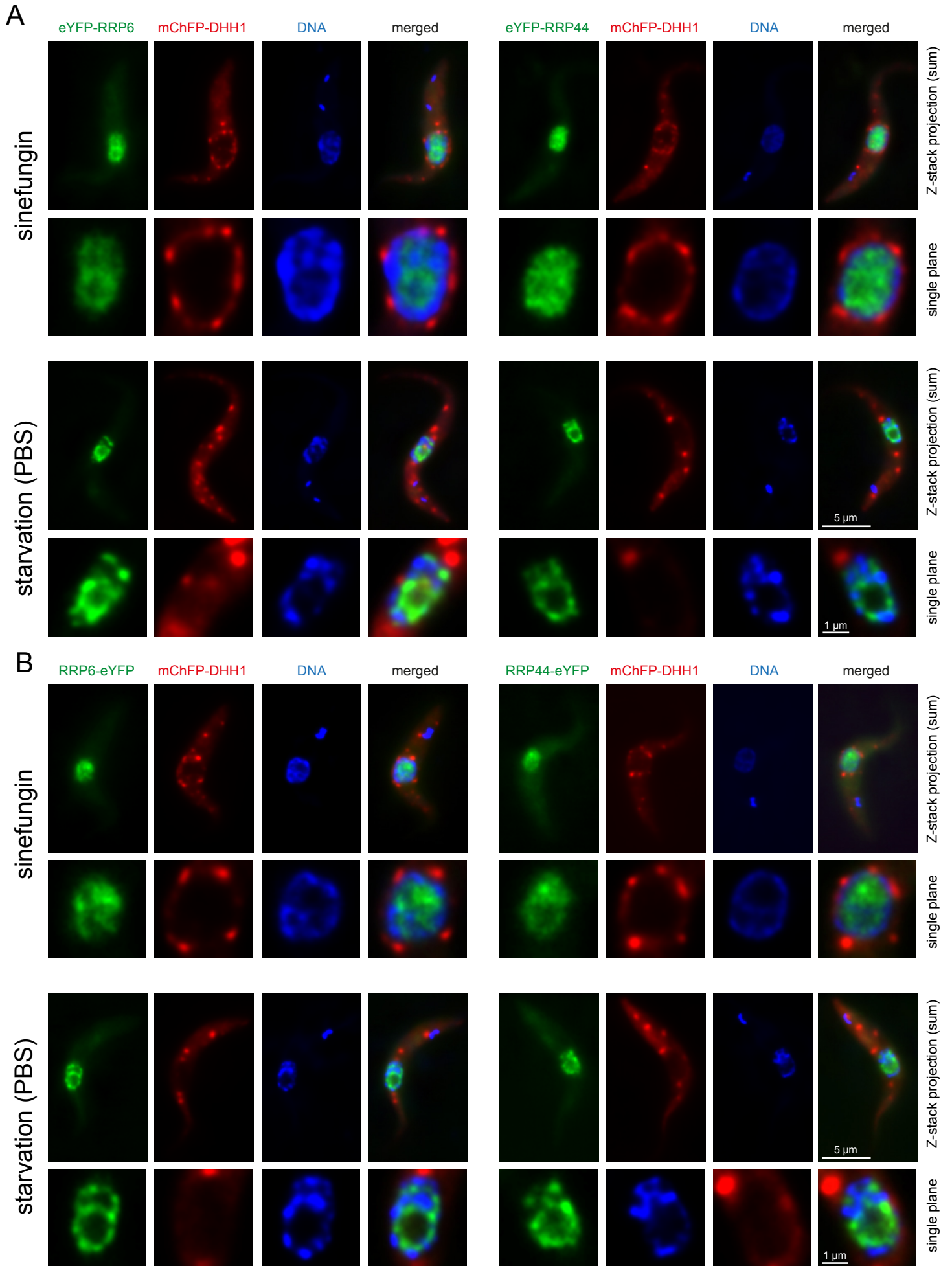


Figure S2

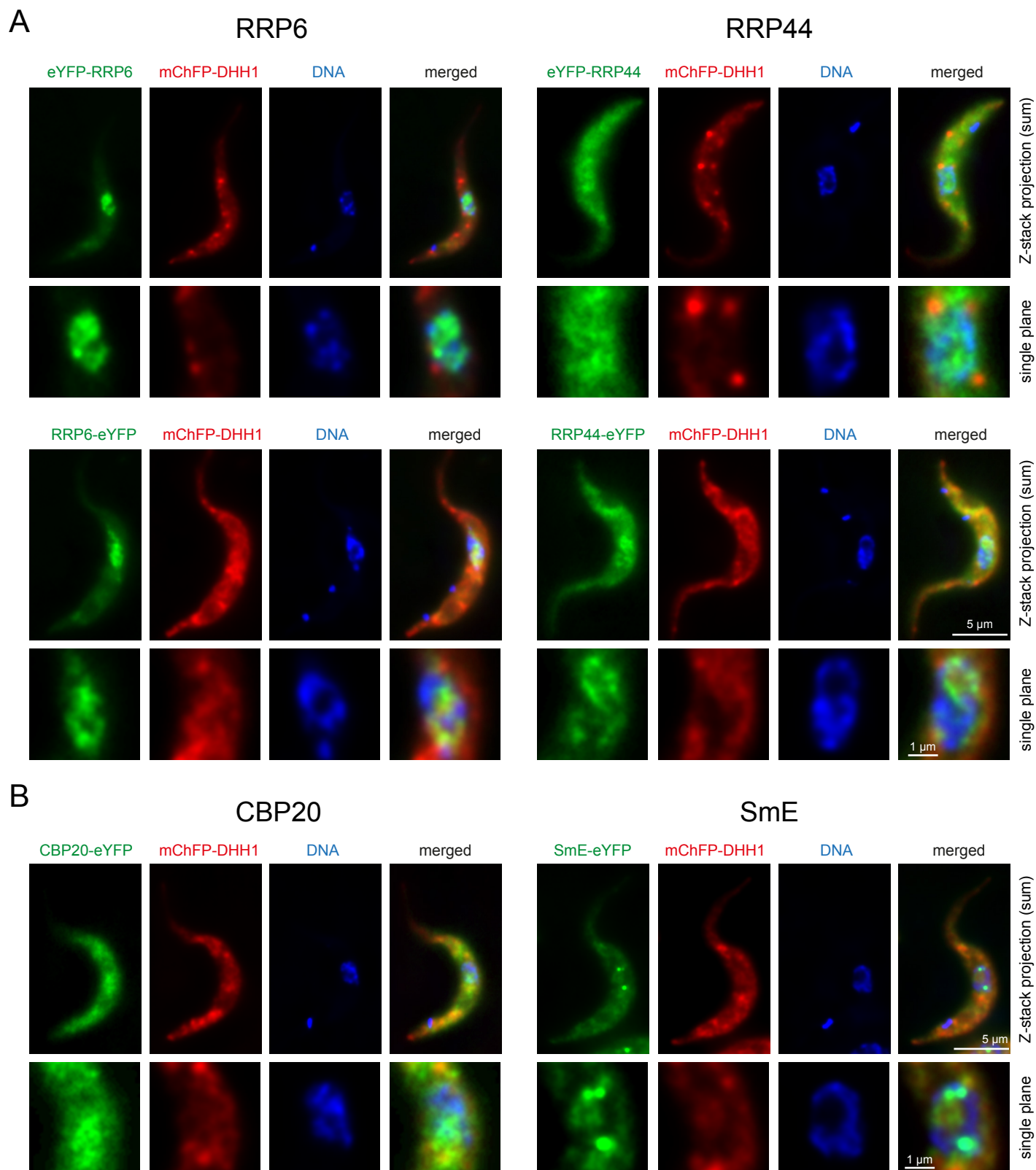


Figure legends

Fig. S1: Localization of N and C-terminal eYFP fusions of RRP6 and RRP44 at starvation or sinefungin treatment

Cells expressing mChFP-DHH1 and RRP6 or RRP44 as either N-terminally **(A)** or C-terminally **(B)** tagged eYFP fusion protein were treated with sinefungin (2 µg/ml, 1 hour) or starved (cultured in PBS for 2 hours).

Z-stacks (100 images, 100-nm spacing) were recorded with a custom-built TILL Photonics iMIC microscope equipped with a 100×, 1.4 numerical aperture objective (Olympus, Tokyo, Japan) and a sensicam qe CCD camera (PCO, Kehlheim, Germany) and deconvolved using Huygens Essential software (SVI, Hilversum, The Netherlands). For each cell line, one representative fluorescent cell is shown as a Z-stack projection (method sum slices). In addition, the nucleus of a deconvolved single plane image is shown enlarged.

Fig. S2: Localization of N and C-terminal eYFP fusions of RRP6 and RRP44 at heat shock treatment

Cells expressing mChFP-DHH1 and RRP6 or RRP44 as either N-terminally or C-terminally tagged eYFP fusion protein **(A)** and, as a control, cells expressing CBP20-eYFP or SmE-eYFP **(B)** were treated with heat shock (2 hours, 41°C).

Z-stacks (100 images, 100-nm spacing) were recorded with a custom-built TILL Photonics iMIC microscope equipped with a 100×, 1.4 numerical aperture objective (Olympus, Tokyo, Japan) and a sensicam qe CCD camera (PCO, Kehlheim, Germany) and deconvolved using Huygens Essential software (SVI, Hilversum, The Netherlands). For each cell line, one representative fluorescent cell is shown as a Z-stack projection (method sum slices). In addition, the nucleus of a deconvolved single plane image is shown enlarged.