

SUPPLEMENTAL INFORMATION

Belonging to the manuscript:

**SepG coordinates sporulation-specific cell division and nucleoid organization
in *Streptomyces coelicolor***

Le Zhang*, Joost Willemse*, Dennis Claessen and Gilles P. van Wezel[#]

Molecular Biotechnology, Institute of Biology, Leiden University, PO Box 9505,
2300RA, Leiden, The Netherlands

* These authors contributed equally to this work

[#]To whom correspondence should be addressed. Tel: +3171527430; email:
g.wezel@biology.leidenuniv.nl

Table S1. Bacterial strains.

Bacterial strains	Genotype	Reference
<i>E. coli</i> JM109	See reference	[1]
<i>E. coli</i> ET12567	See reference	[2]
M145	<i>S. coelicolor</i> A3(2) SCP1- SCP2-	[3]
K202	M145 + KF41	[4]
GAL1	M145Δy/lmG	This work
GAL2	M145 + pGWS755	This work
GAL3	GAL1 + KF41	This work
GAL4	GAL1 + pGWS116	This work
GAL5	GAL1 + pGWS526	This work
GAL7	GAL1 + pGWS755	This work
GSA3	M145ΔssgA(:aadA)	[5]
GSB1	M145ΔssgB(:aac(3)IV)	[6]
GAL93	GAL1 + pGWS771	This work
GAL94	M145 + pGWS791 + pGWS116	This work
GAL95	M145 + pGWS791 + pGWS526	This work
GAL96	M145 + pGWS755 + pGWS529	This work
GAL102	M145 + pGWS526 + pGWS527	This work
GAL103	M145 + pGWS755 + GAPP-mCherry	This work

Table S2. Plasmids and constructs.

Plasmid and constructs	Description	Reference
pGWS116	pHJL401 harbouring SsgA-eGFP	[7]
pGWS526	pHJL401 harbouring SsgB-eGFP	[8]
pGWS527	pSET152 harbouring FtsZ-mCherry behind the native <i>ftsZ</i> promoter	This work
pGWS529	pHJL401 expressing FtsZ-mCherry from the <i>S. coelicolor ftsZ</i> promoter.	This work
pGWS725	pWHM3 with XbaI site removed from the multiple cloning site.	This work
pGWS731	pWHM3 containing around 1.5 kb flanking regions of <i>S. coelicolor SCO2078</i> with apra- <i>loxP</i> inserted in between.	This work
pGWS755	pSET152 harbouring the <i>yImG-egfp</i> under the control of the <i>S. coelicolor ftsZ</i> promoter region	This work
pGWS756	pHJL401 harbouring the <i>yImG-egfp</i> under the control of the <i>S. coelicolor ftsZ</i> promoter region	This work
pGWS771	pSET152 harbouring the +1/+294 region of <i>yImG</i> under the control of the <i>ftsZ</i> promoter region of <i>S. coelicolor</i> .	This work
pGWS791	pSET152 with <i>yImG-mCherry</i> under the control of the <i>ftsZ</i> promoter region of <i>S. coelicolor</i>	This work
pGWS793	pSET152 harbouring <i>sepG-blaM</i> under the control of the <i>S. coelicolor ftsZ</i> promoter region	This work
pGWS794	As pGWS794 but expressing the N-terminal part of SepG fused to BlaM	This work
pGWS795	As pGWS794 but expressing the C-terminal part of SepG fused to BlaM	This work
pGWS796	As pGWS794 but expressing full-length BlaM	This work
pGWS797	As pGWS794 but expressing mature BlaM (no signal peptide)	This work
pHJL401	<i>E. coli/Streptomyces</i> shuttle vector, low copy (<5 copies per chromosome) in <i>Streptomyces</i> and high copy (>100 copies per chromosome) in <i>E. coli</i>	[9]
pKF41	Integrative construct based on pSET152 expressing <i>ftsZ-egfp</i> from the <i>S. coelicolor ftsZ</i> promoter region	[4]
pKT25	BACTH plasmid harboring the T18 fragment of	[10]

	the <i>Bordetella pertussis cya</i> gene.	
pSET152	<i>E. coli/Streptomyces</i> shuttle vector, around 100 copies per chromosome in <i>E. coli</i> and integrative in the φC31 attachment site in <i>Streptomyces</i>	[11]
pUWL-Cre	Plasmid harboring Cre-recombinase	[12]
pWHM3	<i>E. coli/Streptomyces</i> shuttle vector, around 100 copies per chromosome in both <i>E. coli</i> and <i>Streptomyces</i> .	[13]

Table S3. Oligonucleotides.

Name	5'-3' sequence [#]
yImG_LF-1442	GTC <u>GAATT</u> C CGGGACGATGTGCGCATTCTCTCG
yImG_LR+6	GTCAGAAGTTATCCATCAC <u>CT</u> TAGA GCTCATGACCTGTGCTCCCTCTC
yImG_RF+277	GTCAGAAGTTATCGCGCAT <u>CT</u> TAGA CAGCTGTGAGCGATGAGAGAGATAC
yImG_RR+1521	GTC <u>AGGATC</u> CTCGATCAGGAACCCGCGCATCGGC
yImG_F+1	GTCAGAATT <u>AGG</u> C TTGACATGAGCGTGGCTGGATGTC
yImG_R+282	GTCAAAGCTT <u>GGATCC</u> AGCTGGCTCACGATCGAGAT
yImG_R+294	GCT <u>AGGATC</u> CTCATCGCTCACAGCTGGCT
BlaM_F_EB	ctgaGAATT <u>GGATCC</u> GGCGGTTCCACATGCTATAAGTCCAGCCCATTACGCCACCCAGAAACGCTGGTAAA
BlaM_R_XH	ctgaAAGCTT <u>TCTAGA</u> TTACCAATGCTTAATCAGTGAGGC
YImG_R+120	ctgaAAGCTT <u>GGATCC</u> GCCTTGCCGGTTGC
YImG_F+121	ctgaGAATT <u>AGG</u> C TTGACATGGTGGCTGGAGGCCA
BlaM_F+1_ES	ctgaGAATT <u>AGG</u> C TTGACATGAGTATTCAACATTCCGT
BlaM_F+70_ES	ctgaGAATT <u>AGG</u> C TTGACAT <u>G</u> CACCCAGAAACGCTGGTAAA

[#] Restriction sites used for cloning are underlined and in bold face. GGATCC, BamHI; GAATT, EcoRI; AGGCCT, StuI; TCTAGA, XbaI; AAGCTT, HindIII. Sequence shadowed in grey encoded the 12 amino acids linker between SepG and BlaM or FtsZ and BlaM

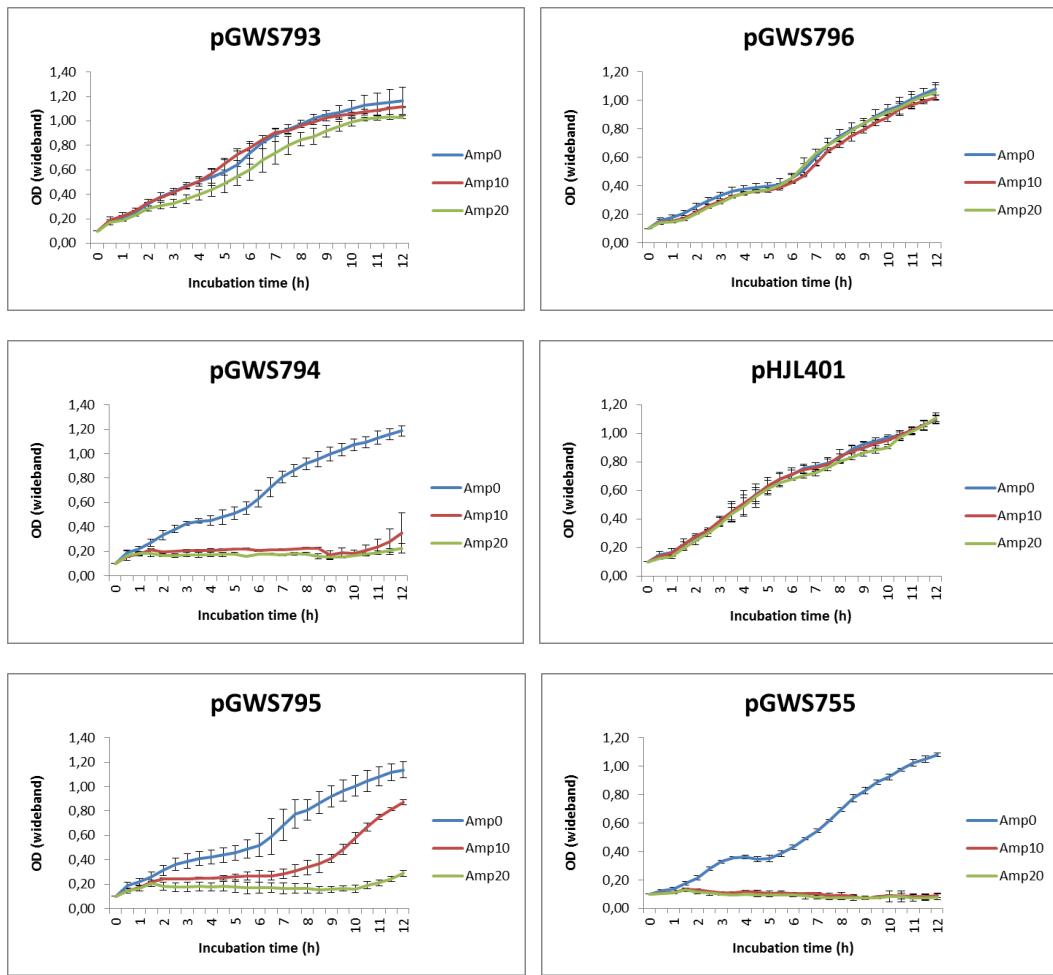


Figure S1. Growth curves of *E. coli* JM109 carrying plasmids expressing different BlaM fusions. The graphs belong to Figure 8 of the manuscript. The BlaM fusions were: full-length SepG-BlaM (pGWS793); N-terminal SepG₁₋₄₀-BlaM (pGWS794; first TM domain of SepG); C-terminal SepG₄₁₋₉₄-BlaM (pGWS795; second TM domain of SepG); BlaM with signal sequence expressed from the ftsZ promoter(s) (pGWS796; positive control); BlaM with signal sequence expressed from its native promoter (pHJL401; positive control); and pGWS755 expressing SepG-eGFP (pGWS755; negative control). Transformants were grown in presence of 0, 10 or 20 µg/ml ampicillin. Growth curve for each strain was measured every 30 minutes for 12 h at 37°C. Cultures were obtained for three independent transformants (and each of these in triplicate) using a Bioscreen C microtitre system with automated biomass reading.

REFERENCES TO SUPPLEMENTAL DATA

1. Sambrook J, Fritsch E, Maniatis T. 1989 *Molecular cloning: a laboratory manual.*: New York: Cold spring harbor laboratory press.
2. MacNeil DJ, Gewain KM, Ruby CL, Dezeny G, Gibbons PH, Maeneil T. 1992 Analysis of *Streptomyces avermitilis* genes required for avermectin biosynthesis utilizing a novel integration vector. *Gene*. **111**, 61-68.
3. Kieser T, Bibb MJ, Buttner MJ, Chater KF, Hopwood DA. 2000 *Practical Streptomyces genetics.*: John Innes Foundation.
4. Grantcharova N, Lustig U, Flärdh K. 2005 Dynamics of FtsZ assembly during sporulation in *Streptomyces coelicolor* A3(2). *Journal of bacteriology*. **187**, 3227-3237.
5. van Wezel GP, van der Meulen J, Kawamoto S, Luiten RG, Koerten HK, Kraal B. 2000 SsgA is essential for sporulation of *Streptomyces coelicolor* A3(2) and affects hyphal development by stimulating septum formation. *Journal of bacteriology*. **182**, 5653-5662.
6. Keijser BJF, Noens EE, Kraal B, Koerten HK, van Wezel GP. 2003 The *Streptomyces coelicolor ssgB* gene is required for early stages of sporulation. *FEMS Microbiol. Lett.* **225**, 59-67.
7. Noens EE, Mersinias V, Willemse J, Traag BA, Laing E, Chater KF, Smith CP, Koerten HK, van Wezel GP. 2007 Loss of the controlled localization of growth stage-specific cell-wall synthesis pleiotropically affects developmental gene expression in an *ssgA* mutant of *Streptomyces coelicolor*. *Mol. Microbiol*. **64**, 1244-1259.
8. Willemse J, Borst JW, de Waal E, Bisseling T, van Wezel GP. 2011 Positive control of cell division: FtsZ is recruited by SsgB during sporulation of *Streptomyces*. *Genes Dev.* **25**, 89-99.
9. Larson JL, Hershberger CL. 1986 The minimal replicon of a streptomycete plasmid produces an ultrahigh level of plasmid DNA. *Plasmid*. **15**, 199-209.
10. Karimova G, Pidoux J, Ullmann A, Ladant D. 1998 A bacterial two-hybrid system based on a reconstituted signal transduction pathway. *Proc. Natl. Acad. Sci. U. S. A.* **95**, 5752-5756.
11. Bierman M, Logan R, O'Brien K, Seno ET, Nagaraja Rao R, Schoner BE. 1992 Plasmid cloning vectors for the conjugal transfer of DNA from *Escherichia coli* to *Streptomyces* spp. *Gene*. **116**, 43-49.
12. Fedoryshyn M, Welle E, Bechthold A, Luzhetsky A. 2008 Functional expression of the Cre recombinase in actinomycetes. *Appl. Microbiol. Biotechnol.* **78**, 1065-1070.
13. Vara J, Lewandowska-Skarbek M, Wang YG, Donadio S, Hutchinson CR. 1989 Cloning of genes governing the deoxysugar portion of the erythromycin biosynthesis pathway in *Saccharopolyspora erythraea* (*Streptomyces erythreus*). *J. Bacteriol.* **171**, 5872-5881.