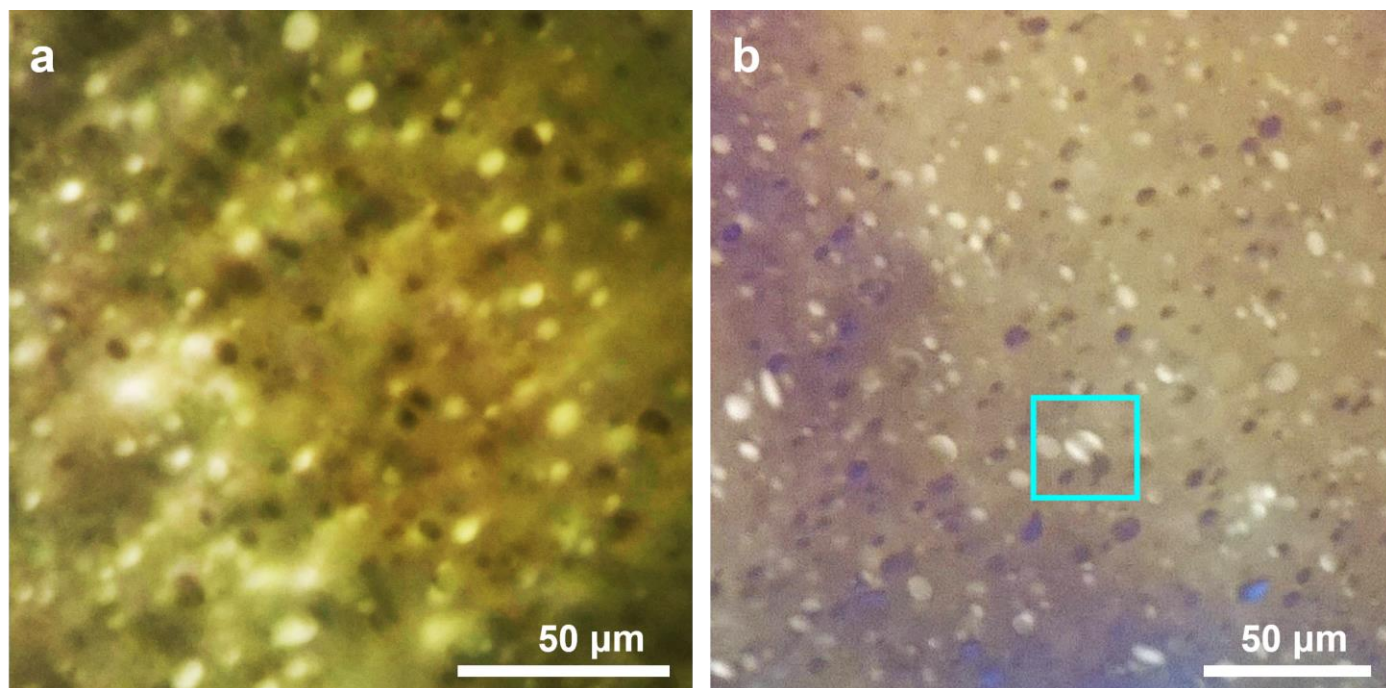
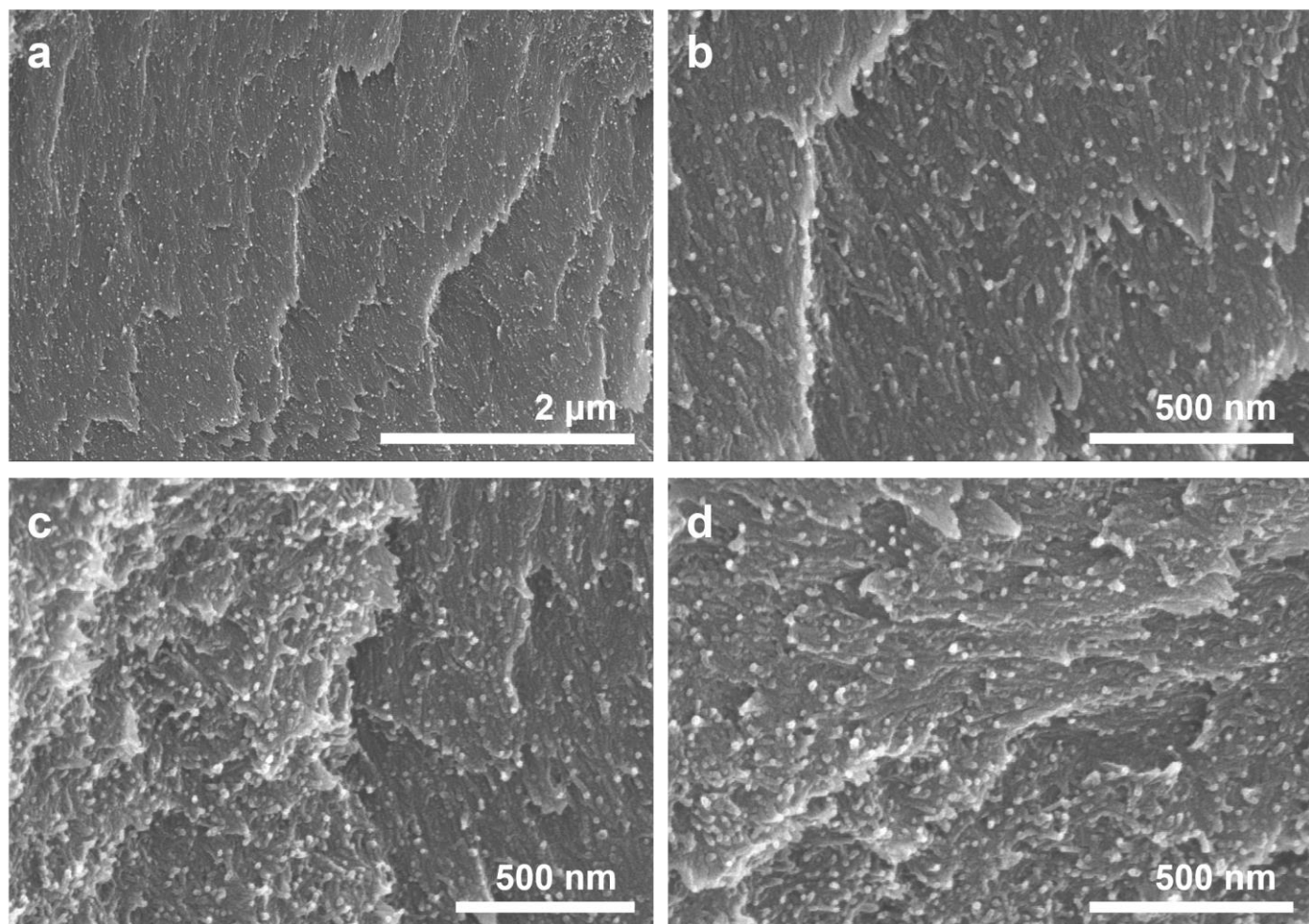


Supplementary Materials

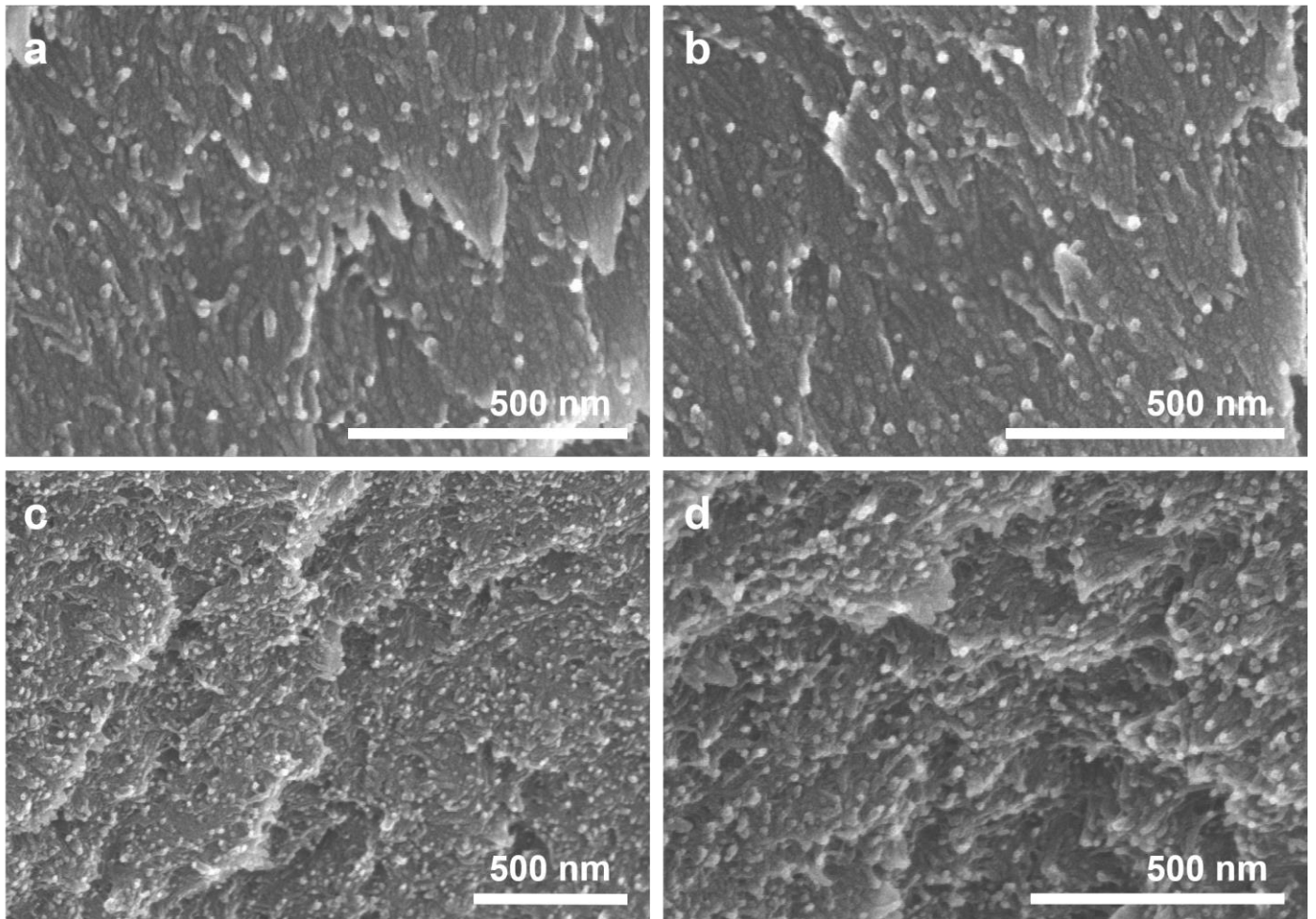
Supplementary Figures



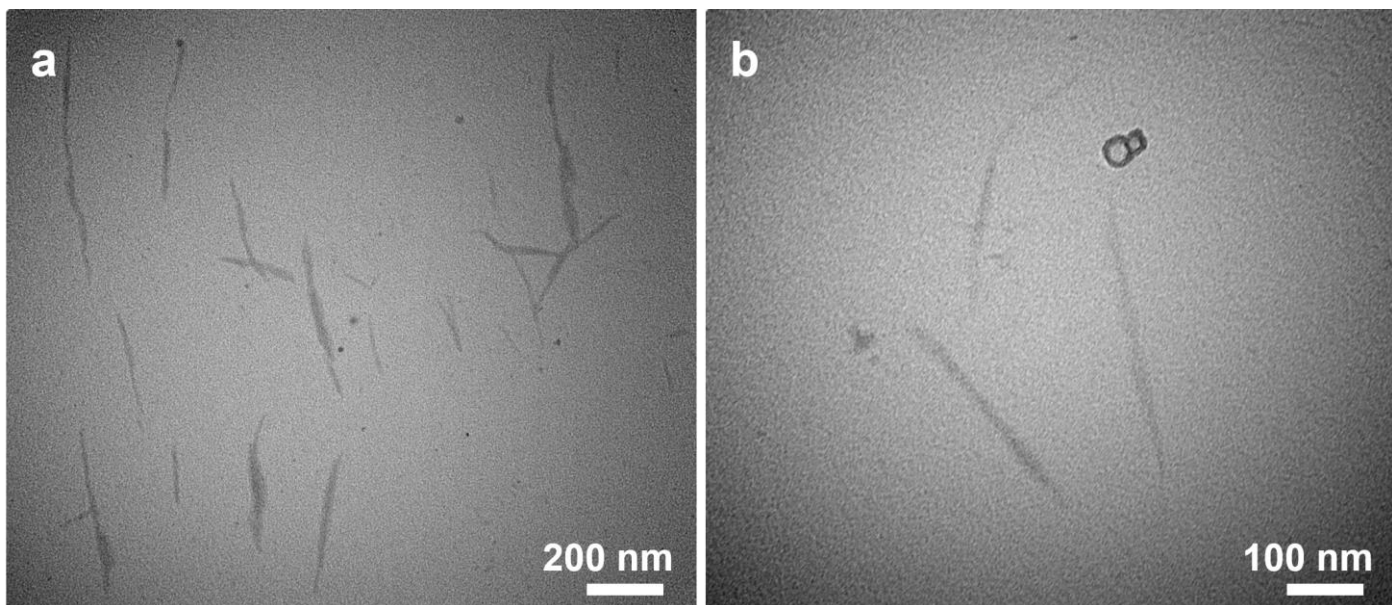
Supplementary Figure 1. POM images for newborn CNC tactoids. (a) POM image of newborn “baby” tactoids in 4 wt% CNC suspension after sonication. (b) POM image showing a tactoid with one birefringent band, which is indicated by a rectangle.



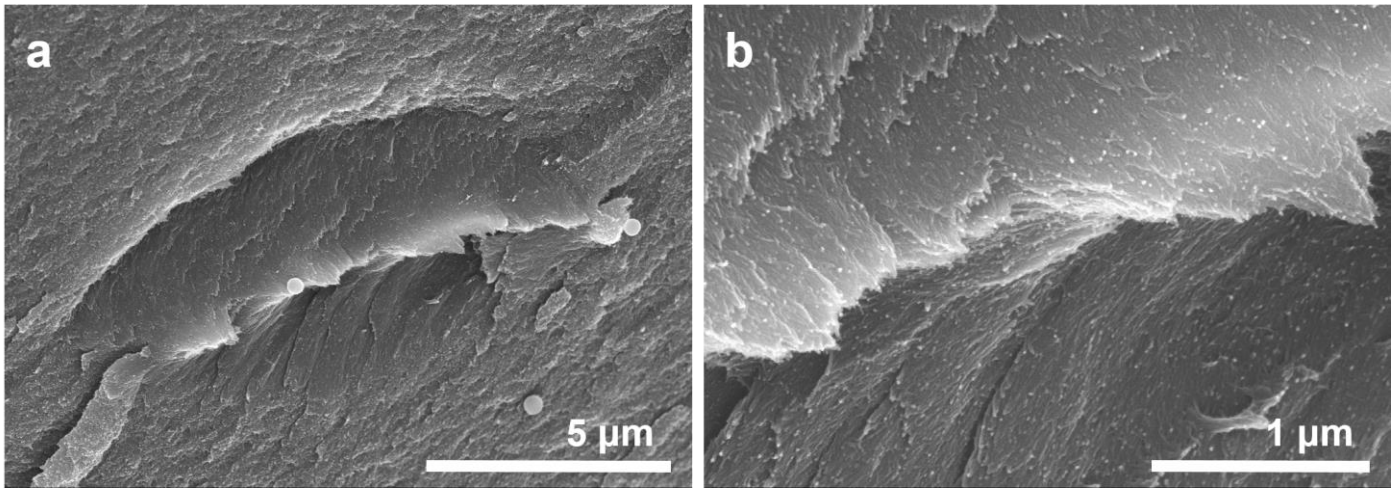
Supplementary Figure 2. Additional SEM images for the tactoid in Figure 2. (a)-(b) SEM images showing the nematic arrangements of CNCs inside this "baby" tactoid. (c) Magnified view for its left edge. (d) Magnified view for its bottom edge.



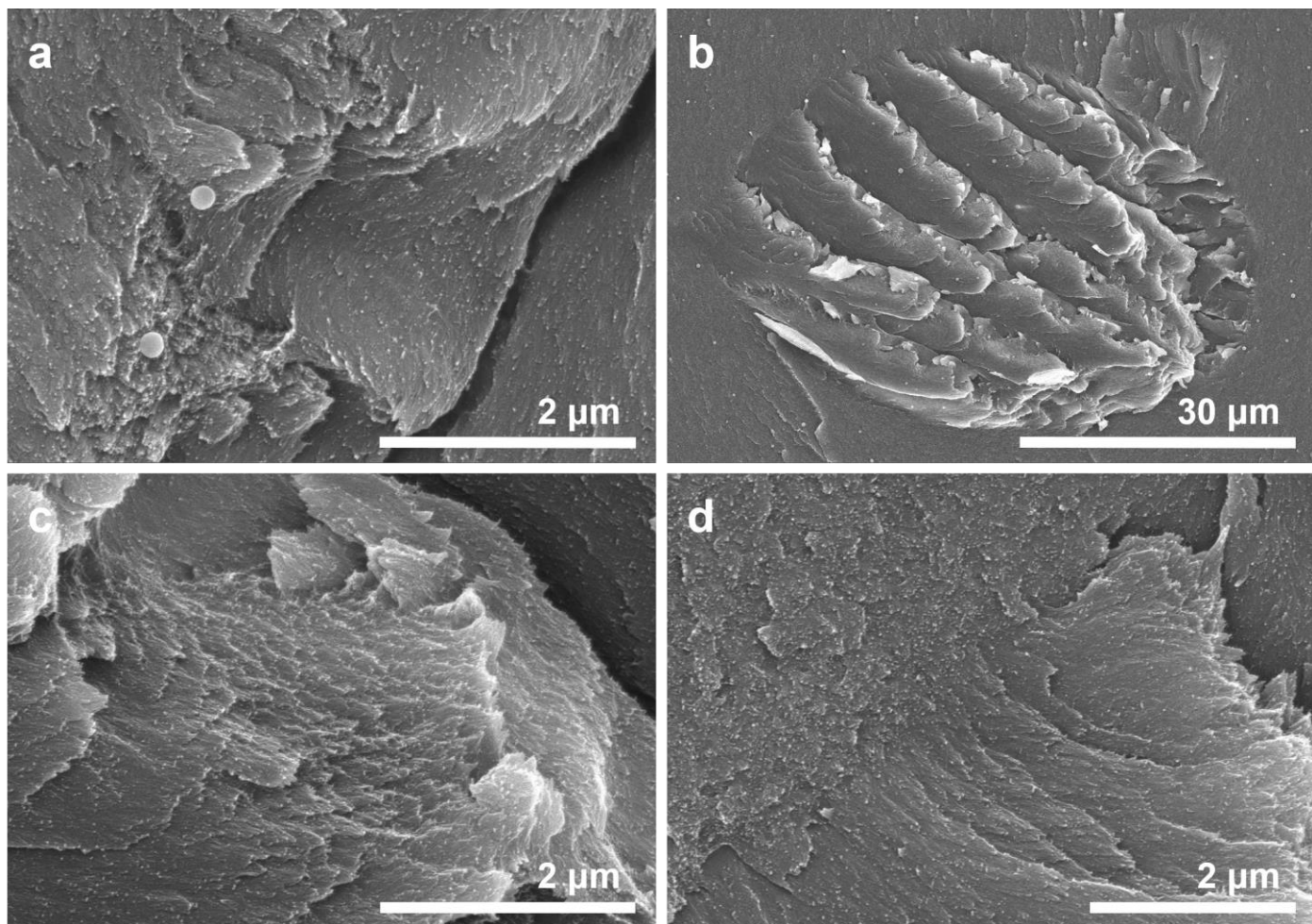
Supplementary Figure 3. Inside and outside the tactoid in Figure 2. (a)-(b) SEM images showing the parallel arranged CNCs inside the tactoid. (c)-(d) The randomly arranged CNCs in nearby isotropic regions outside the tactoid.



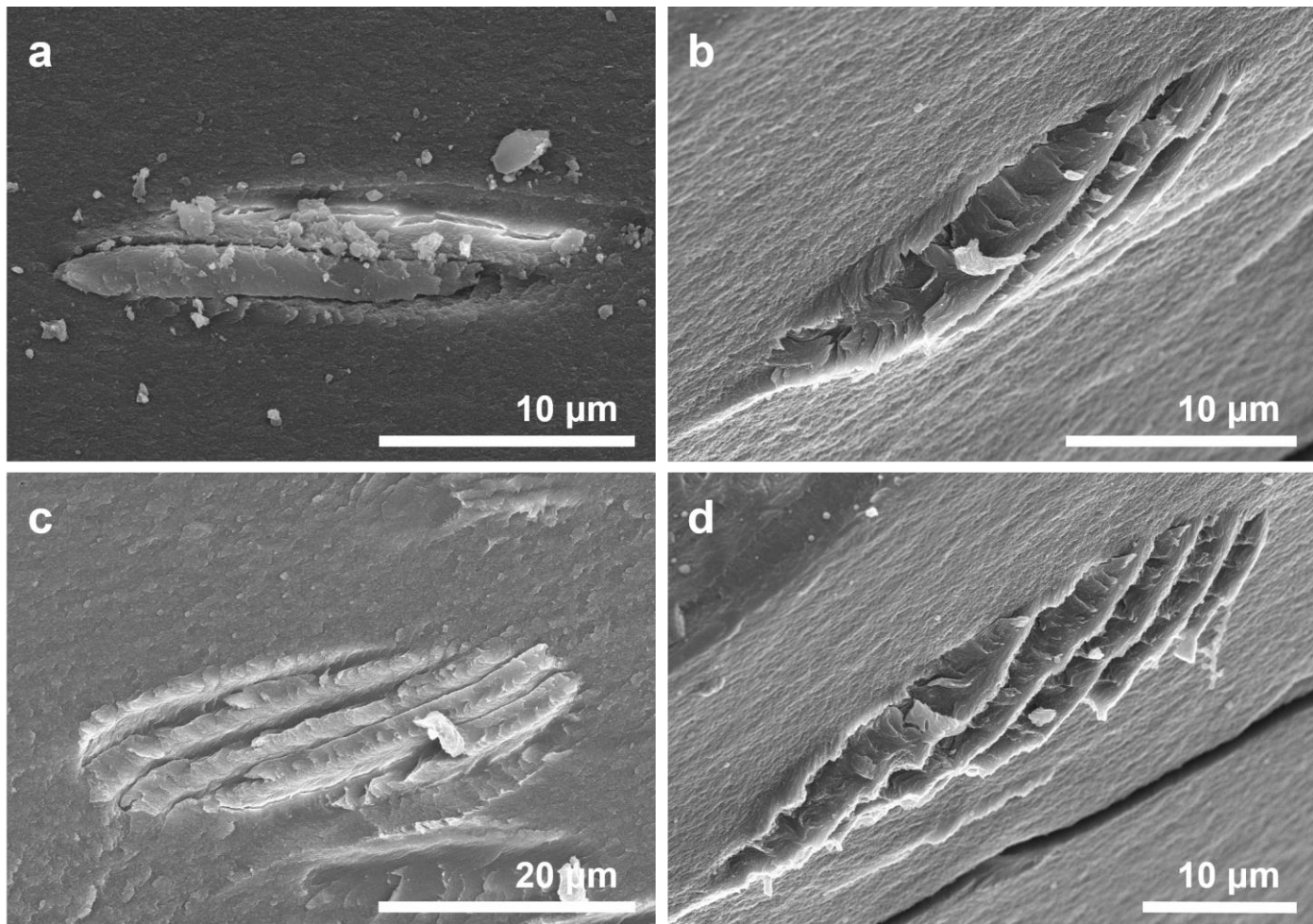
Supplementary Figure 4. TEM images of cellulose nanocrystals used in these studies. (a) Cellulose nanocrystals dispersed from water. **(b)** Cellulose nanocrystals dispersed from water, shown at higher magnification.



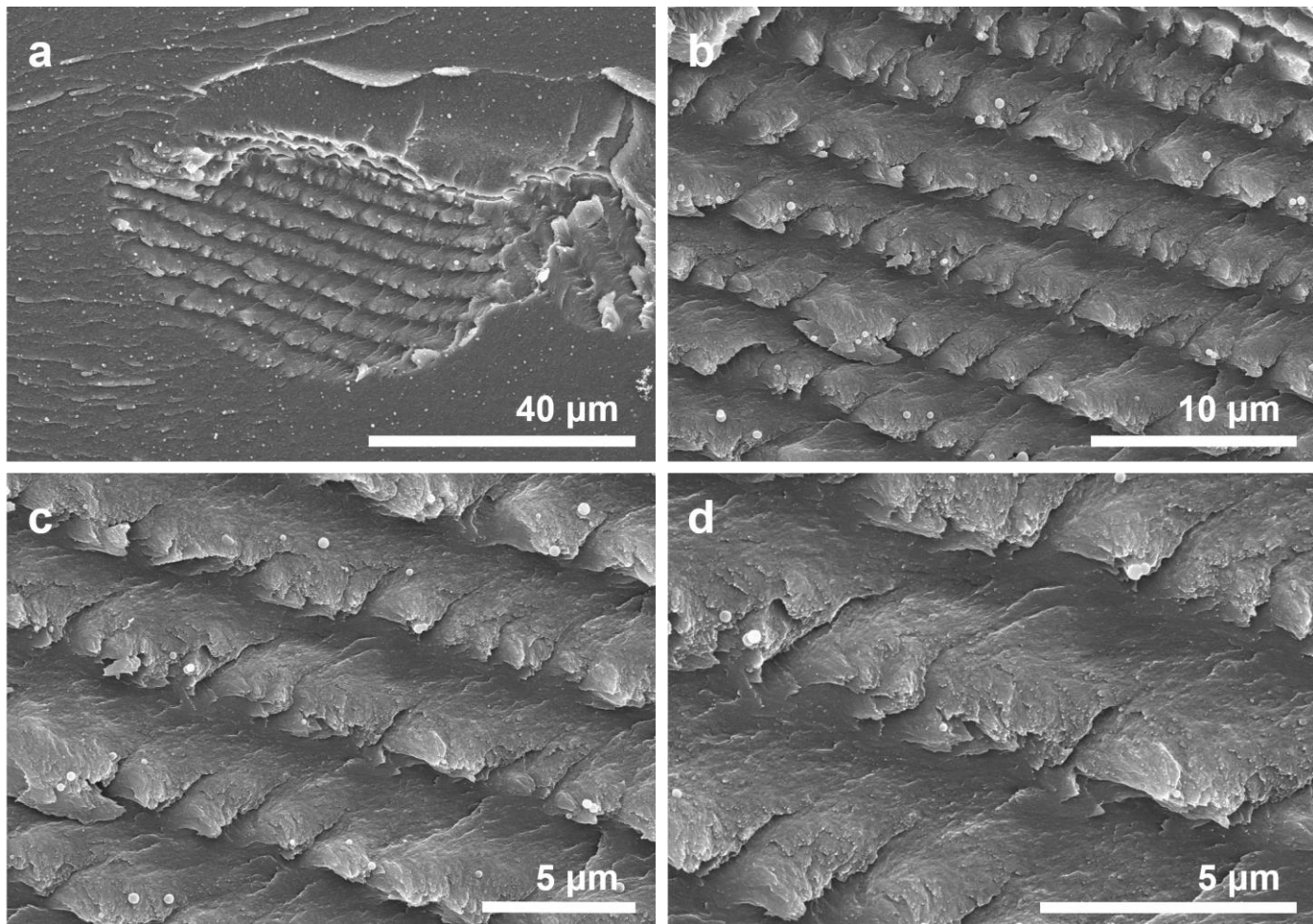
Supplementary Figure 5. A tactoid with one band. (a) SEM image of a tactoid with only one periodic band. (b) Expanded view for its band.



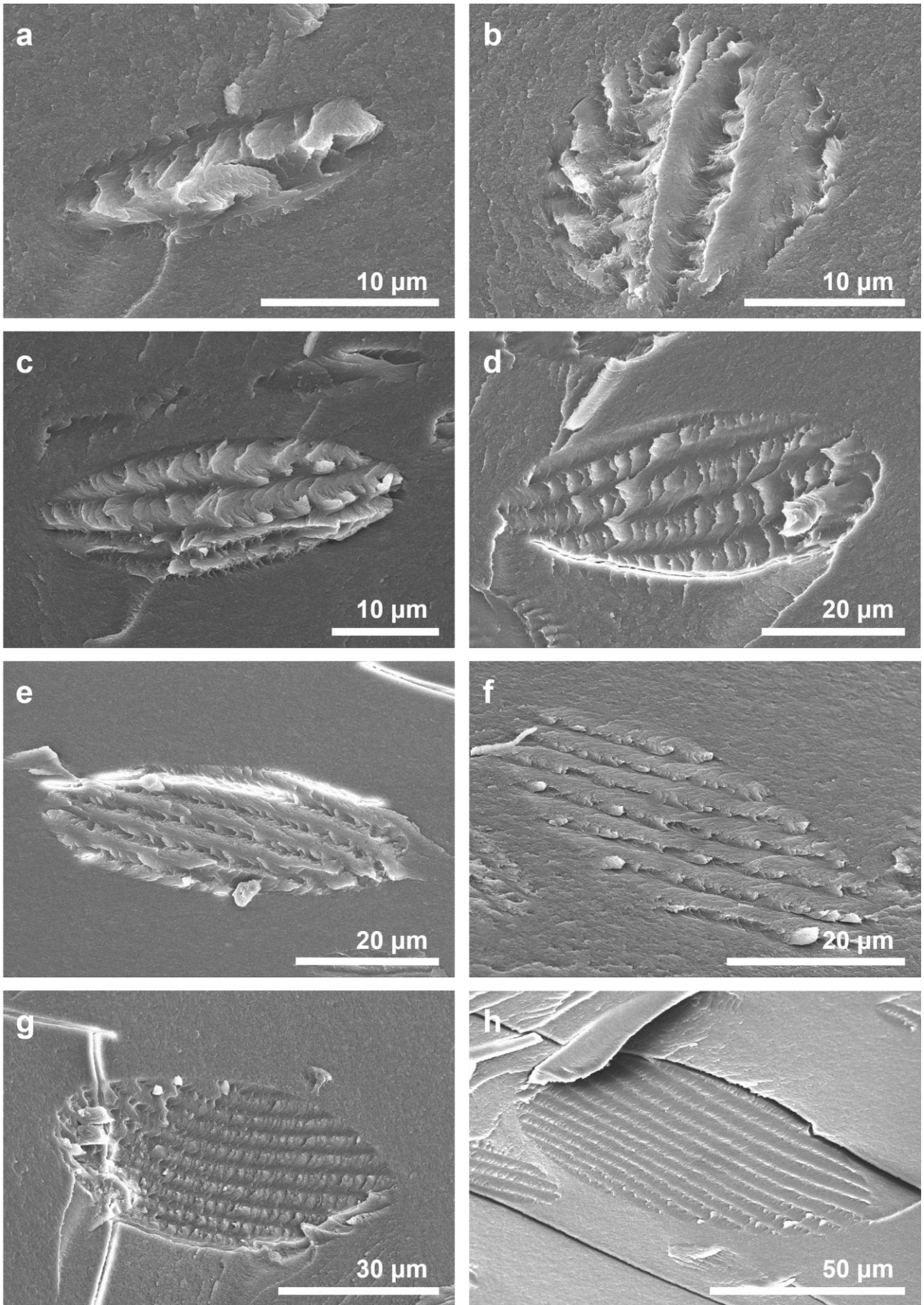
Supplementary Figure 6. Stepped geometry of the periodic bands. (a) SEM image of the same tactoid in **Figure 3c**, a cross-section of one of its periodic bands is shown here. SEM image (b) shows a similar tactoid with 6 periodic bands, a cross-section of which is shown in (c), while an edge of this tactoid is shown in (d).



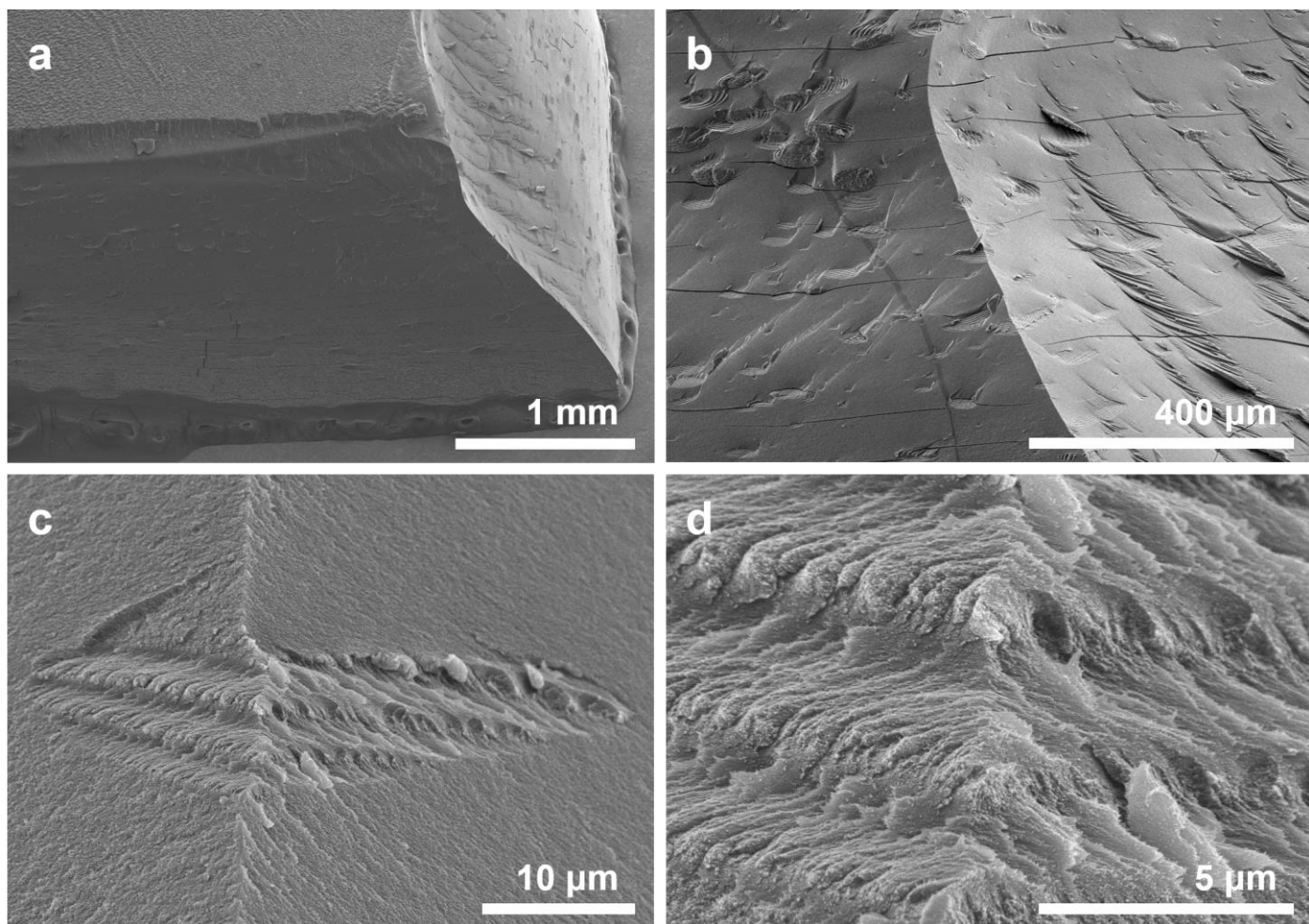
Supplementary Figure 7. SEM images of tactoids with thick sheets. Each of these sheets contains a half helical pitch.



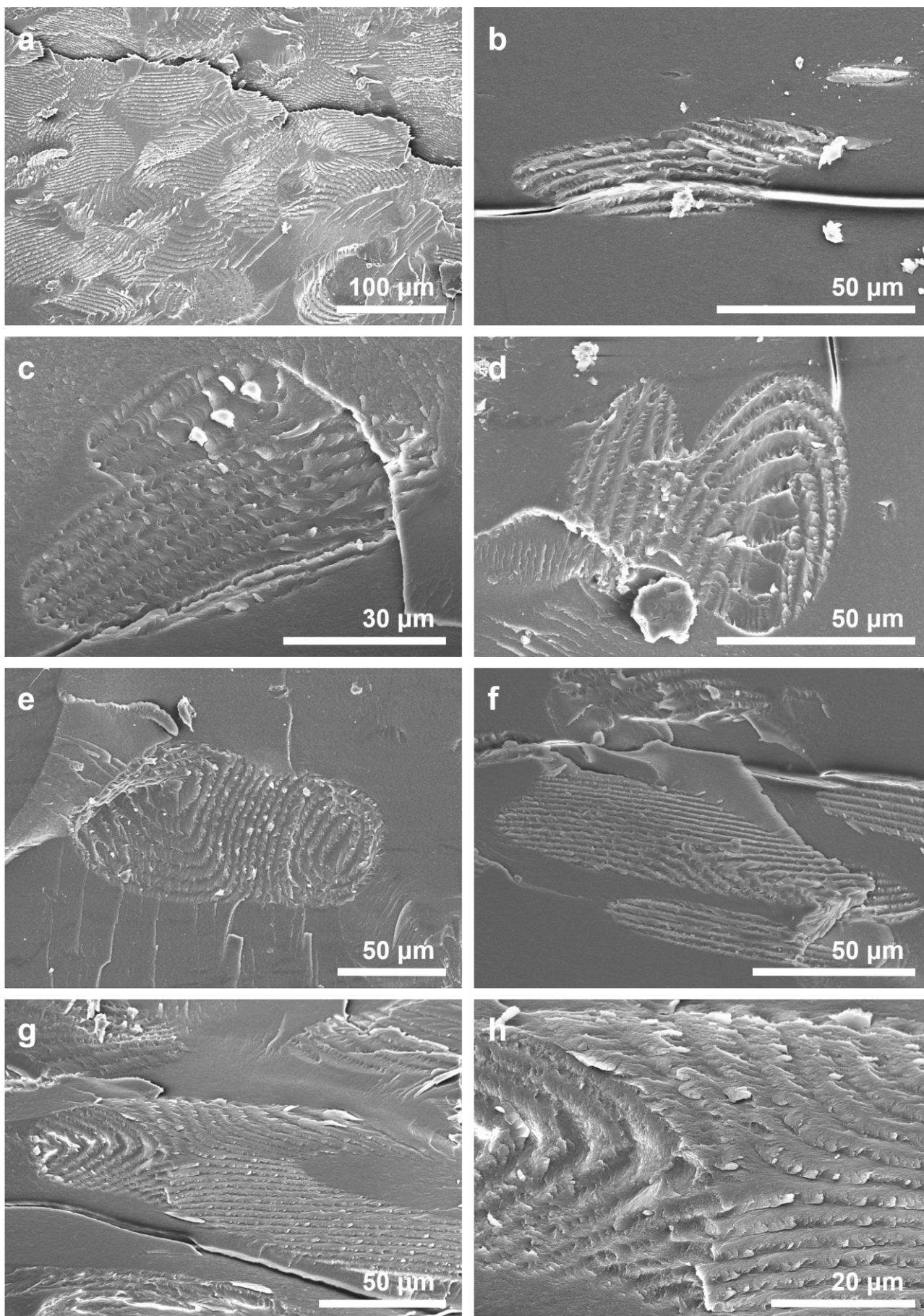
Supplementary Figure 8. Additional SEM images of the tactoid in Figure 3e.



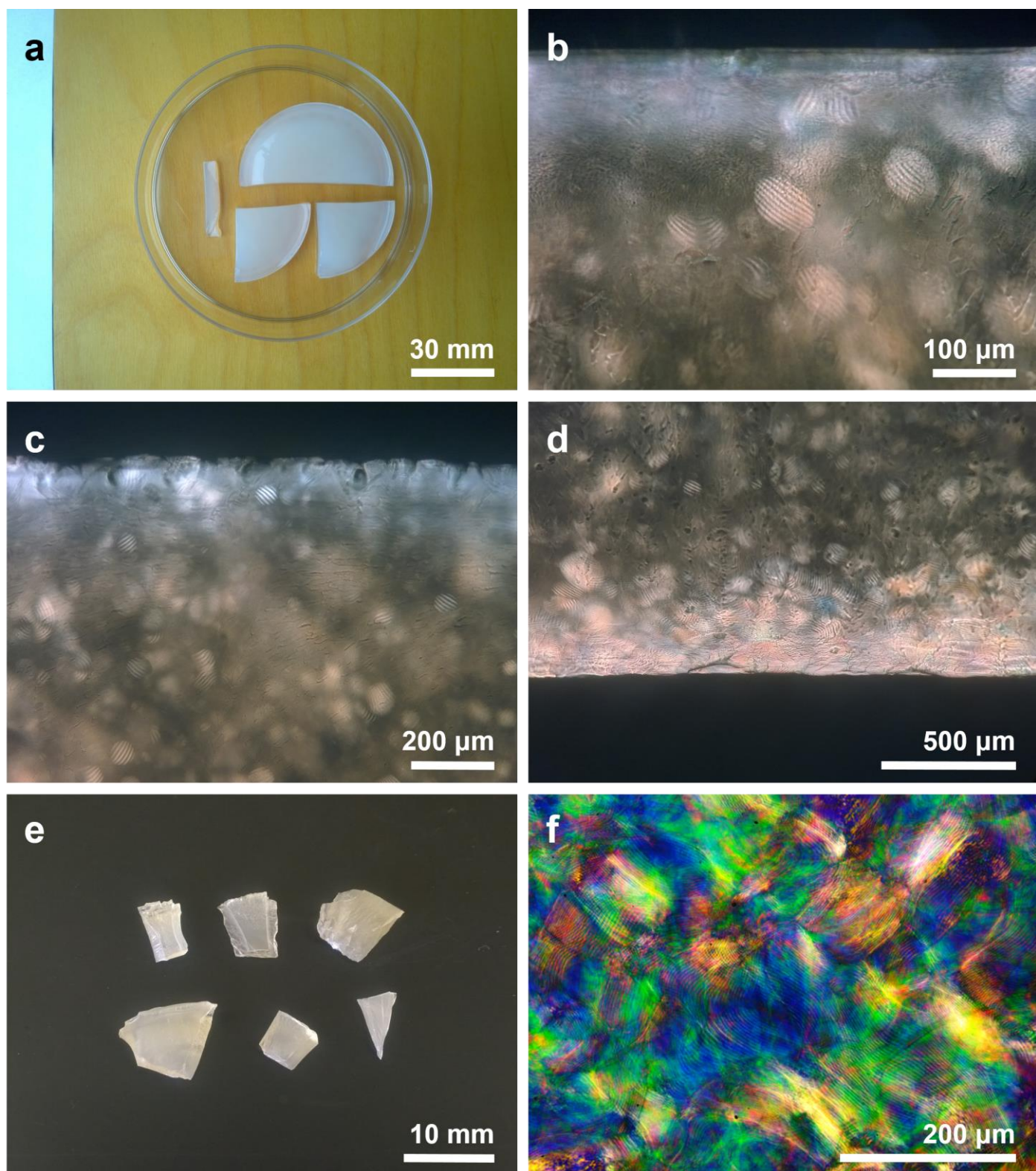
Supplementary Figure 9. Tactoids with different numbers of bands. SEM images of tactoids with (a) 1, (b) 2, (c) 3, (d) 4, (e) 5, (f) 7, (g) 9, and (h) 12 periodic bands.



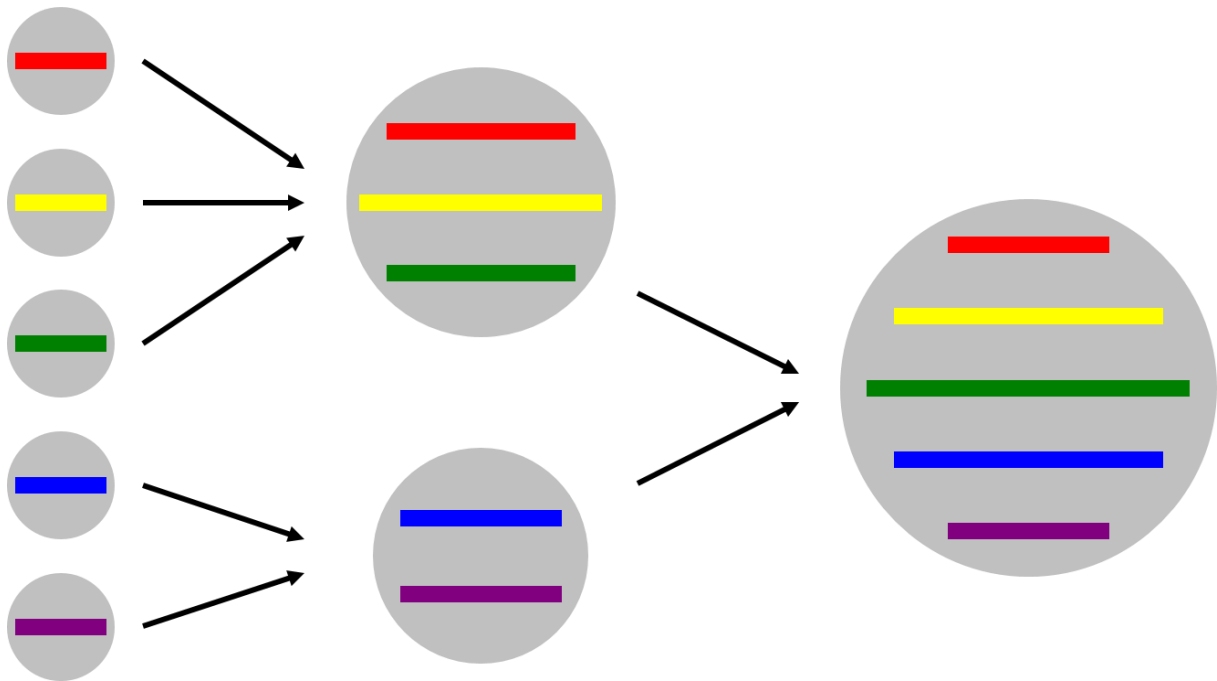
Supplementary Figure 10. Tactoids at a right angled edge. (a) SEM image for a piece of dried gel with a right-angled cross-section, (b) shows a tactoid in the center sitting at this cross-section, which is the same one as that in **Figure 4** and **Figure 5**. (c) is another tactoid at the same right-angled cross-section; its chiral nematic structure is shown with a higher magnification in (d).



Supplementary Figure 11. Fusion of tactoids. (a) SEM image showing the aggregation of tactoids in a region near the bottom of a dried gel. (b)-(d) The collisions between tactoids. (e)-(g) Hybrid tactoids formed by fusions. (h) shows a magnified view of the fusion defect in the hybrid tactoid shown in (g).



Supplementary Figure 12. Additional images for this work. (a) shows pieces of a freshly sliced hydrogel. (b)-(d) are POM images for the section in (a), which show tactoids captured in the hydrogel, and the aggregation of tactoids at the bottom of the CNC suspension. (e) shows some pieces of dried gels obtained with a hammer. (f) is a POM image showing the fingerprints of a dried CNC film.



Supplementary Figure 13. Fusion process of tactoid. A cartoon depiction for an idealized fusion process from five one-band small tactoids to one five-band big tactoid. We hypothesize that different periodic bands in a big tactoid would be collected from different small tactoids through fusion processes.

Supplementary Table 1. Mass decrease and concentration increase of a CNC-PAAm precursor mixture during evaporation.^a

Time	0 h	0.5	1 h	2 h	3 h	4 h	5 h
Mass (g)	10.7397	10.6523	10.5824	10.4382	10.2980	10.1507	9.9993
Conc. (wt.%)	4.00	4.04	4.07	4.13	4.19	4.26	4.33
6 h	7 h 5 min	8 h	9 h	10 h	10 h 40 min	11 h 30 min	12 h
9.8501	9.6832	9.5496	9.3997	9.2383	9.1345	8.9937	8.9137
4.41	4.49	4.56	4.64	4.74	4.80	4.88	4.93

^a In order to determine the approximate concentration of CNCs during the evaporation, we undertook an experiment identical to the ones reported in the paper, weighing the solution as a function of time. Initially, a 4 wt.% CNC suspension (9.9184 g) was mixed with 1.00 g acrylamide, 100 mg cross-linker and 5 mg photo-initiator in a vial. 10.7397 g of this mixture (97.43% of 11.0234 g) was transferred into a 60 mm diameter Petri dish; this mixture contains 0.3865 g CNCs and 9.2766 g water in theory. Based on this information, the concentrations of CNCs at different time points are calculated. The mass of this CNC-PAAm precursor mixture decreased linearly with time, $m = 10.744 - 0.1508t$, $R^2 = 0.9997$ (m in grams, t in hours). The wt% CNCs are based on the wt% CNCs per unit water, neglecting the added PAAm, cross-linker, and initiator, and we assumed that only water evaporates from the mixture over this time period. Also, the experiment was performed on a different day than the ones for the images, but we believe that the humidity was similar (~70%) and the approximate concentrations are correct.

Supplementary Table 2. Average spacing between adjacent periodic bands in 29 CNC tactoids having different numbers of bands.^a

Bands	2	3	3	3	3	4	4	4	4
Spacing (μm)	6.020	4.805	3.664	3.222	3.298	5.405	5.668	2.579	5.122
4	4	4	5	5	6	6	6	6	7
4.199	2.605	2.498	3.590	2.687	2.881	3.729	3.991	2.934	2.322
7	7	7	8	8	9	9	9	10	12
4.750	3.757	2.371	1.966	3.579	3.991	3.071	2.372	1.978	2.871

^a Distances were determined by measuring the total edge-to-edge distance (SEM) and dividing by the number of bands. The spacing between adjacent bands corresponds to one half helical pitch.

Supplementary Methods

Acrylamide (Aldrich, 98%), N,N'-methylenebisacrylamide (Aldrich, 99%), 2,2-diethoxyacetophenone (Acros, 98%), and 2-hydroxy-4'-(2-hydroxyethoxy)-2-methylpropiophenone (Aldrich, 98%) were used without further purification. CNC suspensions (4 wt.% in water, $\sigma = 2.19 \text{ mS}\cdot\text{cm}^{-1}$, pH = 2.44) were provided by CelluForce. The CNCs were obtained from hydrolysis of wood pulp in sulfuric acid using the literature procedure.¹ TEM of these crystals gave dimensions of $19\pm 9 \text{ nm}$ by $245\pm 135 \text{ nm}$ (based on measuring 98 crystallites).

Optical microscopy was performed with an Olympus BX41 microscope. Scanning electron microscopy (SEM) experiments were conducted on a Hitachi S4700 electron microscope at an acceleration voltage of 10 kV. Samples were sputter-coated with 8 nm of platinum-palladium (80:20) alloy before imaging. Transmission electron microscopy (TEM) images were obtained with a Hitachi H7600 electron microscope at an acceleration voltage of 62 kV. Samples for TEM measurements were prepared by diluting an aqueous suspension of CNCs to $\sim 0.002 \text{ wt.}\%$, sonicating the dilute solution for 3 h, then drop-casting the suspension onto carbon-coated copper grids. The thickness of hydrogels and dried gels was measured with a MARATHON Electronic Digital Micrometer. The diameter of dried gels was measured with a ruler. A Durasonix 3 Litre Ultrasonic Cleaner (Power: 120 W; Frequency: 40 kHz) was used in this work for sonication treatments of CNC suspensions.

Supplementary Reference

[1] Hamad, W. Y. & Hu, T. Q. Structure–process–yield interrelations in nanocrystalline cellulose extraction. *Can. J. Chem. Eng.* **88**, 392–402 (2010).