

Tumour-specific metabolic adaptation to acidosis is coupled to epigenetic stability in osteosarcoma cells

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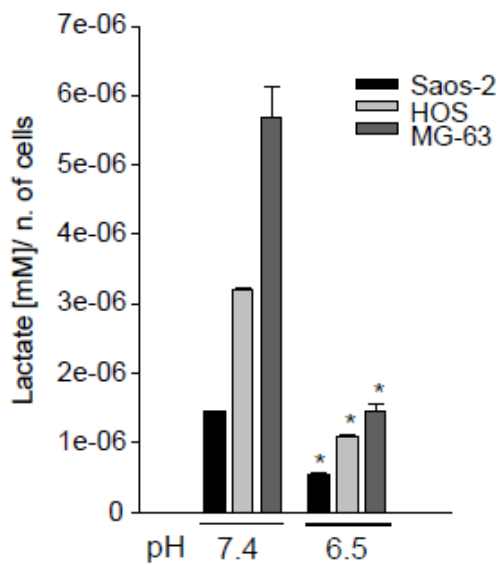
Supplementary Table S1: Heat map that includes the intracellular metabolites related to the identified pathways involved in the response to acute acidosis.

Compound name	m/z	MT	Standardized Relative Area									
			Acidic pH					Neutral pH				
			Sarcomas (OS)			Normals (Fb)		Sarcomas (OS)			Normals (Fb)	
HOS	MG-63	Saos-2	TIG-121	TIG-108	HOS	MG-63	Saos-2	TIG-121	TIG-108			
Dihydroxyacetone phosphate	168.991	12.79	-0.411	-0.665	-0.355	0.026	0.057	-0.022	0.149	-0.666	-0.277	2.492
Phosphocreatine	210.028	12.37	-0.782	-0.894	-0.883	0.070	0.014	-0.474	0.605	-0.968	-0.306	1.569
Glu	149.063	10.93	-0.577	-0.644	-0.169	0.094	0.352	-0.228	-0.050	-0.518	2.407	2.717
Argininosuccinic acid	291.128	9.42	-0.634	-0.587	-0.697	-0.278	-0.184	-0.297	-0.144	-0.532	1.548	2.820
ATP	505.989	11.91	-0.643	-0.680	-0.225	0.347	0.158	-0.594	-0.218	-0.915	-0.028	2.519
GTP	521.985	11.59	-0.516	-0.644	-0.599	-0.133	-0.228	-0.175	0.028	-0.578	-0.236	1.275
CTP	481.977	12.48	-0.232	0.205	-0.543	0.802	0.577	0.115	-0.386	-1.134	0.551	2.756
UTP	482.962	12.70	-0.025	-0.724	-0.813	1.735	1.078	-0.200	0.012	-0.844	0.995	1.422
UDP-glucose, UDP-galactose	565.049	8.73	-0.068	-0.515	-0.658	1.293	1.009	-0.176	-0.352	-0.725	0.951	2.970
CMP-N-acetylneuraminat	613.142	8.26	-0.612	0.215	-0.280	1.980	2.700	-0.467	-0.475	-0.713	1.264	-0.052
NAD⁺	662.103	6.66	-0.781	-1.049	0.113	1.893	1.137	-0.792	-0.822	-0.803	0.782	1.016
N⁸-Acetylspermidine	188.175	6.32	-0.391	-0.217	-0.370	2.665	2.808	-0.416	-0.394	-0.415	-0.421	-0.421
Pelargonic acid	157.123	8.21	-0.709	-0.767	0.574	2.378	1.466	-0.773	-0.733	-0.709	1.765	0.916
Decanoic acid	171.139	8.03	-0.660	-0.767	0.230	1.854	1.986	-0.830	-0.773	-0.775	1.547	1.444
Asp	134.044	11.58	-0.624	-0.501	-0.325	1.874	0.816	-0.382	-0.435	-0.671	1.770	2.444
Gly	77.042	8.30	-1.009	-0.715	0.401	0.549	1.197	-0.935	-0.642	-0.851	1.638	2.022
His	156.077	7.29	-1.055	-0.255	0.295	0.235	0.986	-0.917	-0.480	-1.121	2.092	1.584
Tyr	182.081	11.36	-1.151	-0.367	0.761	0.247	1.087	-1.033	-0.608	-1.179	1.646	1.283
Phe	166.086	11.09	-1.131	-0.418	0.818	0.423	1.376	-1.030	-0.592	-1.175	1.654	1.211
Trp	205.096	11.02	-1.134	-0.254	0.807	0.581	1.603	-1.142	-0.771	-1.277	1.505	0.930
Val	118.086	10.01	-1.020	-0.413	0.989	0.872	1.823	-1.028	-0.734	-1.208	1.231	0.999
Ile	132.102	10.20	-1.005	-0.377	0.805	0.799	1.793	-0.977	-0.669	-1.203	1.588	1.277
Leu	132.102	10.31	-1.022	-0.385	0.909	0.797	1.684	-0.991	-0.696	-1.186	1.659	1.240
Met	150.058	10.71	-1.033	-0.367	0.933	0.442	1.318	-0.955	-0.700	-1.114	1.854	1.821
Lys	147.113	6.87	-0.885	-0.202	0.781	0.440	1.488	-0.812	-0.654	-0.911	1.570	2.278
Arg	175.118	7.10	-0.837	-0.202	0.531	0.525	1.740	-0.826	-0.628	-0.905	1.680	2.259
Cys	122.027	11.32	-0.643	0.495	-0.383	0.895	1.765	-0.706	-0.690	-0.788	1.785	1.595
Pro	116.070	10.81	-0.719	-0.389	1.771	1.604	1.721	-0.875	-0.811	-0.728	1.243	0.578
Methionine sulfoxide	166.052	11.80	-0.975	-0.741	0.785	0.972	1.345	-0.993	-0.866	-0.849	2.091	1.132
Choline	104.107	6.85	-0.842	-0.786	1.135	0.528	1.272	-1.092	-0.642	-0.749	2.026	1.194
Creatinine	114.066	7.29	-0.519	-0.982	1.235	0.434	1.669	-1.100	-0.588	-1.056	0.599	1.217
Hypotaurine	110.027	18.11	-0.879	-0.953	1.260	0.477	1.170	-0.866	-0.627	-0.400	0.237	1.649
Putrescine	89.107	4.69	-0.837	-0.710	0.438	0.936	1.944	-0.431	-0.670	-0.724	2.034	1.525
myo-Inositol 2-phosphate	259.022	10.57	-0.466	-0.630	-0.080	1.525	1.042	-0.634	-0.576	-0.675	0.454	3.024
N-Acetyllysine	189.122	9.78	-0.956	-0.188	0.006	1.116	1.209	-0.753	-0.163	-0.891	1.086	2.655
Diethanolamine	106.086	7.64	-0.387	-0.524	0.181	0.421	2.020	-0.690	-0.756	-0.813	2.461	1.168
Glycerol	93.055	22.14	-0.490	-0.580	0.125	0.520	2.584	-0.606	-0.679	-0.795	1.501	1.752
1-Methylnicotinamide	137.071	7.35	-0.406	-0.441	-0.503	0.402	2.274	-0.455	-0.506	-0.567	1.617	2.370
XCO132	325.159	8.68	-0.518	-0.415	-0.386	1.472	2.753	-0.525	-0.477	-0.500	1.322	1.419
Lauric acid	199.170	7.76	-0.503	-0.639	0.847	1.377	1.789	-0.781	-0.718	-0.590	0.669	2.424
Carnosine	227.113	6.75	-0.708	-0.708	0.456	1.450	2.128	-0.708	-0.571	-0.435	-0.708	2.433
XCO126	310.110	14.98	-0.240	-0.336	0.561	0.860	2.811	-0.746	-0.510	-1.005	0.447	1.610
Pyridoxine	170.081	8.63	-0.414	-0.521	0.453	0.881	2.398	-0.762	-0.646	-0.900	0.890	1.374
Adenosine 5'-phosphosulfate	426.013	10.22	-0.111	-0.051	0.901	2.170	2.007	-0.962	-0.770	-0.688	1.012	-0.221
GABA	104.070	7.69	-0.501	-0.467	1.388	2.345	2.150	-0.738	-0.284	-0.198	0.154	0.692
Morpholine	88.076	6.62	-0.672	-0.561	0.038	2.168	2.005	-0.722	-0.669	-0.884	0.850	1.174
Hexanoic acid	115.076	9.02	-0.723	-0.691	0.585	2.845	0.297	-0.764	-0.715	-0.654	1.795	0.875
p-Toluic acid	135.045	9.24	-0.751	-0.751	0.514	3.087	0.904	-0.751	-0.751	-0.560	1.349	0.339
N-Acetylneuraminic acid	308.100	7.29	-0.641	-0.639	-0.158	2.575	1.623	-0.639	-0.655	-0.649	1.201	-0.532
Streptomycin sulfate_+H ₂ O_divalent	300.644	6.86	-0.384	-0.372	-0.259	3.580	1.398	-0.431	-0.433	-0.441	0.210	0.184
Fumaric acid	115.004	25.85	-0.582	-0.816	-0.591	-0.245	-0.329	0.080	0.030	-0.214	-0.816	2.110
Malic acid	133.014	21.69	-0.547	-0.760	-0.610	-0.248	-0.421	0.148	0.090	-0.233	-0.494	1.490
Nicotinamide	123.055	7.40	-0.379	0.006	0.344	0.950	3.144	-0.896	-0.693	-0.950	0.854	0.510
Glycerophosphocholine	258.109	21.70	0.132	0.147	-0.283	1.602	3.099	-0.685	-0.611	-0.709	-0.596	-0.648

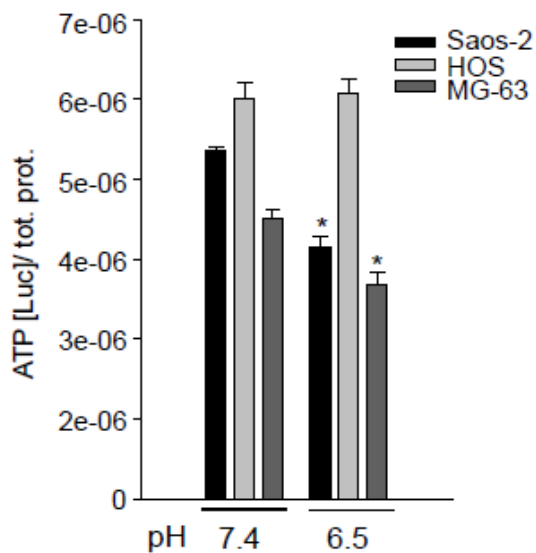
The heat map represents the fold change of metabolite concentration. Standardization of the heat map was performed as follows: the mean for each metabolite within the group of the values obtained for the different cell models (HOS, MG-63, Saos-2, TIG-121, TIG-108) in the acidic condition or in the neutral condition was calculated and considered as 0. Then, the average deviation was shown as +1 or -1 for increment (red colour) or decrease (blue colour), respectively, compared with the mean.

Supplementary Table S2: Heat map of transcripts of genes related to the metabolic pathways identified by metabolomics analysis and involved in the response to acute acidosis.

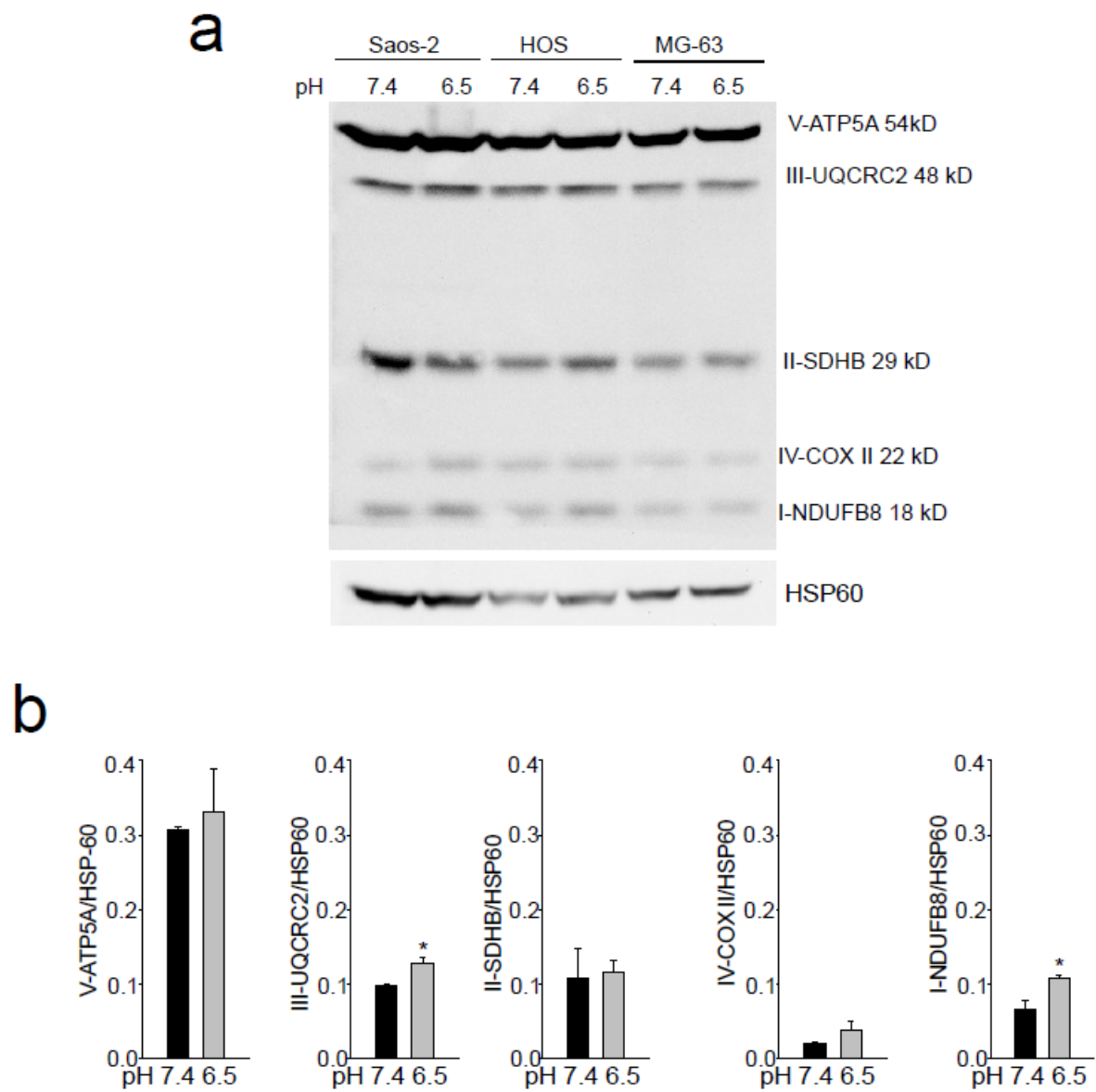
Gene Symbol	Entrez ID	Ensembl ID	Acidic pH					Neutral pH				
			Sarcomas (OS)			Normals (Fb)		Sarcomas (OS)			Normals (Fb)	
			HOS	MG-63	Saos-2	TIG-121	TIG-108	HOS	MG-63	Saos-2	TIG-121	TIG-108
Glycolysis												
HK1	3098	ENSG00000156515	12.409199	11.253214	11.562604	11.318684	11.224559	12.044444	11.522645	11.815693	12.082178	12.370876
HK2	3099	ENSG00000159399	11.342816	10.264063	11.334791	12.502488	12.189524	10.674194	10.743927	10.924035	10.725984	10.640933
HK3	3101	ENSG00000160883	0	0	0	0	0	0	1.1116893	0	0	0
PFKL	5211	ENSG00000141959	9.766287	10.308735	10.523505	10.369005	10.582753	9.62831	10.069268	10.940122	10.271774	10.717787
PFKM	5213	ENSG00000152556	10.75107	10.875085	9.812639	10.123131	9.851434	11.006119	10.538072	10.420568	10.633866	10.769643
PFKP	5214	ENSG00000067057	11.692869	11.247765	10.415174	10.299332	10.840413	11.914386	11.091313	10.782291	11.031064	11.509333
ALDOA	226	ENSG00000149925	12.826196	13.137805	12.493885	12.499638	12.47743	12.928181	12.990125	12.531553	12.566268	12.466882
ALDOB	229	ENSG00000136872	0	2.3689234	2.1955543	0.76722986	2.719281	0	4.2200866	1.6892521	1.6951414	0
ALDOC	230	ENSG00000109107	9.843989	3.3684845	9.716939	8.018367	8.4635935	8.802816	4.840587	8.633072	7.170773	6.682957
TP1	7167	ENSG00000111669	11.35943	12.36279	11.096159	11.006119	10.8421335	11.458347	11.883163	11.19116	11.548942	11.460541
GAPDH	2597	ENSG00000111640	12.845823	13.766205	12.8784485	12.662054	12.797461	13.0022955	13.440406	12.827934	12.87597	12.857994
GAPDH5	2630	ENSG00000105679	8.443409	7.7832227	6.8617463	8.138038	8.830945	7.421616	7.9533796	6.497021	6.9308434	7.2491875
PGK1	5230	ENSG00000102144	12.153183	12.7832575	12.823092	12.190403	12.019995	12.367074	12.481405	12.881366	12.703712	12.506502
PGK2	5232	ENSG00000170950	0	0	1.2442105	0.8200225	0	0	0	0	0.06469393	0
PGAM1	5223	ENSG00000171314	8.505315	9.618388	8.191748	8.633442	8.458765	6.880887	8.483391	8.056486	8.542646	8.778131
PGAM2	5224	ENSG00000164708	0	0	0	0	0	0	0	0	0	0
PGAM4	441531	ENSG00000226784	0	1.2442105	0	1.0026534	1.282275	0	1.7861416	0.11121831	1.9980683	0
PGAM5	192111	ENSG00000176894	10.943783	10.763016	10.527151	9.290789	8.598321	10.333752	10.382821	10.873635	9.653443	9.164069
ENO1	2023	ENSG00000074800	13.575606	14.060843	14.089886	12.588876	12.216739	13.6991415	14.116754	14.149594	13.367027	13.3909
ENO2	2026	ENSG00000111674	9.656764	10.425203	8.362465	9.67768	9.676269	9.597572	10.150731	8.368593	9.470452	10.866586
ENO3	2027	ENSG00000108515	6.4588904	6.6666303	4.346933	5.2690516	6.0184245	6.470815	6.4919186	3.9816904	4.917551	5.722017
ENO4	387712	ENSG00000186316	5.6089706	2.2519855	4.3200617	1.2381275	3.536486	3.9477217	4.2412043	2.342518	3.477151	4.5170555
PKLR	5313	ENSG00000143627	0	0	0.06988551	0	0	0	0	0	0	0
PKM2	5315	ENSG00000067225	13.703678	14.10248	12.863498	12.583481	13.107946	14.040522	14.127852	13.240816	13.668372	13.816241
LDHA	3939	ENSG00000134333	12.797461	12.863498	12.847786	12.75372	12.713863	13.14687	12.856271	12.805337	13.065542	13.232918
LDHAL6A	160287	ENSG00000166800	1.6134268	0	0	0.5267878	0.33389223	2.7795272	2.3645723	3.321858	3.0302413	2.9824562
LDHAL6B	92483	ENSG00000171989	1.8835616	3.312686	2.600904	1.7075523	3.377551	2.0342543	1.7235659	2.5791528	1.6340747	3.8760486
LDHB	3945	ENSG00000111716	12.810263	13.518546	12.65571	12.417956	11.99684	13.018645	13.2833805	12.720022	12.469393	12.374005
LDHC	3948	ENSG00000166796	0	0	0	0	0	0	1.3688694	0.9233714	0	1.1581382
LDHD	197257	ENSG00000166816	0	3.2344809	0.7475067	1.0792903	6.060171	0	2.6591272	0.36865154	3.1395314	4.222813
PDHA1	5160	ENSG00000131828	11.195777	11.345575	10.45147	10.156827	10.377528	11.248066	10.981101	10.508164	10.494298	10.213851
PDHA2	5161	ENSG00000163114	6.5416756	7.352835	6.053615	6.0844575	5.806809	5.7422595	6.04653	5.7851199	5.564605	4.6303153
PDHB	5162	ENSG00000168291	11.151771	10.787737	10.01857	10.189819	10.388985	11.545805	10.989304	10.010316	11.223675	10.659366
PDHX	8050	ENSG00000110435	11.469987	11.069379	10.798909	11.0383415	10.878394	11.018299	10.266986	9.940872	10.120102	9.42201
PDK1	5163	ENSG00000152256	9.173602	11.090658	9.899037	11.15062	10.462985	9.06427	9.361494	9.3715315	9.372674	8.934001
PDK2	5164	ENSG00000005892	6.306005	7.9820004	8.818203	8.370209	8.300196	7.780081	8.296767	9.375695	9.043879	9.461926
PDK3	5165	ENSG00000067992	8.446831	6.1376824	9.52045	6.9146137	4.635946	9.675262	5.68029	9.095707	7.0415764	6.6787095
PDK4	5166	ENSG00000004799	7.6518774	8.032203	11.511249	11.819046	12.233117	5.739105	5.982508	8.313715	6.9271545	7.760661
TCA cycle & Glutaminolysis												
CS	1431	ENSG00000062485	12.104154	12.021648	11.546583	10.678837	10.871778	12.1156435	11.830169	11.618069	11.229054	11.537178
ACO1	48	ENSG00000122729	11.587562	10.381481	10.606644	11.573494	11.407493	11.581986	10.318593	10.8283825	11.790245	11.535990
ACO2	50	ENSG00000100412	10.506633	10.818874	10.057255	10.767778	10.978623	10.597727	10.450865	10.106592	10.363483	10.414825
IDH1	3417	ENSG00000138413	10.595347	11.01756	10.460838	10.687077	10.902105	11.718281	11.673391	11.204786	11.644097	11.660861
IDH2	3418	ENSG00000182654	11.077193	10.961503	10.41898	9.969523	10.418154	10.688006	10.532778	9.9948845	9.937695	9.433058
IDH3A	3419	ENSG00000116041	11.032922	10.788222	10.939995	10.768717	10.128427	11.290143	10.7333145	10.960981	10.556621	10.378237
IDH3B	3420	ENSG00000101365	11.246002	11.426627	9.633114	9.766799	9.731436	11.145845	10.172446	9.77124	10.073116	9.870022
IDH3G	3421	ENSG00000067829	8.646579	9.6368475	9.483188	9.242747	9.570819	8.41706	7.9897714	9.819309	8.999907	8.99914
OGDH	4967	ENSG00000105953	11.244468	10.544521	10.400756	10.427435	11.404189	11.651203	11.589277	10.963297	11.014926	11.717787
OGDHL	5573	ENSG00000197444	7.161757	5.225427	5.5602875	4.660977	3.2740405	7.903981	2.3115678	6.4633293	2.357908	3.3727562
SUCLA2	8803	ENSG00000136143	10.113728	10.807966	10.407981	10.005362	10.270010	10.675802	10.400082	10.516266	10.890263	10.539910
SUCLG1	8802	ENSG00000163541	10.498468	11.132253	11.093054	9.97737	9.808286	10.491657	10.69309	10.95031	10.803535	10.206887
SUCLG2	8801	ENSG00000172340	9.950822	11.320187	9.990201	10.591742	10.760133	10.025218	10.652701	8.982019	10.740157	10.780341
SDHA	6389	ENSG00000005378	11.6333275	10.944166	11.536349	10.424515	10.637836	11.3883705	10.622279	11.294328	10.035286	10.27693
SDHAF1	644096	ENSG00000205135	7.8281655	8.214627	7.5337143	8.502333	8.400414	6.370119	7.297434	7.2360277	6.83329	6.5152802
SDHAF2	54949	ENSG00000167985	9.564266	8.146522	9.127642	9.551334	9.255624	9.358638	8.99914	9.279241	9.578248	9.065778
SDHB	6390	ENSG00000117118	10.841224	11.636481	11.129066	10.4654875	10.07992	10.830645	10.953857	11.033436	10.471801	10.104429
SDHC	642502	ENSG00000143252	8.821783	10.378237	9.102445	8.874095	8.920991	9.074144	9.313252	8.948969	9.551334	9.134911
SDHD	ENSG00000204370	10.205	11.215267	10.500593	10.576683	10.382821	10.454075	9.94469	9.67723	10.193846	10.058991	
FH	2271	ENSG00000194483	11.337451	11.773607	10.07992	10.934129	10.280787	11.115621	11.463231	9.927909	11.116801	11.01886
MDH1	4190	ENSG00000114641	11.485329	12.17761	12.511225	11.288983	11.303142	11.923249	11.82592	12.288181	11.910987	11.740643
MDH1B	130752	ENSG00000138400	0.64805865	6.4000435	4.4412627	1.8758872	0.26045844	4.063967	5.0481176	5.435817	3.7656691	3.2227116
MDH2	4191	ENSG00000146701	11.66197	11.607603	10.77409	10.79865	10.690191	11.504586	11.131183	10.785915	10.97756	10.955593
PC	5091	ENSG00000173599	8.937276	8.106986	8.480386	6.2737064	8.17619	10.168089	8.329315	9.256029	6.797544	8.089377
GLS	2744	ENSG00000115419	12.872203	12.721584	13.473581	13.13611	12.915356	13.236686	12.918546	13.116251	13.542142	13.272663
GLS2	27165	ENSG00000135423	6.2819786	4.7095656	5.321238	3.579793	4.3548245	5.7892957	6.364008	5.875963	5.0412135	4.8090204
GLUD1	2746	ENSG00000148672	11.947237	12.342611	11.486347	11.605607	11.831195	11.673391	11.855499			



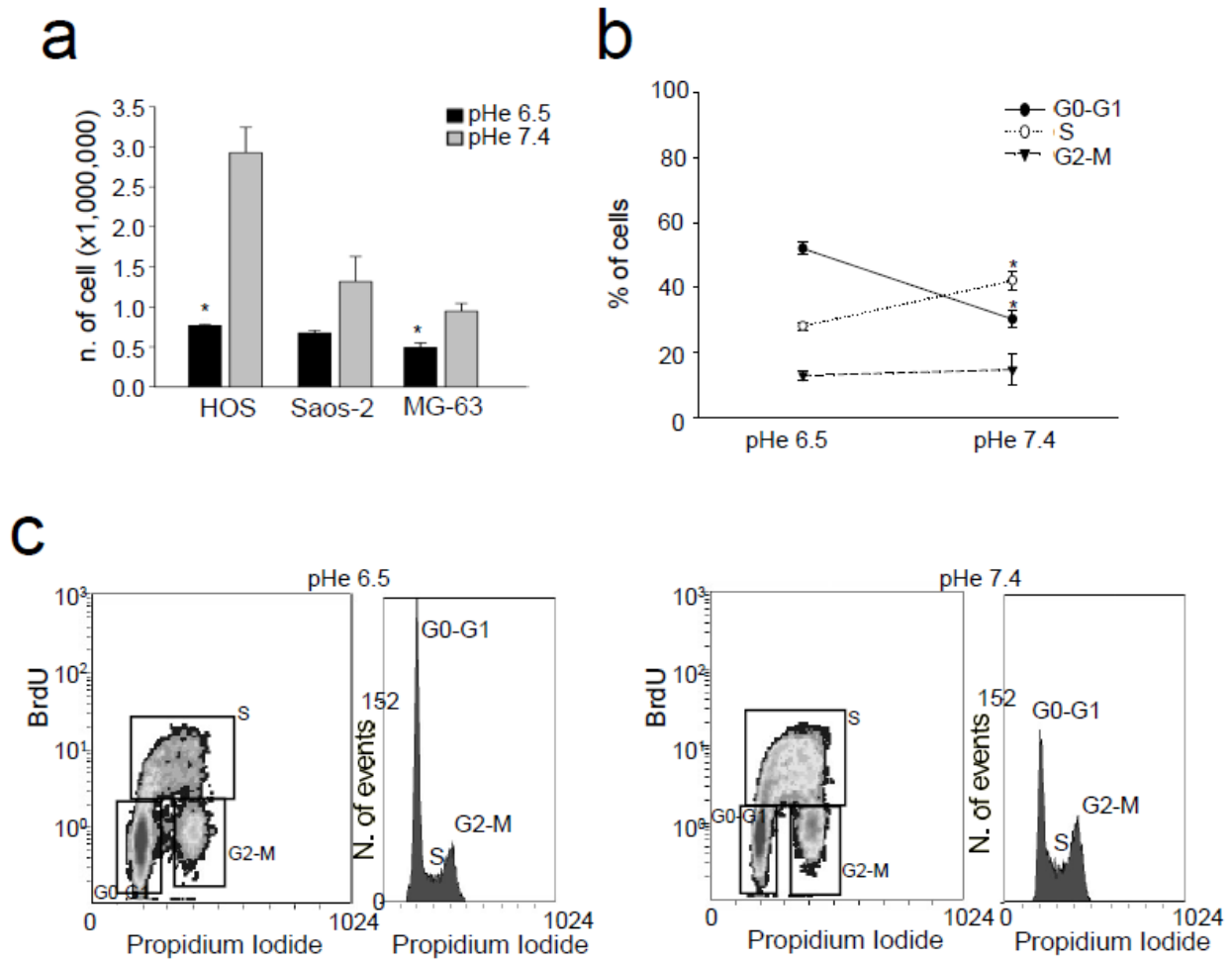
Supplementary Figure S1: Lactate concentration in the supernatant of OS cells maintained in neutral and acidic medium. Lactate concentration is significantly decreased under acidity in all the OS cell lines supernatants, mean \pm SE (*p < 0.05, n = 3).



Supplementary Figure S2: Intracellular ATP content in OS cells cultured under acidic medium. Mean \pm SE (*p < 0.05, n = 3). ATP values were significantly reduced only up to at 80-100% in Saos-2 and MG-63 cells and unchanged in HOS cells.



Supplementary Figure S3: Protein expressions of complexes of the respiratory chain (OXPHOS) in OS cell lines under both neutral and acidic medium. (a) Western blot analysis of the expression of OXPHOS complexes under different pHs. Representative image. (b) Bar graph of the quantification of the expression of OXPHOS complexes showed in panel A, mean \pm SE (* $p < 0.05$, $n = 3$).



Supplementary Figure S4: Proliferation under different pH. (a) Cell count of OS cells cultured at different pHs, soon after the time population doubling; (b) cell cycle analysis of OS cells cultured at different pH; (c) representative graph of the cell cycle analysis showed in panel B.

Supplementary methods

Lactate assay. Two $\times 10^5$ cells of OS were seeded into 6-well plates and allowed to attach overnight. After 24 h the media were replaced with 3 ml of freshly neutral (pH 7.3) or acidic (pH 6.5) DMEM containing 2% NuSerum, a low protein alternative to FBS (BD Biosciences). After additional 24h, lactate concentration in the supernatant was measured by lactate Assay kit (EnzyChrome, BioAssay Systems), according to the manufacturer's instruction. The assay was repeated three times in quadruplicate. Lactate concentration was normalized to total cell number.

ATP assay. OS cells were seeded into 6-well plates at a density of 2×10^5 cells/well. After 24 h media were replaced with 2 ml of freshly neutral (pH 7.4) or acidic (pH 6.5) DMEM high glucose (4.5 g/L) containing 10% FBS. The cells were incubated at 37 °C and 5% CO₂ for 24 h. For the analysis of ATP content, cells were lysed by using boiling ATP lysis buffer [0.1 M Tris (hydroxymethyl) aminomethane (Tris base) (Sigma) and 2 mM EDTA adjusted to pH 7.75 with acetic acid (Sigma) and 2.5% Dodecyltrimethylammonium bromide (Sigma)]. The ATP content was measured with the ATP determination kit (Molecular Probes). The luciferase signal was normalized with the total protein concentration measured by Bradford assay (Bio-Rad Laboratories). All experiments were performed in triplicates.

Western Blotting. To detect ATP synthase subunit- α , Complex II subunit 30 kDa, Complex III subunit Core 2, V0c and V1B2 subunits of the V-ATPase protein, and heat shock protein 60 (HSP60) in total lysates, the cells were harvested in lysis buffer (RIPA buffer) containing protease and phosphatase inhibitors and centrifuged at 13,000x g for 10 min at 4°C to remove insoluble debris. Protein concentrations were analyzed using Bradford assay (Biorad). 30 μ g of proteins were loaded and separated by SDS-PAGE and transferred to a 0.2 μ m nitrocellulose membrane (Fisher Scientific). After blocking for 1 h in TBST (10 mM Tris-HCl pH 8.0, 150 mM NaCl, 0.5% Tween-20) with 5% non-fat dry milk, membranes were incubated with the primary antibody for 1 h or overnight, washed and incubated for 1 h with horseradish peroxidase-conjugated secondary antibodies. The membranes were washed and incubated with an enhanced chemi-luminescence substrate (ECL; Thermo Scientific). Blots were probed with MitoProfile total oxidative phosphorylation (OXPHOS) human antibody Cocktail 1:300 (Abcam, Cambridge, UK), anti-HSP60 (1:2000, Sigma) polyclonal antibodies.

Cell cycle analysis. In order to analyze cell cycle distribution after exposure to acidic conditions, DNA content and bromodeoxyuridine (BrdU) incorporation were determined by simultaneous analysis of propidium iodide and fluorescein isothiocyanate (FITC)-conjugated anti-BrdU fluorescence. Cells were seeded at low density in complete medium, and after 24 h the medium was changed with complete medium at pH 6.5 or 7.4. After additional 48 h, cells were incubated with 10 mM BrdU (Sigma) for 60 min before harvesting. Cells were collected by trypsinization, followed by fixation in 40% ethanol for 20 min. Partial DNA denaturation was performed by incubating cells in HCl, followed by neutralization with sodium tetraborate. Samples were then exposed to a monoclonal anti-BrdU FITC antibody (BD Biosciences, Erembodegem, Belgium), washed, and finally stained with 2.5 mg/ml propidium iodide (Sigma). Flow cytometric analysis was performed with a Coulter EPICS1 XL Flow Cytometer (Coulter Corporation, Beckman Coulter, Milan, Italy). Monoparametric and biparametric analyses were performed using the WinMDI 2.7 software. The experiment was repeated three times.

RNA extraction and RNA-seq. analysis. Total RNA from each cell was extracted using guanidinium thiocyanate-phenol-chloroform. The total RNA was quantified with Bioanalyzer (Agilent, Santa Clara, CA) following the manufacturer's instructions. RIN (RNA Integrity Number) and A260/A280 ratio of the prepared total RNA were all 10, and over 1.8, respectively.

The library of template molecules for high throughput DNA sequencing was converted from the total RNA using TruSeq RNA Sample Prep Kit v2 (Illumina, San Diego, CA) following the manufacturer's protocol. The library was quantified with Bioanalyzer (Agilent) following the manufacturer's instruction. Library (7 pM) was subjected to cluster amplification to cluster generation on a Single Read Flow Cell v4 with a cluster generation instrument (Illumina). Sequencing was performed on a Genome Analyzer GAIIx for 76 cycles using Cycle Sequencing v4 reagents (Illumina)..

Image analysis and base calling were performed using Off-Line Basecaller Software 1.6 (Illumina). Reads were aligned using ELAND v2 of CASAVA Software 1.7 with the sequence data sets. Human genome build 19 (hg19) were downloaded from University of California, Santa Cruz genome browser (<http://genome.ucsc.edu/>) as the analytic reference. Transcript coverage for every gene locus was calculated from the total number passing filter reads that mapped, by ELAND-RNA, to exons. These analyses were performed using default parameters. The advanced analysis for quantification with Quantile normalization algorithm was performed using Avadis NGS software (version1.5, Strand Scientific Intelligence Inc., San Francisco, CA). The filtering was performed using default parameters. All new data has been deposited in DDBJ/EMBL/GenBank under DRA004087.