

## Supplement

### **N-Terminal Ile-Orn- and Trp-Orn-Motif Repeats Enhance Membrane Interaction and Increase the Antimicrobial Activity of Apidaecins against *Pseudomonas aeruginosa***

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Table S1: MIC-values in  $\mu\text{g/mL}$  of Api137, Api794, and Api795 determined against *P. aeruginosa* DSM 1117 and *E. coli* DSM 1103 in 50% MHB and 33% TSB, respectively, using an inoculum of  $5 \times 10^8$  cells/mL corresponding to the conditions applied for electron microscopy.

Peptide	<i>P. aeruginosa</i> DSM1117			<i>E. coli</i> DSM 1103		
	0.25x MIC	MIC	4x MIC	0.25x MIC	MIC	4x MIC
Api137	32	128	512	8	32	128
Api794	16	64	256	16	64	256
Api795	4	16	64	8	32	128

Table S2: Quantification of morphological changes of *P. aeruginosa* DSM 1117 cells [%].

Conc. [ $\mu\text{g/mL}$ ]	Membrane		Cells		DNA/ribosomes relocalization				Ribosome clustering				Vesicle release	dissociated Fragments				
	i	b	r	spongy	-	+	++	+++	-	+	++	+++		-	+	++	LV	
<b>Water</b> -	100	0	0	0	90	10	0	0	87	13	0	0	13	87	13	0	0	
<b>Api137</b>	32	100	0	0	40	43	17	0	37	63	0	0	43	87	13	0	0	
	128	100	0	0	47	37	17	0	43	57	0	0	47	87	10	3	0	
	512	100	0	0	47	40	13	0	70	30	0	0	37	97	0	3	0	
<b>Api795</b>	4	100	0	0	40	37	23	0	30	70	0	0	67	93	3	3	0	
	16	100	0	0	50	40	10	0	70	30	0	0	13	100	0	0	0	
	64	90	10	3	10	10	67	13	10	77	7	7	50	97	0	3	0	
	512	20	80	63	10	0	0	0	100	0	0	7	93	7	87	0	13	0
<b>Api794</b>	16	100	0	0	40	37	23	0	20	80	0	0	87	77	10	13	0	
	64	20	80	47	0	3	3	10	83	3	13	10	73	20	17	20	63	30
	256	34	67	67	80	0	0	0	100	0	0	0	100	3	7	13	80	93
	512	17	13	17	87	0	0	0	100	0	0	7	93	0	40	0	60	97

i, intact; b, broken; r, ruptured; DNA/ribosome relocalization and ribosome clustering: -, no; +, starting; ++ relocalized/clustering; +++, extremely relocalized/clustering; fragments: -, no; +, small and thin; ++, long and thin; LV, large vesicles

Tab. S3: Quantification of morphological changes of *E. coli* DSM 1103 cells [%].

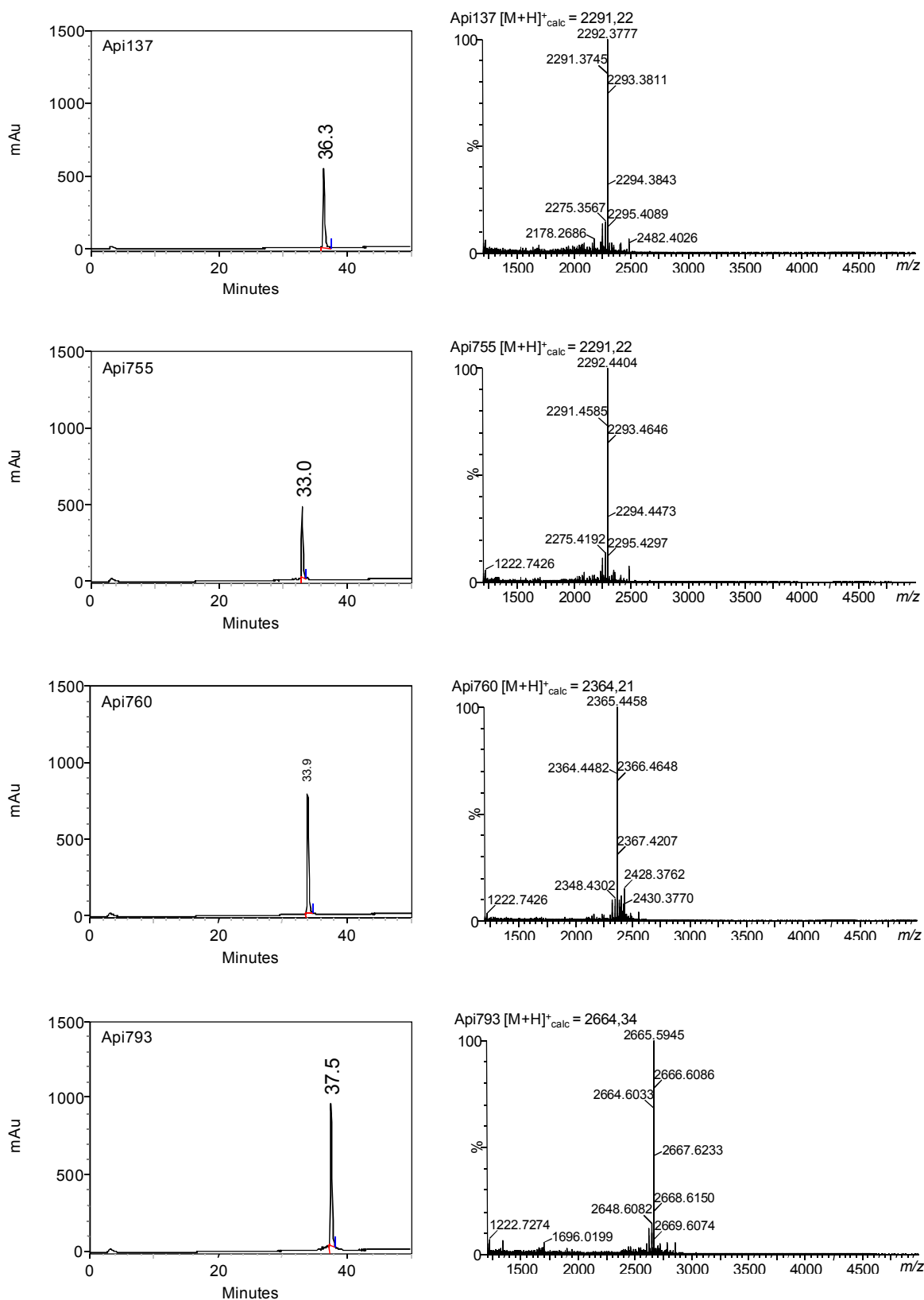
	Conc. [µg/mL]	Membrane		Cell r	DNA/Ribosomes relocalized	Vesicle release		Fragments				
		i	w			weak	strong	-	+	++	+++	LV
<b>water</b>	-	13	87	0	0	13	-	33	63	3	0	0
<b>Api137</b>	8	100	0	0	17	-	100	43	30	27	0	0
	32	97	3	0	30	-	100	37	40	23	0	0
	128	4	96	0	0	35	-	4	54	42	0	0
	512	32	68	0	0	50	-	11	61	29	0	0
<b>Api795</b>	8	3	97	0	0	30	-	13	63	23	0	0
	32	7	93	0	0	43	-	33	57	10	0	0
	128	17	83	0	0	47	-	17	53	30	0	0
	512	33	67	0	30	60	-	0	63	37	0	0
<b>Api794</b>	16	33	67	0	0	-	100	37	40	23	0	0
	64	100	0	0	0	-	97	17	27	33	23	3
	256	3	97	0	0	-	97	17	43	37	3	93
	512	93	7	3	30	-	93	17	37	30	17	97

i, intact; w, wrinkled; r, ruptured; fragments: -, no; +, small and thin; ++, long and thin; +++ long and thick; LV, large vesicles

Table S4: Survival rates of HeLa cells after incubating them with Cf-labeled apidaecin peptides for 10 min or 6 h. The fluorescence corresponding to 5(6)-carboxyfluorescein and eFluor660 to stain dead cells were measured by flow cytometry. Dead cells were excluded from the population. Experiments were performed twice (Exp1/Exp2).

Time	Water	Cf-Api137	Cf-Api794	Cf-Api795
10 min	90% / 87%	89% / 85%	91% / 85%	90% / 85%
6 h	93% / 86%	93% / 91%	85% / 84%	93% / 86%

Figure S1: RP-HPLC and MALDI-TOF-MS of all purified apidaecin analogs used in this study. RP-HPLC relied on a Jupiter C<sub>18</sub>-column and a linear aqueous acetonitrile gradient from 21-33% (v/v) acetonitrile in 30 min in the presence of TFA (0.1% v/v; left). Mass spectra were recorded on a Synapt G2Si mass spectrometer equipped with a MALDI-source using CHHA as matrix (right).



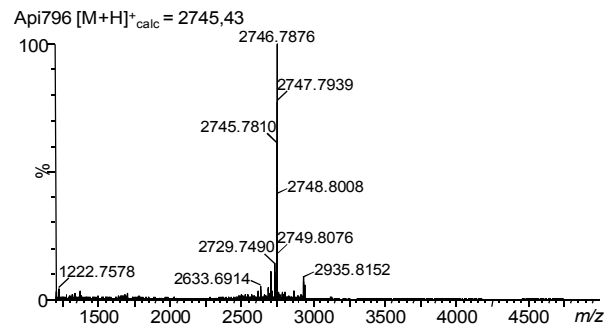
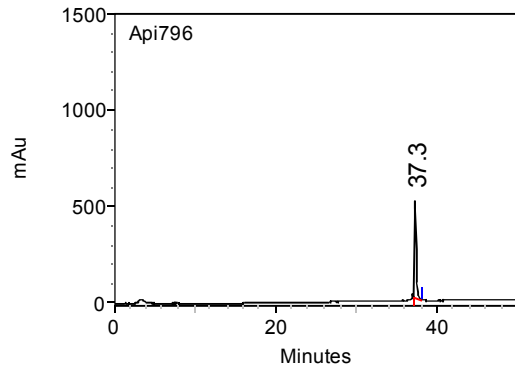
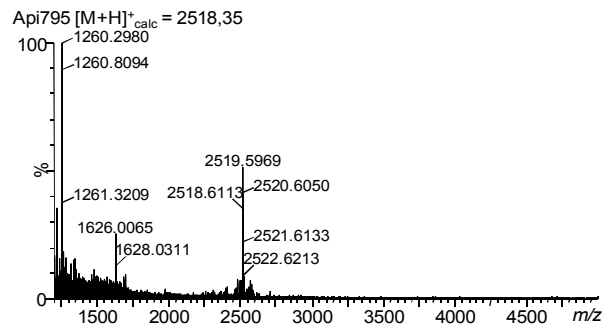
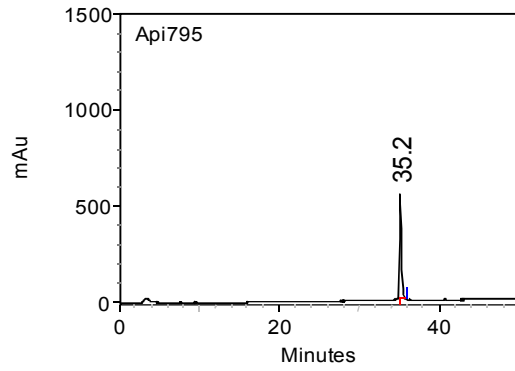
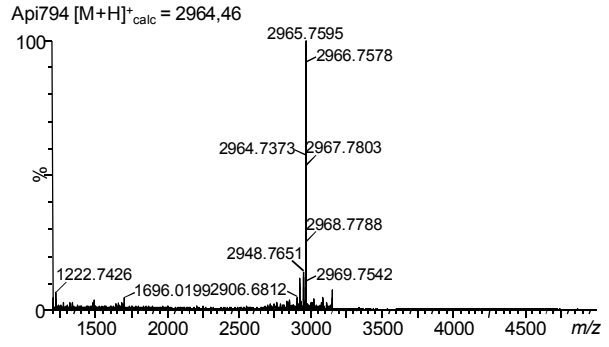
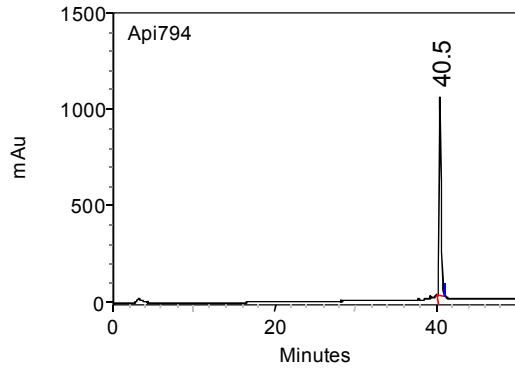


Figure S2: RP-HPLC and MALDI-TOF-MS of all purified 5(6)-carboxyfluorescein labeled apidaecin peptides used in this study. RP-HPLC relied on a Jupiter C<sub>18</sub>-column and a linear aqueous acetonitrile gradient from 31.5-49.5% (v/v) acetonitrile in 30 min in the presence of TFA (0.1% v/v; left). Mass spectra were recorded on a Synapt G2Si mass spectrometer equipped with a MALDI-source using CHHA as matrix (right).

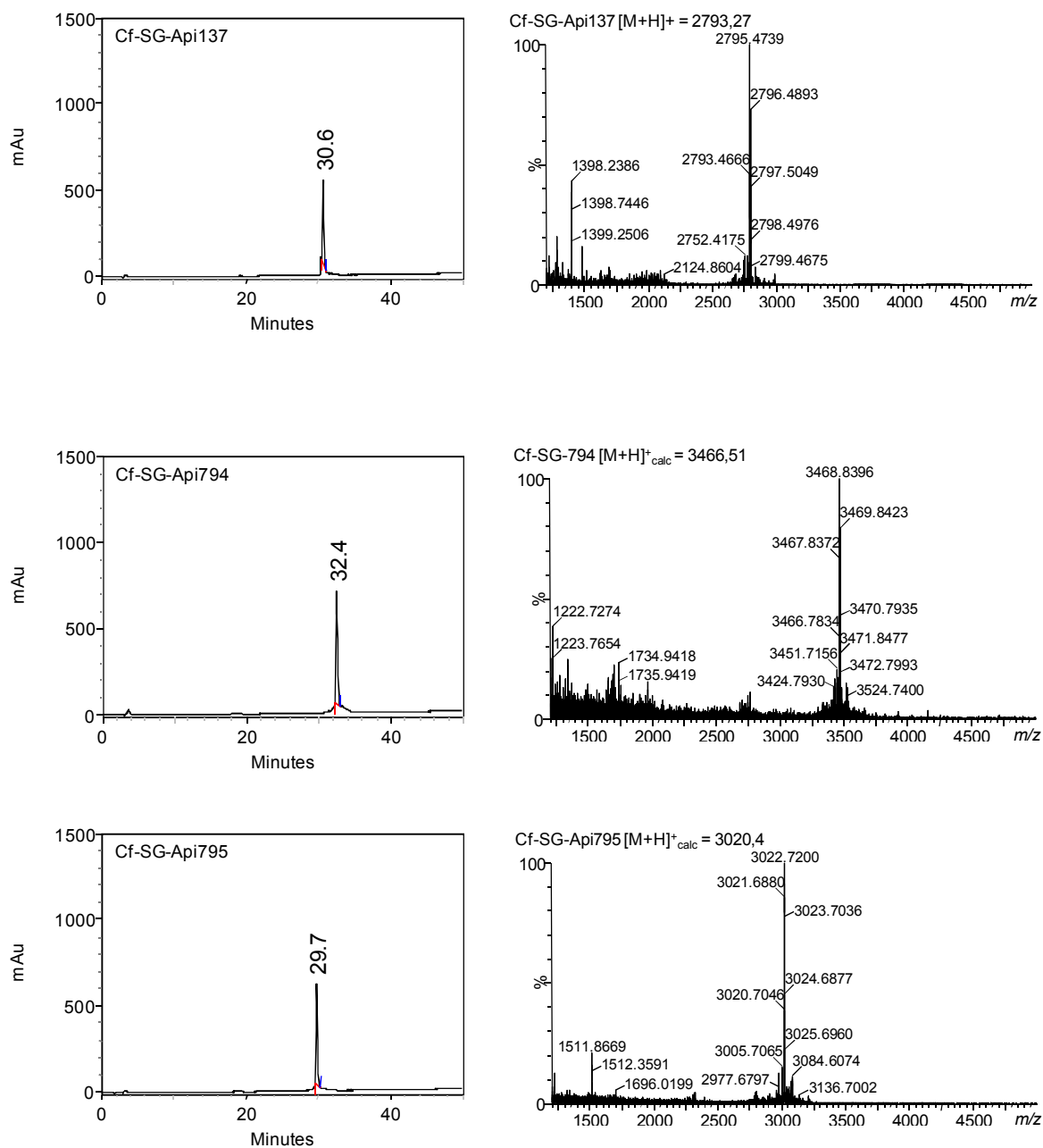


Figure S3: Electron microscopy images of *P. aeruginosa* DSM 1117 incubated with Api137 (A), Api795 (B), and Api794 (C) at concentrations corresponding to 0.25x MIC, MIC, and 4x MIC for 1 h using cell densities of  $5 \times 10^8$  CFU/mL. Black bars represent 200 nm.

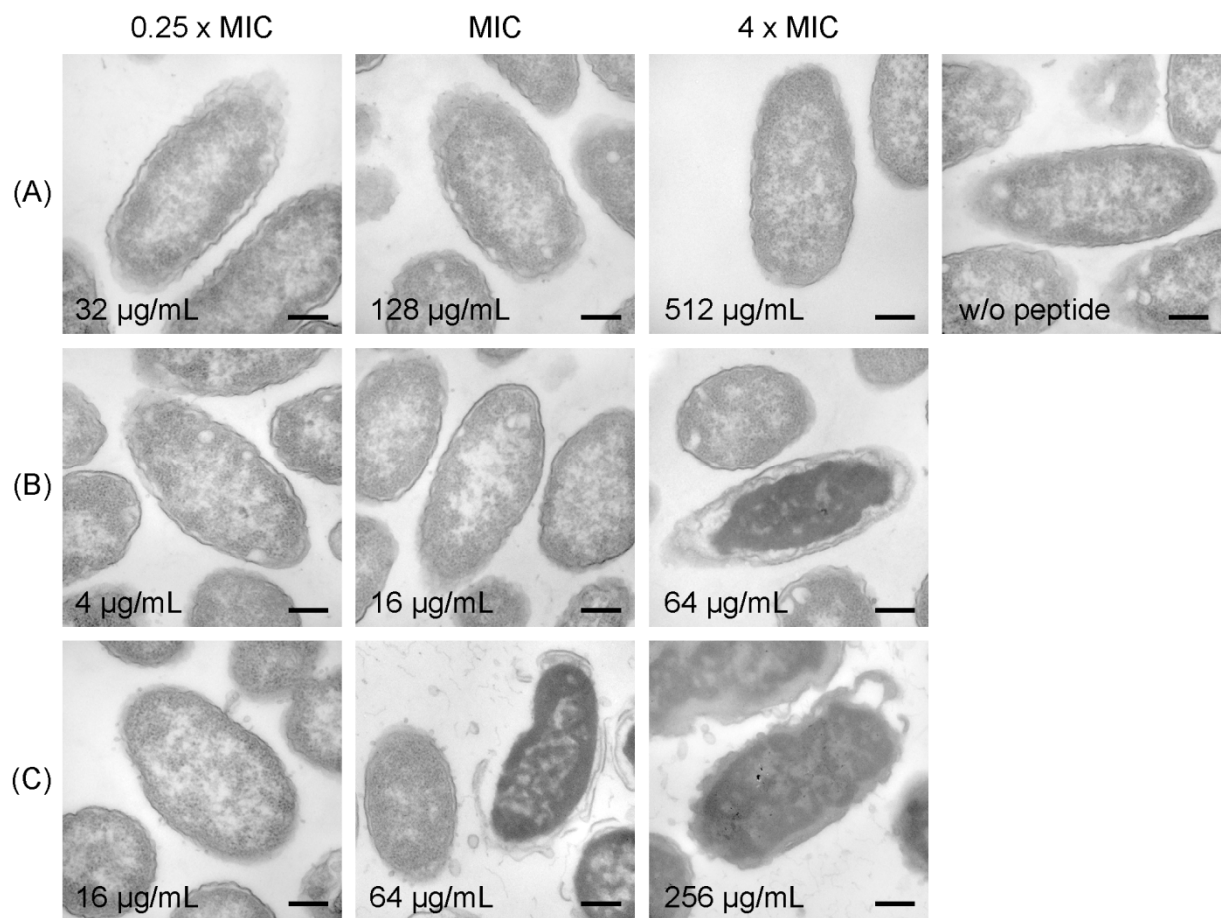


Figure S4: Electron microscopy images of *E. coli* DSM 1103 incubated with Api137 (A), Api795 (B), and Api794 (C) at concentrations corresponding to 0.25x MIC, MIC, and 4x MIC for 1 h using cell densities of  $5 \times 10^8$  CFU/mL. Black bars represent 200 nm.

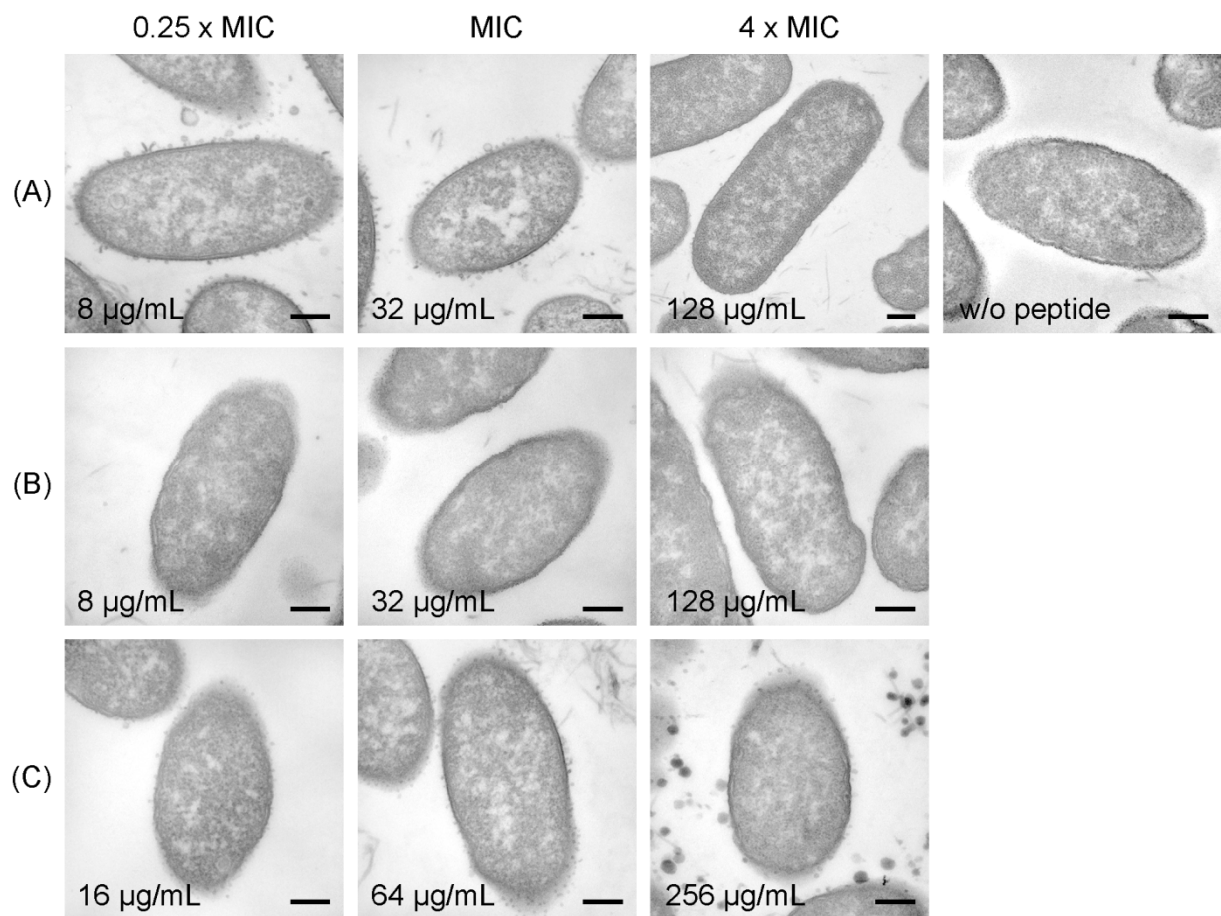




Figure S5: Electron microscopy images of *P. aeruginosa* DSM 1117 (A) and *E. coli* DSM 1103 (B) incubated with the indicated apidaecin analogs (512  $\mu\text{g/mL}$ ) for 1 h using cell densities of  $5 \times 10^8$  CFU/mL. Black bars refer to 500 nm.

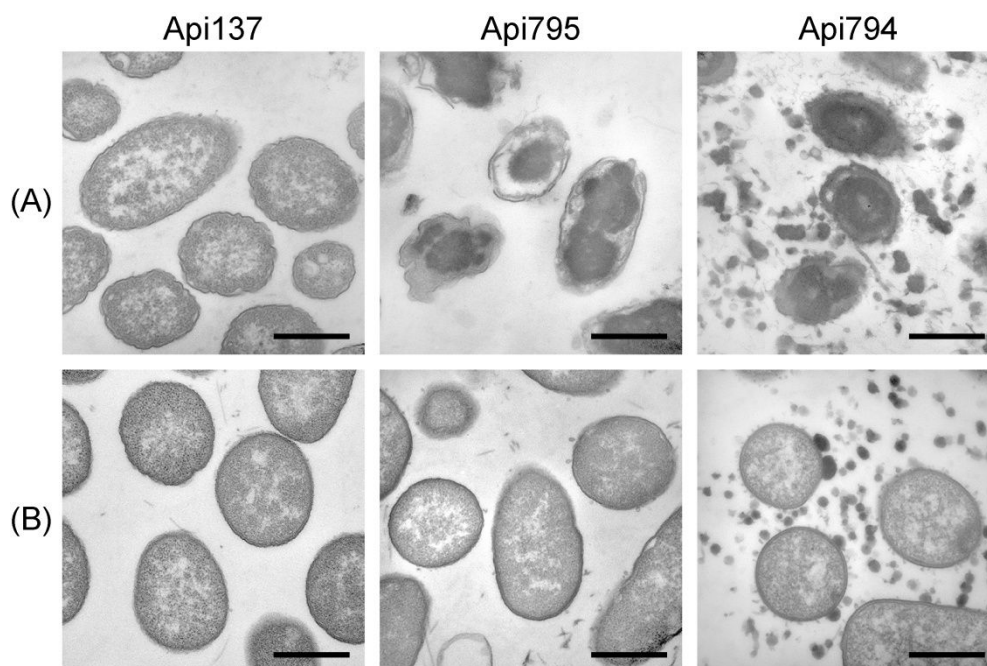


Figure S6: RP-chromatograms of LL-37, Api137, Api794, and Api795 before and after incubation at a concentration of 0.5 g/L with *P. aeruginosa* elastase (12 units/mL) in Tris-HCl buffer (pH 8.3) at 37 °C.

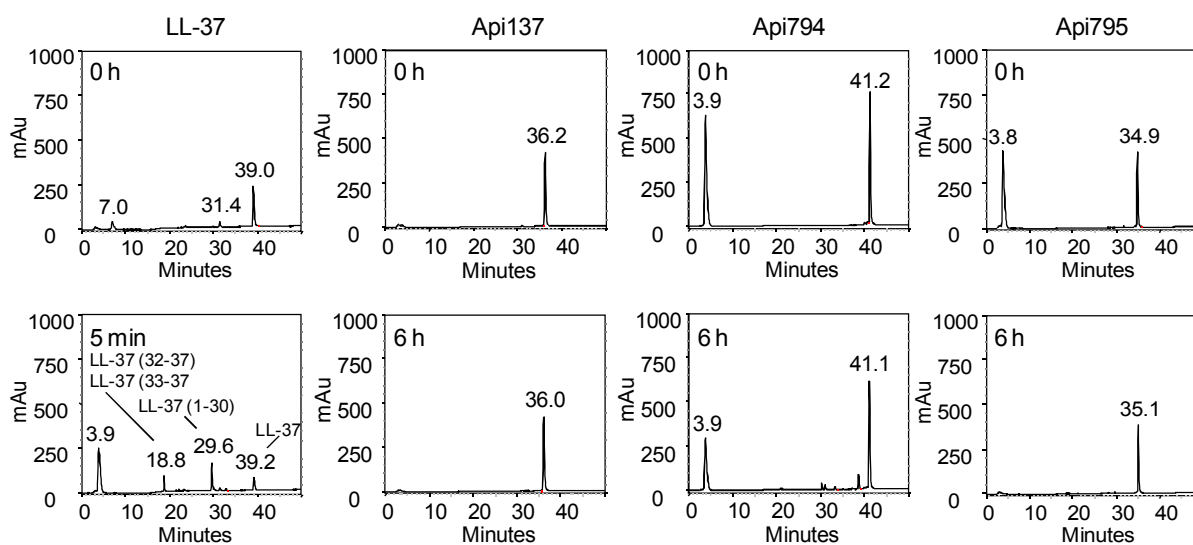


Figure S7: Confocal microscopy images of HeLa cells incubated with Cf-labeled apidaecin peptides (green). Cells were treated without (A) or with dynasore (B); 0.2 mmol/L) for 45 min prior to peptide treatment (40  $\mu$ mol/L, 30 min). The cells' nuclei were visualized by the use of Hoechst 33324 (blue). Bars refer to 20  $\mu$ m. Arrows indicate examples of areas with endosomes (white) and stained cytosol (red).

