

Supporting Information

1. Syntheses and characterization of **1b**·PF₆ and [(η⁶-p-cymene)Ru(*k*NPh,*k*²NO)] (**3a**).

[(η⁶-p-cymene)Ru(*k*NHBn,*k*NOH)Cl]PF₆ (1b**·PF₆):** KPF₆ (0.05 g, 0.26 mmol) was added to a CH₂Cl₂ (15.0 mL) solution of **1b** (0.15 g, 0.26 mmol), and the mixture was stirred overnight. The resulting suspension was filtered to remove insoluble KCl and the solvent was evaporated to dryness from the orange solution to afford an orange solid, which was identified as compound **1b**·PF₆ (yield 0.16 g, 90 %). C₂₇H₃₈ClF₆N₂OPRu (688.09): calcd. C 47.13, H 5.57, N 4.07; found C 46.80, H 4.89, N 3.32. IR (KBr): $\nu = 3300\text{-}3100$ (NH/OH), 1646, 1614 (C=N) cm⁻¹. ¹H NMR (400.1 MHz, 293 K, CDCl₃): $\delta = 7.41$ (overlapped, 5H, -C₆H₅), 5.85, 5.81 (both d, each 1H, ³J_{HH} = 6, p-cymene-C₆H₄), 5.46, 5.44 (both d, each 1H, ³J_{HH} = 8, p-cymene-C₆H₄), 4.78 (s, 1H, =CH₂), 4.61 (m, 2H, -CH₂Ph) 4.53 (br, 1H, =CH₂), 3.92 (m, 1H, NH), 3.61 (d, 1H, ²J_{HH} = 14, -CH₂⁶), 2.58 (spt, 1H, ³J_{HH} = 6, p-cymene-CHMe₂), 2.55 (br, 1H, -CH⁵), 2.32 (dd, 1H, ²J_{HH} = 16, ³J_{HH} = 6, -CH₂⁶), 1.97 (overlapped, 4H, p-cymene-CH₃ + CH₂^{3,4}), 1.81 (m, 1H, -CH₂^{3,4}), 1.64 (m, 1H, -CH₂^{3,4}), 1.66 (s, 3H, CH₃-C=), 1.36 (m, 1H, -CH₂^{3,4}), 1.19, 0.99 (both d, each 3H, ³J_{HH} = 6, p-cymene-CH(CH₃)₂) ppm. ³¹P NMR (161.9 MHz, 293 K, CDCl₃): $\delta = -144.0$ (spt, J_{P-F} = 692 Hz, PF₆) ppm. ¹⁹F NMR (376.5 MHz, 293 K, CDCl₃): $\delta = -72.5$ (d, J_{P-F} = 692 Hz, PF₆) ppm.

[(η⁶-p-cymene)Ru(*k*NPh,*k*²NO)] (3a**):** A THF solution of **1a** (0.25 g, 0.44 mmol) was treated with NaOMe (0.05 g, 0.98 mmol) at room temperature. After stirring of the mixture during 2 hours, evaporation of the THF and extraction from the solid residue with toluene affords an orange solution from which an orange solid was isolated, washed with hexane (5x2 mL) and fully characterized as derivative **3a** (yield 0.18 g, 83 %). C₂₆H₃₄N₂ORu (491.63): calcd. C 63.52, H 6.97, N 5.70; found C 63.99, H 6.90, N, 5.53. IR (KBr): $\nu = 1622, 1590$ (C=N) cm⁻¹. ¹H NMR (plus HSQC, plus HMBC, plus COSY, 400.1 MHz, 293 K, CDCl₃): $\delta = 7.28$ (m, 2H, C₆H₅^{o/m}), 7.16 (m, 1H, C₆H₅^p), 7.10 (m, 2H, C₆H₅^{o/m}), 5.26 (d, 1H, ³J_{HH} = 6, p-cymene-C₆H₄), 5.12 (1H, =CH₂), 5.10, 5.02, 4.81 (both d, each 1H, ³J_{HH} = 6, p-cymene-C₆H₄), 4.73 (br, 1H, =CH₂), 3.88 (d, 1H, ²J_{HH} = 16, -CH₂⁶), 2.63 (spt, 1H, ³J_{HH} = 6, CHMe₂), 2.38 (br, 1H, -CH⁵), 2.12 (s, 3H, p-cymene-CH₃), 1.85 (dd, 1H, ²J_{HH} = 16, ³J_{HH} = 6, -CH₂⁶), 1.61 (overlapped, 4H, CH₃-C= + -CH₂⁴), 1.47 (overlapped, 2H, 1-CH₂⁴ + 1-CH₂³), 1.25, 1.23 (overlapped, both d, each 3H, ³J_{HH} = 6, CH(CH₃)₂), 1.17 (m, 1H, -CH₂³), 1.05 (s, 3H, HNC-CH₃) ppm. ¹³C-NMR (plus APT, plus gHSQC, plus HMBC, 100.6 MHz, 293 K, CDCl₃): $\delta = 163.4$ (-, C=N-O), 158.3 (-, C_{ipso}-Ph), 144.9 (-, C=CH₂), 128.0, 126.6, 125.2 (+, -C₆H₅^{o,m,p}), 112.3 (-, =CH₂), 100.3, 89.9 (-, C_{ipso}-p-cymene), 83.1, 82.9, 82.6, 80.8 (+, p-cymene-C₆H₄), 77.7 (-, C-N-Ru), 42.1 (+, -CH⁵), 33.8 (-, -CH₂³), 31.4 (+, p-cymene-CHMe₂), 26.2 (-, -CH₂⁴), 25.3 (-CH₂⁶), 24.1, 24.0 (both +, p-cymene-CH(CH₃)₂), 23.8 (+, CH₃-CNH), 22.8 (+, CH₃-C=), 20.0 (+, p-cymene-CH₃) ppm. ¹⁵N NMR (gHMBC, 40.5 MHz, 293 K, CDCl₃): $\delta = 329.0$ (=NO), 263.5 (Ru-NPh) ppm.

Preligands a, b, c and compounds 1a, 1b and 2b

Syntheses and characterization of amino-oxime derivatives (2S,5R)-[NHR,NOH]¹ (R = a, b, c) and ruthenium compounds [(η^6 -p-cymene)Ru(*k*NHR,*k*NOH)Cl]Cl (R = Ph **1a**, Bn **1b**)² and [(η^6 -p-cymene)Ru(*k*NHBn,*k*²NO)Cl] (**2b**)² was reported elsewhere. During this work, we performed the IR and ¹⁵N-¹H HMBC spectra of all these compounds, as well as the spectroscopic NMR characterization in methanol-*d*₄ of compounds **1a** and **1b**, since their NMR spectra in CDCl₃ changed dramatically with the concentration (see Dilution Spectra, page S9). Solubility in water of **1a**, **1b** and **2b** is given in mM concentration. Data are given in the following section:

(2S,5R)-[NHPh,NOH] (a). IR (KBr): ν = 3100-3300 (NH/OH), 1644, 1602 (C=N) cm⁻¹. ¹⁵N NMR (gHMBC, 40.5 MHz, 293 K, CDCl₃): δ = 343.3 (=NOH), 84.1 (NHPh) ppm.

(2S,5R)-[NHBn,NOH] (b). IR (KBr): ν = 3100-3320 (NH/OH), 1644, 1602 (C=N) cm⁻¹. ¹⁵N NMR (gHMBC, 40.5 MHz, 293 K, CDCl₃): δ = 340.0 (=NOH), 60.0 (NHBn) ppm.

(2S,5R)-[NH(2-pic),NOH] (c). IR (KBr): ν = 3086-3314 (NH/OH), 1650, 1595 (C=N) cm⁻¹. ¹⁵N NMR (gHMBC, 40.5 MHz, 293 K, CDCl₃): δ = 343.3 (=NOH), 305.3 (=Npic), 51.8 (NH(2-pic)) ppm.

[(η^6 -p-cymene)Ru(*k*NHPh,*k*NOH)Cl]Cl (1a**)**: IR (KBr): ν = 3391-3151 (NH/OH), 1642, 1597 (C=N) cm⁻¹. Solubility in H₂O at 24 °C (mM): 21 ± 2. Value of pH ([9.0 mM]) in H₂O at 24 °C: 4.62.

¹H NMR (plus HMBC, gHSQC, plus COSY, 400.1 MHz, 293 K, CD₃OD): δ = 7.89, 7.65, 7.52, 7.41, 7.02 (all br, each 1H, C₆H₅), 6.19, 5.86, 5.52 (all d, each 1H, ³J_{HH} = 6, C₆H₄), 5.39 (s, 1H, NH), 4.85, 4.85 (overlapped with CD₃OD, 2H, C₆H₄ + =CH₂), 4.60 (br, 1H, =CH₂), 3.66 (d, 1H, ²J_{HH} = 15, CH₂⁶), 2.71 (spt, 1H, ³J_{HH} = 7, p-cymene-CHMe₂), 2.53 (m, 1H, -CH⁵), 2.56 (m, 1H, -CH₂⁶), 2.27 (s, 3H, p-cymene-CH₃), 1.78 (m, 1H, -CH₂⁴), 1.79 (s, 3H, CH₃-C=), 1.67 (s, 3H, PhNC-CH₃), 1.55 (m, 3H, -CH₂⁴+ -CH₂³), 1.13, 0.62 (both d, each 3H, ³J_{HH} = 7, p-cymene-CH(CH₃)₂) ppm. ¹³C NMR (plus APT, plus gHSQC, plus HMBC, 100.6 MHz, 293 K, CD₃OD): δ = 170.2 (-, C=NO), 146.3 (-, =C-Me), 144.1 (-, C_{ipso}-Ph), 130.8, 130.8, 129.2, 125.6, 126.2, 114.2 (-, =CH₂), 106.3 (-, C_{ipso}, p-cymene-CCHMe₂), 98.8 (-, C_{ipso}, p-cymene-CMe), 87.3, 86.1, 87.9, 81.9 (all +, p-cymene-C₆H₄), 71.6 (-, C-NH), 49.9 (+, -CH⁵), 36.0 (-, -CH₂³), 31.9 (+, p-cymene-CHMe₂), 28.9 (-, -CH₂⁶), 24.1 (+, p-cymene-CH(CH₃)₂), 24.7 (-CH₂⁴), 22.6 (+, CH₃-C=), 22.2 (+, CH₃-CNPh), 19.0 (+, p-cymene-CH(CH₃)₂), 18.6 (+, p-cymene-CH₃) ppm. ¹⁵N NMR (gHMBC, 40.5 MHz, 293 K, CDCl₃): δ = 272.1 (=NOH), 68.1 (NHPh) ppm. ¹⁵N NMR (gHMBC, 40.5 MHz, 293 K, CD₃OD): δ = 266.5 (=NOH), 68.2 (NHPh) ppm.

[(η^6 -p-cymene)Ru(*k*NHBn,*k*NOH)Cl]Cl (1b**)**: IR (KBr): ν = 3400-3040 (NH/NOH), 1643, 1600 (C=N) cm⁻¹. Solubility in H₂O at 24 °C (mM): 28 ± 4. Value of pH ([9.0 mM]) in H₂O at 24 °C: 4.70.

¹ Carman, R. M.; Mathew, P. C.; Saraswathi, G. N.; Singaram, B.; Verghese, J. *Aust. J. Chem.* **1977**, *30*, 1323

² Ibn El Alami, M. S.; El Amrani, M. A.; Dahdouh, A.; Roussel, P.; Suisse, I.; Mortreux, A. *Chirality* **2012**, *24*, 675-682 and references therein.

^1H NMR (plus gHSQC, plus HMBC, plus COSY, 400.1 MHz, 293 K, CD_3OD): δ = 7.45 (m, 5H, C_6H_5), 5.85, 5.84, 5.46, 5.33 (d, each 1H, $^3J_{\text{HH}} = 6$, p-cymene- C_6H_4), 4.79 (m, 1H, $=\text{CH}_2$), 4.62 (second order system, 2H, $-\text{CH}_2\text{Ph}$), 4.60 (br, 1H, $=\text{CH}_2$), 4.02 (br, 1H, NH), 3.60 (d, 1H, $^2J_{\text{HH}} = 16$, CH_2^6), 2.53 (overlapped, 3H, p-cymene- $\text{CHMe}_2 + \text{CH}_2^6 + \text{CH}^5$), 2.14 (m, 1H, $-\text{CH}_2^3$), 1.99 (s, 3H, p-cymene- CH_3), 1.83 (m, 2H, $-\text{CH}_2^4$), 1.66, 1.65 (both s, each 3H, $\text{BnNC-CH}_3 + \text{CH}_3\text{-C=}$), 1.39 (m, 1H, $-\text{CH}_2^3$), 1.22, 1.02 (both d, each 3H, $^3J_{\text{HH}} = 8$, p-cymene- $\text{CH}(\text{CH}_3)_2$) ppm. ^{13}C - NMR (plus APT, plus gHSQC, 100.6 MHz, 293 K, CD_3OD): δ = 170.8 (-, C=N), 145.9 (-, $=\text{C-CH}_3$), 137.1 (-, $\text{C}_{\text{ipso}}\text{-Ph}$), 130.1, 129.5, 129.4 (all +, - $\text{C}_6\text{H}_5^{(\text{o,m,p})}$), 113.2 (-, $=\text{CH}_2$), 108.9, 98.8 (both -, $\text{C}_{\text{ipso}}\text{-p-cymene}$), 87.5, 84.8, 83.3, 83.2 (all +, $-\text{C}_6\text{H}_4$), 70.5 (-, C-NH), 55.9 (-, CH_2Ph), 39.4 (+, $-\text{CH}^5$), 35.4 (-, $-\text{CH}_2^3$), 32.5 (+, p-cymene- CHMe_2), 29.1 (-, $-\text{CH}_2^6$), 25.1 ($-\text{CH}_2^4$), 23.9 (+, p-cymene- $\text{CH}(\text{CH}_3)_2$), 22.3 (+, $\text{CH}_3\text{-C-NH}$), 20.8 (+, p-cymene- $\text{CH}(\text{CH}_3)_2$), 20.7 (+, $\text{CH}_3\text{-C=}$), 18.4 (+, p-cymene- CH_3) ppm. ^{15}N NMR (gHMBC, 40.5 MHz, 293 K, CDCl_3): δ = 272.0 (=NOH), 50.4 (NHBn) ppm. ^{15}N NMR (gHMBC, 40.5 MHz, 293 K, CD_3OD): δ = 266.7 (=NOH), 50.0 (NHBn) ppm.

$[(\eta^6\text{-p-cymene})\text{Ru}(\kappa\text{NHBn}, \kappa\text{NO})\text{Cl}]$ (2b). IR (KBr): ν = 3400-3060 (NH/NOH), 1700, 1643 (C=N) cm^{-1} . Solubility in H_2O at 24 $^\circ\text{C}$ (mM): 20.0 ± 4 . ^{15}N NMR (gHMBC, 40.5 MHz, 293 K, CDCl_3): δ = 291.7 (=NOH), 50.4 (NHBn) ppm.

2. ¹H NMR dilution data

Dilutions experiments were carried out from an initial stock solution. This initial stock solution (500 μL) was diluted with 100 μL of CDCl₃ and the dilution process sequentially repeated for the next 9 samples.

OriginLab Software was used for least-squares curve fitting to the theoretical equation:

Dimerization model, Eq.1: $\delta = \delta_m + ((\delta_d - \delta_m) * (1 + ((1 - ((8 * K_a * C) + 1)^{1/2}))) / (4 * K_a * C))$ or

Infinite association model, Eq.1: $\delta = \delta_m + ((\delta_a - \delta_m) * (1 + ((1 - ((4 * K_a * C) + 1)^{1/2}))) / (2 * K_a * C))$

Where δ_m = calculated chemical shift of the monomer, δ_d = calculated chemical shift of the dimer; K_a = association constant; C = Total concentration; δ_a = average chemical shift of aggregates.

1a. ¹H NMR dilution data of 1a and 1b in CDCl₃

Table S1. Tabulated ¹H NMR dilution data for the –CH₂⁶ protons of 1a at 22 °C

Conc ^a	δ -CH ₂ ⁶
65,185	3,752
32,592	3,780
21,728	3,799
16,296	3,810
13,037	3,818
10,864	3,824
9,312	3,829
8,148	3,831
7,243	3,836
6,518	3,839
δ_m^b	3.871±0.003
δ_d^c	3.619±0.009
K_a^d	13.27±1.5
R^e	0.99915

^aTotal millimolar concentration of compound. ^bcalculated chemical shifts for the monomer. ^ccalculated chemical shifts for the dimer. ^d Association constant of dimerization (M⁻¹). ^e goodness-of-fit value

Figure S1. Concentration dependence of ^1H NMR chemical shifts for the $-\text{CH}_2^{\delta}$ proton of **1a** in CDCl_3

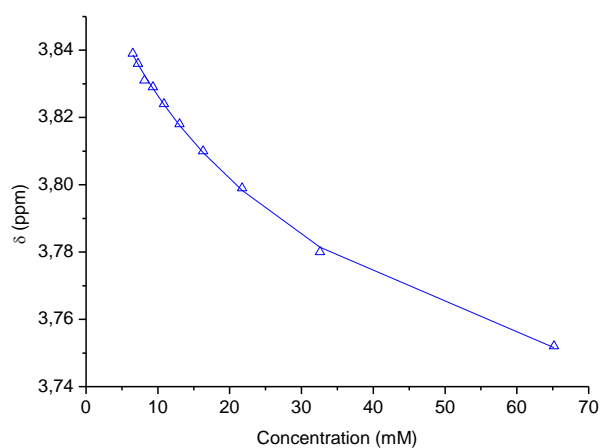
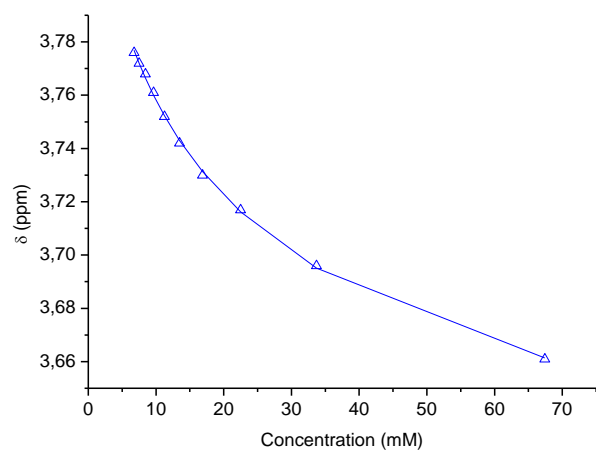


Table S2. Tabulated ^1H NMR dilution data for the $-\text{CH}_2^{\delta}$ protons of **1b** at 22 °C

Conc ^a	δ - CH_2^{δ}
67.406	3.661
33.703	3.696
22.469	3.717
16.851	3.730
13.481	3.742
11.234	3.752
9.629	3.761
8,426	3.768
7,489	3.772
6,741	3.776
δm^b	3.849 ± 0.006
δd^c	3.542 ± 0.007
Ka^d	29.95 ± 3.8
R^e	0.99931

^aTotal millimolar concentration of compound. ^b Calculated chemical shifts for the monomer. ^c Calculated chemical shifts for the dimer. ^d Association constant of dimerization (M^{-1}). ^e goodness-of-fit value

Figure S2. Concentration dependence of ^1H NMR chemical shifts for the $-\text{CH}_2^6$ proton of **1b** in CDCl_3



3. DOSY NMR data

DOSY experiments were acquired in a Bruker Ultra Shield 400 spectrometer, using the ledbpgp2s pulse program. The gradient strength (g) was the variable parameter, while Δ (diffusion time) and δ (diffusion gradient length) were kept constant during the 2D-DOSY study. Appropriate Δ and δ values were selected for each sample by optimization of the attenuation of the ^1H NMR signals in 1D-versions of the diffusing ledbpgp1s pulse program. The values of Δ and δ were 40-100 ms and 1.5-2.5 ms, respectively; depending on the sample and the solution concentration (eddy current delay was set to 5 ms in all the experiments). The pulse gradients (g) were incremented from 2 to 95% of the maximum gradient strength in a linear ramp. The diffusion dimension was processed with Bruker topspin T1/T2 software.

Table S3. Full data of diffusion coefficients (Dt , $\text{m}^2 \cdot \text{s}^{-1}$) for solutions of TMSS (1mM) and TMSO (1mM) at various concentrations (C, mM) of compounds **1a-c**, **1b**·**PF₆**, **2b** and **3b** at 295 K.

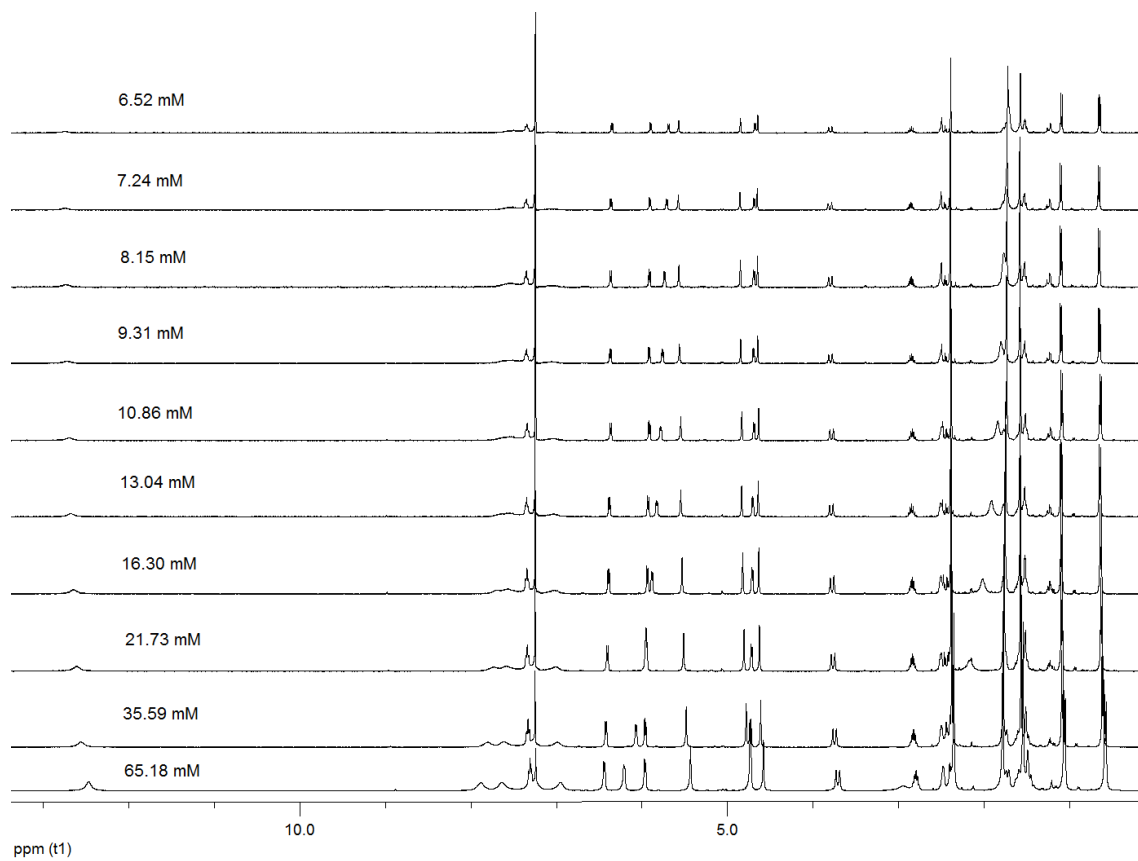
Compound	Solvent	C	$10^{10} \cdot Dt_{(s)}$	$10^{10} \cdot Dt_{(\text{TMSS})}$	$10^{10} \cdot Dt_{(\text{TMSO})}$
1a	CDCl_3	11.7	6.957	11.31	10.76
1a	CDCl_3	63.0	5.180	11.06	10.56
1a	$\text{CDCl}_3:\text{C}_6\text{D}_6^a$	5.31	6.685	10.88	10.13
1a	$\text{CDCl}_3:\text{C}_6\text{D}_6^a$	65.5	3.676	10.62	9.952
1a	CD_3OD	14.5	6.966	9.648	9.063
1a	CD_3OD	76.5	6.280	9.265	8.661
1a	$(\text{CD}_3)_2\text{CO}$	7.75	11.67	18.49	16.72
1a	$(\text{CD}_3)_2\text{CO}$	64.9	10.02	17.08	15.59
1c	$(\text{CD}_3)_2\text{CO}$	3.25	10.59	19.61	17.70
1c	$(\text{CD}_3)_2\text{CO}$	62.1	10.42	19.61	17.70
1b	CDCl_3	6.92	7.894	12.62	11.74
1b	CDCl_3	30.7	4.442	10.72	10.13
1b	CDCl_3	62.5	3.362	10.58	9.848
1b · PF₆	CDCl_3	6.10	5.473	10.71	10.17
1b PF₆	CDCl_3	52.3	4.346	10.62	10.07
2b	CDCl_3	7.38	5.739	11.03	10.46
2b	CDCl_3	81.2	4.830	11.03	10.27
3b	CDCl_3	5.82	8.542	11.49	10.66
3b	CDCl_3	87.1	7.341	10.67	9.914

^a $\text{CDCl}_3:\text{C}_6\text{D}_6 = 8:2$

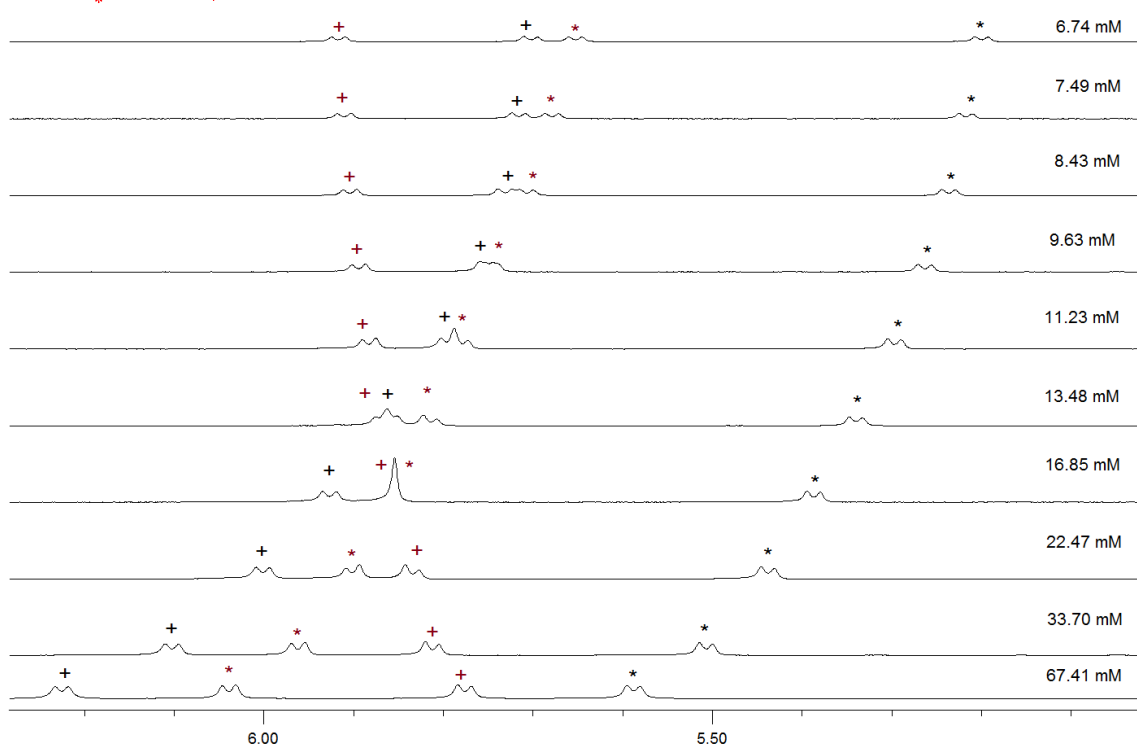
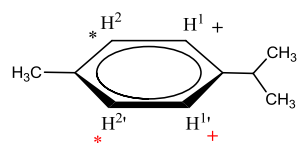
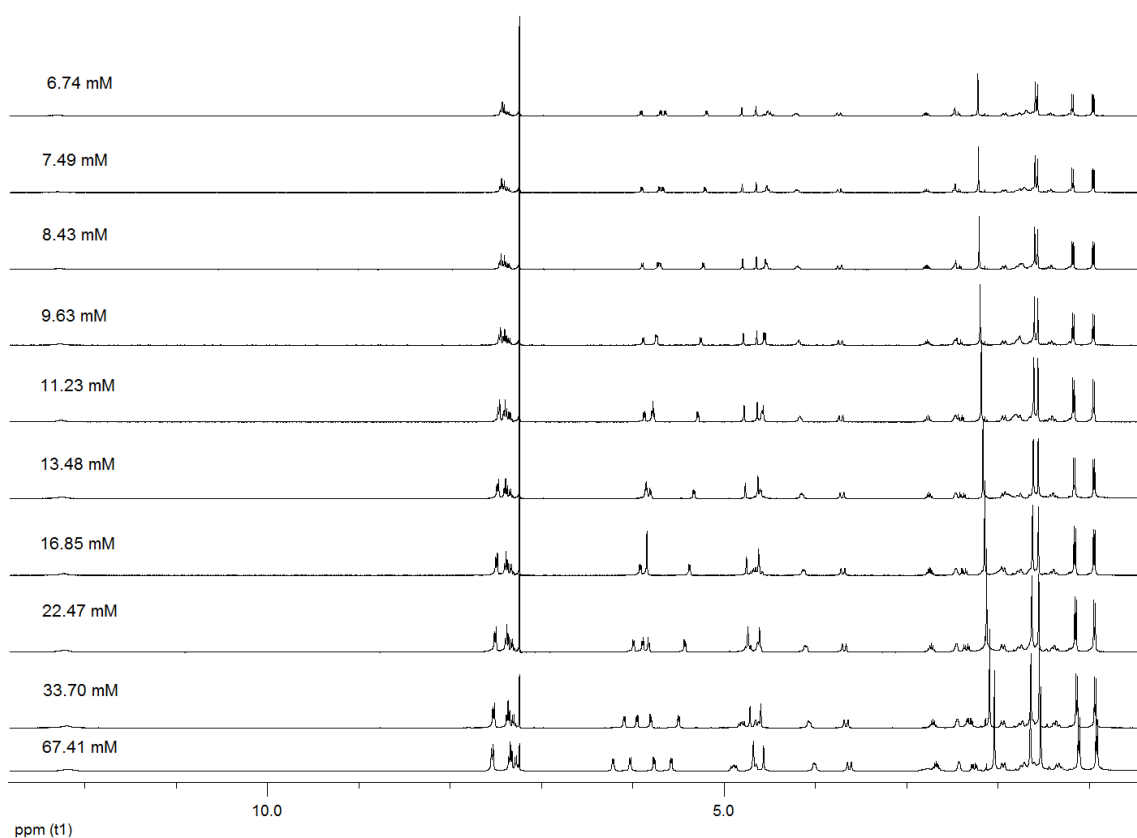
4. Selected NMR spectra

4a. Dilution Spectra

a) ^1H NMR of **1a** in CDCl_3 at different concentrations (25 °C)

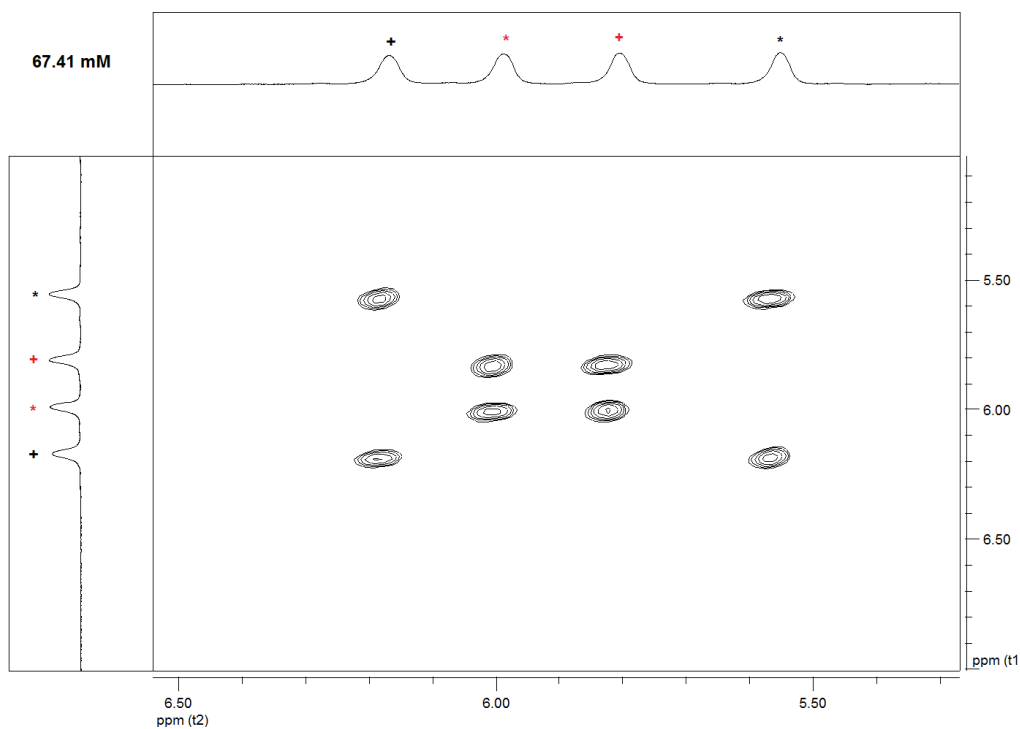


b) ^1H NMR (full and expansion) of **1b** in CDCl_3 at different concentrations (25 $^\circ\text{C}$)

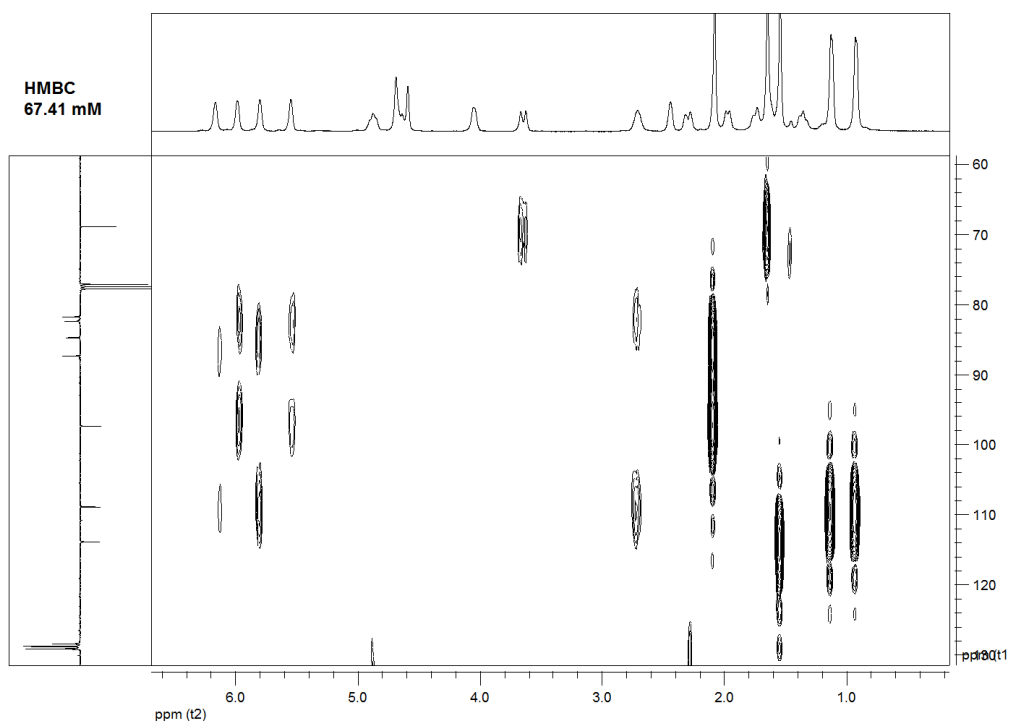


Assignment of p-cymene proton resonances of **1b** in CDCl₃

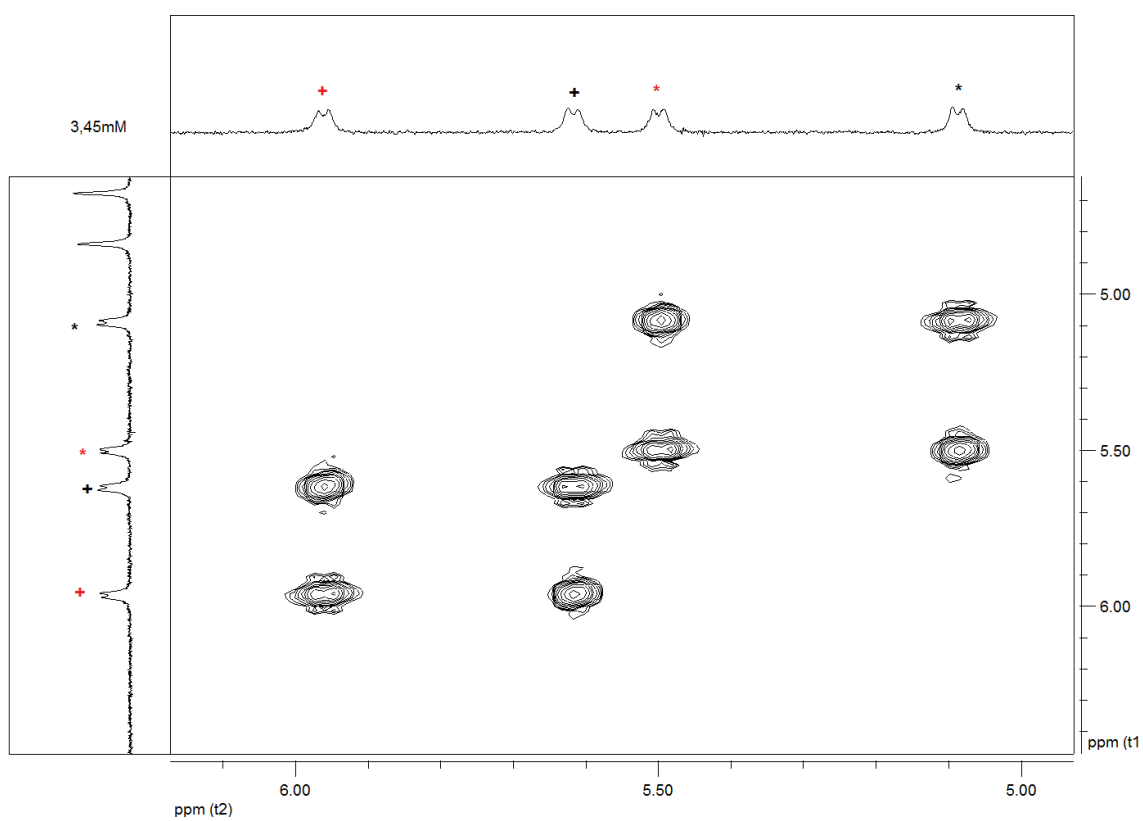
c) Expansion of ¹H-¹H COSY NMR spectrum of **1b** in CDCl₃ (67.41 mM)



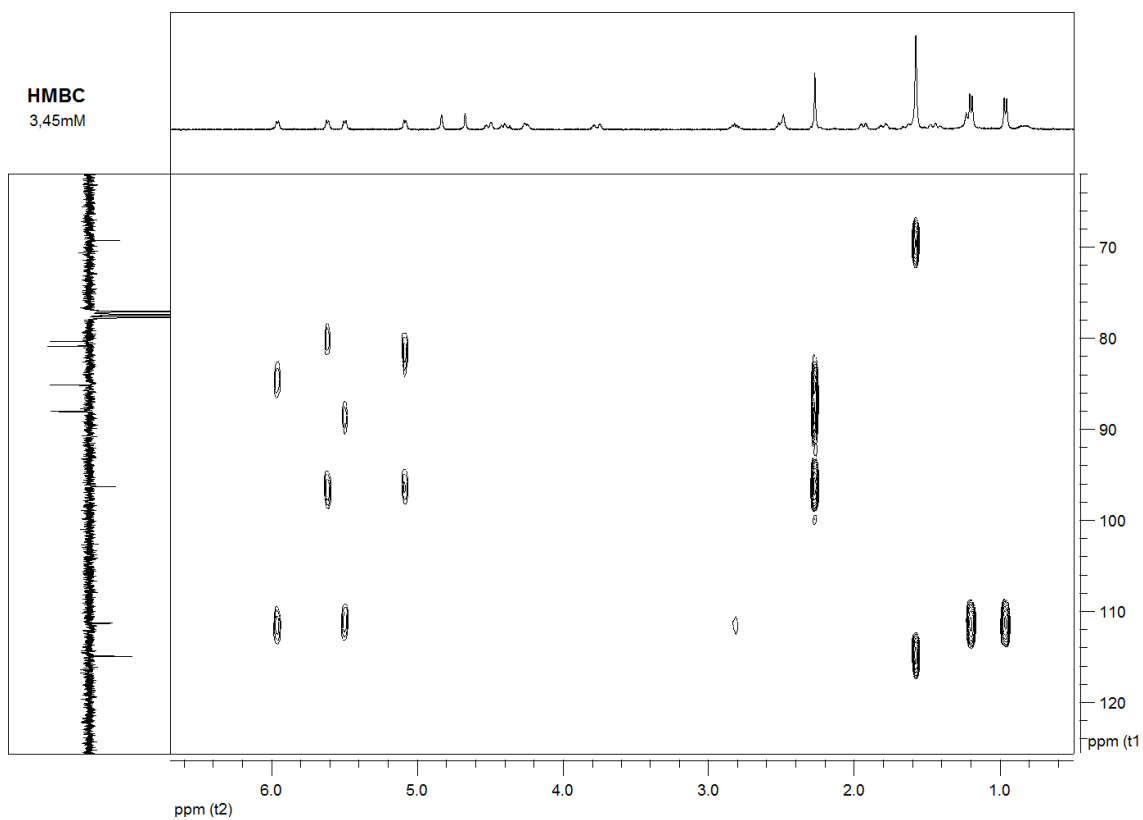
d) Expansion of ¹H-¹³C HMBC NMR spectrum of **1b** in CDCl₃ (67.41 mM)



e) Expansion of ^1H - ^1H COSY NMR spectrum of **1b** in CDCl_3 (3.45 mM)

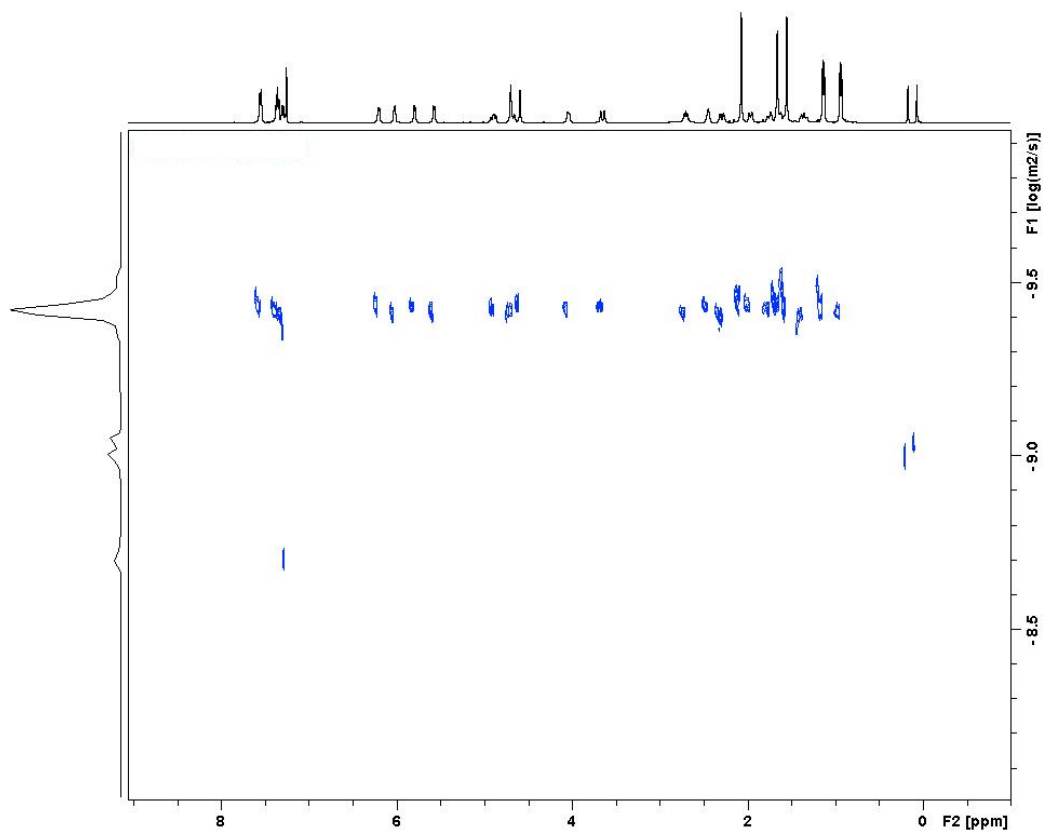


f) Expansion of ^1H - ^{13}C HMBC NMR spectrum of **1b** in CDCl_3 (3.45 mM)

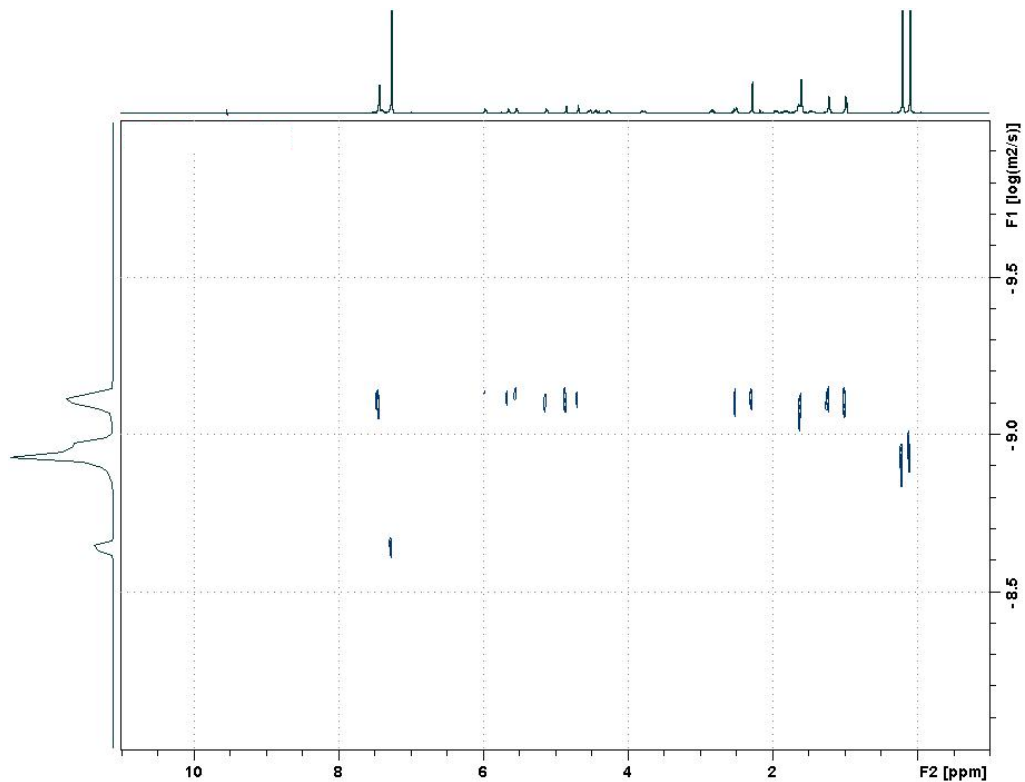


4b. DOSY spectra

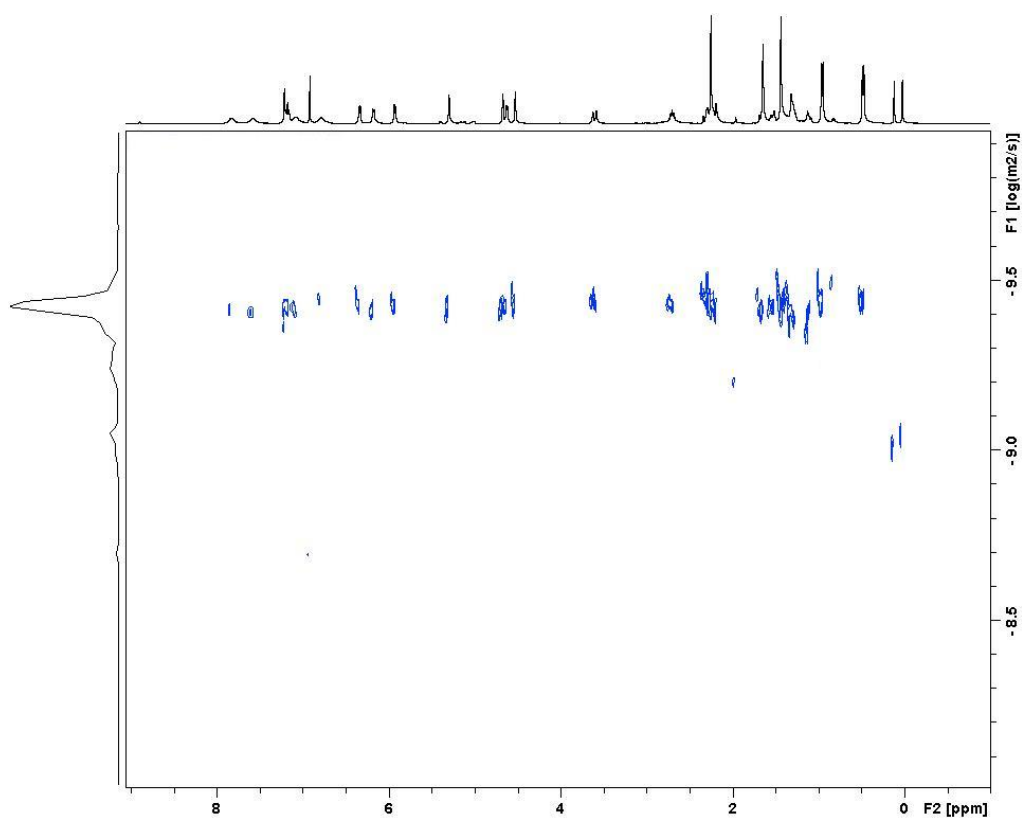
a) DOSY NMR of **1b**, (62,5mM), TMSS (1mM) and TMSO (1mM) in CDCl₃



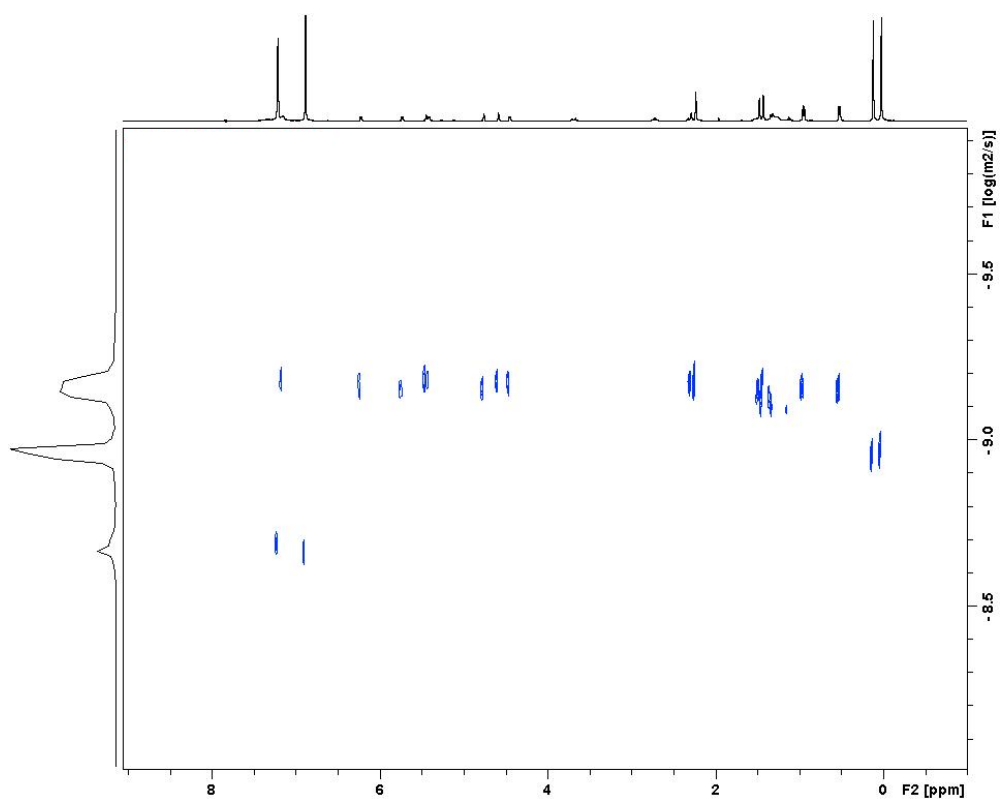
b) DOSY NMR of **1b**, (6.92 mM), TMSS (1mM) and TMSO (1mM) in CDCl₃



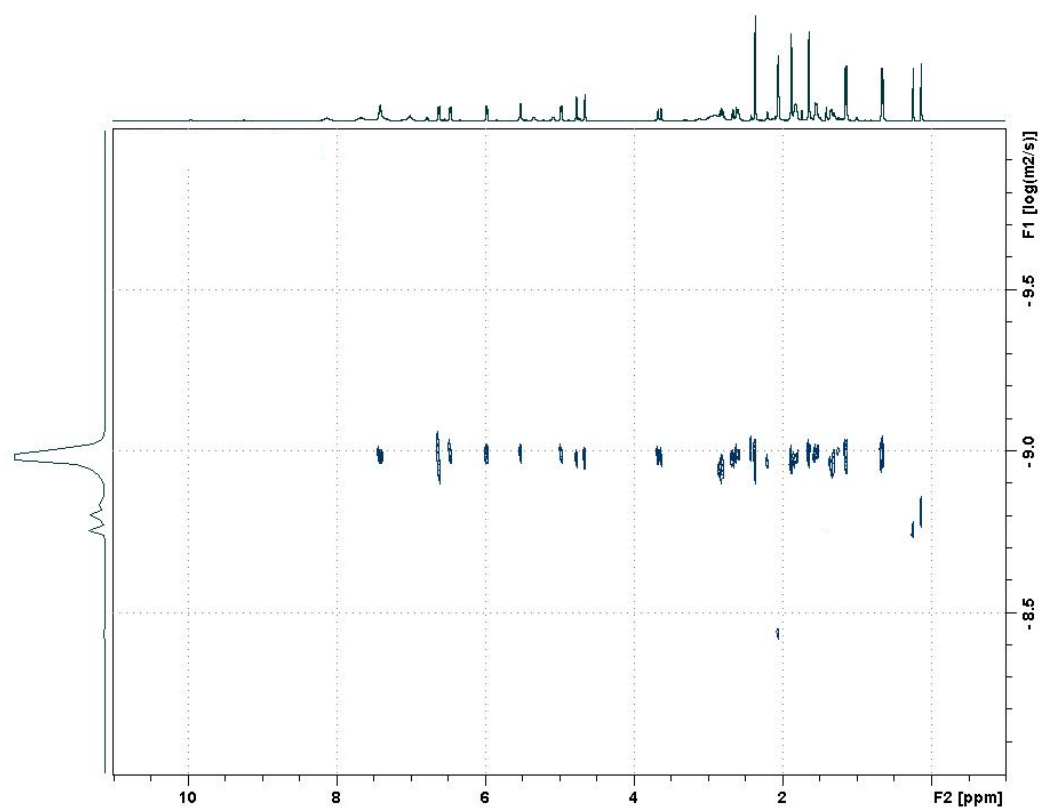
c) DOSY NMR of **1a**, (65.5 mM), TMSS (1mM) and TMSO (1mM) in C₆D₆:CDCl₃ (8:2)



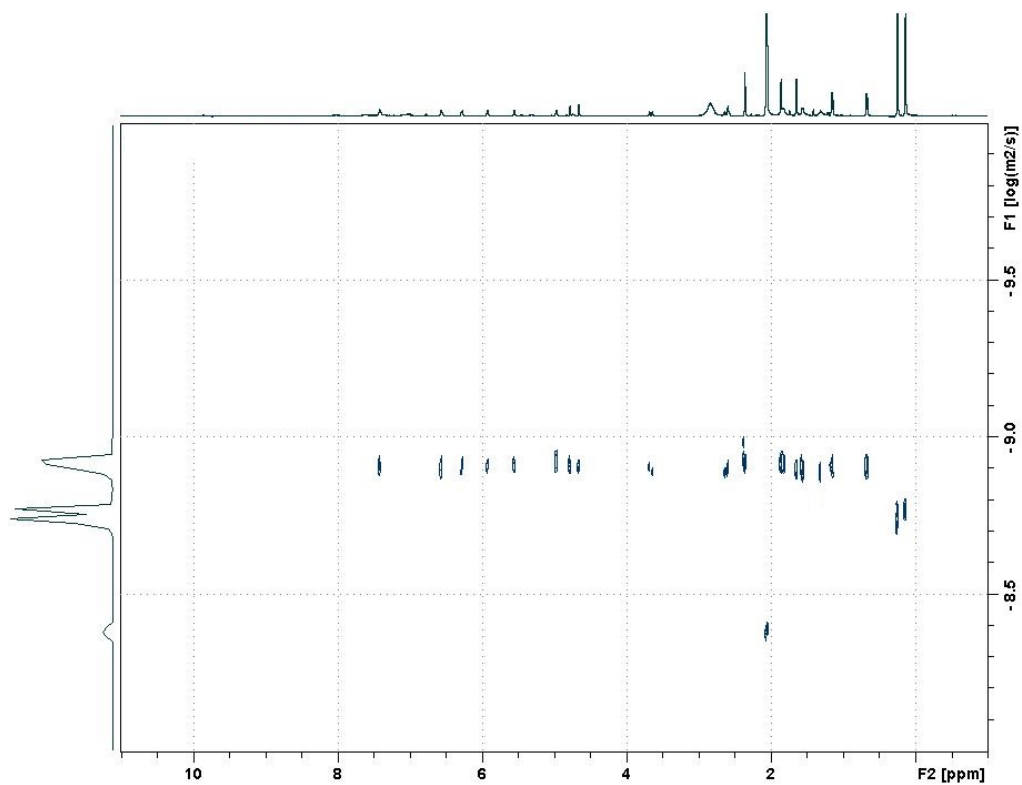
d) DOSY NMR of **1a**, (5.31 mM), TMSS (1mM) and TMSO (1mM) in C₆D₆:CDCl₃ (8:2)



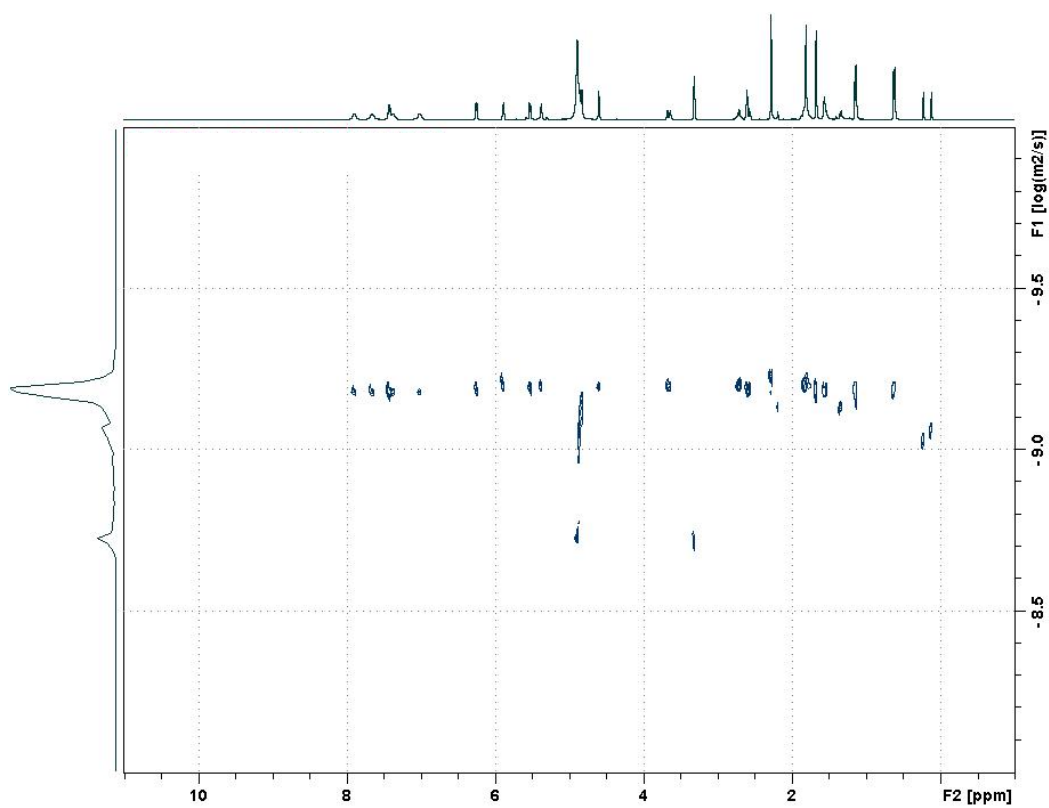
e) DOSY NMR of **1a**, (64.9 mM), TMSS (1mM) and TMSO (1mM) in acetone- d_6



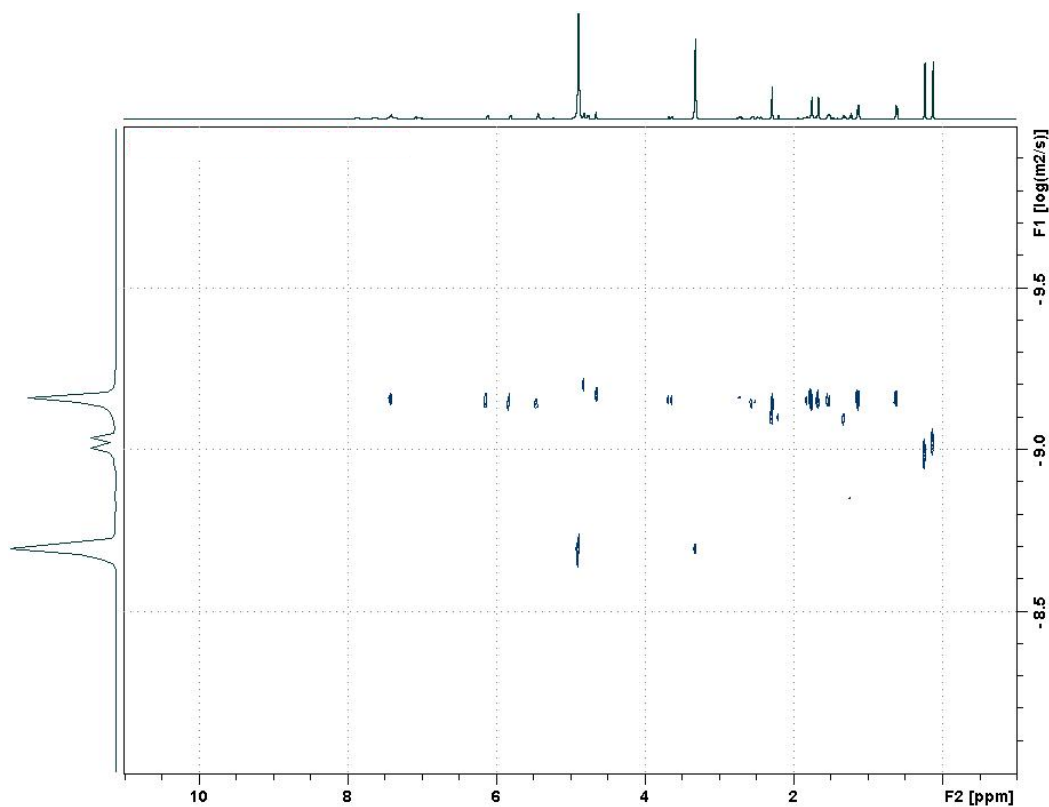
f) DOSY NMR of **1a**, (7.75 mM), TMSS (1mM) and TMSO (1mM) in acetone- d_6



g) DOSY NMR of **1a**, (76.5 mM), TMSS (1mM) and TMSO (1mM) in methanol-d₄

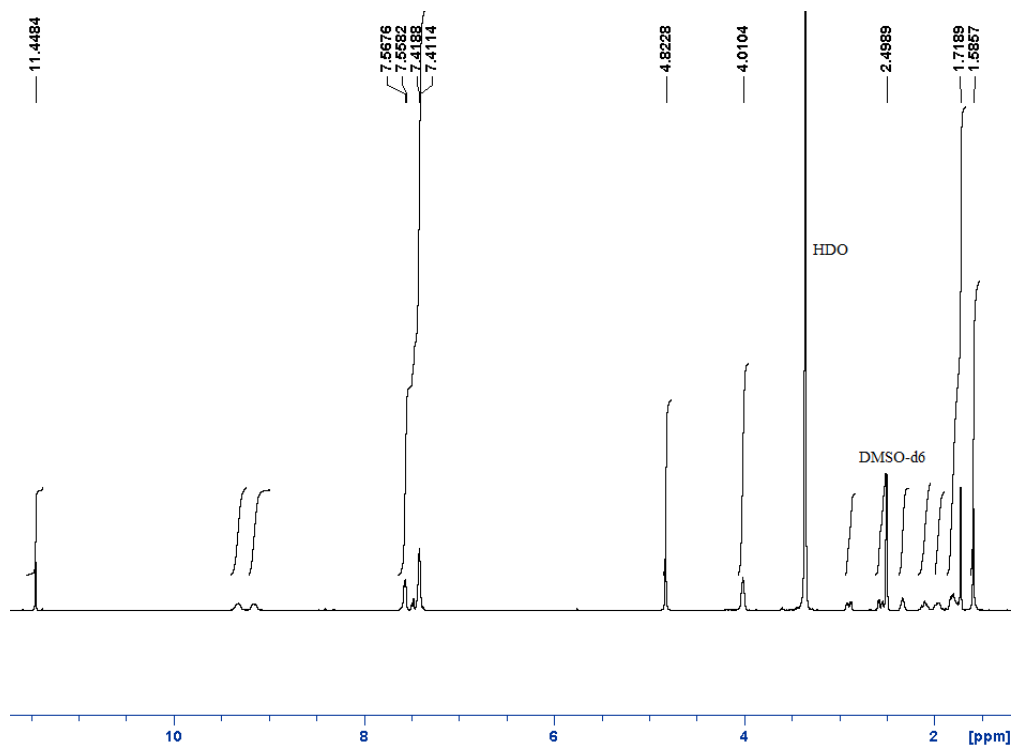


h) DOSY NMR of **1a**, (14.5 mM), TMSS (1mM) and TMSO (1mM) in methanol-d₄

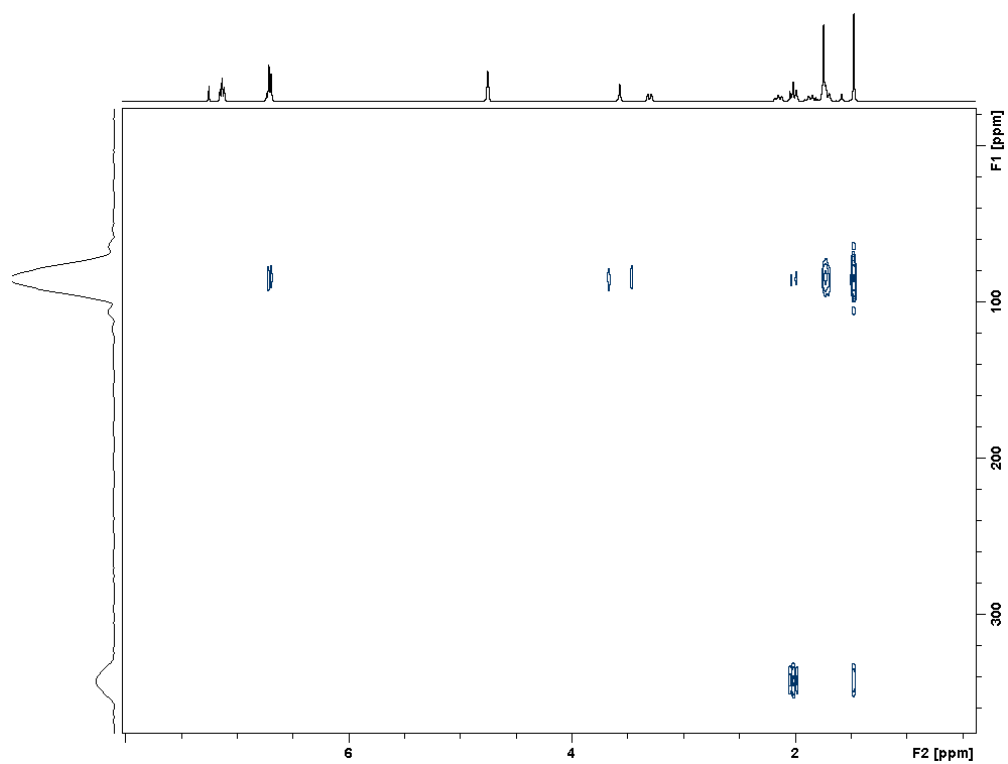


4c. Selected NMR spectra of amino-oxime compounds a, b, b-HCl, c and corresponding ruthenium compounds 1a-c, 2b, 3a and 3b.

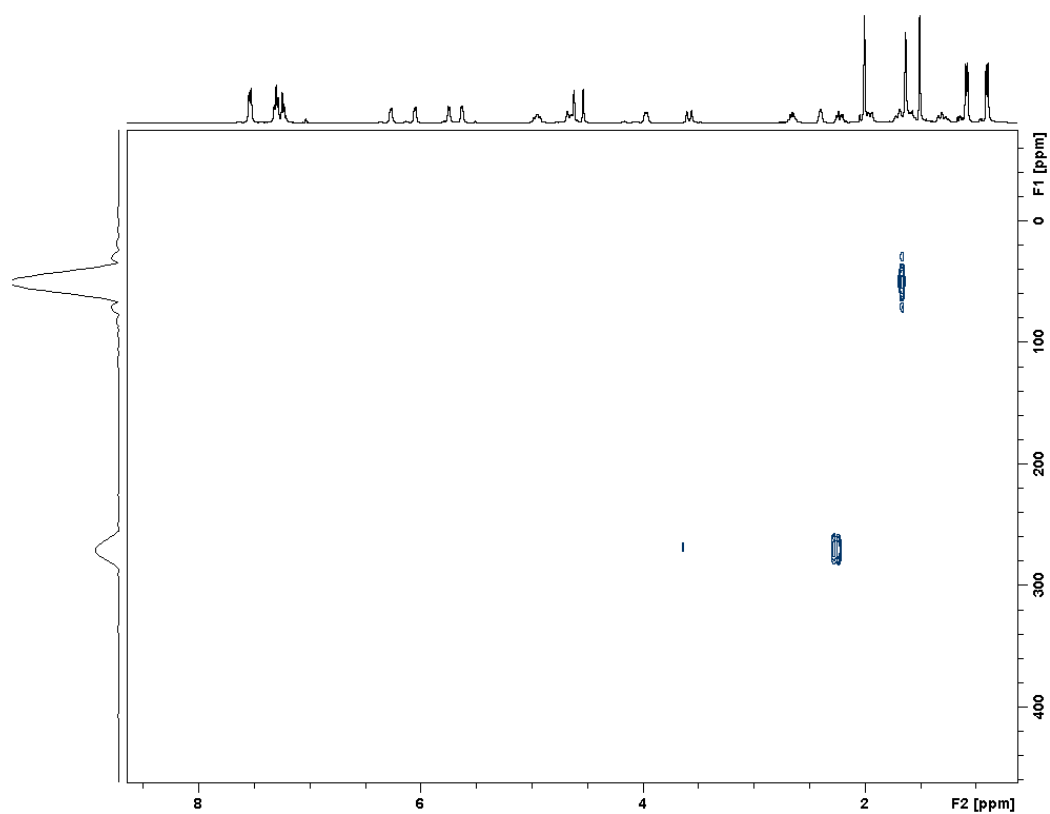
1. ^1H NMR of (2S,5R)-[ClH·NHBn,NOH] (**b-HCl**) in $\text{DMSO-}d_6$



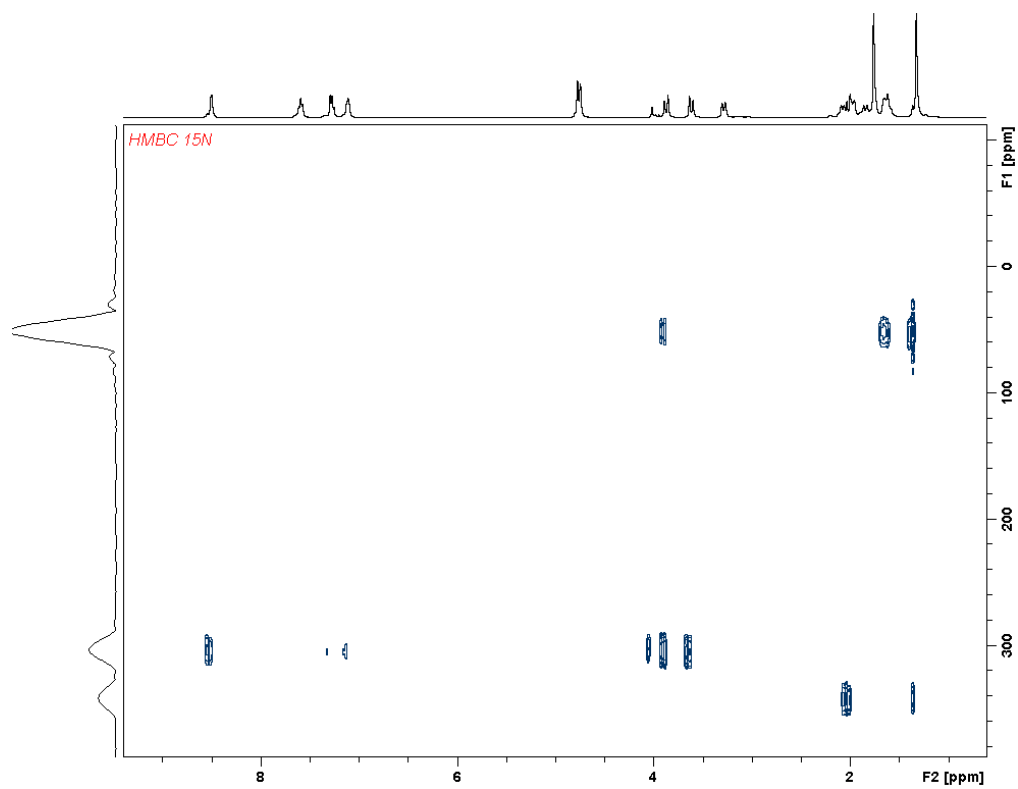
2. $^1\text{H-}^{15}\text{N}$ HMBC NMR of (2S,5R)-[NHPH,NOH] (**a**) in CDCl_3



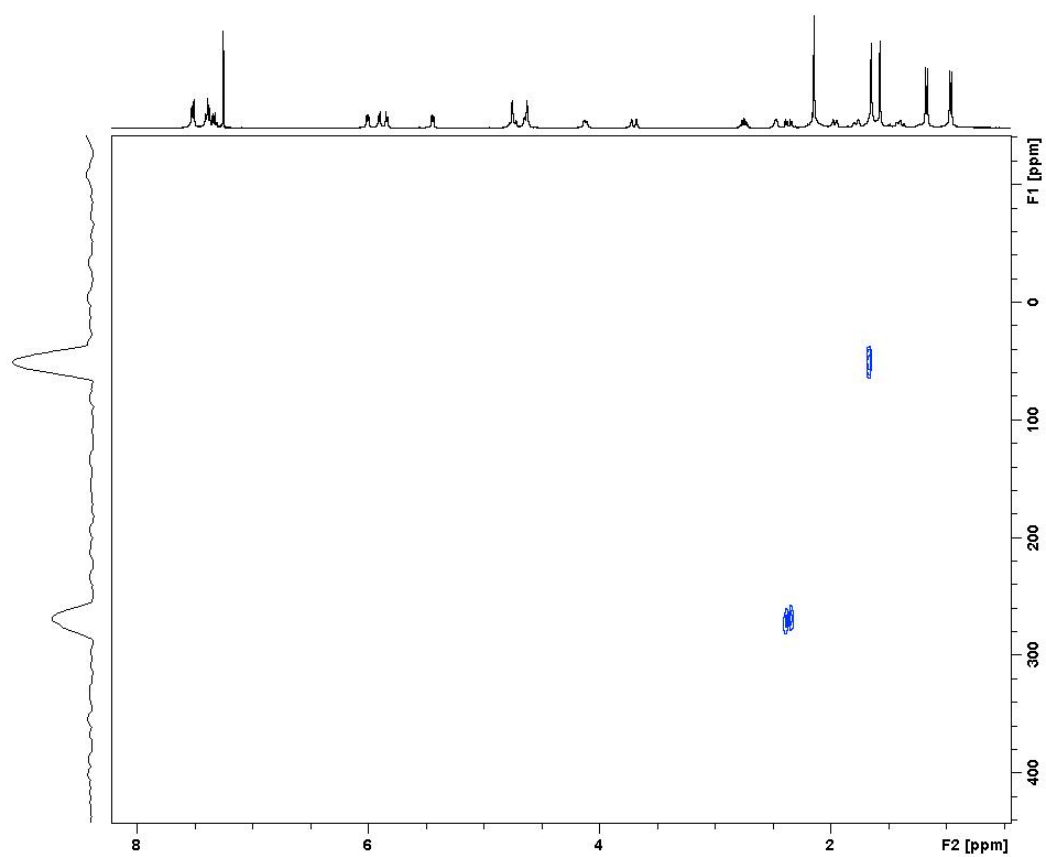
3. ^1H - ^{15}N HMBC NMR of (2S,5R)-[NHBn,NOH] (**b**) in CDCl_3



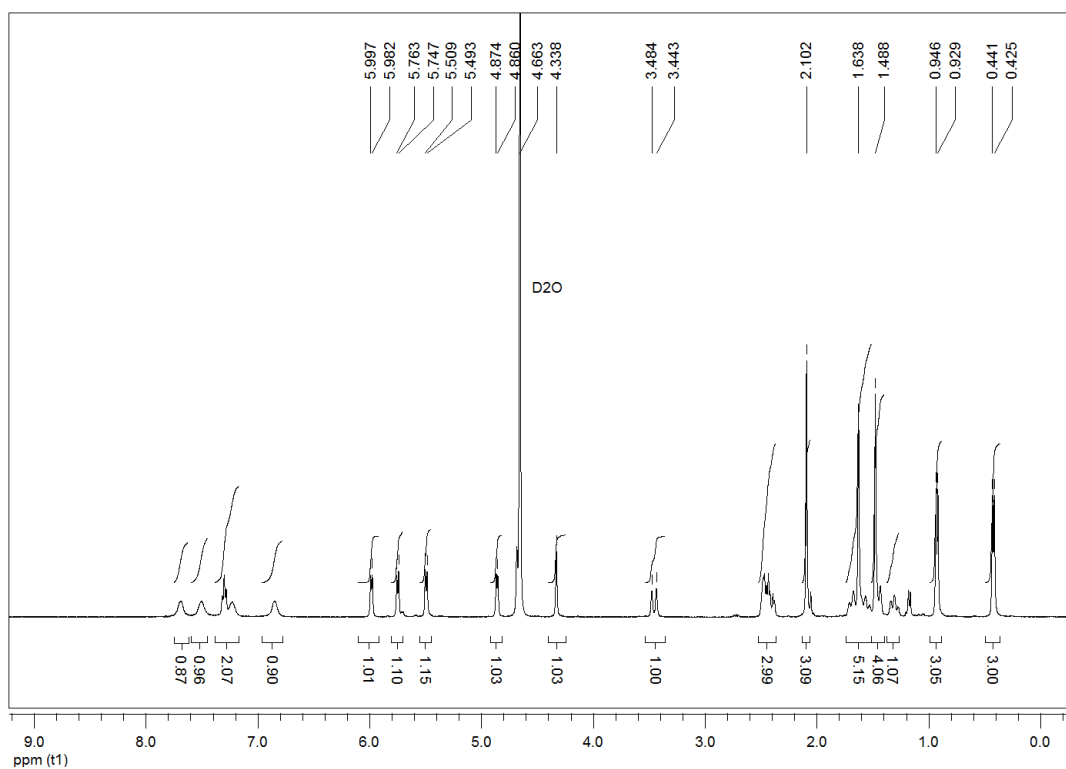
4. ^1H - ^{15}N HMBC NMR of (2S,5R)-[NH(2-pic),NOH] (**c**) in CDCl_3



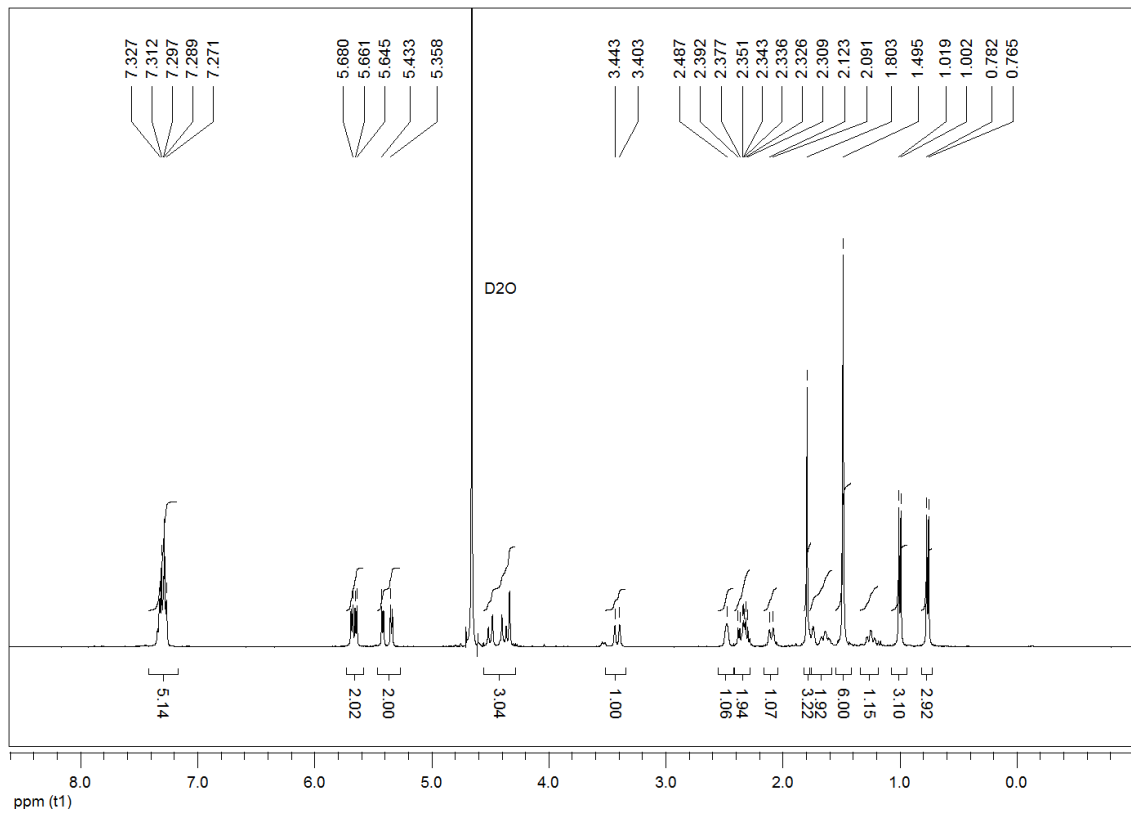
5. ^1H - ^{15}N HMBC NMR of **1b** in CDCl_3



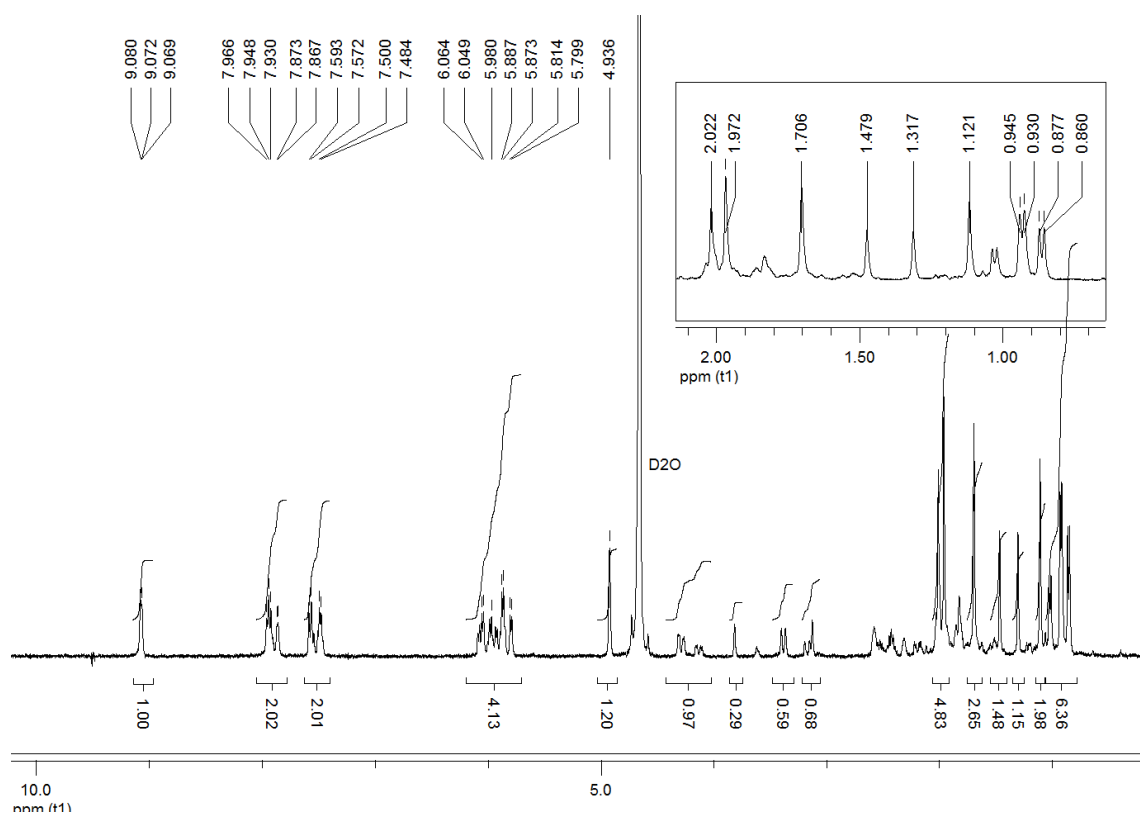
6. ^1H NMR of **1a** in D_2O



7. ^1H NMR of **1b** in D_2O

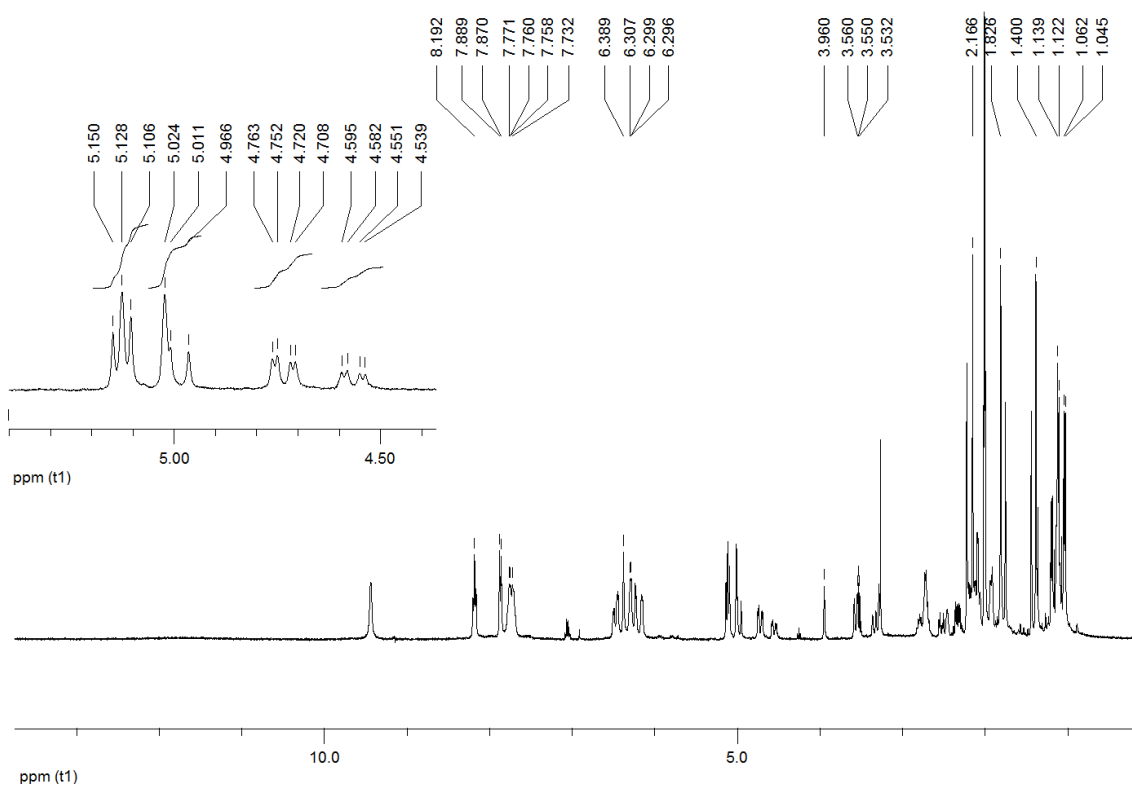


8. ^1H NMR of **1c** in D_2O

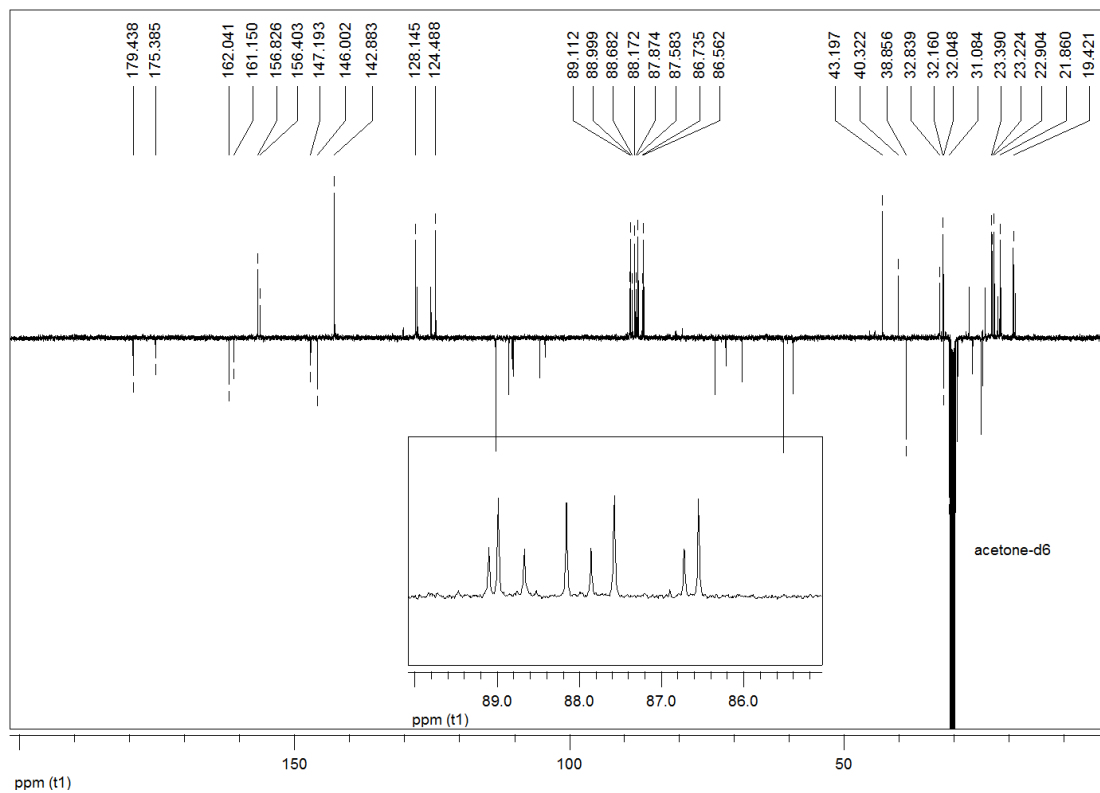


NMR spectra of 1c in acetone-d₆

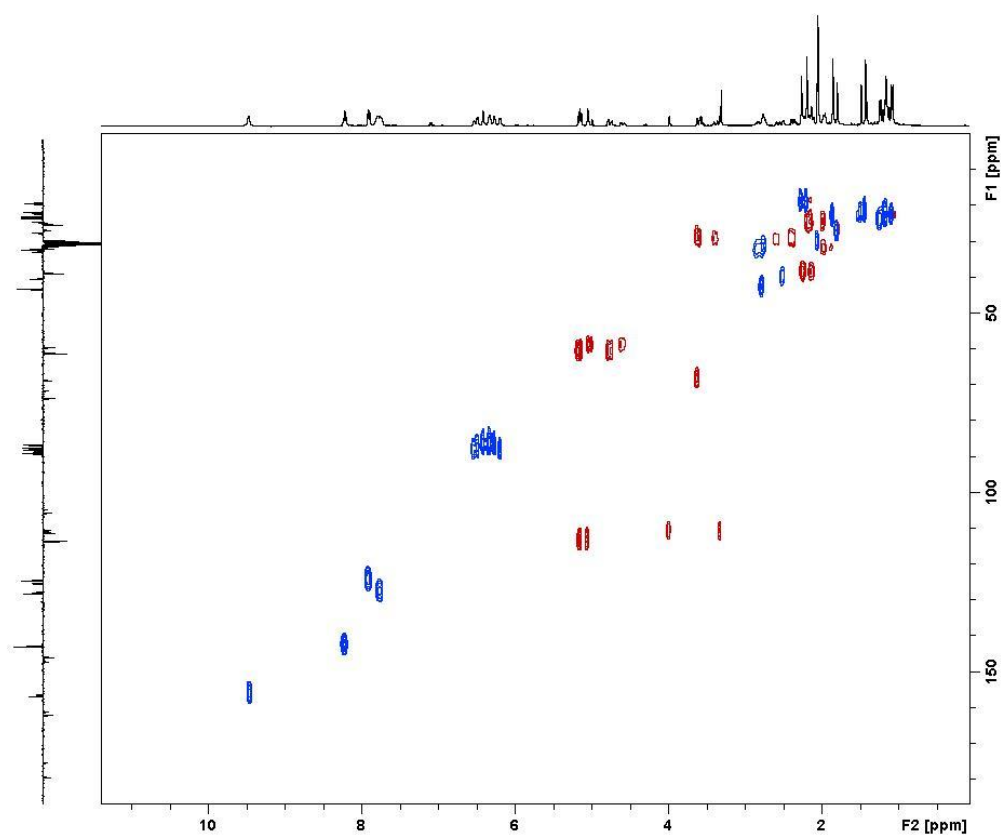
1. ¹H NMR of 1c



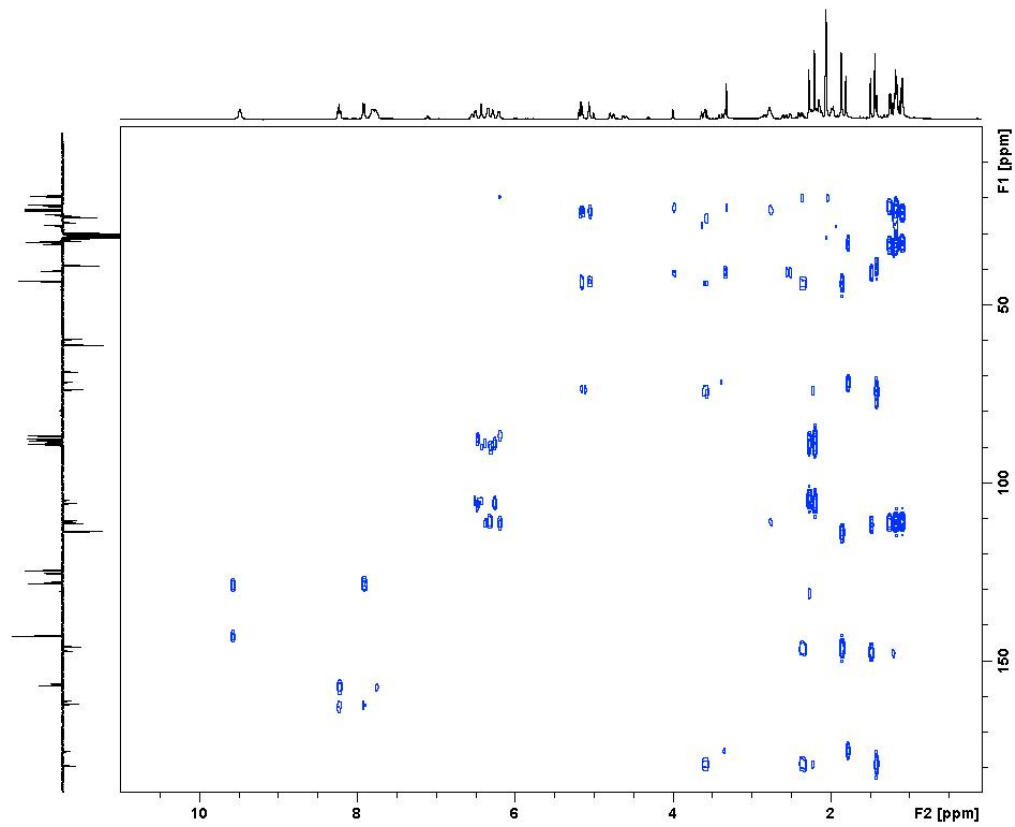
2. ¹³C APT-NMR of 1c



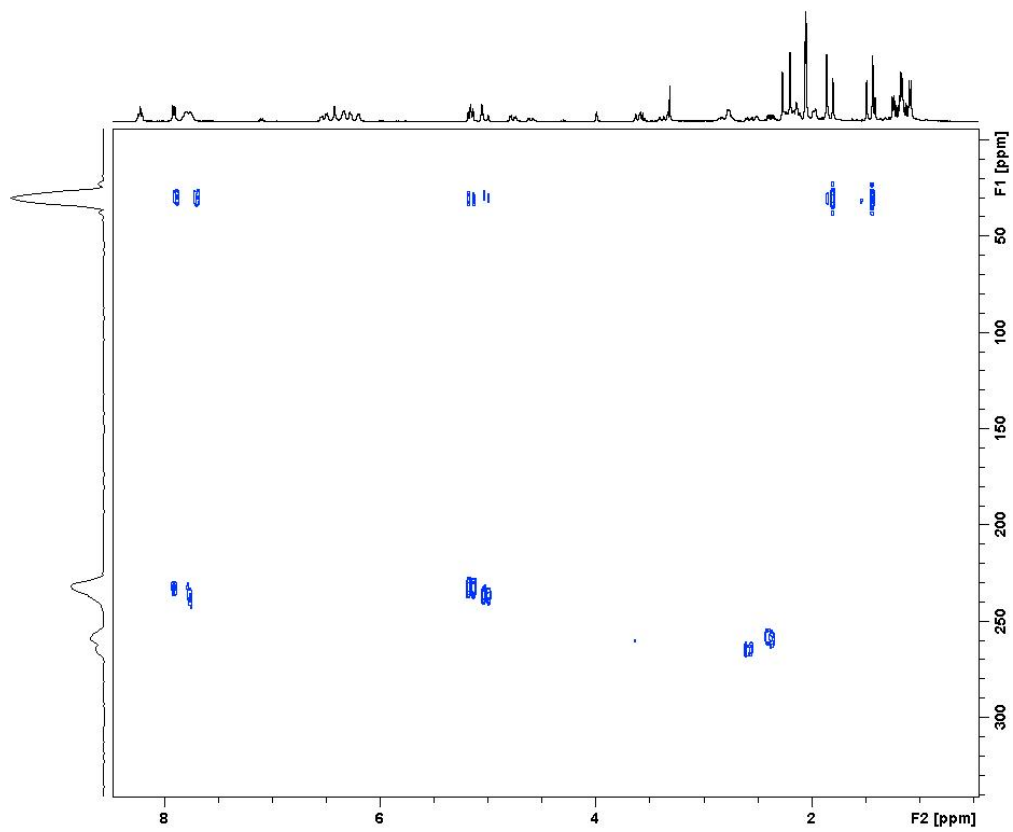
3. ^1H - ^{13}C CHSQC NMR of **1c**



4. ^1H - ^{13}C HMBC NMR of **1c**

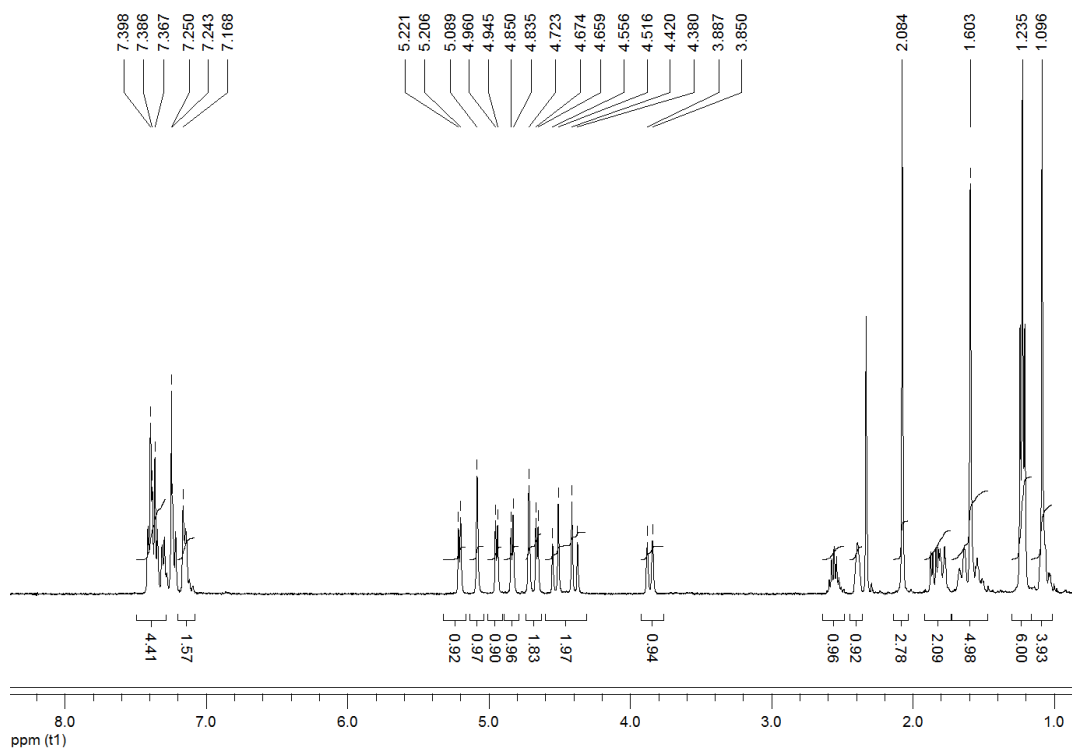


5. ^1H - ^{15}N HMBC NMR of **1c**

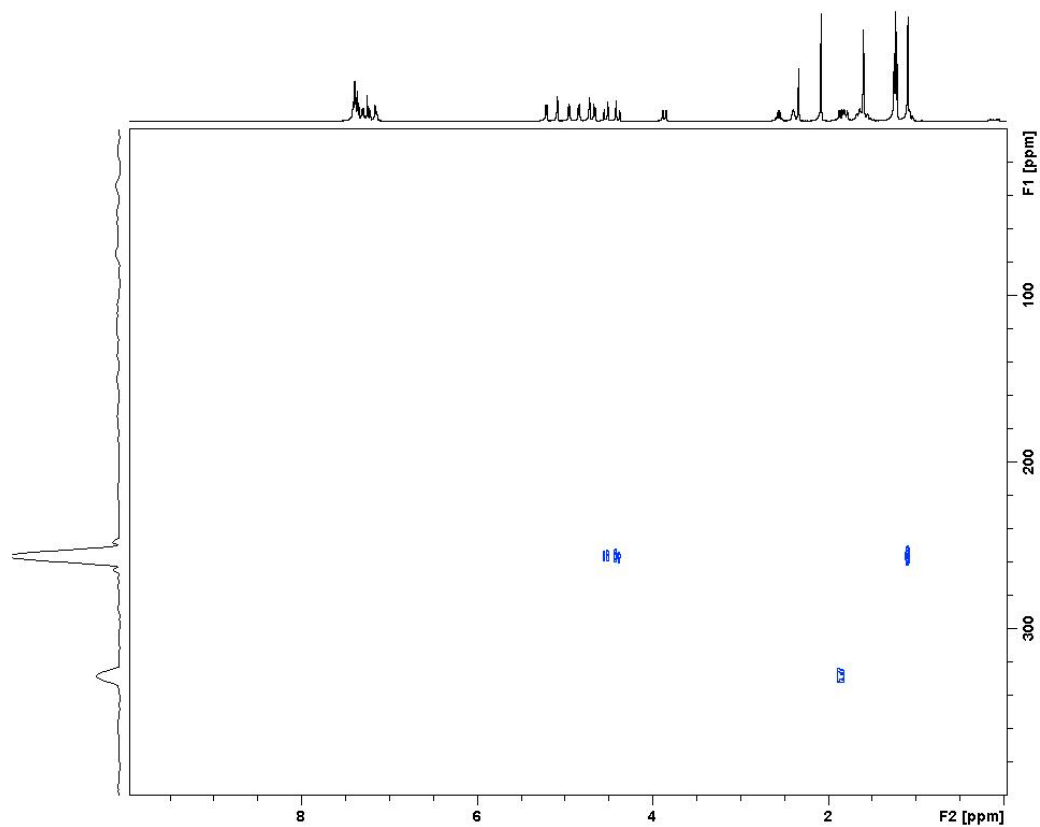


NMR spectra of 3b in CDCl₃

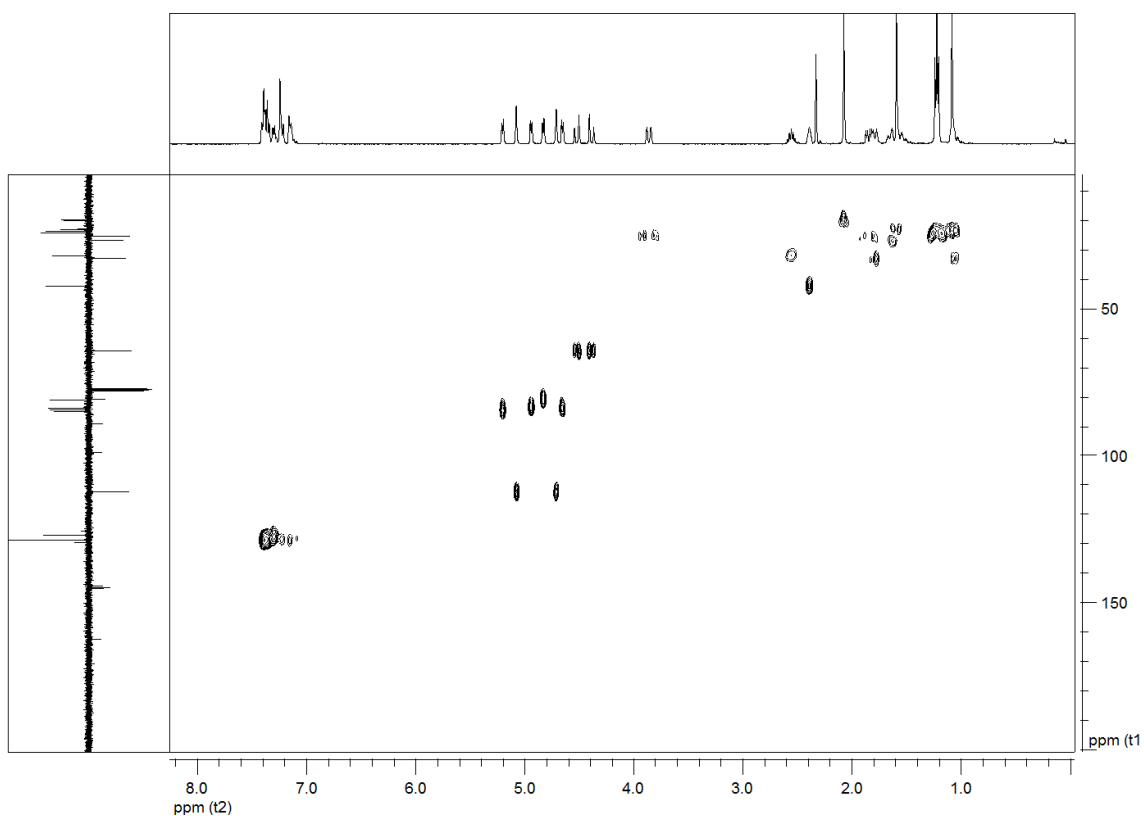
1. ¹H NMR of 3b



2. ¹H-¹⁵N HMBC NMR of 3b

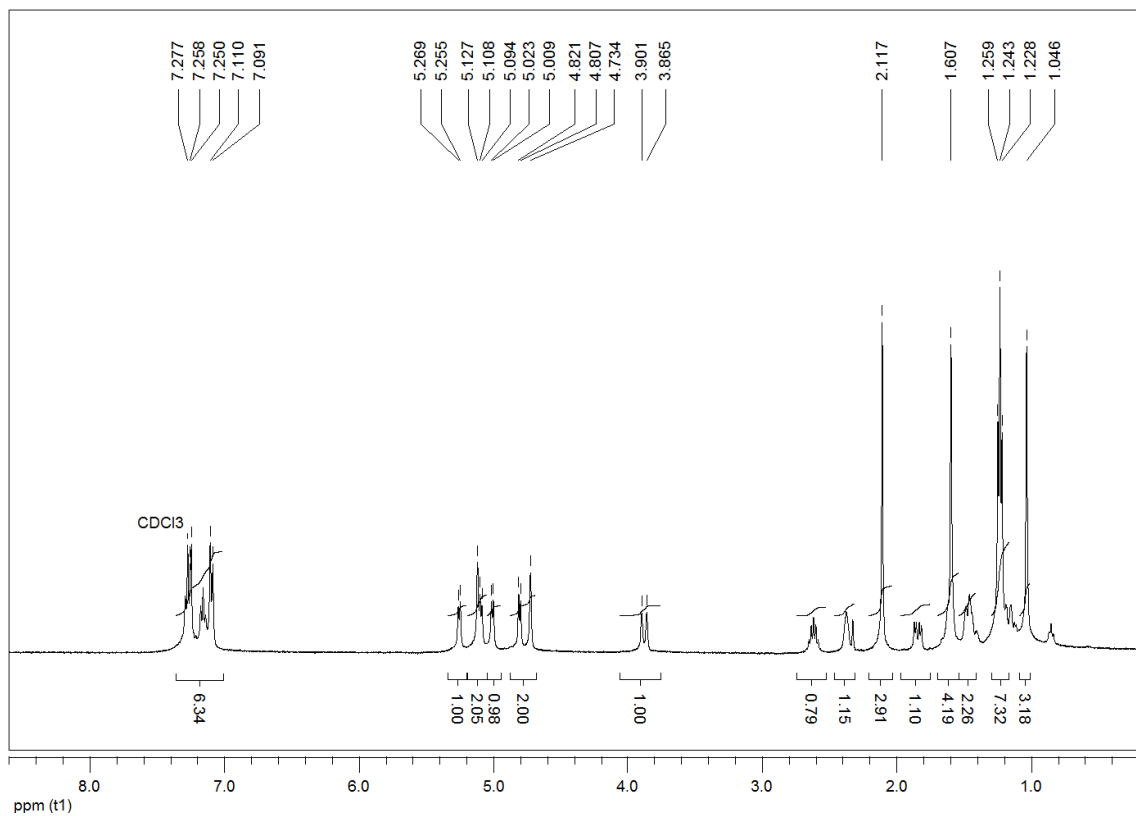


1. ^{13}C - ^1H HSQC NMR of **3b**

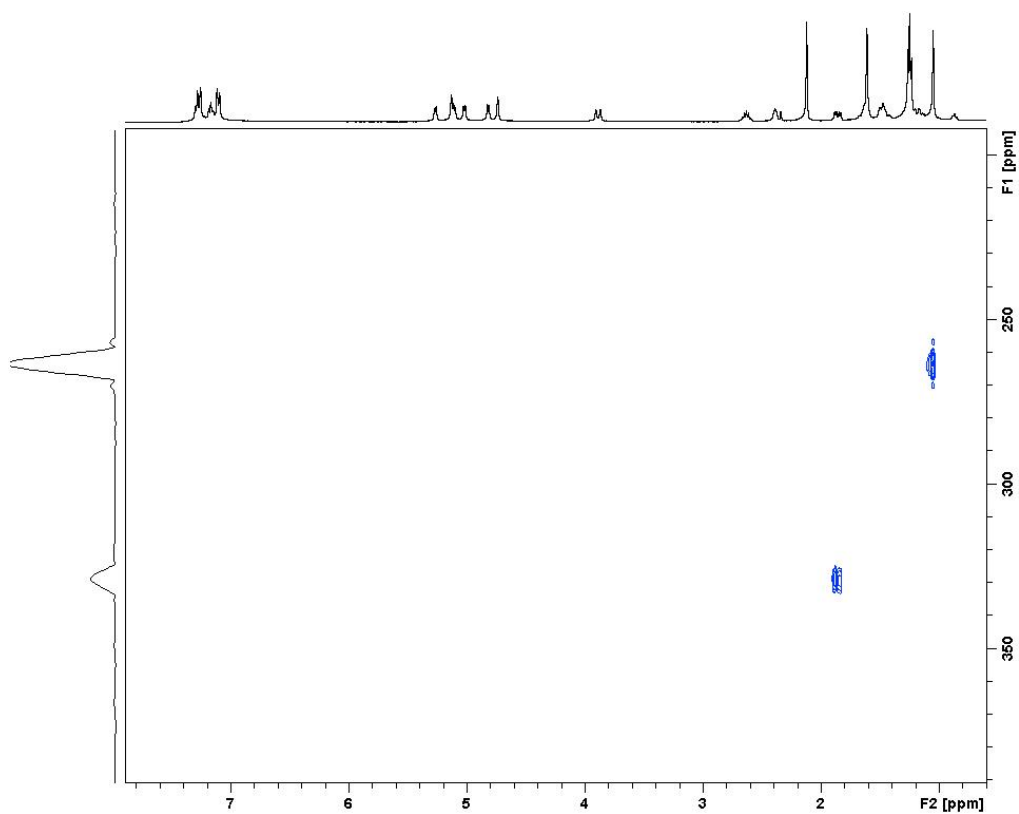


NMR spectra of **3a** in CDCl_3

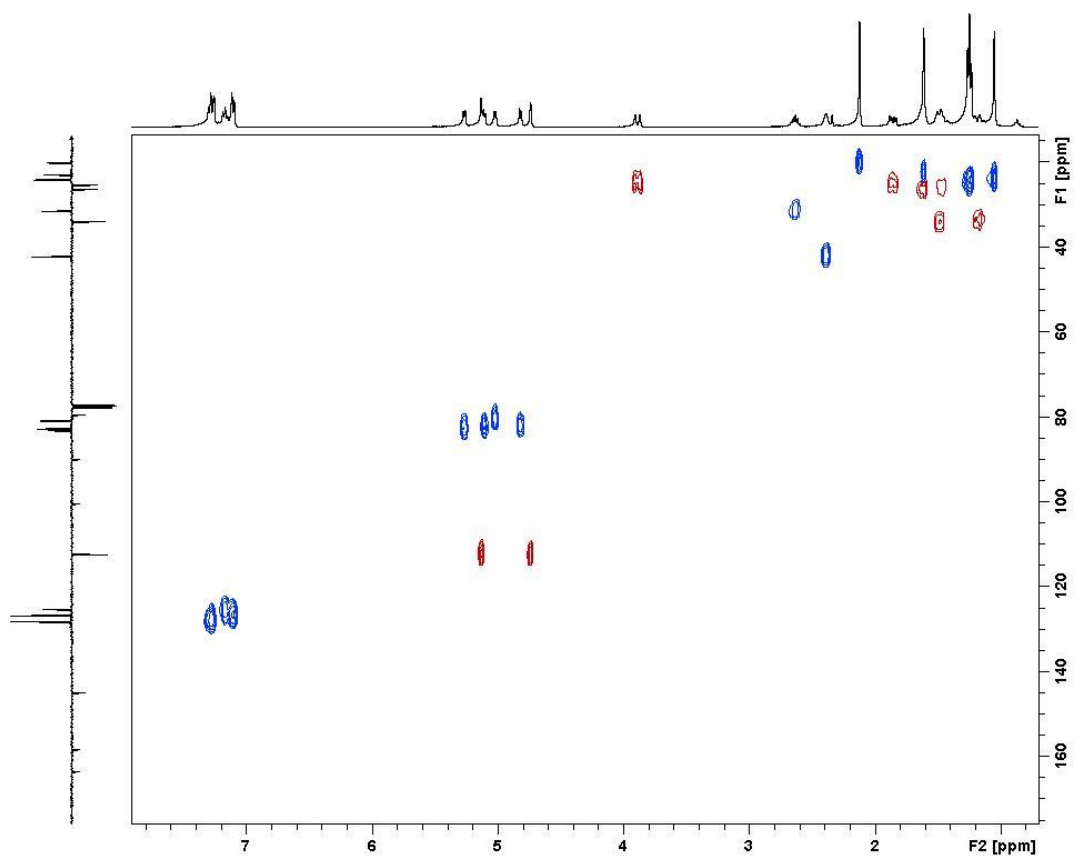
1. ^1H NMR of **3a**.



2. ^1H - ^{15}N HMBC NMR of **3a**.



3. ^1H - ^{13}C HSQC NMR of **3a**.



5. NMR experiments under physiological conditions.

Phosphate buffer saline (PBS) was prepared according to Cold Spring Harbor Protocols (<http://cshprotocols.cshlp.org/content/2006/1/pdb.rec8247>) using NaCl, KCl, Na₂HPO₄ and KHPO₄ in D₂O. Adjustment of pH to 7.40 was carried out using a DCl solution in D₂O (0.01M) with the help of a HANNA HI208 pHmeter. Compounds **1a-1c** were then dissolved in 2500 μL of the freshly prepared PBS (6-9 mM of ruthenium compounds), final pH measured (7.36-7.38) and time-dependent ¹H NMR spectra of 500 μL aliquots of final solutions were carried out at 25 °C.

Figure S3. ¹H NMR spectrum (25 °C) of: **A)** **1b** in D₂O, **B)** Phosphate buffer saline (PBS) in D₂O, **C)** **1b** in PBS solution after 24h at r.t. **D)** **1b** in PBS solution after 48h at r.t. **E)** **1b** in PBS solution after 72 h at r.t., **F)** **1b** in PBS solution after 5 days at room temperature.

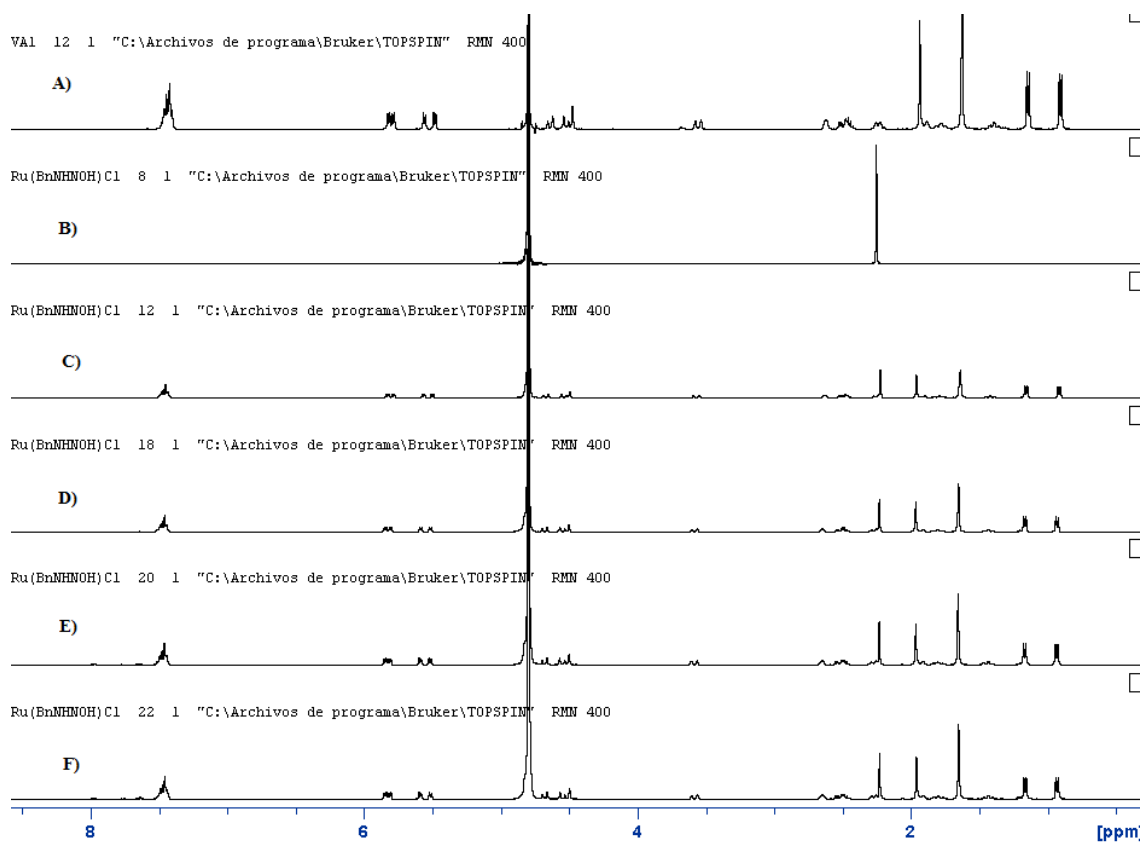
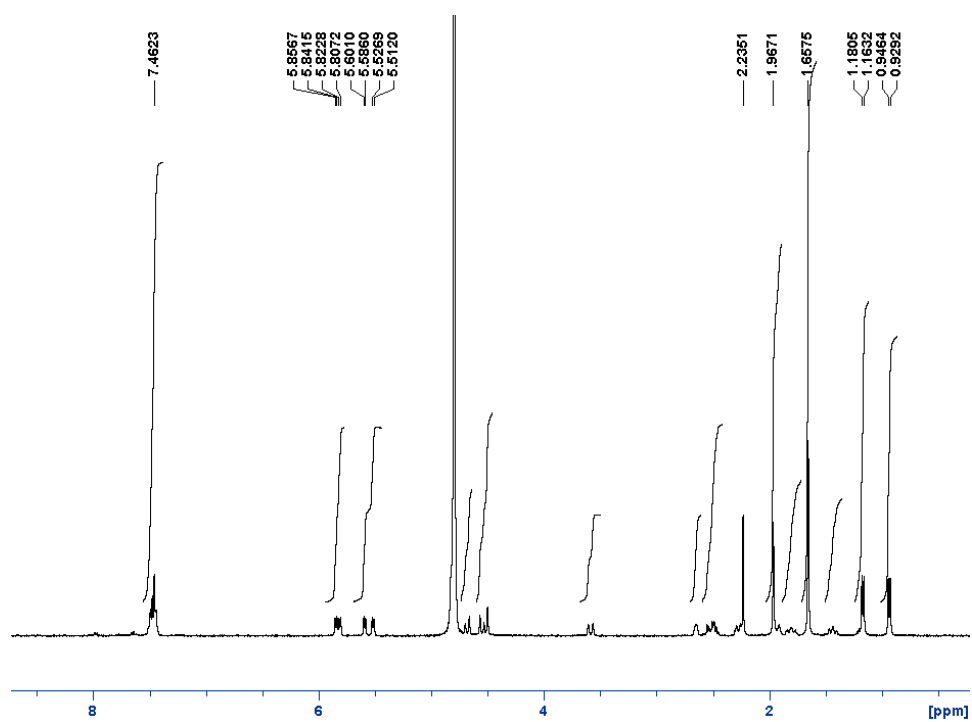
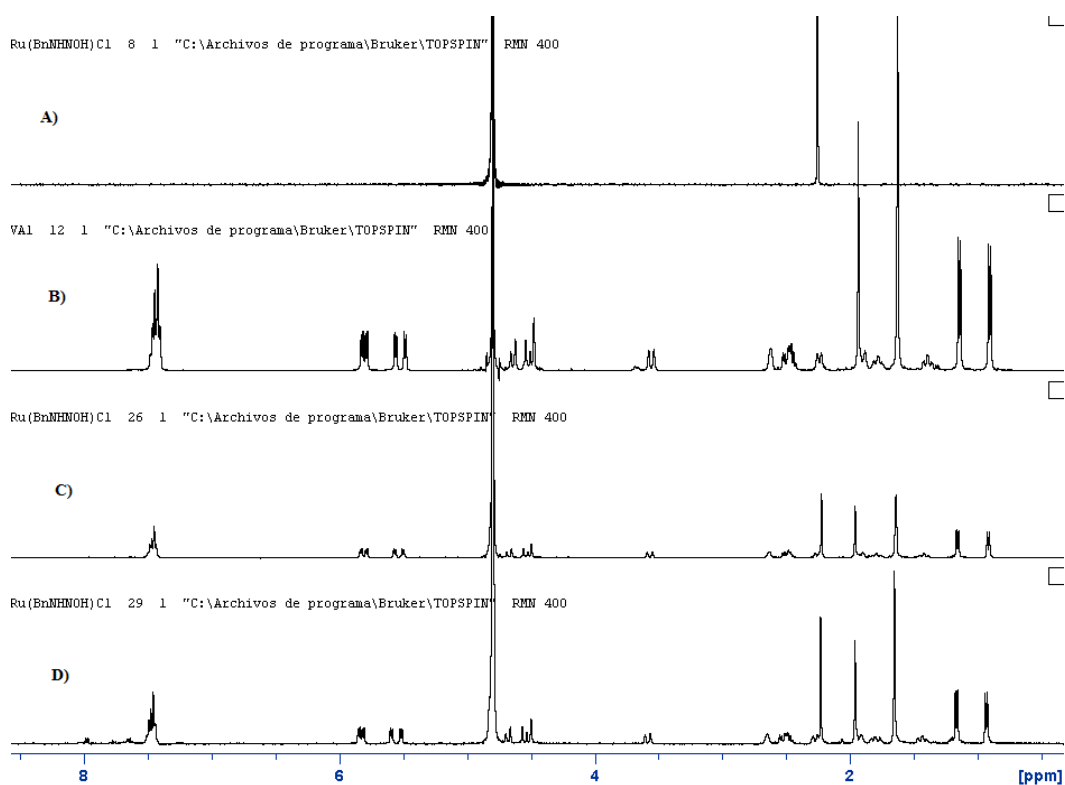


Figure S4. Full ^1H NMR spectrum (25 $^\circ\text{C}$) of **1b** in PBS solution after 72 h at r.t.



A 500 μL aliquot of **1b** in PBS (pH = 7.4), prepared as described above, was warmed up to ca. 36 $^{\circ}\text{C}$, and time-dependent ^1H NMR spectra were carried out at 25 $^{\circ}\text{C}$. Spectra of this experiment are shown in Figure S5.

Figure S5. ^1H NMR spectrum (25 $^{\circ}\text{C}$) of: **A)** Phosphate buffer saline (PBS) in D_2O , **B)** **1b** in D_2O , **C)** **1b** in PBS solution after 24h at 36 $^{\circ}\text{C}$ **D)** **1b** in PBS solution after 72h at 36 $^{\circ}\text{C}$.



6. *In vitro* assays.

Figure S6. Effect of derivatives A) **1a**, B) **1b** and C) **1c** on PC3 cells viability, compared to that of ammonium-oxime compounds **a-HCl** and ruthenium dimer $[(\eta^6\text{-p-cymene})\text{RuCl}_2]_2$. Cells were treated with increasing doses of organic and ruthenium compounds for 3 hours. Cell viability was measured by means of MTT assay. The results are expressed as a percentage of live cells compared to control. Data are the mean \pm S.E.M. of at least three experiments. *, $P < 0.05$; **, $P < 0.01$; ***, $P < 0.001$ versus Control.

