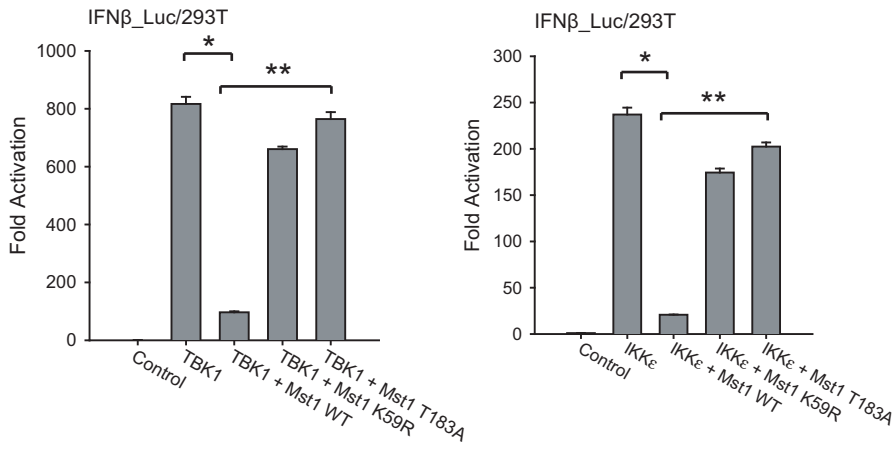


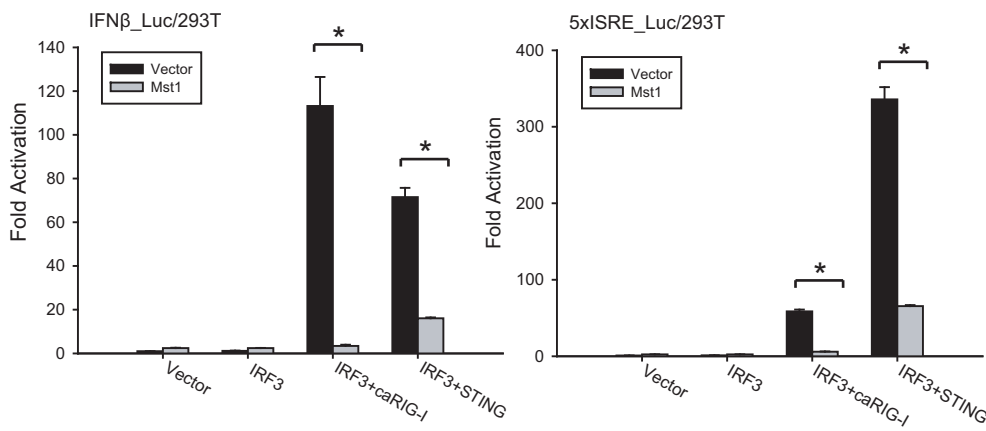
Supplementary Materials:

- 1) Supplemental Figures 1 - 4;
- 2) Supplemental Figure Legends;
- 3) Oligo sequence of qPCR.

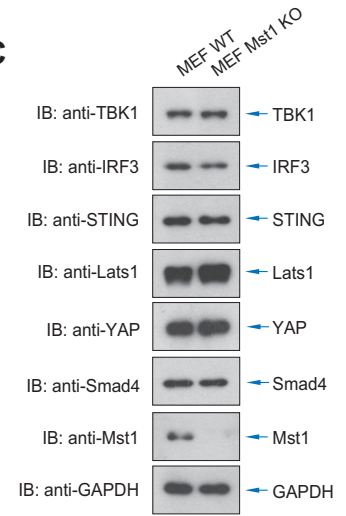
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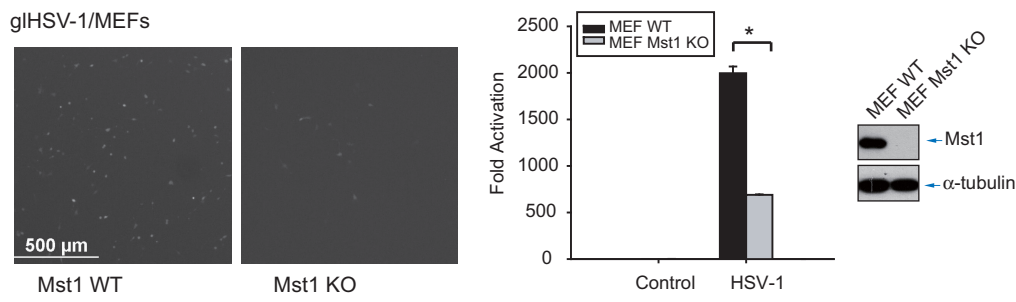
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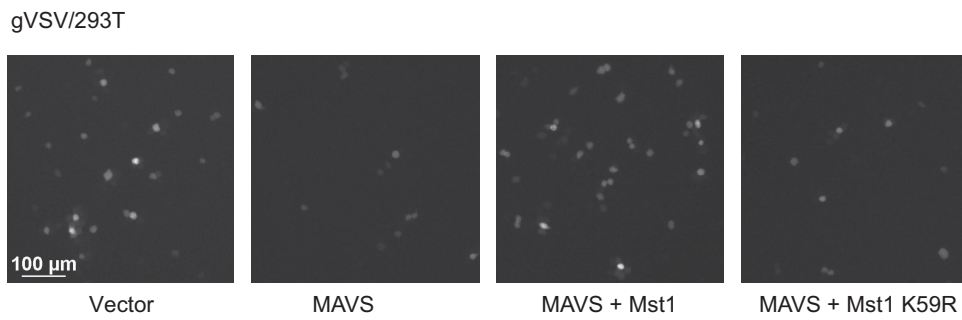
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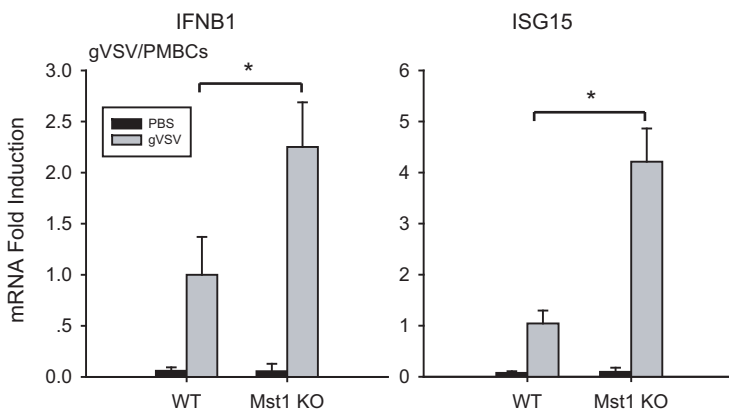
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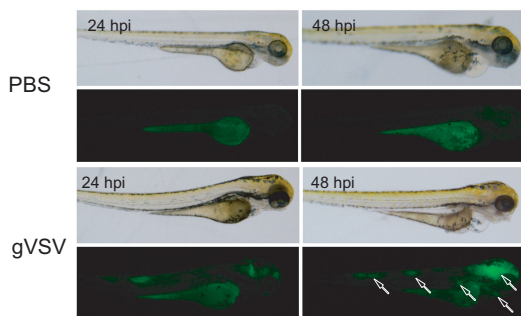
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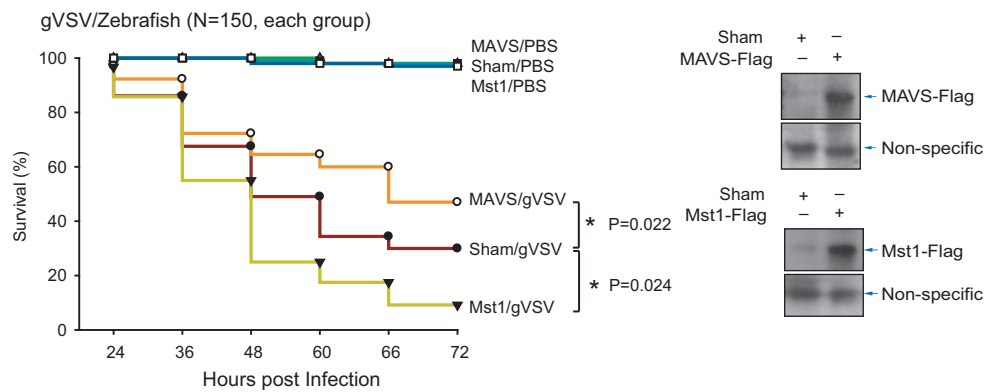
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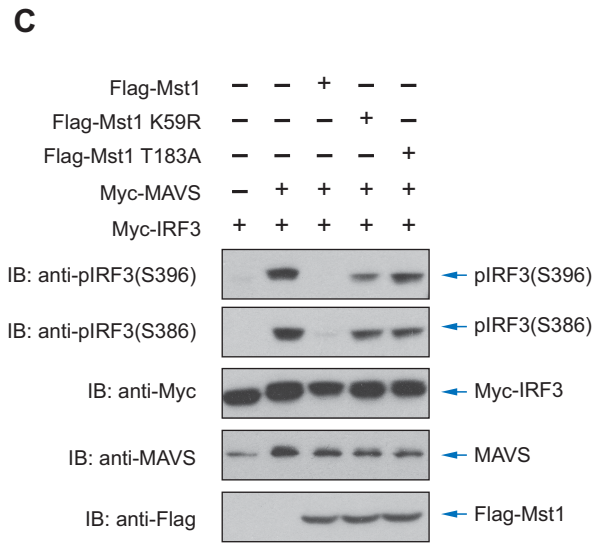
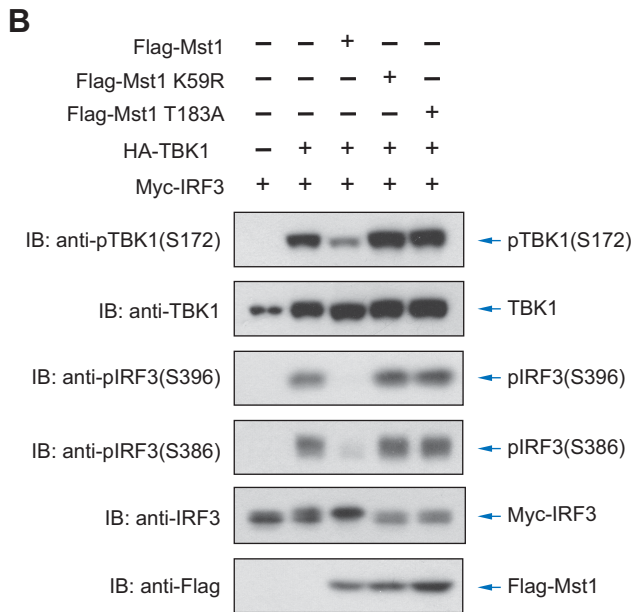
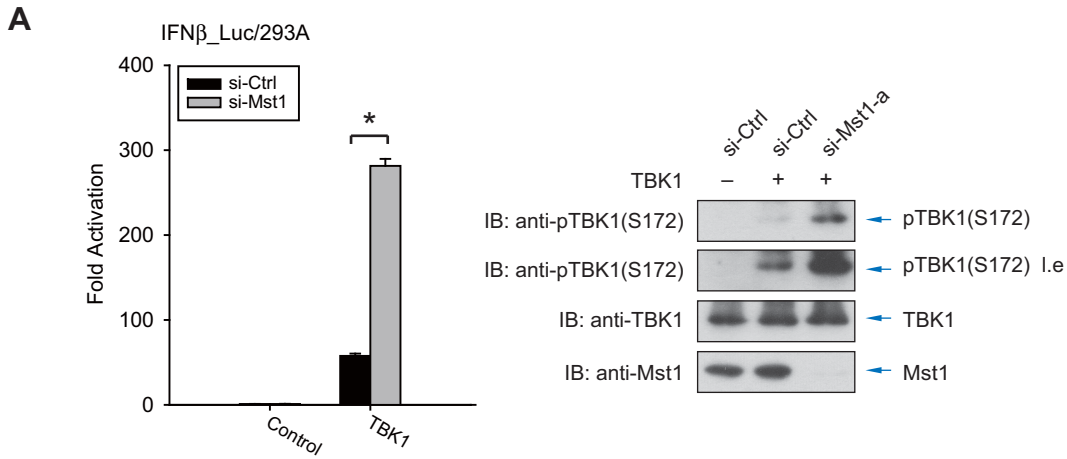


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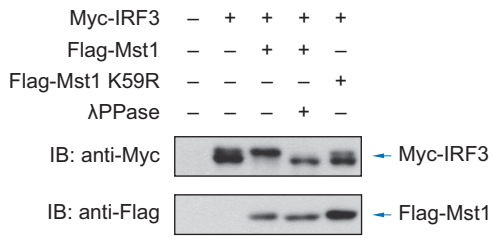


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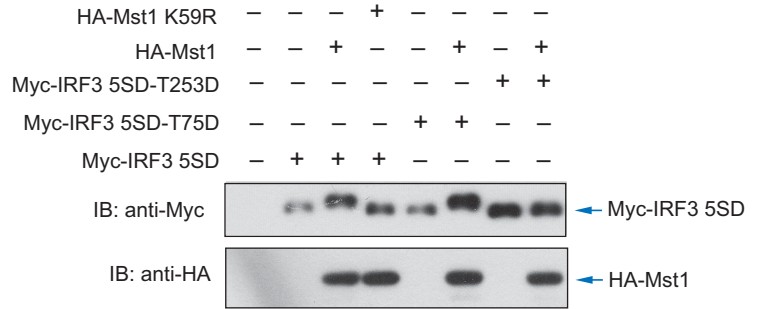




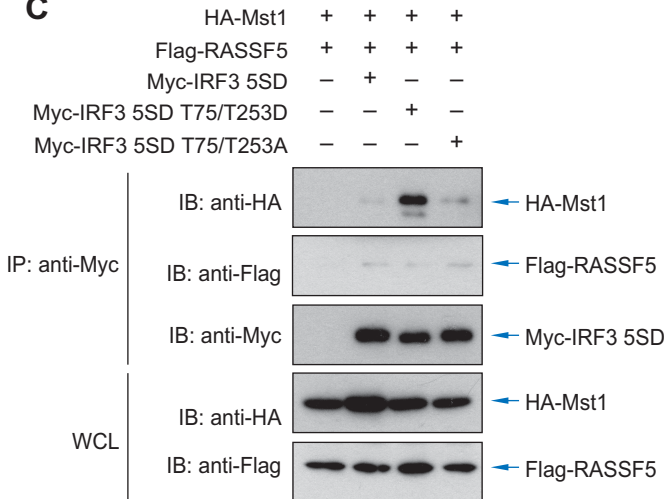
A



B



C



SUPPLEMENTAL FIGURE LEGENDS

sFig. 1: (A), Transfection of TBK1 (left panel) or IKK ϵ (right panel) induced robust activation of IRF3-responsive IFN β promoter, which was similarly blocked by cotransfection of wild-type Mst1. Kinase-dead mutants of Mst1 had only marginal effects on TBK1/IKK ϵ -induced IRF3 activation. N=3 experiments. * and **, P < 0.001, compared with control or wild-type Mst1, by Student's t-test. (B), Transfection of activated RIG-I (caRIG-I or RIG-I-N) or STING exerted a robust activation of IRF3-responsive promoters. Mst1 cotransfection, while strongly suppressed the stimulated IRF3 transactivation, did not inhibit both IFN β (left panel) and 5xISRE (right panel) promoters in resting cells. N=3 experiments. *, P < 0.001, compared with control, by Student's t-test. (C), Immunoblotting revealed protein levels of endogenous IRF3, TBK1, STING, Lats1, YAP and Smad4 in wild-type and Mst1 KO MEFs, without showing profound changes with Mst1 ablation.

sFig. 2: (A), Mst1^{-/-} MEFs displayed a resistance to HSV-1 infection, seeing by reduced virus-containing GFP⁺ cells (left panel), and lower amount of luciferase activity (right panel). Both GFP and luciferase ORFs were integrated into viral genome of HSV-1 (gIHSV). N=3 experiments, *, P < 0.01, compared with control MEF, by Student's t-test. (B), Expression of MAVS enhanced cellular resistance against gVSV infection in 293T cells, as displayed by reduced level of GFP⁺ cells. Cotransfection of wild-type Mst1, but not its kinase-dead form K59R, negated the MAVS-endowed viral resistance. (C), Enhanced antiviral response was detected in PBMCs from gVSV-infected Mst1^{-/-} mice, revealing by induction of antiviral proteins including IFNB1 and ISG15 at 6 hpi. N=5 mice for each group. *, P < 0.05, compared with control Mst1^{+/+} group, by Student t-test. (D), gVSV was microinjected into yolk of zebrafish embryos, which elicited a robust virus infection, occurring strongly at brain and also visible at muscle and gut tissues. (E), Zebrafish embryos were microinjected with mammal MAVS or Mst1 mRNA to gain ectopic expression of MAVS or Mst1 protein, as detected by immunoblottings (right panel). Zebrafish were then infected with VSV, and a vulnerable phenotype of Mst1 expressing embryos and

a resistance phenotype of MAVS expressing embryos were detected by VSV challenge. N=150 embryos for each group, * and **, P < 0.05, compared with sham group, by paired Student t-test.

sFig. 3: (A), siRNA-mediated depletion of Mst1 in HEK293A cells potentiated TBK1-induced IRF3 transactivation (left panel), and the enhanced TBK1 activation that is evidenced by TBK1 phospho-Ser172 immunoblotting (right panel). N=3 experiments, *, P < 0.01, compared with control siRNA, by Student's t-test. (B), IRF3 mobility shift by Mst1 cotransfection was revealed by immunoblotting with anti-IRF3 antibody, similar as experiment in Fig. 3C. (C), Loss of IRF3 phosphorylation and IRF3 mobility shift were also observed when IRF3 was cotransfected with MAVS in the presence of wild-type Mst1.

sFig. 4: (A), Mst1-induced IRF3 mobility shift was eliminated by λ phosphatase treatment, suggesting it is caused by the phosphorylation modification(s). (B), SDS-PAGE of phospho-mimetic mutants of IRF3 5SD showed that phospho-Thr253 was main modification by Mst1 to change IRF3 migration. (C), Co-immunoprecipitation assay by differential tags showed a weak interaction between IRF3 and wild-type or T75A/T253A Mst1, while an enhanced association with T75D/T253D mutant, implying its conformational changes after phosphorylation.

Oligo sequence:

Q-PCR primer sequence (mouse) 5'->3'	
IFIT1-Forward	GCCTATCGCCAAGATTTAGATGA
IFIT1-Reverse	TTCTGGATTTAACCGGACAGC
ISG15-Forward	GGTGTCCGTGACTAACTCCAT
ISG15-Reverse	CTGTACCACTAGCATCACTGTG
IFNB1-Forward	AGCTCCAAGAAAGGACGAACA
IFNB1-Reverse	GCCCTGTAGGTGAGGTTGAT
IRF7-Forward	GCGTACCCTGGAAGCATTTC
IRF7-Reverse	GCACAGCGGAAGTTGGTCT

Q-PCR primer sequence (VSV) 5'->3'	
VSV -Forward	GATAGTACCGGAGGATTGACGACTA
VSV -Reverse	TCAAACCATCCGAGCCATTC