

# Total Synthesis of (–)-Chromodorolide B

Daniel J. Tao, Yuriy Slutskyy, and Larry E. Overman\*

*Department of Chemistry, University of California, Irvine, California 92697-2025*

## Supporting Information – Table of Contents

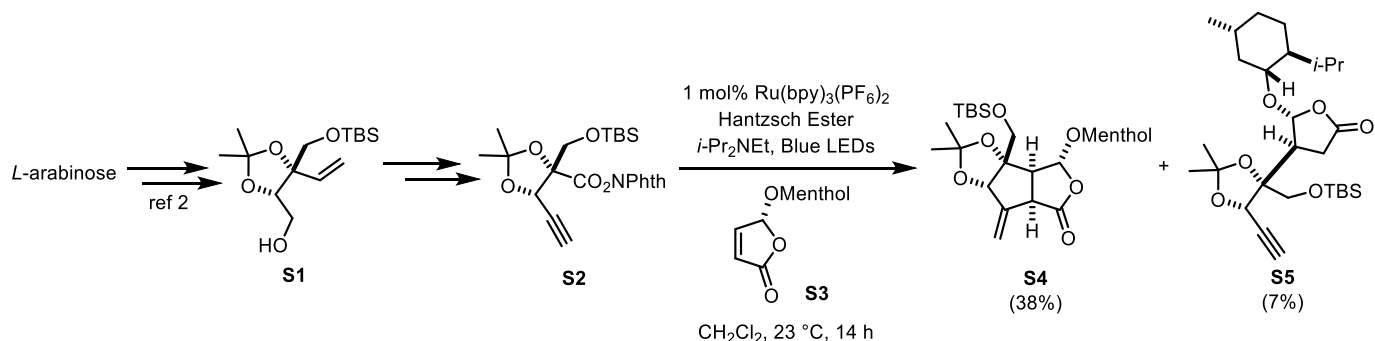
<b>Materials and Methods.....</b>	<b>S2–S3</b>
<b>Model System Study and Synthetic Procedures.....</b>	<b>S4–S8</b>
<b>Synthetic Procedures for Total Synthesis of (–)-Chromodorolide B.....</b>	<b>S9–S30</b>
<b>Comparison Table for Synthetic and Natural (–)-Chromodorolide B....</b>	<b>S31–S32</b>
<b>Optimization Tables of ACF Cascade.....</b>	<b>S33</b>
<b>Proposed Sequence to form ACF Cascade Products.....</b>	<b>S34</b>
<b>References.....</b>	<b>S35</b>
<b>Spectral Data.....</b>	<b>S36—S85</b>

## **Materials and Methods**

Unless stated otherwise, reactions were conducted in oven-dried glassware under an atmosphere of nitrogen or argon. Tetrahydrofuran (THF), diethyl ether, toluene, benzene, dichloromethane, methanol (MeOH), pyridine, DIPEA, and triethylamine were dried by passage through activated alumina. Benzyloxymethyl chloride (BOM-Cl) distilled under Ar from CaH directly before use. 1,1,3,3-Tetramethylguanidine was distilled under Ar from barium oxide directly before use. Thionyl chloride was distilled from quinoline under Ar. Tributylphosphine was distilled under reduced pressure and stored in a Schlenk flask. All other commercial reagents were used as received unless otherwise noted. Hantzsch ester<sup>1a</sup> (**HE**) and its 4-dideutero derivative<sup>1b</sup> were prepared according to literature procedures. Reaction temperatures were controlled using a temperature modulator, and unless stated otherwise, reactions were performed at room temperature (rt, approximately 23 °C). Thin-layer chromatography (TLC) was conducted with silica gel 60 F254 pre-coated plates, (0.25 mm) and visualized by exposure to UV light (254 nm) or by *p*-anisaldehyde, ceric ammonium molybdate, and potassium permanganate staining. Silica gel 60 (particle size 0.040–0.063 mm) was used for flash column chromatography. <sup>1</sup>H NMR spectra were recorded at 500 or 600 MHz and are reported relative to deuterated solvent signals. Data for <sup>1</sup>H NMR spectra are reported as follows: chemical shift (δ ppm), multiplicity, coupling constant (Hz), and integration. For late-stage intermediates, integrations for the upfield protons (δ2.00–0.50) of the hydrindane fragment were assigned based on 2D NMR techniques and chemical intuition. <sup>13</sup>C NMR spectra were recorded at 125 or 150 MHz. Data for <sup>13</sup>C NMR spectra are reported in terms of chemical shift. IR spectra were recorded on a FT-IR spectrometer and are reported in terms of frequency of absorption (cm<sup>-1</sup>). High-resolution mass spectra were obtained with a LCT spectrometer. Blue LEDs (30 cm, 1 watt) were purchased from <http://www.creativelightings.com> (product code CL-FRS5050-12WP-12V) and powered by 8 AA

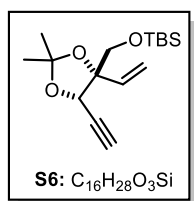
batteries. See JOC Standard Abbreviations and Acronyms for abbreviations (available at [http://pubs.acs.org/userimages/ContentEditor/1218717864819/joc\\_eah\\_abbreviations.pdf](http://pubs.acs.org/userimages/ContentEditor/1218717864819/joc_eah_abbreviations.pdf)).

## Model System Study and Synthetic Procedures



### (+)-Tert-butyl(((4*S*,5*S*)-5-ethynyl-2,2-dimethyl-4-vinyl-1,3-dioxolan-4-

yl)methoxy)dimethylsilane (**S6**): To a suspension of known alcohol **S1**<sup>2</sup> (0.732 g, 2.42 mmol) and



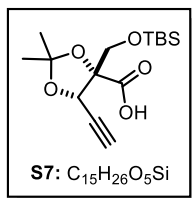
solid  $\text{NaHCO}_3$  (1.01 g, 12.1 mmol) in  $\text{CH}_2\text{Cl}_2$  (4 mL) was added Dess-Martin periodinane (1.23 g, 2.90 mmol). The reaction was vigorously stirred for 2 h, at which point the suspension was filtered through Celite and concentrated *in vacuo*.

The residue was then washed with pentanes (4 x 8 mL), and the combined organic washes were filtered through Celite and concentrated *in vacuo* to afford the crude aldehyde as a yellow oil which was carried forward immediately.

The crude aldehyde and dimethyl (1-azoacetyl)phosphonate (0.558 g, 2.90 mmol) were dissolved in MeOH (9 mL). Solid  $\text{K}_2\text{CO}_3$  (0.669 g, 4.84 mmol) was then added, and the suspension was vigorously stirred for 2 h. Celite (~5 g) was added to the reaction vessel, and the reaction was concentrated *in vacuo*. Purification by flash column chromatography (5% EtOAc in hexanes to 7% EtOAc in hexanes) afforded alkyne **S6** (0.550 g, 1.86 mmol, 77% yield) as a clear oil, which solidified upon standing.  $R_f$  0.90 (20% EtOAc in hexanes; visualized with  $\text{KMnO}_4$ ).  $^1\text{H NMR}$  (500 MHz,  $\text{CDCl}_3$ )  $\delta$  6.12 (dd,  $J = 17.2, 10.9$  Hz, 1H), 5.53 (dd,  $J = 17.5, 1.6$  Hz, 1H), 5.30 (dd,  $J = 10.9, 1.6$  Hz, 1H), 4.99 (d,  $J = 2.1$  Hz, 1H), 3.58 (d,  $J = 10.6$  Hz, 1H), 3.55 (d,  $J = 10.7$  Hz, 1H),

2.60 (d,  $J = 2.2$  Hz, 1H), 1.54 (s, 3H), 1.43 (s, 3H), 0.89 (s, 9H), 0.07 (s, 6H);  $^{13}\text{C}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  136.46, 116.73, 110.27, 85.75, 79.25, 77.10, 69.70, 65.41, 27.87, 27.01, 26.00, 18.44, -5.23, -5.49; IR (thin film) 3312, 2988, 2955, 2858, 1741, 1378, 1253  $\text{cm}^{-1}$ ;  $[\alpha]_{\text{D}}^{25}$ : +0.79 ( $c = 2.5$ ,  $\text{CH}_2\text{Cl}_2$ ); HRMS (ESI) calculated for  $\text{C}_{16}\text{H}_{29}\text{O}_3\text{Si}$  ( $\text{M}+\text{H}$ ) 297.1887, observed 297.1890; mp 39–41  $^\circ\text{C}$ .

**(-)-(4*S*,5*S*)-4-(((tert-butyldimethylsilyl)oxy)methyl)-5-ethynyl-2,2-dimethyl-1,3-dioxolane-4-carboxylic acid (S7):** A solution of alkyne **S6** (0.553 g, 1.87 mmol) in methanol (8 mL) was



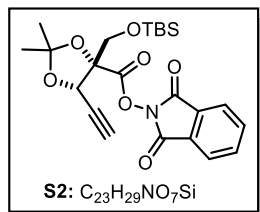
cooled to  $-78$   $^\circ\text{C}$ . Ozone from an ozone generator was bubbled through the solution until a pale blue color was observed ( $\sim 5$  min). The solution was then sparged with oxygen until the pale blue color disappeared. Dimethyl sulfide (0.31

mL, 4.3 mmol) was added to the solution, which was maintained at  $-78$   $^\circ\text{C}$  for 1 h. The reaction vessel was allowed to warm to  $23$   $^\circ\text{C}$  and concentrated *in vacuo*. The residue was then redissolved in a 3:1 solution of *t*-BuOH/ $\text{H}_2\text{O}$  (8 mL). A solution of 2-methyl-2-butene (2.0 mL, 19 mmol) was added to the mixture, followed by  $\text{NaH}_2\text{PO}_4$  (1.80 g, 15.0 mmol) and  $\text{NaClO}_2$  (0.845 g, 9.35 mmol). The reaction was maintained at  $23$   $^\circ\text{C}$  for 2 h, at which point  $\text{H}_2\text{O}$  (4 mL) was added. This mixture was washed with EtOAc (3 x 10 mL), and the combined organic layers were washed with aq. NaOH (5 mL of 0.5 M soln). The aqueous layer was then acidified with aq. HCl (7 mL of 0.5 M soln). The aqueous layer was then washed with EtOAc (3 x 10 mL), and the combined organic layers were washed with brine (1 x 10 mL), dried over  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated *in vacuo* to provide acid **S7** as a colorless oil (0.450 g, 1.43 mmol, 76% yield).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  4.97 (d,  $J = 2.3$  Hz, 1H), 3.94 (d,  $J = 11.0$  Hz, 1H), 3.92 (d,  $J = 11.0$  Hz, 1H), 2.63 (d,  $J = 2.2$  Hz, 1H), 1.67 (s, 3H), 1.47 (s, 3H), 0.90 (s, 9H), 0.09 (s, 3H), 0.08 (s, 3H);  $^{13}\text{C}$  NMR (500 MHz,

CDCl<sub>3</sub>)  $\delta$  173.25, 113.33, 88.23, 77.69, 69.04, 63.64, 26.99, 26.97, 25.95, 18.48, 14.32, -5.26, -5.48; IR (thin film) 3505, 3277, 2990, 2931, 2858, 1731, 1379 cm<sup>-1</sup>; [ $\alpha$ ]<sub>D</sub><sup>25</sup> : -30.0 (c = 2.1, CH<sub>2</sub>Cl<sub>2</sub>); HRMS (ESI) calculated for C<sub>15</sub>H<sub>25</sub>O<sub>5</sub>Si (M-H) 313.1471, observed 313.1467.

**(+)-1,3-dioxoisindolin-2-yl(4*S*,5*S*)-4-(((tert-butyldimethylsilyl)oxy)methyl)-5-ethynyl-2,2-**

**dimethyl-1,3-dioxolane-4-carboxylate (S2):** Acid S7 (0.453 g, 1.44 mmol) was charged into a



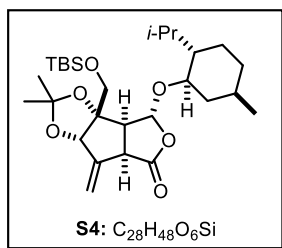
flask with THF (8 mL). *N*-hydroxyphthalimide (0.399 g, 2.45 mmol), *N,N'*-dicyclohexylcarbodiimide (0.446 g, 2.16 mmol), and 4-dimethylaminopyridine (9 mg, 0.07 mmol) were added to the reaction

vessel, which was maintained at 23 °C for 20 h. Hexanes (5 mL) was added to the reaction, and the resulting suspension was filtered through Celite. The yellow filtrate was concentrated *in vacuo* and then purified by flash column chromatography (10% EtOAc in hexanes to 15% EtOAc in hexanes) to provide *N*-acyloxyphthalimide S2 (0.539 g, 1.18 mmol, 82% yield) as a colorless crystalline solid. R<sub>f</sub> 0.25 (20% EtOAc in hexanes; visualized with ceric ammonium molybdate).

<sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  7.88 (m, 2H), 7.78 (m, 2H), 5.14 (d, *J* = 2.2 Hz, 1H), 4.10 (d, *J* = 11.5 Hz, 1H), 4.07 (d, *J* = 11.3 Hz, 1H), 2.78 (d, *J* = 2.2 Hz, 1H), 1.71 (s, 3H), 1.50 (s, 3H), 0.93 (s, 9H), 0.13 (s, 3H), 0.12 (s, 3H); <sup>13</sup>C NMR (500 MHz, CDCl<sub>3</sub>)  $\delta$  166.77, 161.37, 134.87, 129.09, 124.07, 113.53, 88.06, 78.80, 76.25, 68.81, 62.80, 26.7, 26.64, 26.00, 18.54, -5.18, -5.49; IR (thin film) 3283, 2930, 2855, 2360, 2340, 2118, 1789, 1748 cm<sup>-1</sup>; [ $\alpha$ ]<sub>D</sub><sup>25</sup> : +38.3 (c = 2.0, CH<sub>2</sub>Cl<sub>2</sub>); HRMS (ESI) calculated for C<sub>23</sub>H<sub>29</sub>NO<sub>7</sub>SiNa(M+Na) 482.1611, observed 482.1612; mp 105–109 °C.

**(-)-(3a*S*,3b*S*,4*R*,6a*R*,7a*S*)-3a-(((tert-butyldimethylsilyl)oxy)methyl)-4-(((1*R*,2*S*,5*R*)-2-isopropyl-5-methylcyclohexyl)oxy)-2,2-dimethyl-7-methylenehexahydro-6*H*-**

**furo[3',4':3,4]cyclopenta[1,2-d][1,3]dioxol-6-one (S4):** To a vial charged with *N*-

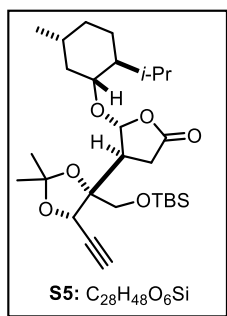


acyloxyphthalimide **S2** (100 mg, 0.218 mmol) was added CH<sub>2</sub>Cl<sub>2</sub> (2 mL) that had been separately sparged with argon. Butenolide **S3**<sup>3</sup> (78 mg, 0.33 mmol), Hantzsch ester (82 mg, 0.63 mmol), Ru(bpy)<sub>3</sub>(PF<sub>6</sub>)<sub>2</sub> (2 mg, 0.02 mmol), and Hünig's base (80 μL, 0.48 mmol) were then added to the

reaction. The vial was then vigorously stirred while being irradiated by a single strip of blue LED lights (450 nm) at 23 °C. After 6 h, the reaction mixture was diluted with hexanes (2 mL) and filtered through Celite. The resulting solution was then concentrated *in vacuo* and separated by flash column chromatography (3% EtOAc in hexanes to 5% EtOAc in hexanes) to provide lactone **S4** (42 mg, 0.083 mmol, 38% yield) as a colorless, crystalline solid and addition product **S5** (7.5 mg, 0.015 mmol, 7% yield) as an oil. A single crystal X-ray structure of lactone **S4** was obtained after recrystallization in MeOH/hexanes to confirm structural assignment.<sup>4</sup> R<sub>f</sub> for **S4**: 0.65 (10% EtOAc in hexanes; visualized with ceric ammonium molybdate). R<sub>f</sub> for **S5**: 0.60 (10% EtOAc in hexanes; visualized with ceric ammonium molybdate).

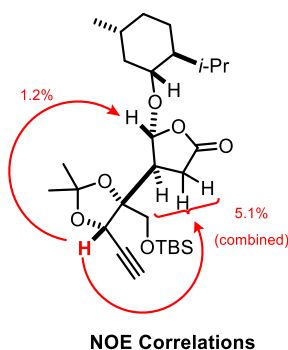
**S4** for <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 5.90 (d, *J* = 2.6 Hz, 1H), 5.62 (dd, *J* = 2.3, 1.0 Hz, 1H), 5.50 (dd, *J* = 2.7, 0.8 Hz, 1H), 4.73 (app s, 1H), 3.96–3.92 (m, 1H), 3.86 (d, *J* = 10.7 Hz, 1H), 3.77 (d, *J* = 10.8 Hz, 1H), 3.51 (dt, *J* = 10.6, 4.3 Hz, 1H), 3.07 (dd, *J* = 10.3, 2.3 Hz, 1H), 2.14–2.03 (m, 2H), 1.69–1.60 (m, 2H), 1.47 (s, 3H), 1.40–1.32 (m, 4H), 1.27–1.16 (m, 2H), 1.04–0.93 (m, 1H), 0.92 (d, *J* = 6.5 Hz, 3H), 0.90–0.80 (m, 13H), 0.77 (d, *J* = 6.9 Hz, 3H), 0.09 (s, 3H), 0.08 (s, 3H); <sup>13</sup>C NMR (500 MHz, CDCl<sub>3</sub>) δ 175.05, 143.32, 116.61, 112.69, 99.71, 90.76, 86.88, 77.22, 64.36, 57.42, 48.22, 47.87, 40.01, 34.42, 31.51, 28.37, 27.04, 25.93, 25.51, 23.20, 22.39, 21.03, 18.41,

15.81, -5.45, -5.50; IR (thin film) 2953, 2929, 2858, 1779, 1461  $\text{cm}^{-1}$ ;  $[\alpha]_D^{25}$  : -133 (c = 1.9,  $\text{CH}_2\text{Cl}_2$ ); HRMS (ESI) calculated for  $\text{C}_{28}\text{H}_{48}\text{O}_6\text{SiNa}$  (M+Na) 531.3118, observed 531.3126; mp 136–142  $^\circ\text{C}$ .



(-)-**S5** for  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  5.82 (d,  $J = 2.2$  Hz, 1H), 4.59 (d,  $J = 2.2$  Hz, 1H), 3.98 (d,  $J = 10.7$  Hz, 1H), 3.75 (d,  $J = 10.7$  Hz, 1H), 3.52 (dt,  $J = 10.9, 4.4$  Hz, 1H), 2.82–2.65 (m, 3H), 2.58 (d,  $J = 2.3$  Hz, 1H), 2.17–2.04 (m, 2H), 1.69–1.59 (m, 2H), 1.50 (s, 3H), 1.38 (s, 3H), 1.37–1.32 (m, 1H), 1.27–1.17 (m, 2H), 0.99 (app qd,  $J = 12.5, 3.3$  Hz, 1H), 0.91 (d,  $J = 6.9$  Hz,

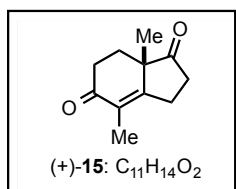
3H), 0.90 (s, 9H), 0.87 (d,  $J = 7.0$  Hz, 3H), 0.86–0.81 (m, 1H), 0.77 (d,  $J = 7.0$  Hz, 3H), 0.09 (s, 6H);  $^{13}\text{C}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  175.88, 110.04, 100.76, 83.81, 78.81, 77.21, 76.74, 71.62, 64.38, 47.88, 46.89, 39.88, 34.46, 31.50, 29.82, 28.24, 27.07, 26.04, 25.56, 23.22, 22.42, 21.02, 18.36, 15.80, -5.44, -5.46; IR (thin film) 3311, 3262, 2955, 2929, 2858, 1791, 1462, 1374, 1252  $\text{cm}^{-1}$ ;  $[\alpha]_D^{25}$  : -93.7 (c = 1.2,  $\text{CH}_2\text{Cl}_2$ ); HRMS (ESI) calculated for  $\text{C}_{28}\text{H}_{48}\text{O}_6\text{SiNa}$  (M+Na) 531.3118, observed 531.3131.





## Synthetic Procedures for Total Synthesis of (-)-Chromodorolide B

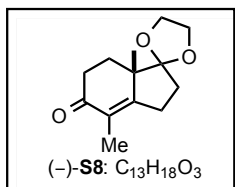
(+)-(S)-4,7a-dimethyl-2,3,7,7a-tetrahydro-1H-indene-1,5(6H)-dione (15): (S)-enone was



prepared according to a literature procedure.<sup>5</sup> A 100 mL round-bottom flask was charged with 2-methyl-2-(3-oxopentyl)cyclopetane-1,3-dione (39.6 g, 202 mmol), followed by the addition of *L*-phenylalanine (10 g, 61 mmol),

PPTS (25.3 g, 101 mmol), and DMSO (14 mL, 200 mmol). The heterogeneous reaction mixture was then sonicated for 36 h at 50 °C. The mixture was transferred into a separatory funnel with EtOAc (500 mL), followed by the addition of H<sub>2</sub>O (500 mL). The two layers were separated and the aqueous phase was extracted with EtOAc (3 x 500 mL). The combined organic extracts were washed sequentially with aq. HCl (1 x 500 mL of 1 M soln), sat. aq. NaHCO<sub>3</sub> (1 x 500 mL), and brine (1 x 500 mL). The organic phase was dried over MgSO<sub>4</sub> and concentrated *in vacuo* to yield a viscous red oil. The resulting oil was filtered through a silica gel plug (100 g SiO<sub>2</sub>) with 30% EtOAc in hexanes (1 L) to afford 32.7 g of crude (+)-15 as an orange oil. R<sub>f</sub> 0.23 (30% EtOAc in hexanes; visualized with *p*-anisaldehyde). The resulting oil was crystallized from Et<sub>2</sub>O (23 mL) at -20 °C utilizing a seed crystal (obtained via crystallization from Et<sub>2</sub>O/benzene at -20 °C) to provide (+)-15 as an off-white crystalline solid (21.3 g, 120 mmol, 59% yield, 99% *ee*). Spectral data were consistent with reported values.<sup>5</sup> The enantiomeric excess was determined by chiral stationary-phase HPLC analysis (Chiracel OB-H column; flow: 2.0 mL/min, 20% isopropanol:*n*-hexane; λ = 254 nm; minor enantiomer t<sub>R</sub> = 5.86 min, major enantiomer t<sub>R</sub> = 7.25 min).

**(-)-(S)-4,7a-dimethyl-2,3,7,7a-tetrahydrospiro[indene-1,2-[1,3]dioxolan]-5(6H)-one (S8):** A

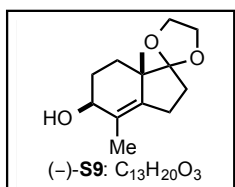


2 L round-bottom flask was charged with enone (+)-**15** (7.7 g, 43 mmol), followed by the addition of benzene (860 mL), ethylene glycol (2.9 mL, 52 mmol), and *p*-TsOH•H<sub>2</sub>O (1.6 g, 8.6 mmol). A Dean-Stark apparatus was

fitted to the flask, and the homogenous reaction mixture was maintained at reflux overnight. Upon completion of the reaction, as indicated by TLC analysis (40% EtOAc in hexanes; visualized with *p*-anisaldehyde), the mixture was cooled to 0 °C and sat. aq. NaHCO<sub>3</sub> (300 mL) was added. The resulting biphasic mixture was separated and the aqueous layer was extracted with Et<sub>2</sub>O (3 x 200 mL). The combined organic layers were washed with brine (1 x 500 mL), dried over MgSO<sub>4</sub>, and concentrated *in vacuo* to yield a yellow oil. The crude residue was purified by flash column chromatography (20% EtOAc in hexanes) to yield **S8** as a yellow oil (9.6 g, 43 mmol, 100% yield).

R<sub>f</sub> 0.45 (40% EtOAc in hexanes; visualized with *p*-anisaldehyde). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ 4.04–3.91 (m, 4H), 2.58–2.51 (m, 2H), 2.43 (dd, *J* = 5.4, 3.6 Hz, 1H), 2.28 (td, *J* = 13.2, 5.4 Hz, 1H), 2.20–2.16 (m, 1H), 1.95–1.91 (m, 1H), 1.68 (s, 3H), 1.61–1.58 (m, 1H), 1.22 (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 198.63, 167.10, 128.96, 118.04, 65.94, 65.05, 47.68, 33.23, 31.92, 26.80, 25.99, 20.49, 10.83; IR (thin film) 2953, 2881, 1660, 1451, 1153 cm<sup>-1</sup>; [α]<sub>D</sub><sup>21</sup>: -7.79 (c = 1.6, CH<sub>2</sub>Cl<sub>2</sub>); HRMS (ESI) calculated for [C<sub>13</sub>H<sub>18</sub>O<sub>3</sub>Na]<sup>+</sup> (M+Na) 245.1154, observed 245.1163.

**(-)-(5S,7aS)-4,7a-dimethyl-2,3,5,6,7,7a-hexahydrospiro[indene-1,2'-[1,3]dioxolan]-5-ol (S9):**

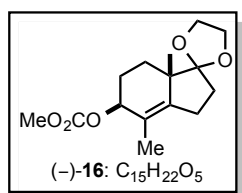


A 1 L round-bottom flask was charged with enone **S8** (9.6 g, 43 mmol), followed by the addition of Et<sub>2</sub>O (310 mL). The solution was cooled to -78 °C. A solution of LiAlH<sub>4</sub> (65 mL of 1 M in Et<sub>2</sub>O, 65 mmol) was added dropwise.

The homogenous solution was warmed to 0 °C and maintained at that temperature until TLC

analysis (40% EtOAc in hexanes; visualized with *p*-anisaldehyde) indicated complete consumption of the starting material, typically 10–20 min. Upon completion of the reaction, sat. aq. Rochelle's salt (150 mL) was slowly added. The biphasic mixture was stirred vigorously for 30 min at 0 °C. The solution was then transferred to a separatory funnel and extracted with Et<sub>2</sub>O (3 x 150 mL). Combined organic layers were washed with brine (1 x 500 mL), dried over MgSO<sub>4</sub>, and concentrated *in vacuo* to yield a colorless solid. The crude residue was purified by flash column chromatography (30% EtOAc in hexanes) to yield **S9** as a colorless, crystalline solid (9.5 g, 42 mmol, 98% yield). R<sub>f</sub> 0.25 (40% EtOAc in hexanes; visualized with *p*-anisaldehyde). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 4.14–4.12 (m, 1H), 3.96–3.86 (m, 4H), 2.33–2.30 (m, 2H), 2.13–2.06 (m, 2H), 1.87–1.78 (m, 2H), 1.71–1.62 (m, 4H), 1.35–1.31 (m, 2H), 1.14 (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 141.44, 127.91, 118.50, 71.73, 65.73, 64.94, 46.72, 32.07, 30.05, 26.64, 23.98, 22.51, 14.57; IR (thin film) 3411, 2947, 2873, 1642, 1149 cm<sup>-1</sup>; [α]<sub>D</sub><sup>21</sup>: -32.0 (c = 1.8, CH<sub>2</sub>Cl<sub>2</sub>); HRMS (ESI) calculated for [C<sub>13</sub>H<sub>20</sub>O<sub>3</sub>] (M) 224.1413, observed 224.1406; mp 86–88 °C.

**(-)-(5*S*,7*aS*)-4,7*a*-dimethyl-2,3,5,6,7,7*a*-hexahydrospiro[indene-1,2'-[1,3]dioxolan]-5-yl**

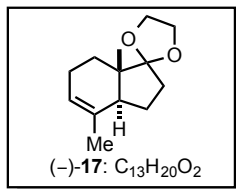


**methyl carbonate (16)**: A 1 L round-bottom flask was charged with allylic alcohol **S9** (9.5 g, 42 mmol), followed by the addition of DMAP (15.0 g, 126 mmol), CH<sub>2</sub>Cl<sub>2</sub> (420 mL), and methyl chloroformate (13 mL, 170 mmol). The

resulting homogenous solution was maintained at 35 °C until TLC analysis (30% EtOAc in hexanes; visualized with *p*-anisaldehyde) indicated complete consumption of the starting material, typically 30–90 min. Upon completion of the reaction, the mixture was cooled to room temperature, followed by the addition of sat. aq. NH<sub>4</sub>Cl (200 mL). Next, the solution was transferred to a separatory funnel and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 x 200 mL). The combined organic

layers were washed with brine (1 x 500 mL), dried over MgSO<sub>4</sub>, and concentrated *in vacuo* to yield a colorless solid. The crude residue was purified by flash column chromatography (10% EtOAc in hexanes) to yield **16** as a colorless, crystalline solid (11.4 g, 40.4 mmol, 96% yield). R<sub>f</sub> 0.35 (20% EtOAc in hexanes; visualized with *p*-anisaldehyde). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) δ 5.19 (app t, *J* = 7.8 Hz, 1H), 3.96–3.87 (m, 4H), 3.79 (s, 3H), 2.34 (t, *J* = 7.8 Hz, 2H), 2.24–2.20 (m, 1H), 2.12–2.07 (m, 1H), 1.89 (td, *J* = 14.4, 3.0 Hz, 1H), 1.84–1.77 (m, 2H), 1.57 (s, 3H), 1.36 (dt, *J* = 12.7, 3.8 Hz, 1H), 1.15 (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>) δ 156.27, 144.24, 123.87, 118.39, 78.66, 65.76, 64.93, 54.80, 46.47, 31.90, 26.34, 25.76, 24.11, 22.15, 14.69; IR (thin film) 2954, 2878, 1742, 1442, 1260 cm<sup>-1</sup>; [α]<sub>D</sub><sup>22</sup>: -41.32 (c = 1.1, CH<sub>2</sub>Cl<sub>2</sub>); HRMS (ESI) calculated for [C<sub>15</sub>H<sub>22</sub>O<sub>5</sub>Na]<sup>+</sup> (M+Na) 305.1365, observed 305.1370; mp 56–58 °C.

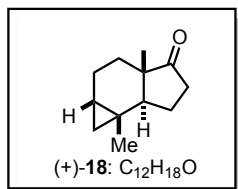
**(-)-(3a*S*,7a*S*)-4,7a-dimethyl-2,3,3a,6,7,7a-hexahydrospiro[indene-1,2'-[1,3]dioxolane] (17):**



A 200 mL round-bottom flask was charged with Pd(acac)<sub>2</sub> (490 mg, 1.6 mmol), followed by the addition of benzene (21 mL), PBu<sub>3</sub> (0.4 mL, 1.6 mmol). The homogenous mixture was maintained at room temperature for 5 min. Ammonium formate (4.2 g, 67 mmol) was finely crushed with a mortar and pestle and added to the reaction mixture in one portion. The resulting heterogeneous solution was stirred vigorously for 10 min. Next, a solution of carbonate **16** (3.0 g, 10.6 mmol) in benzene (32 mL) was added dropwise. The heterogeneous mixture was stirred vigorously overnight at room temperature. Upon completion of the reaction, as indicated by TLC analysis (10% EtOAc in hexanes; visualized with *p*-anisaldehyde), the mixture was filtered through a silica gel plug (10%EtOAc in hexanes) to provide a brown oil. The crude residue was purified by flash column chromatography (350 g SiO<sub>2</sub>, 1 L 100% hexanes, 1 L 0.5% EtOAc in hexanes, 5 L 1% EtOAc in hexanes) to yield (-)-**17** (1.72

g, 8.26 mmol, 78% yield) as a yellow oil that contained ~1% tributylphosphine oxide as an impurity.  $R_f$  0.2 (2% EtOAc in hexanes, visualized with *p*-anisaldehyde). *Note*: It is helpful to develop the TLC plate 2 times to visualize two more polar impurities with similar  $R_f$  values. If desired, the yellow oil can be purified further by Kugelrohr short-pass distillation (135 °C, 0.6 torr) to yield **17** (1.69 g, 8.11 mmol, 77% yield) as a colorless oil.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  5.20 (s, 1H), 3.94–3.84 (m, 4H), 2.46 (br s, 1H), 2.10–2.05 (m, 3H), 1.91–1.84 (m, 1H), 1.78–1.69 (m, 2H), 1.62 (s, 3H), 1.46–1.40 (m, 2H), 0.84 (s, 3H);  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ )  $\delta$  134.88, 119.96, 119.36, 65.44, 64.73, 46.38, 45.69, 35.21, 28.47, 24.08, 22.04, 20.45, 13.84; IR (thin film) 2944, 2878, 1436, 1376, 1044  $\text{cm}^{-1}$ ;  $[\alpha]_D^{21}$ :  $-62.3$  ( $c = 1.0$ ,  $\text{CH}_2\text{Cl}_2$ ); HRMS (ESI) calculated for  $[\text{C}_{13}\text{H}_{20}\text{O}_2]$  (M) 208.1463, observed 208.1460.

**(+)-(1a*S*,3a*S*,6a*S*,6b*R*)-3a,6b-dimethyloctahydrocyclopropa[*e*]inden-4(1*H*)-one (18)**: A 500

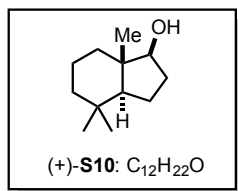


mL round-bottom was charged with ketal **17** (6.2 g, 30 mmol), followed by the addition of  $\text{CH}_2\text{Cl}_2$  (150 mL). The solution was cooled to 0 °C, and a solution of  $\text{Et}_2\text{Zn}$  (60 mL of 1 M in hexanes, 60 mmol) was added dropwise.

After 10 min at 0 °C, chloriodomethane (8.7 mL, 120 mmol) was added. After 2 h at 0 °C, the heterogeneous mixture was allowed to warm to room temperature and stirred overnight while shielded from light. Upon cooling the reaction mixture to 0 °C, conc. HCl (7.6 mL) in MeOH (115 mL) was added dropwise. Upon complete deprotection of the ketal (typically 10–30 min), as indicated by TLC analysis (5% EtOAc in hexanes, visualized with *p*-anisaldehyde), the mixture was transferred to a separatory funnel.  $\text{H}_2\text{O}$  (150 mL) was added, and the resulting biphasic mixture was extracted with  $\text{CH}_2\text{Cl}_2$  (3 x 200 mL). The combined organic layers were washed with brine (1 x 500 mL), dried over  $\text{MgSO}_4$ , and concentrated *in vacuo* to yield a yellow oil. The crude

residue was purified by flash column chromatography (0% to 2% EtOAc in hexanes) to yield cyclopropane **18** as a colorless oil (4.9 g, 28 mmol, 92% yield).  $R_f$  0.15 (5% EtOAc in hexanes; visualized with *p*-anisaldehyde). Alternatively, cyclopropane **18** can be purified by Kugelrohr short-pass distillation (100 °C, 0.4 torr).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  2.50–2.45 (m, 1H), 2.10–2.04 (m, 2H), 2.01–1.97 (m, 1H), 1.91–1.85 (m, 1H), 1.81 (dd,  $J = 14.4, 6.9$  Hz, 1H), 1.66 (dd,  $J = 13.4, 7.8$  Hz, 1H), 1.37 (dd,  $J = 13.3, 6.0$  Hz, 1H), 1.10 (s, 3H), 0.90–0.85 (m, 4H), 0.67–0.64 (m, 1H), 0.56 (dd,  $J = 9.5, 4.3$  Hz, 1H), 0.01 (app t,  $J = 5.3$  Hz, 1H);  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ )  $\delta$  221.17, 50.66, 47.34, 36.62, 27.99, 23.29, 22.96, 22.21, 19.73, 17.84, 16.30, 13.33; IR (thin film) 3051, 2945, 2864, 1737, 1445, 1024  $\text{cm}^{-1}$ ;  $[\alpha]_D^{22}$ : +118 ( $c = 1.0, \text{CH}_2\text{Cl}_2$ ); HRMS (ESI) calculated for  $[\text{C}_{12}\text{H}_{18}\text{O}]$  (M) 178.1358, observed 178.1358.

**(+)-(1*S*,3*aS*,7*aS*)-4,4,7*a*-trimethyloctahydro-1*H*-inden-1-ol (S10)**: A 20 mL vial was charged

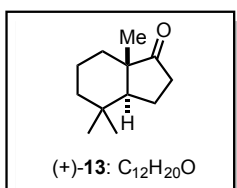


with ketone **18** (1.1 g, 6.0 mmol), followed by the addition of AcOH (6.0 mL),  $\text{PtO}_2 \cdot \text{H}_2\text{O}$  (270 mg, 1.2 mmol). The flask was then placed in a Parr high pressure vessel that was subsequently filled with  $\text{H}_2$  (10 atm). The vessel was

placed on top of an IKA magnetic plate and stirred overnight. The reaction mixture was filtered through Celite into a separatory funnel, followed by the addition of EtOAc (50 mL). The resulting solution was washed with  $\text{H}_2\text{O}$  (3 x 50 mL), sat. aq.  $\text{NaHCO}_3$  (1 x 50 mL), and brine (1 x 50 mL). The organic layer was then dried over  $\text{MgSO}_4$  and concentrated *in vacuo* to yield a colorless solid. The crude residue was purified by flash column chromatography (2% to 6% EtOAc in hexanes) to yield **S10** as a colorless solid (1.02 g, 5.59 mmol, 93% yield) that contained ~5% of an impurity.  $R_f$  0.20 (10% EtOAc in hexanes; visualized with *p*-anisaldehyde). Recrystallization from hot *n*-

hexanes (50 mL) yielded **S10** as colorless needles (820 mg, 4.5 mmol, 81% recovery). Spectral data were consistent with reported values.<sup>6</sup>

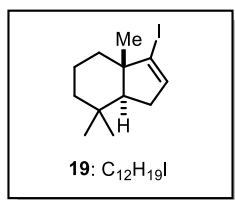
**(+)-(3a*S*,7a*S*)-4,4,7a-trimethyloctahydro-1*H*-inden-1-one (13)**: A 100 mL round-bottom flask



was charged with alcohol **S10** (0.80 g, 4.4 mmol), followed by the addition of PCC (2.0 g, 9.4 mmol), Celite (2.0 g), and CH<sub>2</sub>Cl<sub>2</sub> (22 mL). The resulting heterogenous solution was stirred vigorously at room temperature, until

TLC analysis (10% EtOAc in hexanes, visualized with *p*-anisaldehyde) indicated complete consumption of the starting material (typically 60–90 min). Hexanes (22 mL) was added to the reaction mixture, which was subsequently gravity filtered. The reaction vessel and filtrate were washed with 10% EtOAc in hexanes (4 x 25 mL). The combined organic washes were concentrated *in vacuo* to yield an orange oil. The crude residue was purified by flash column chromatography (0% to 2% EtOAc in hexanes) to provide **13** (0.75 g, 4.2 mmol, 95% yield, 98.5% *ee*) as a colorless oil, which solidified upon standing. R<sub>f</sub> 0.29 (5% EtOAc in hexanes, visualized with *p*-anisaldehyde). Alternatively, ketone **13** can be purified by Kugelrohr short-pass distillation (130 °C, 0.8 torr). Spectral data were consistent with reported values.<sup>6</sup> The enantiomeric excess of the corresponding trisyl hydrazone<sup>7</sup> was determined by chiral stationary-phase HPLC analysis (Chiracel OD-H column; flow: 1.0 mL/min, 1% isopropanol:*n*-hexane; λ = 254 nm; minor enantiomer t<sub>R</sub> = 13.65 min, major enantiomer t<sub>R</sub> = 21.34 min). *Note*: The reaction is readily scalable. In a separate experiment, crude alcohol **S10** (4.46 g, 24.5 mmol) was oxidized according to the above procedure to yield ketone **13** (4.33 g, 24.0 mmol, 96% yield) as a colorless oil. The material contained ~5% impurity that was carried through from the previous step. Therefore, it is important to recrystallize alcohol **S10** prior to oxidation to obtain pure ketone **13**.

**(-)-(3a*S*,7a*S*)-3-iodo-3a,7,7-trimethyl-3a,4,5,6,7,7a-hexahydro-1*H*-indene (19):** Hydrazine

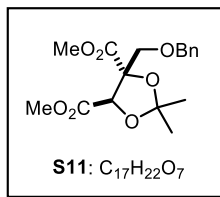


hydrate (20 mL) and NEt<sub>3</sub> (16.3 mL, 118 mmol) were added to a solution of ketone **13** (1.06 g, 5.88 mmol) in EtOH (45 mL). The reaction was heated to reflux for 20 h; upon cooling to 23 °C, CH<sub>2</sub>Cl<sub>2</sub> (100 mL) and H<sub>2</sub>O (150 mL)

were added. The aqueous layer was washed with CH<sub>2</sub>Cl<sub>2</sub> (3 x 100 mL), and the combined organic layers were dried over Mg<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated *in vacuo*. The remaining white solid (excess hydrazine) was removed by filtration using hexanes. Concentration *in vacuo* provided the crude hydrazone as a yellow oil, which was carried forward without further purification.

A solution of 1,1,3,3-Tetramethylguanidine (5.15 mL, 41.2 mmol) in THF (30 mL) was added dropwise over 10 min to a solution of I<sub>2</sub> (3.28 g, 12.9 mmol) in THF (30 mL). The hydrazone (5.88 mmol) in THF (6 mL) was then added dropwise over 10 min, and the reaction was maintained for 30 min. The dark red solution was then concentrated *in vacuo*, and the resulting red oil was heated neat at 90 °C for 5 h with a reflux condenser attached. The reaction was then cooled to 23 °C, diluted with Et<sub>2</sub>O (60 mL), and concentrated *in vacuo* over silica gel (~10 g). Purification by flash column chromatography (100% hexanes) provided light-sensitive vinyl iodide **19** (1.33 g, 4.58 mmol, 78%) as a colorless, crystalline solid. Spectral data were consistent with reported values.<sup>6,7</sup>

**(-)-Dimethyl (4*R*,5*R*)-4-((benzyloxy)methyl)-2,2-dimethyl-1,3-dioxolane-4,5-dicarboxylate**

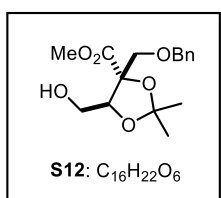


**(S11):** The procedure for the preparation of diester **S11** was a slight modification from the literature procedure.<sup>8</sup> Dimethyl 2,3,-*O*-isopropylidene-*L*-tartrate (3.83 g, 17.6 mmol) was dissolved in THF (67 mL) and cooled to –

78 °C. HMPA (13 mL) was added, followed by BOM-Cl (5.6 mL, 40 mmol). Freshly prepared



LDA (17.7 mmol) in THF (50 mL) was then added to the reaction flask via cannula over ~30 min. The reaction was maintained for 5 h at  $-78\text{ }^{\circ}\text{C}$ , before warming to  $0\text{ }^{\circ}\text{C}$ . After 3 h, the reaction was quenched with sat. aq.  $\text{NH}_4\text{Cl}$  solution (50 mL). The organic layer was washed with  $\text{H}_2\text{O}$  (3 x 40 mL) and brine (1 x 40 mL), dried over  $\text{MgSO}_4$ , and concentrated *in vacuo*. Unreacted dimethyl 2,3,-*O*-isopropylidene-*L*-tartrate was distilled from the crude product ( $120\text{ }^{\circ}\text{C}$ , 0.3 torr). The remaining oil was purified by flash column chromatography (8% EtOAc in hexanes to 15% EtOAc in hexanes) to provide diester **S11** (2.75 g, 8.14 mmol, 46%) as a light yellow oil. This reaction could be run on larger scale (~5x) with similar yields (41–43%).  $R_f$  0.80 (40% EtOAc in hexanes; visualized with ceric ammonium molybdate).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  7.34–7.23 (m, 5H), 5.12 (s, 1H), 4.52 (d,  $J = 12.2$  Hz, 1H), 4.47 (d,  $J = 12.2$  Hz, 1H), 3.82 (s, 3H), 3.73 (d,  $J = 9.8$  Hz, 1H), 3.70 (d,  $J = 9.8$  Hz, 1H), 3.63 (s, 3H), 1.59 (s, 3H), 1.42 (s, 3H);  $^{13}\text{C}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  170.80, 168.80, 137.60, 128.43, 127.77, 127.53, 112.71, 85.27, 77.56, 73.71, 70.11, 53.17, 52.40, 27.44, 25.95; IR (thin film) 2989, 2950, 1743, 1442, 1436, 1391, 1382, 1256, 1211  $\text{cm}^{-1}$ ;  $[\alpha]_D^{25}$ :  $-33.8$  ( $c = 1.7$ ,  $\text{CH}_2\text{Cl}_2$ ); HRMS (ESI) calculated for  $\text{C}_{17}\text{H}_{22}\text{O}_7\text{Na}$  ( $\text{M}+\text{Na}$ ) 361.1263, observed 361.1271.



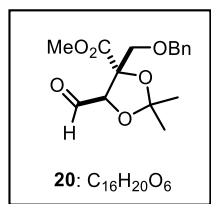
**(–)-Methyl (4*R*,5*S*)-4-((benzyloxy)methyl)-5-(hydroxymethyl)-2,2-dimethyl-1,3-dioxolane-4-carboxylate (S12)**: The procedure for the preparation of alcohol **S12** was a slight modification from the literature

procedure.<sup>8</sup> Diester **S11** (17.7 g, 52.3 mmol) was dissolved in THF (450 mL) and cooled to  $-78\text{ }^{\circ}\text{C}$ . DIBAL-H (14 mL, 79 mmol) was added dropwise to the reaction. After 5 min, the reaction was warmed to  $0\text{ }^{\circ}\text{C}$ . After 1 h, a saturated solution of Rochelle's salt (250 mL) and EtOAc (100 mL) were added. The reaction was allowed to warm to  $23\text{ }^{\circ}\text{C}$ , and the heterogeneous mixture was

extracted with EtOAc (4 x 150 mL). The combined organic layers were dried over MgSO<sub>4</sub>, filtered, and concentrated *in vacuo*. The crude residue was then purified by flash column chromatography (20% EtOAc in hexanes to 50% EtOAc in hexanes) to provide recovered diester **17** (6.34 g, 18.6 mmol, 36%) as a light yellow oil and alcohol **S12** (7.44 g, 23.9 mmol, 46%) as a clear oil. R<sub>f</sub> 0.35 (40% EtOAc in hexanes; visualized with ceric ammonium molybdate). <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 7.36–7.25 (m, 5H), 4.56–4.51 (m 3H), 3.91 (dd, *J* = 12.1, 5.3 Hz, 1H), 3.85 (dd, *J* = 12.2, 5.5 Hz, 1H), 3.80 (s, 3H), 3.72 (d, *J* = 9.4 Hz, 1H), 3.65 (d, *J* = 9.4 Hz, 1H), 2.37 (bs, 1H), 1.47 (s, 3H), 1.40 (s, 3H); <sup>13</sup>C NMR (500 MHz, CDCl<sub>3</sub>) δ 171.80, 137.24, 128.70, 128.07, 127.89, 110.16, 83.83, 73.94, 70.51, 60.65, 52.91, 27.75, 25.34; IR (thin film) 3500, 2989, 2937, 2871, 1743, 1454, 1380 cm<sup>-1</sup>; [α]<sub>D</sub><sup>25</sup>: -2.17 (c = 1.2); HRMS (ESI) calculated for C<sub>16</sub>H<sub>22</sub>O<sub>6</sub>NH<sub>4</sub> (M+NH<sub>4</sub>) 328.1760, observed 328.1754.

**(-)-Methyl**

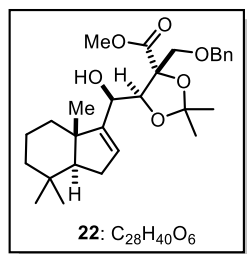
**(4*R*,5*R*)-4-((benzyloxy)methyl)-5-formyl-2,2-dimethyl-1,3-dioxolane-4-**



**carboxylate (20)**: To a stirring suspension of alcohol **S12** (4.80 g, 15.5 mmol) and NaHCO<sub>3</sub> (6.50 g, 77.4 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (40 mL) was added Dess-Martin periodinane (7.87 g, 18.6 mmol) in two portions over 5 min. After 2 h, the

reaction mixture was diluted with Et<sub>2</sub>O (40 mL) and filtered through a cotton plug to remove solid NaHCO<sub>3</sub>. The filtrate was concentrated *in vacuo*, resulting in a white solid. The solid was then washed with hexanes (6 x 30 mL), and the combined hexane washes were filtered through Celite. Upon concentration, aldehyde **20** (4.33 g, 14.0 mmol, 91%) was obtained as a colorless oil. Notes: 1) Aldehyde **20** was found to decompose within 14 h upon its formation (at room temperature or in the freezer), possibly due to self-aldol polymerization. Therefore, it was always carried forward *immediately* into the next reaction. 2) Aldehyde **20** did not appear unstable to column

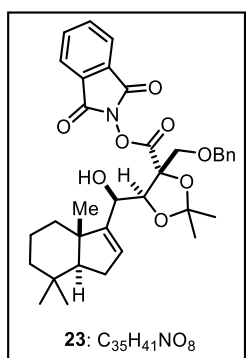
chromatography, but it could not be purified in that manner. 3) Aqueous washes diminished the yields, possibly from hydrate formation.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  9.69 (s, 1H); 7.36–7.24 (m, 5H), 4.89 (s, 1H), 4.47 (d,  $J = 12.1$  Hz, 1H), 4.44 (d,  $J = 12.1$  Hz, 1H), 3.82 (s, 3H), 3.66 (d,  $J = 10.0$  Hz, 1H), 3.63 (d,  $J = 10.0$  Hz, 1H), 1.59 (s, 3H), 1.42 (s, 3H);  $^{13}\text{C}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  197.23, 170.67, 137.15, 128.54, 127.84, 127.68, 112.75, 86.30, 82.95, 73.47, 69.28, 53.27, 27.30, 25.77; IR (thin film) 2991, 2937, 2868, 1740, 1454, 1374  $\text{cm}^{-1}$ ;  $[\alpha]_D^{25}$ :  $-2.37$  ( $c = 2.5$ ,  $\text{CH}_2\text{Cl}_2$ ); HRMS (ESI) calculated for  $[\text{C}_{16}\text{H}_{20}\text{O}_6\text{NH}_4]^+$  ( $\text{M}+\text{NH}_4$ ) 326.1604, observed 326.1612.



(–)-Methyl (4*R*,5*S*)-4-((benzyloxy)methyl)-5-((*R*)-hydroxy((3*aS*,7*aS*)-3*a*,7,7-trimethyl-3*a*,4,5,6,7,7*a*-hexahydro-1*H*-inden-3-yl)methyl)-2,2-dimethyl-1,3-dioxolane-4-carboxylate (**22**): *L*-Oxazoline **21**<sup>9</sup> (2.86 g, 9.65 mmol) and  $\text{CrCl}_2$  (1.19 g, 9.65 mmol) were dissolved in THF (20 mL) in the

glove box, and  $\text{NEt}_3$  (1.34 mL, 9.65 mmol) was then added. The suspension was vigorously stirred for 6 h, and then  $\text{NiCl}_2$  (36 mg, 0.28 mmol) was added, followed by a solution of vinyl iodide **19** (0.80 g, 2.8 mmol) and aldehyde **20** (1.30 g, 4.21 mmol) in THF (10 mL). Vigorous stirring was maintained for 20 h before removing the flask from the glovebox and cooling the solution to 0 °C. Ethylene diamine (2 mL) was added to quench the reaction. After stirring for 30 min,  $\text{H}_2\text{O}$  (40 mL) and  $\text{Et}_2\text{O}$  (40 mL) were added. The aqueous layer was extracted with  $\text{EtOAc}$  (4 x 20 mL), and the combined organic layers were washed with sat. aq.  $\text{NaHCO}_3$  solution (40 mL) and brine (1 x 40 mL), dried over  $\text{MgSO}_4$ , filtered, and concentrated *in vacuo*. Purification by flash column chromatography (5%  $\text{EtOAc}$  in hexanes to 11%  $\text{EtOAc}$  in hexanes) provided a single diastereomer, alcohol **22** (0.860 g, 1.82 mmol, 66%) as a clear oil. *L*-Oxazoline **21** was recovered during flash column chromatography (60–80% recovery) and recrystallized from  $\text{Et}_2\text{O}$ /hexanes for reuse.  $R_f$  0.50 for **22** (20%  $\text{EtOAc}$  in hexanes; visualized with ceric ammonium molybdate). Diagnostic

peaks for  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  7.36–7.24 (m, 5H), 5.78–5.74 (m, 1H), 4.61 (d,  $J = 12.6$  Hz, 1H), 4.55 (d,  $J = 12.6$  Hz, 1H), 4.44 (d,  $J = 9.0$  Hz, 1H), 4.37 (s, 1H), 3.97 (d,  $J = 9.8$  Hz, 1H), 3.80 (d,  $J = 9.8$  Hz, 1H), 3.77 (s, 3H), 2.58 (d,  $J = 9.0$  Hz, 1H), 2.12–1.99 (m, 2H), 1.74–1.70 (m, 2H), 1.58–1.56 (m, 1H), 1.54 (s, 3H), 1.5–1.43 (m, 2H), 1.42 (s, 3H), 1.25–1.19 (m, 1H), 1.11–1.01 (m, 1H), 0.98 (s, 3H), 0.95 (s, 3H), 0.88 (s, 3H);  $^{13}\text{C}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  172.59, 155.05, 137.89, 128.42, 127.76, 127.72, 126.03, 110.24, 80.41, 73.71, 71.94, 65.50, 59.97, 52.69, 41.51, 35.53, 33.29, 32.93, 28.76, 27.61, 25.46, 21.45, 20.15, 18.15; IR (thin film) 3527, 2989, 2926, 2848, 1741, 1454, 1380  $\text{cm}^{-1}$ ;  $[\alpha]_D^{25}$ :  $-4.85$  ( $c = 1.5$ ,  $\text{CH}_2\text{Cl}_2$ ); HRMS (ESI) calculated for  $\text{C}_{28}\text{H}_{40}\text{O}_6\text{NH}_4$  ( $\text{M}+\text{NH}_4$ ) 490.3169, observed 490.3165.



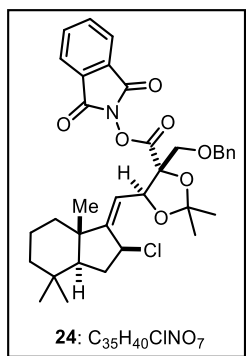
**(-)-1,3-Dioxoisindolin-2-yl (4*S*,5*R*)-4-((benzyloxy)methyl)-5-((*R*)-hydroxy((3*aS*,7*aS*)-3*a*,7,7-trimethyl-3*a*,4,5,6,7,7*a*-hexahydro-1*H*-inden-3-yl)methyl)-2,2-dimethyl-1,3-dioxolane-4-carboxylate (23):** Alcohol **22** (0.850 g, 1.80 mmol) was dissolved in a mixture of MeOH (10 mL) and  $\text{H}_2\text{O}$  (10 mL). Potassium hydroxide pellets (0.807 g, 14.4 mmol) were then added,

and the reaction was warmed to 50 °C. After 3 h, TLC analysis confirmed starting material was consumed; and the reaction was cooled to 23 °C. Aqueous HCl (18 mL of 1 M soln) was added to the flask, and the heterogeneous mixture was extracted with EtOAc (5 x 15 mL). The combined organic layers were washed with brine (1 x 20 mL), dried over  $\text{MgSO}_4$ , filtered, and concentrated *in vacuo* to provide the crude acid as a clear oil which was carried forward without further purification.

The crude acid was dissolved in THF (20 mL) to which *N*-hydroxyphthalimide (0.881 g, 5.40 mmol), DCC (0.483 g, 2.34 mmol), and DMAP (11 mg, 0.090 mmol) were added. The

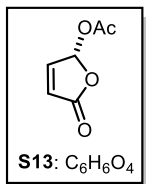
reaction was maintained for 3 h at 23 °C, at which point Celite (~2 g) was added. The reaction mixture was concentrated *in vacuo*, and the resulting residue was purified by flash column chromatography using pH 7 silica gel (10% EtOAc in hexanes to 20% EtOAc in hexanes) to provide *N*-acyloxyphthalimide **23** as a colorless solid. Recrystallization from acetone/hexanes afforded *N*-acyloxyphthalimide **23** (0.750 g, 1.24 mmol, 69%) as colorless needles.  $R_f$  0.25 (20% EtOAc in hexanes; visualized with ceric ammonium molybdate).  $^1\text{H}$  NMR (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.91–7.88 (2H, m), 7.81–7.78 (2H, m), 7.42–7.26 (m, 5H), 5.80 (s, 1H), 4.75 (d,  $J = 12.4$  Hz, 1H), 4.71 (d,  $J = 12.4$  Hz, 1H), 4.69 (s, 1H), 4.49 (d,  $J = 9.7$  Hz, 1H), 4.14 (d,  $J = 9.9$  Hz, 1H), 3.97 (d,  $J = 10.1$  Hz, 1H), 2.47 (d,  $J = 9.6$  Hz, 1H), 2.11–2.00 (m, 2H), 1.74–1.64 (m, 2H), 1.59 (s, 3H), 1.56–1.51 (m, 2H), 1.54 (s, 3H), 1.43 (app d,  $J = 13.3$  Hz, 1H), 1.27 (app td,  $J = 12.5, 3.7$  Hz, 1H), 1.13 (app td,  $J = 13.5, 4.3$  Hz, 1H), 0.98 (s, 3H), 0.95 (s, 3H), 0.88 (s, 3H);  $^{13}\text{C}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  169.29, 161.58, 154.82, 137.69, 134.94, 129.11, 128.46, 128.03, 127.75, 126.32, 124.17, 111.30, 85.13, 80.61, 74.20, 71.82, 65.56, 59.87, 47.34, 41.49, 35.42, 33.28, 32.93, 28.79, 27.39, 25.05, 21.48, 20.13, 18.18; IR (thin film) 3524, 2989, 2928, 2862, 1813, 1788, 1747, 1454, 1373  $\text{cm}^{-1}$ ;  $[\alpha]_D^{25}$  :  $-7.60$  ( $c = 1.6, \text{CH}_2\text{Cl}_2$ ); HRMS (ESI) calculated for  $\text{C}_{35}\text{H}_{41}\text{NO}_8\text{Na}$  ( $\text{M}+\text{Na}$ ) 626.2730, observed 626.2712; mp 139–141 °C.

(+)-1,3-dioxisoindolin-2-yl (4*R*,5*S*)-4-((benzyloxy)methyl)-5-(((2*S*,3*aS*,7*aS*,*Z*)-2-chloro-4,4,7*a*-trimethyloctahydro-1*H*-inden-1-ylidene)methyl)-2,2-dimethyl-



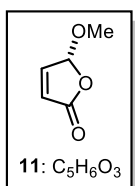
1,3-dioxolane-4-carboxylate (**24**): *N*-acyloxyphthalimide **23** (0.223 g, 0.369 mmol) was dissolved in a 10:1 mixture of  $\text{Et}_2\text{O}$ /pyridine (3.5 mL) and cooled to  $-45$  °C. A solution of  $\text{SOCl}_2$  (54  $\mu\text{L}$ , 0.74 mmol) in a 10:1 mixture of  $\text{Et}_2\text{O}$ /pyridine (0.5 mL) was then added dropwise to the reaction over 5

min. The reaction was maintained at  $-45\text{ }^{\circ}\text{C}$  until full conversion of starting material was observed by TLC analysis ( $\sim 45$  min). Saturated aq.  $\text{NaHCO}_3$  solution (2 mL) was added, and the reaction was allowed to warm to  $23\text{ }^{\circ}\text{C}$ . The mixture was then diluted with  $\text{H}_2\text{O}$  (2 mL) and washed with EtOAc (3 x 3 mL). The combined organic layers were washed with brine (1 x 2 mL), dried over  $\text{MgSO}_4$ , filtered, and concentrated *in vacuo* onto Celite ( $\sim 1$  g). Purification by flash column chromatography using pH 7 silica gel (5% EtOAc in hexanes to 11% EtOAc in hexanes) provided allylic chloride **24** as a colorless solid. Recrystallization from acetone/hexanes afforded allylic chloride **24** (0.143 g, 0.229 mmol, 62%) as colorless needles.  $R_f$  0.40 (20% EtOAc in hexanes; visualized with ceric ammonium molybdate).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  7.95-7.88 (m, 2H), 7.83-7.78 (m, 2H), 7.40-7.26 (m, 5H), 5.59 (d,  $J = 9.6$  Hz, 1H), 5.18 (d,  $J = 9.6$  Hz, 1H), 4.96 (app t,  $J = 7.6$  Hz, 1H), 4.70 (d,  $J = 12.3$  Hz, 1H), 4.64 (d,  $J = 12.3$  Hz, 1H), 3.78 (d,  $J = 10.0$  Hz, 1H), 3.71 (d,  $J = 10.0$  Hz, 1H), 2.32 (app quint,  $J = 6.4$  Hz, 1H), 1.84 (td,  $J = 13.7$  Hz, 7.5 Hz, 1H), 1.74 (app d,  $J = 12.6$  Hz, 1H), 1.67-1.59 (m, 1H), 1.59 (s, 6H), 1.55-1.48 (m, 1H), 1.41 (app d,  $J = 13.6$  Hz, 1H), 1.13 (s, 3H), 0.99-0.84 (m, 3H), 0.87 (s, 3H), 0.77 (s, 3H);  $^{13}\text{C}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  169.07, 161.68, 161.30, 137.64, 134.99, 129.09, 128.45, 127.95, 127.80, 124.25, 114.32, 111.61, 84.96, 76.44, 74.04, 71.07, 54.60, 54.10, 45.10, 41.19, 37.03, 34.08, 32.24, 32.81, 27.55, 24.78, 21.25, 21.11, 19.49; IR (thin film) 2986, 2928, 2866, 2350, 2336, 1813, 1787, 1747,  $1459\text{ cm}^{-1}$ ;  $[\alpha]_{\text{D}}^{25}$ : +83.2 ( $c = 1.8$ ,  $\text{CH}_2\text{Cl}_2$ ); HRMS (ESI) calculated for  $\text{C}_{35}\text{H}_{40}\text{ClNO}_7\text{Na}$  ( $\text{M}+\text{Na}$ ) 644.2391, observed 644.2383; mp  $154\text{--}158\text{ }^{\circ}\text{C}$ .



**(-)-(R)-5-acetoxypyrrolidin-2(1H)-one (S13):** The procedure for preparation of acetoxypyrrolidinone **S13** was a slight modification from the literature procedure.<sup>10</sup> 5-

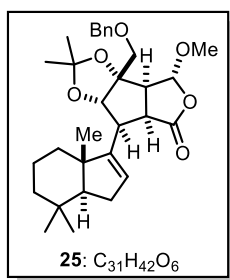
Hydroxypyrrolidin-2(1H)-one<sup>3</sup> (2.90 g, 28.9 mmol) was dissolved in vinyl acetate (30 mL). Amano lipase AK (2.00 g) was then added, and the suspension was stirred for 8 days at 23 °C. The suspension was then filtered through Celite, and the filtrate was concentrated *in vacuo*. Purification of the residue by flash column chromatography (40% EtOAc in hexanes) provided (-)-5-acetoxypyrrolidin-2(1H)-one **S13** (3.58 g, 25.3 mmol, 87% yield) as a yellow oil.  $R_f$  0.35 (40% EtOAc in hexanes; visualized with  $KMnO_4$ ). Spectral data were consistent with reported values.<sup>10</sup> The enantiomeric excess was determined to be 92% *ee* by known methods.<sup>10</sup>



**(-)-(R)-5-methoxypyrrolidin-2(1H)-one (11):** Acetoxypyrrolidinone **S13** (1.23 g, 8.65 mmol) was dissolved in MeOH (35 mL), and  $Pd(PPh_3)_4$  (0.500 g, 0.433 mmol) was added to the solution. The solution, which turned a deep red, was maintained at 23

°C for 50 min. Upon TLC analysis confirming consumption of starting material (TLC, 10% acetone in hexanes and running the TLC plate 3x), the reaction solution was directly filtered through a silica gel plug (250 mL of 40% acetone in hexanes). The eluent was concentrated *in vacuo*, and the residue was distilled (0.8 torr, 110 °C) to provide methoxypyrrolidinone **11** and a trace amount of AcOH. Removal of AcOH upon further concentration *in vacuo* afforded methoxypyrrolidinone **11** (0.705 g, 6.18 mmol, 71% yield) as a clear oil. Spectral data were consistent with reported values.<sup>11</sup> HPLC analysis was used to determine the enantiomeric ratio to be 92:8 (Chiracel AS column; flow: 2.0 mL/min, 10% isopropanol:*n*-hexane;  $\lambda$  = 210 nm; major enantiomer  $t_R$  = 8.70 min, minor enantiomer  $t_R$  = 11.60 min);  $[\alpha]_D^{25}$  : -124 ( $c$  = 1.2,  $CH_2Cl_2$ ).

(-)-(3a*S*,3b*S*,4*R*,6a*S*,7a*S*)-3a-((benzyloxy)methyl)-4-methoxy-2,2-dimethyl-7-((3a*S*,7a*S*)-



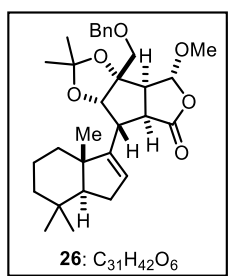
**3a,7,7-trimethyl-3a,4,5,6,7,7a-hexahydro-1H-inden-3-yl)hexahydro-6H-furo[3',4':3,4]cyclopenta[1,2-d][1,3]dioxol-6-one (25):** Allylic chloride **24** (70 mg, 0.11 mmol), methoxy butenolide **11** (51 mg, 0.45 mmol), D<sub>2</sub>-Hantzsch ester (43 mg, 0.17 mmol), and [Ru(bpy)<sub>3</sub>](PF<sub>6</sub>)<sub>2</sub> (1 mg, 0.001 mmol)

were charged into a vial. Acetonitrile (1.1 mL) was added, and the solution was sparged with Ar. The vial was then vigorously stirred while being irradiated by a single strip of blue LED lights (450 nm) at 23 °C. After 6 h, the reaction mixture was concentrated *in vacuo*, and the residue was dissolved in EtOAc (1 mL) and washed with aq. HCl (4 x 2 mL of 4 M soln) followed by H<sub>2</sub>O (2 x 2 mL). The organic layer was dried over MgSO<sub>4</sub>, filtered, and concentrated *in vacuo*. <sup>1</sup>H NMR analysis of the crude residue using an internal standard (dimethoxybenzene) showed 28% yield of **25**, 37% yield of **26**, 8% yield of **27**, and 13% yield of **28**. Purification of the crude residue by flash column chromatography (0% acetone in hexanes to 5% acetone in hexanes) provided **25** (15 mg, 0.030 mmol, 27%) as a clear oil. R<sub>f</sub> for **25**: 0.55 (20% acetone in hexanes; visualized with ceric ammonium molybdate). Flash column chromatography under separate conditions of the remaining mixed fractions from the first purification (4% EtOAc in hexanes to 10% EtOAc in hexanes) provided epimeric product **26** (20 mg, 0.039 mmol, 35%) as a clear oil. R<sub>f</sub> for **26**: 0.45 (20% acetone in hexanes; visualized with ceric ammonium molybdate).

Desired ACF product **25** for <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 7.38–7.28 (m, 5H), 5.48 (app s, 1H), 5.38 (app s, 1H), 4.62 (d, *J* = 12.0 Hz, 1H), 4.59 (d, *J* = 12.1 Hz, 1H), 4.41 (d, *J* = 7.5 Hz, 1H), 3.64 (d, *J* = 10.5 Hz, 1H), 3.50 (d, *J* = 10.3 Hz, 1H), 3.43 (app t, *J* = 8.7 Hz, 1H), 3.38 (s, 3H), 3.07 (app d, *J* = 8.7 Hz, 1H), 3.00 (app t, *J* = 8.2 Hz, 1H), 2.10 (ddd, *J* = 14.9, 6.3, 3.0 Hz, 1H), 2.02 (app t, *J* = 13.3 Hz, 1H), 1.76 (dd, *J* = 11.7, 6.3 Hz, 1H), 1.60–1.50 (m, 2H), 1.58 (s,

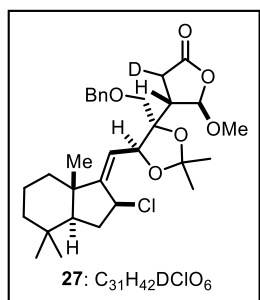


3H), 1.50 (s, 3H), 1.45–1.39 (m, 1H), 1.19 (td,  $J = 13.2, 3.4$  Hz, 1H), 0.95 (s, 3H), 0.92–0.82 (m, 2H), 0.89 (s, 3H), 0.87 (s, 3H);  $^{13}\text{C}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  174.65, 150.69, 137.18, 128.71, 128.38, 128.30, 124.70, 113.17, 103.64, 89.95, 86.77, 73.97, 70.81, 58.24, 56.76, 55.12, 47.84, 45.94, 43.72, 41.46, 34.73, 33.05, 32.90, 20.22, 29.38, 29.16, 21.45, 20.18, 17.70;  $[\alpha]_{\text{D}}^{25}$ :  $-84.9$  ( $c = 1.0, \text{CH}_2\text{Cl}_2$ ); IR (thin film) 2993, 2934, 2862, 1785, 1636, 1455, 1371, 1234, 1215  $\text{cm}^{-1}$ ; HRMS (ESI) calculated for  $\text{C}_{31}\text{H}_{42}\text{O}_6\text{NH}_4$  ( $\text{M}+\text{NH}_4$ ) 528.3325, observed 528.3331.



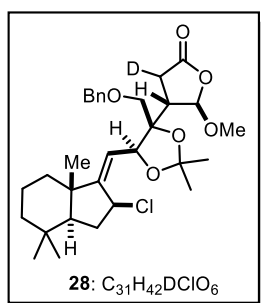
Epimer ( $-$ )-**26** for  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  7.39–7.29 (m, 5H), 5.75 (app s, 1H), 5.53 (d,  $J = 4.8$  Hz, 1H), 4.57 (d,  $J = 12.3$  Hz, 1H), 4.55 (d,  $J = 12.3$  Hz, 1H), 4.47 (d,  $J = 3.7$  Hz, 1H), 3.82 (d,  $J = 9.6$  Hz, 1H), 3.61 (d,  $J = 9.7$  Hz, 1H), 3.50 (app t,  $J = 10.8$  Hz, 1H), 3.41 (s, 3H), 2.88 (dd,  $J = 9.7, 4.8$  Hz, 1H),

2.72 (app d,  $J = 11.3$  Hz, 1H), 2.13–2.06 (m, 2H), 1.76–1.65 (m, 1H), 1.61–1.52 (m, 2H), 1.43 (s, 3H), 1.36–1.27 (m, 1H), 1.28 (s, 3H), 1.15 (td,  $J = 13.6, 4.1$  Hz, 1H), 0.96 (s, 3H), 0.92–0.81 (m, 2H), 0.89 (s, 3H), 0.83 (s, 3H);  $^{13}\text{C}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  175.80, 148.52, 137.12, 128.69, 128.26, 128.05, 127.25, 111.58, 105.02, 89.60, 87.36, 73.83, 72.05, 59.28, 58.09, 57.57, 47.71, 47.54, 45.77, 41.61, 35.68, 33.09, 32.97, 29.17, 27.80, 26.00, 21.47, 20.22, 17.36;  $[\alpha]_{\text{D}}^{25}$ :  $-88.2$  ( $c = 2.0, \text{CH}_2\text{Cl}_2$ ); IR (thin film) 2988, 2929, 2861, 1775, 1454, 1373, 1246  $\text{cm}^{-1}$ ; HRMS (ESI) calculated for  $\text{C}_{31}\text{H}_{42}\text{O}_6\text{Na}$  ( $\text{M}+\text{Na}$ ) 533.2879, observed 533.2897.

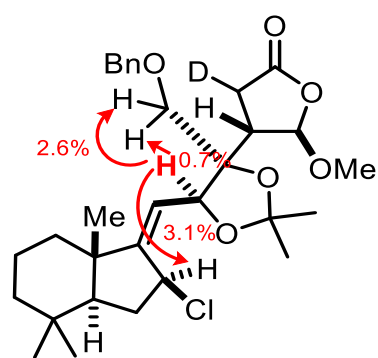
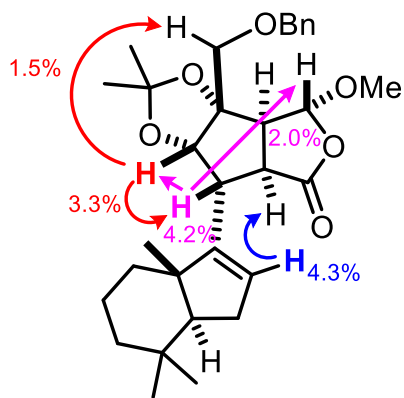
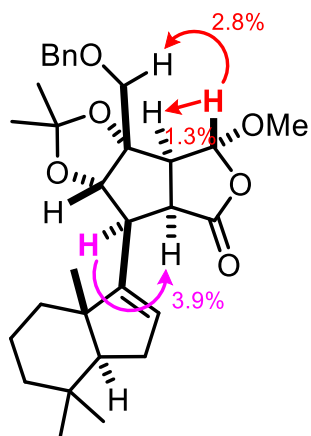


An analytical sample of clean product ( $+$ )-**27** was obtained from flash column chromatography (0% acetone in hexanes to 4% acetone in hexanes).  $R_f$ : 0.60 (20% acetone in hexanes; visualized with ceric ammonium molybdate).  $^1\text{H}$  NMR (600 MHz,  $\text{CDCl}_3$ )  $\delta$  7.36–7.27 (m, 5H), 5.61 (s, 1H),

5.18 (d,  $J = 9.6$ , 1H), 5.09 (dd,  $J = 9.6$ , 1.6 Hz, 1H), 4.57 (td,  $J = 8.0$  Hz, 1.6 Hz, 1H), 4.46 (d,  $J = 10.4$  Hz, 1H), 4.41 (d,  $J = 10.4$  Hz, 1H), 3.63 (d,  $J = 9.6$  Hz, 1H), 3.53 (d,  $J = 9.6$  Hz, 1H), 3.51 (s, 3H), 2.68 (d,  $J = 2.1$  Hz, 1H), 2.50 (d,  $J = 2.1$  Hz, 1H), 1.93–1.86 (m, 1H), 1.76–1.51 (m, 4H), 1.48 (s, 3H), 1.42 (s, 3H), 1.09 (s, 3H), 1.04–0.95 (m, 2H), 0.91–0.83 (m, 1H), 0.85 (s, 3H), 0.74 (s, 3H), 0.60 (dd,  $J = 14.4$ , 6.0 Hz, 1H);  $^{13}\text{C}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  176.81, 159.67, 137.02, 128.73, 128.34, 128.26, 115.56, 109.03, 106.81, 82.60, 78.24, 74.46, 74.39, 56.90, 54.57, 54.53, 45.17, 44.93, 41.17, 36.72, 33.93, 33.15, 32.96, 29.85, 27.58, 26.19, 21.49, 21.15, 19.49;  $[\alpha]_{\text{D}}^{25}$ : +109.1 ( $c = 0.57$ ,  $\text{CH}_2\text{Cl}_2$ ); IR (thin film) 2986, 2931, 2864, 2359, 2342, 1787, 1455, 1370, 1252  $\text{cm}^{-1}$ ; HRMS (ESI) calculated for  $\text{C}_{31}\text{H}_{42}\text{DClO}_6\text{Na}$  ( $\text{M}+\text{Na}$ ) 570.2709, observed 570.2702.

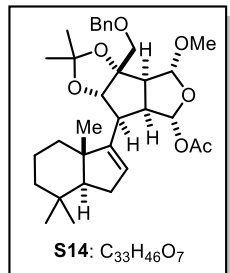


Diagnostic peaks of addition product **28** for  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  5.43 (d,  $J = 2.9$  Hz, 1H), 5.21 (d,  $J = 9.6$ , 1.7 Hz, 1H), 4.75 (d,  $J = 9.6$  Hz, 1H), 4.59 (app t,  $J = 7.0$  Hz, 1H).



(-)-(3aS,3bS,4R,6R,6aS,7S,7aS)-3a-((benzyloxy)methyl)-4-methoxy-2,2-dimethyl-7-  
((3aS,7aS)-3a,7,7-trimethyl-3a,4,5,6,7,7a-hexahydro-1H-inden-3-yl)hexahydro-4H-

furo[3',4':3,4]cyclopenta[1,2-d][1,3]dioxol-6-yl acetate (**S14**): Product **25** (40 mg, 0.078 mmol)



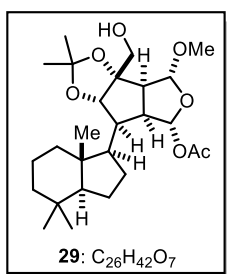
was charged into a flask with toluene (1.4 mL) and then cooled to  $-78\text{ }^{\circ}\text{C}$ . A solution of DIBAL-H (18  $\mu\text{L}$ , 0.10 mmol) in toluene (0.2 mL) was added dropwise to the reaction vessel, keeping the temperature near  $-78\text{ }^{\circ}\text{C}$ . After 45 min, TLC analysis showed some remaining starting material, and an additional

solution of DIBAL-H (5  $\mu\text{L}$ , 0.03 mmol) in toluene (0.05 mL) was added. After 45 min, a solution of DMAP (19 mg, 0.16 mmol), pyridine (20  $\mu\text{L}$ , 0.23 mmol), and  $\text{CH}_2\text{Cl}_2$  (0.2 mL) was added, followed by  $\text{Ac}_2\text{O}$  (44  $\mu\text{L}$ , 0.47 mmol). The reaction was maintained at  $-78\text{ }^{\circ}\text{C}$  for 12 h, at which point it was allowed to warm to  $23\text{ }^{\circ}\text{C}$ . An aqueous solution saturated with Rochelle's salt (3 mL) was added, and the aqueous layer was extracted with EtOAc (3 x 2 mL). The combined organic layers were washed with brine (1 x 5 mL), dried over  $\text{MgSO}_4$ , filtered, and concentrated *in vacuo*. Purification of the residue by flash column chromatography (6% EtOAc in hexanes to 10% EtOAc in hexanes) provided a single diastereomer, diacetal **S14** (36 mg, 0.065 mmol, 83%), as a colorless oil.  $R_f$  0.35 (20% EtOAc in hexanes; visualized with ceric ammonium molybdate).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  7.38–7.27 (m, 5H), 5.92 (d,  $J = 4.1$  Hz, 1H), 5.60 (app s, 1H), 5.17 (s, 1H), 4.61 (d,  $J = 12.3$  Hz, 1H), 4.58 (d,  $J = 12.3$  Hz, 1H), 4.30 (d,  $J = 8.8$  Hz, 1H), 3.60 (d,  $J = 10.4$  Hz, 1H), 3.54 (d,  $J = 10.5$  Hz, 1H), 3.27 (s, 3H), 3.20 (app td,  $J = 7.8, 4.0$  Hz, 1H), 3.03 (d,  $J = 8.0$  Hz, 1H), 2.92 (app t,  $J = 8.1$  Hz, 1H), 2.05 (s, 3H), 2.06–2.00 (m, 2H), 1.72–1.51 (m, 2H), 1.51 (s, 3H), 1.46 (app d,  $J = 13.6$  Hz, 1H), 1.35 (s, 3H), 1.28–1.24 (m, 1H), 1.20 (td,  $J = 12.7, 3.9$  Hz, 1H), 0.98 (td,  $J = 13.7, 4.4$  Hz, 1H), 0.93 (s, 3H), 0.90–0.85 (m, 1H), 0.84 (s, 6H);  $^{13}\text{C}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  170.46, 151.77, 137.54, 128.61, 128.36, 128.13, 125.03, 113.25, 107.80, 99.65, 90.22, 85.91,

73.84, 70.54, 50.88, 47.64, 43.33, 42.11, 36.75, 35.58, 33.29, 32.95, 31.72, 30.64, 29.72, 29.01, 24.81, 22.79, 21.25, 21.17, 20.09, 17.24, 14.27;  $[\alpha]_D^{25}$ :  $-46.3$  ( $c = 2.1$ ,  $\text{CH}_2\text{Cl}_2$ ); IR (thin film) 2991, 2930, 2861, 1748, 1455, 1367  $\text{cm}^{-1}$ ; HRMS (ESI) calculated for  $\text{C}_{33}\text{H}_{46}\text{O}_7\text{Na}$  ( $\text{M}+\text{Na}$ ) 577.3141, observed 577.3127.

**(-)-(3a*S*,3b*S*,4*R*,6*R*,6a*S*,7*R*,7a*S*)-3a-(hydroxymethyl)-4-methoxy-2,2-dimethyl-7-**

**((1*R*,3a*S*,7a*R*)-4,4,7a-trimethyloctahydro-1*H*-inden-1-yl)hexahydro-4*H*-**



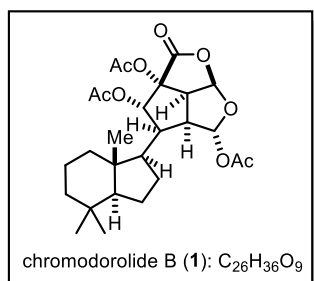
**furo[3',4':3,4]cyclopenta[1,2-d][1,3]dioxol-6-yl acetate (29):** Diacetal **S14**

(28 mg, 0.050 mmol) and 10% Pd/C (28 mg) were charged into a flask with MeOH (1.0 mL). The reaction vessel was then evacuated and refilled with Ar (**3x**). Formic acid (50  $\mu\text{L}$ ) was then added dropwise to the vigorously stirring

suspension. After 2 h, TLC analysis showed full consumption of starting material. The reaction mixture was diluted with MeOH (1 mL), filtered through Celite, and concentrated *in vacuo* to provide the crude alcohol, which was carried forward to the subsequent step.

To a flask containing the crude alcohol (0.050 mmol) was added  $\text{PtO}_2$  (12 mg, 0.050 mmol) and EtOAc (1.0 mL). The reaction vessel was then evacuated and refilled with  $\text{H}_2$  (**3x**, 1 atm  $\text{H}_2$ ). The reaction was maintained under 1 atm of  $\text{H}_2$  for 12 h at 23  $^\circ\text{C}$ , at which point the reaction vessel was refilled first with Ar and then air. Filtration of the suspension through Celite, concentration of the filtrate *in vacuo*, and purification of the residue by flash column chromatography (30% EtOAc in hexanes) provided alcohol **29** (20 mg, 0.043 mmol, 86%) as a colorless oil.  $R_f$  0.25 (30% EtOAc in hexanes; visualized with ceric ammonium molybdate).  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  6.13 (d,  $J = 3.5$  Hz, 1H), 5.31 (s, 1H), 3.86 (d,  $J = 9.6$  Hz, 1H), 3.65 (bs, 2H), 3.31 (s, 3H), 3.19 (app td,  $J = 7.4, 3.6$  Hz, 1H), 2.87 (app d,  $J = 7.6$  Hz, 1H), 2.30 (app dt,  $J = 10.1, 7.3$  Hz, 1H), 2.16 (bs, 1H),

2.05 (s, 3H), 1.80–1.66 (m, 2H), 1.63–1.56 (m, 2H), 1.53 (s, 3H), 1.43 (s, 6H), 1.36–1.28 (m, 1H), 1.11–0.93 (m, 2H), 0.90–0.86 (m, 1H), 0.85 (s, 3H), 0.83 (s, 3H), 0.76 (s, 3H), 0.76–0.69 (m, 1H);  $^{13}\text{C}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  170.84, 112.73, 106.84, 97.67, 90.98, 88.07, 63.62, 57.74, 56.04, 54.85, 52.29, 50.68, 44.98, 42.90, 41.39, 40.00, 33.60, 33.26, 30.71, 30.49, 29.84, 25.75, 21.20, 20.99, 20.94, 20.22, 13.86;  $[\alpha]_D^{25}$ :  $-17.6$  ( $c = 1.7$ ,  $\text{CH}_2\text{Cl}_2$ ); IR (thin film) 3490, 2951, 2931, 2873, 1745, 1459, 1368  $\text{cm}^{-1}$ ; HRMS (ESI) calculated for  $\text{C}_{26}\text{H}_{42}\text{O}_7\text{Na}$  ( $\text{M}+\text{Na}$ ) 489.2828, observed 489.2813.



**(–)-Chromodorolide B (1):** Alcohol **29** (9.0 mg, 0.019 mmol) and Dess-Martin periodinane (12 mg, 0.029 mmol) were charged into a flask with  $\text{CH}_2\text{Cl}_2$  (0.3 mL). The reaction mixture was maintained at 23  $^\circ\text{C}$  for 5 h, at which point it was diluted with hexanes (0.5 mL), filtered

through Celite, and concentrated *in vacuo*. The residue was dissolved in hexanes (1 mL) and filtered through Celite. The filtrate was then concentrated *in vacuo* to afford the crude aldehyde which was carried forward into the next step.

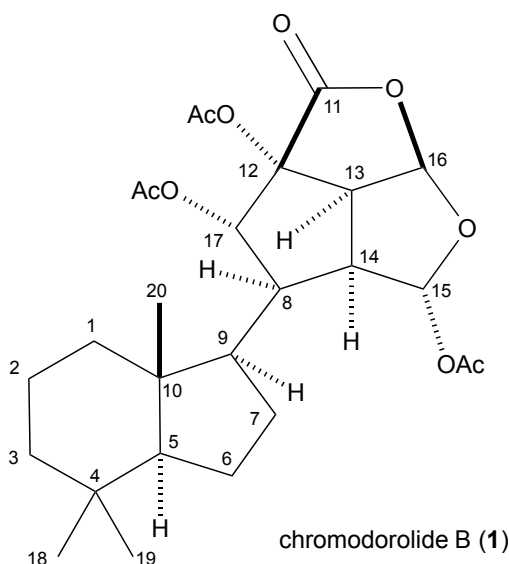
To a solution of crude aldehyde in THF (0.1 mL) was added *t*-BuOH (0.1 mL),  $\text{H}_2\text{O}$  (0.1 mL), 2-methyl-2-butene (50  $\mu\text{L}$ ),  $\text{NaH}_2\text{PO}_4$  (25 mg, 0.21 mmol), and  $\text{NaClO}_2$  (14 mg, 0.15 mmol). The reaction was maintained at 23 $^\circ\text{C}$  for 12 h and then diluted with  $\text{H}_2\text{O}$  (1 mL). The solution was washed with EtOAc (3  $\times$  1 mL); and the combined organic layers were dried over  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated *in vacuo* to provide crude carboxylic acid **30**.

This crude acid **30** was then dissolved in a solution of THF (0.3 mL) and aq. HCl (0.3 mL of 4 M soln), which was maintained at 23  $^\circ\text{C}$  for 72 h. The reaction was then diluted with  $\text{H}_2\text{O}$  (1 mL), and the solution was washed with EtOAc (3  $\times$  1 mL). The combined organic layers were

washed with brine (1 x 1 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated *in vacuo* to afford crude lactol **31**.

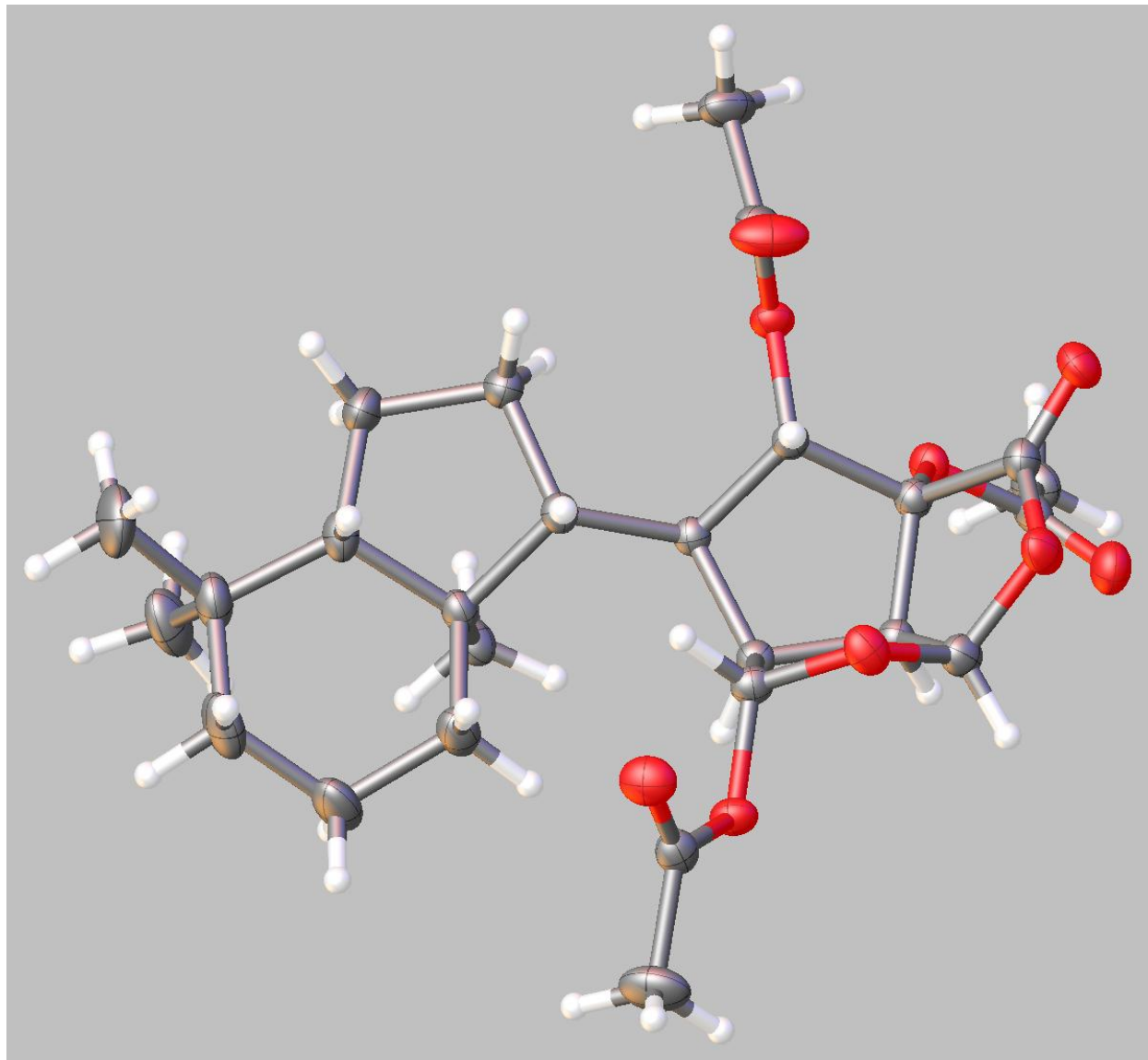
Crude lactol **31** was then dissolved in CH<sub>2</sub>Cl<sub>2</sub> (0.3 mL). Next, DMAP (2 mg, 0.019 mmol) and pyridine (31 μL, 0.38 mmol) were added, followed by Ac<sub>2</sub>O (28 μL, 0.29 mmol). The reaction was maintained at 23 °C for 24 h, at which point it was diluted with H<sub>2</sub>O (2 mL), and the heterogeneous solution was washed with EtOAc (3 x 2 mL). The combined organic layers were washed with brine (1 x 3 mL), dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated *in vacuo*. Purification by flash column chromatography (20% EtOAc in hexanes to 30% EtOAc in hexanes) provided **1** (4.7 mg, 0.010 mmol, 49% over 4 steps) as a colorless solid. Recrystallization of the solid from acetone/hexanes afforded colorless needles. The NMR data matched that of the isolation data.<sup>12</sup>  $[\alpha]_{\text{D}}^{25}$ : -66.8 (c = 0.12, CH<sub>2</sub>Cl<sub>2</sub>) compared to isolation sample  $[\alpha]_{\text{D}}^{25}$ : -95 (c = 0.10, CH<sub>2</sub>Cl<sub>2</sub>)<sup>12</sup>; IR (thin film) 2948, 2876, 1813, 1752, 1370, 1214, 1093, 1000, 964 cm<sup>-1</sup>; HRMS (ESI) calculated for C<sub>26</sub>H<sub>36</sub>O<sub>9</sub>Na (M+Na) 515.2257, observed 515.2260; mp 236–238 °C (decomp).

**Table S1.** Synthetic and natural (–)-Chromodorolide B (**1**) in CDCl<sub>3</sub>.

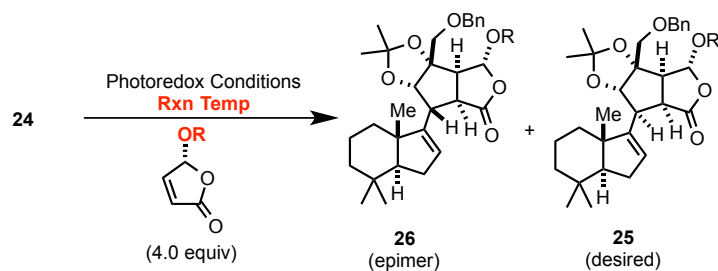


Pos.	Literature (400 MHz, CDCl <sub>3</sub> )		Synthetic (500 MHz, CDCl <sub>3</sub> )	
	δH (multiplicity, <i>J</i> / Hz)	δC	δH (multiplicity, <i>J</i> / Hz)	δC
1	1.03 (dt, <i>J</i> = 12.3, 3.6)	40.9	1.03 (dt, <i>J</i> = 12.3, 3.6)	41.06
1a	1.38 (m)		1.39 (m)	
2	1.55 (m, 2H)	21.1	1.56 (m, 2H)	21.27
3	0.95 (m)	39.1	0.95 (dt, <i>J</i> = 12.6, 3.6)	39.26
3a	1.51 (m)		1.52 (m)	
4		33.1		33.31
5	1.09 (dd, <i>J</i> = 13.1, 6.6)	57.0	1.10 (dd, <i>J</i> = 13.7, 3.7)	57.10
6	1.40 (m)	19.9	1.42 (m)	20.08
6a	1.56 (m)		1.57 (m)	
7	1.48 (m)	25.2	1.50 (m)	25.36
7a	1.58 (m)		1.61 (m)	
8	2.57 (ddd, <i>J</i> = 12.1, 11.6, 7.9)	48.0	2.58 (ddd, <i>J</i> = 12.2, 11.1, 7.6)	48.13
9	1.69 (bdd, <i>J</i> = 12.0, 9.9)	50.3	1.71 (q, <i>J</i> = 10.0)	50.43
10		43.9		44.00
11		169.1		169.24
12		81.4		81.48
13	3.79 (dd, <i>J</i> = 8.9, 6.1)	50.4	3.80 (dd, <i>J</i> = 9.0, 6.0)	50.56
14	2.93 (bt, <i>J</i> = 8.2)	45.6	2.94 (bt, <i>J</i> = 8.2)	45.74
15	6.50 (bs)	97.8	6.51 (bs)	97.88
16	6.08 (d, <i>J</i> = 6.1)	103.4	6.09 (d, <i>J</i> = 6.0)	103.51
17	5.30 (d, <i>J</i> = 11.6)	73.9	5.31 (d, <i>J</i> = 12.4)	74.06
18	0.79 (s, 3H)	33.4	0.79 (s, 3H)	33.55
19	0.83 (s, 3H)	21.0	0.84 (s, 3H)	21.11
20	0.84 (s, 3H)	13.7	0.85 (s, 3H)	13.82
OAc	2.04 (s, 3H)	20.8	2.05 (s, 3H)	20.96
		169.2		169.32
OAc	2.11 (s, 3H)	20.8	2.12 (s, 3H)	20.98
		170.0		170.13
OAc	2.19 (s, 3H)	20.9	2.21 (s, 3H)	21.05
		170.2		170.38

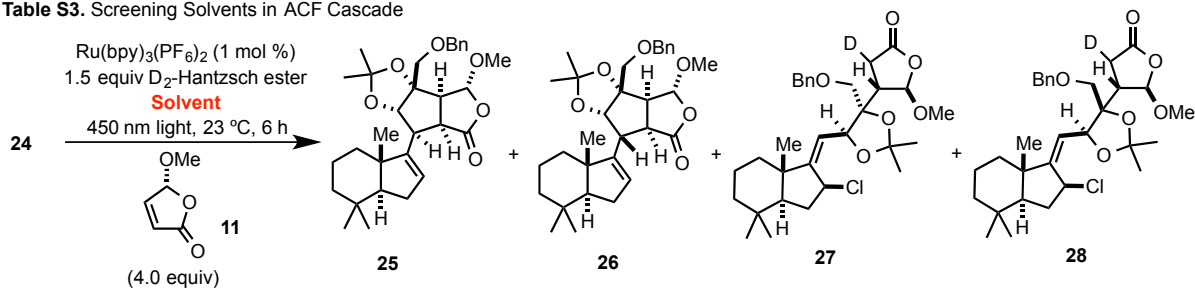
X-ray image of (-)-Chromodorolide B (CCDC1446027)





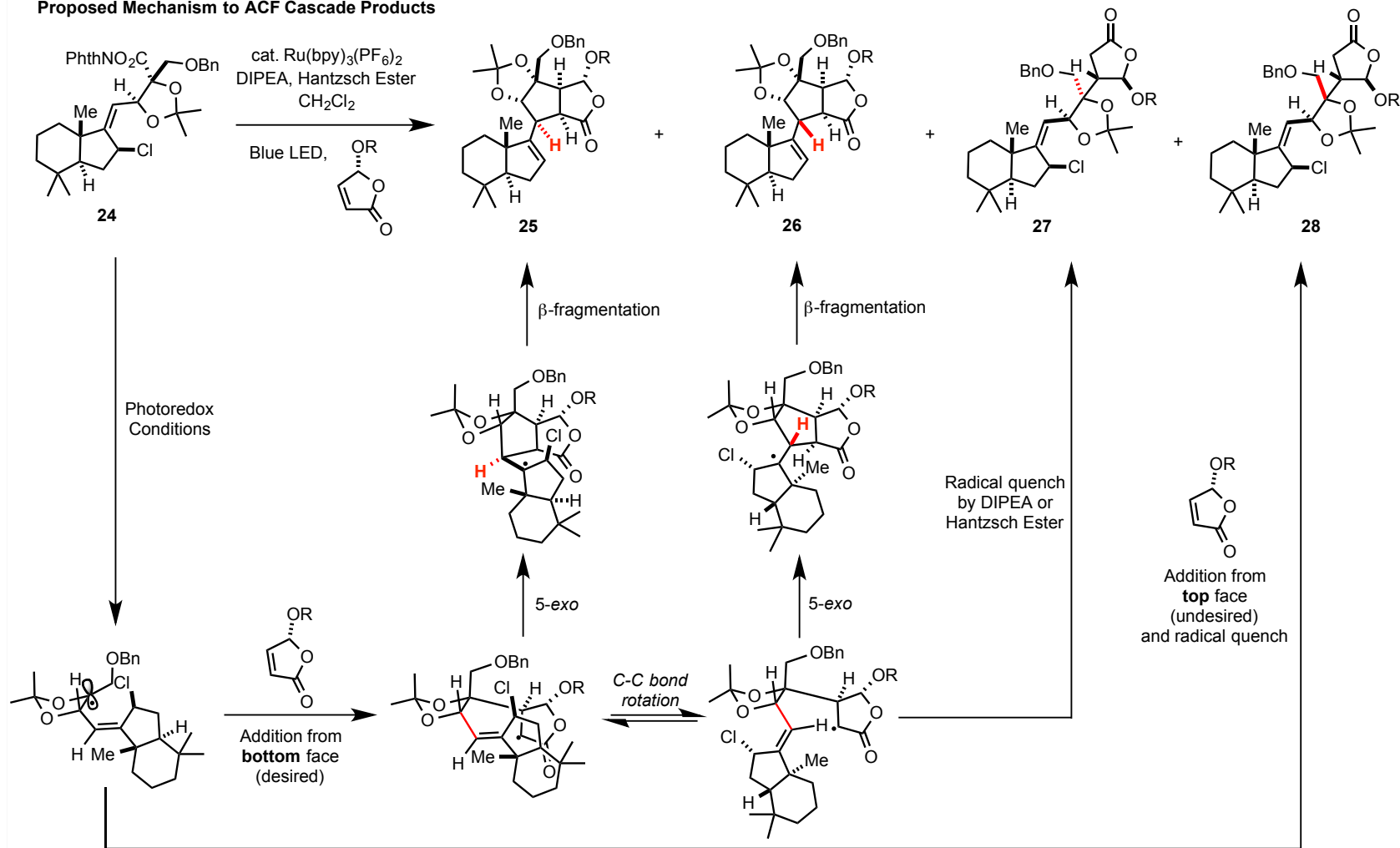
**Table S2.** Effects of Butenolide Substitution and Temperature on Diastereoselectivity

<u>Entry</u>	<u>R</u>	<u>Rxn Temp</u>	<u>Diastereoselectivity (26:25)</u>
1	Menthol	23 °C	3.1 : 1.0
2	Ac	23 °C	2.7 : 1.0
3	Me	23 °C	1.8 : 1.0
4	Me	38 °C	1.8 : 1.0
5	Me	0 °C	1.5 : 1.0

**Table S3.** Screening Solvents in ACF Cascade

<u>Entry</u>	<u>Solvent</u>	<u>Respective Yields by <sup>1</sup>H NMR (with internal standard)</u>	<u>dr 26:25</u>
1	CH <sub>2</sub> Cl <sub>2</sub>	25% : 45% : 6% : 10%	1.8 : 1
2	DME	23% : 36% : 4% : 9%	1.6 : 1
3	THF	21% : 34% : 5% : 8%	1.6 : 1
4	MeCN	28% : 37% : 8% : 13%	1.3 : 1
5	MeOH	12% : 24% : 15% : 12%	2.0 : 1
6	DMSO	17% : 20% : 17% : 15%	1.2 : 1

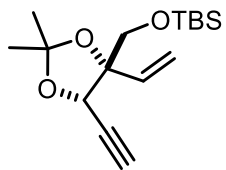
**Proposed Mechanism to ACF Cascade Products**



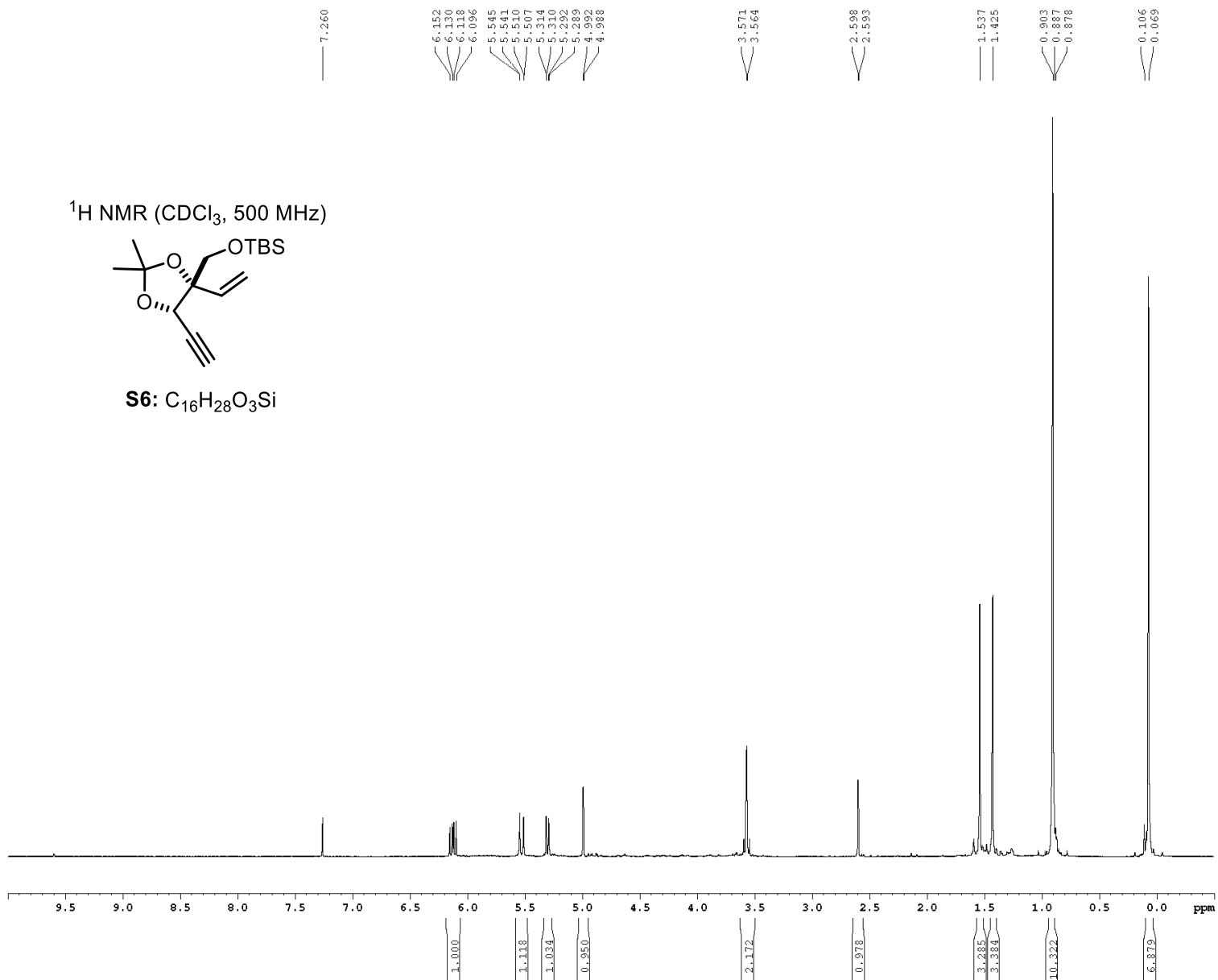
## **References:**

1. (a) Eey, S. T. C.; Lear, M. J. *Org. Lett.* **2010**, *12*, 5510–5513. (b) Larraufie, M.-H.; Pellet, R.; Fensterbank, L.; Goddard, G.-P.; Lacote, E.; Malacria, M.; Ollivier, C. *Angew. Chem., Int. Ed.* **2001**, *50*, 4463–4466.
2. Kim, H.-J.; Ricardo, A.; Illangkoon, H. I.; Kim, M. J.; Carrigan, M. A.; Frye, F.; Benner, S. A. *J. Am. Chem. Soc.* **2011**, *133*, 9457–9468.
3. Moradei, O. M.; Paquette, L. A. *Org. Synth.* **2003**, *80*, 66.
4. X-ray coordinates were deposited with the Cambridge Crystallographic Data Centre: 1447146.
5. Shigehisa, H.; Mizutani, T.; Tosaki, S.-Y.; Ohshima, T.; Shibasaki, M. *Tetrahedron*, **2005**, *61*, 5057–5065.
6. Brady, T. P.; Kim, S. H.; Wen, K.; Kim, C.; Theodorakis, E. A. *Chem. Eur. J.* **2005**, *11*, 7175–7190.
7. Granger, K.; Snapper, M. L. *Eur. J. Org. Chem.* **2012**, 2308–2311.
8. Crich, D.; Hao, X. *J. Org. Chem.* **1999**, *64*, 4016–4024.
9. (a) Wan, Z.-K.; Choi, H. W.; Kang, F.-A.; Nakajima, K.; Demeke, D.; Kishi, Y. *Org. Lett.* **2002**, *4*, 4431–4434. (b) Choi, H. W.; Nakajima, K.; Demeke, D.; Kang, F.-A.; Jun, H.-S.; Wan, Z.-K.; Kishi, Y. *Org. Lett.* **2002**, *4*, 4435–4438.
10. Morita, Y.; Tokuyama, H.; Fukuyama, T. *Org. Lett.* **2005**, *7*, 4337–4340.
11. Feringa, B. L.; De Lange, B. *Tetrahedron* **1988**, *44*, 7213–7222.
12. MorGris, S. A.; Dilip de Silva, E.; Andersen, R. J. *Can. J. Chem.* **1991**, *69*, 768–771.

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)



**S6:** C<sub>16</sub>H<sub>28</sub>O<sub>3</sub>Si



```

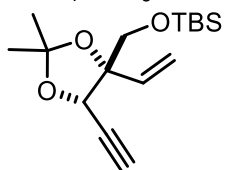
Current Data Parameters
NAME      DJF-II-295
EXPNO    3
PROCNO   1

F2 - Acquisition Parameters
Date_    20131128
Time     15.24
INSTRUM  cryo500
PROBHD   5 mm CPTCI LH-
PULPROG  zg30
TD        81728
SOLVENT  CDCl3
NS        8
DS        2
SWH       8012.820 Hz
FIDRES    0.098043 Hz
AQ        5.0998273 sec
RG         8
DW         62.400 usec
DE         6.00 usec
TE         298.0 K
D1         0.10000000 sec
MREST     0 sec
NEWXK     0.01500000 sec

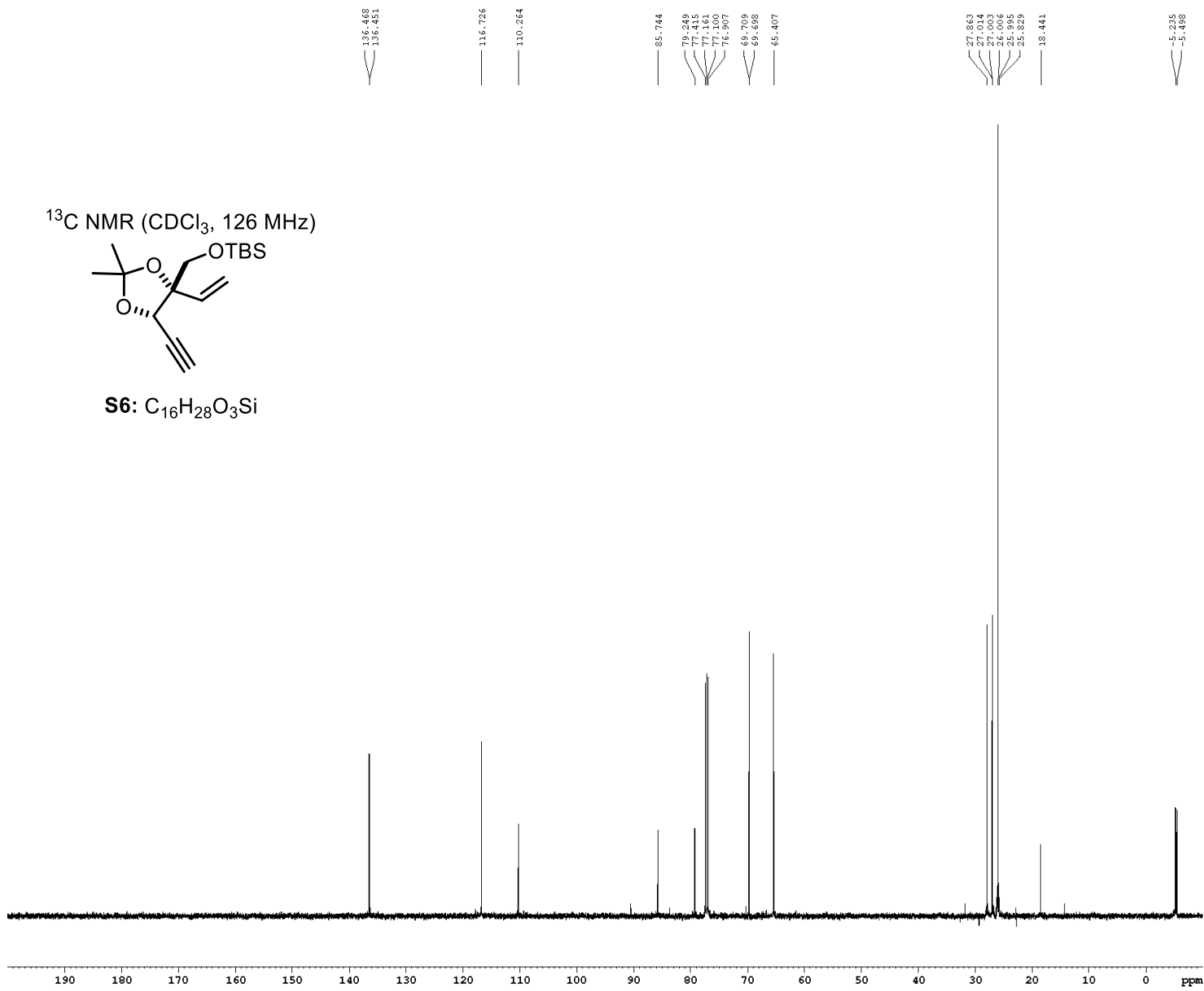
===== CHANNEL f1 =====
NUC1      1H
P1         7.50 usec
PL1        1.60 dB
SFO1      500.225015 MHz

F2 - Processing parameters
SI         6536
SF         500.2200308 MHz
WDW        no
SSB        0
LB         0 Hz
GB         0
PC         4.00
    
```

<sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz)



S6: C<sub>16</sub>H<sub>28</sub>O<sub>3</sub>Si



```
Current Data Parameters
NAME      DJT-II-295
EXPNO     4
PROCNO    1

F2 - Acquisition Parameters
Date_     20131130
Time      15.34
INSTRUM   cryo500
PROBHD    5 mm CPTCI 1H-
PULPROG   SpinEchoq30gp.prd
TD         65536
SOLVENT   CDCl3
NS         385
DS         16
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813440 sec
RG         14596.5
DM         16.500 usec
DE         6.00 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
D16        0.00020000 sec
d17        0.00019600 sec
MCKREST    0 sec
MCWRK      0.01500000 sec
P2         31.00 usec

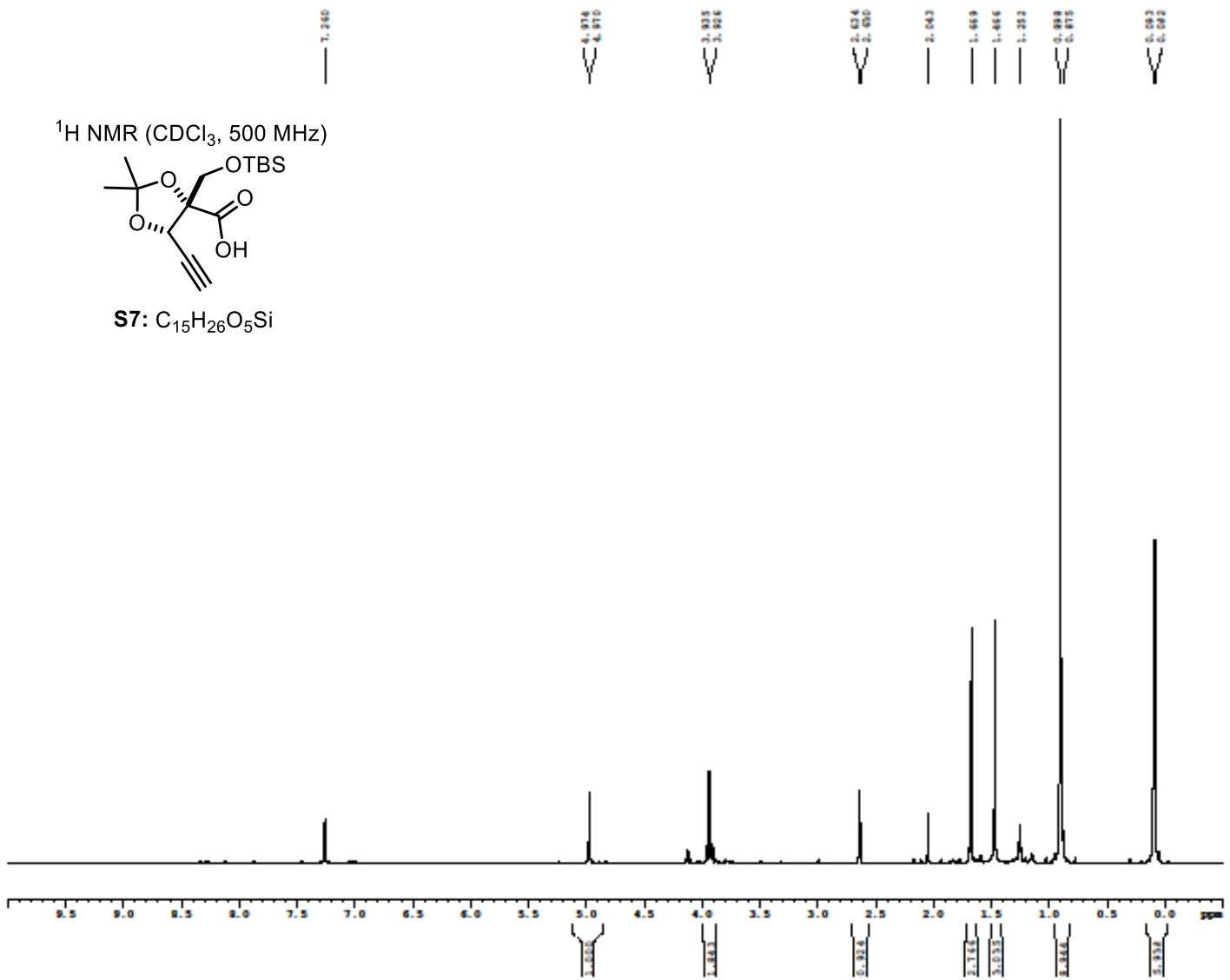
===== CHANNEL f1 =====
NUC1       13C
P1         15.50 usec
P11        500.00 usec
P12        2000.00 usec
PL0        120.00 dB
PL1        -1.00 dB
SFO1       125.7942548 MHz
SP1        3.20 dB
SP2        3.20 dB
SPNAM[1]   Crp60,0.5,20.1
SPNAM[2]   Crp60comp.4
SPOFF1     0 Hz
SPOFF2     0 Hz

===== CHANNEL f2 =====
CPDPRG[2]  waltz16
NUC2       1H
PCPD2      100.00 usec
PL2        1.60 dB
PL12       24.60 dB
SFO2       500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM[1]   SINE.100
GPNAM[2]   SINE.100
GPX1       0 %
GPX2       0 %
GPY1       0 %
GPY2       0 %
GF1        30.00 %
GF2        50.00 %
p15        500.00 usec
p16        1000.00 usec

F2 - Processing parameters
SI         65536
SF         125.7804076 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00
```

**<sup>1</sup>H spectrum**



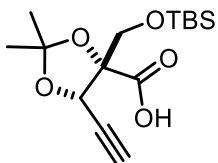
Chemical Data Parameters  
 NAME: 027-117-042  
 EXPNO: 2  
 PROCNO: 1

F2 - Acquisition Parameters  
 Date\_: 20121229  
 Time: 15.30  
 INSTRUM: spect  
 PULPROG: zgpg30  
 PROCNO: 3 on CDCl3 10  
 F2: 500.136260  
 F1: 62.825050  
 SOLVENT: CDCl3  
 NS: 2  
 DS: 2  
 SWH: 2012.822 Hz  
 FIDRES: 0.000413 Hz  
 AQ: 0.096273 sec  
 RG: 313  
 IN: 32.000 umax  
 DE: 4.00 umax  
 TE: 296.2 K  
 D1: 0.1000000 sec  
 ACQNT: 2 sec  
 NS2: 0.0100000 sec

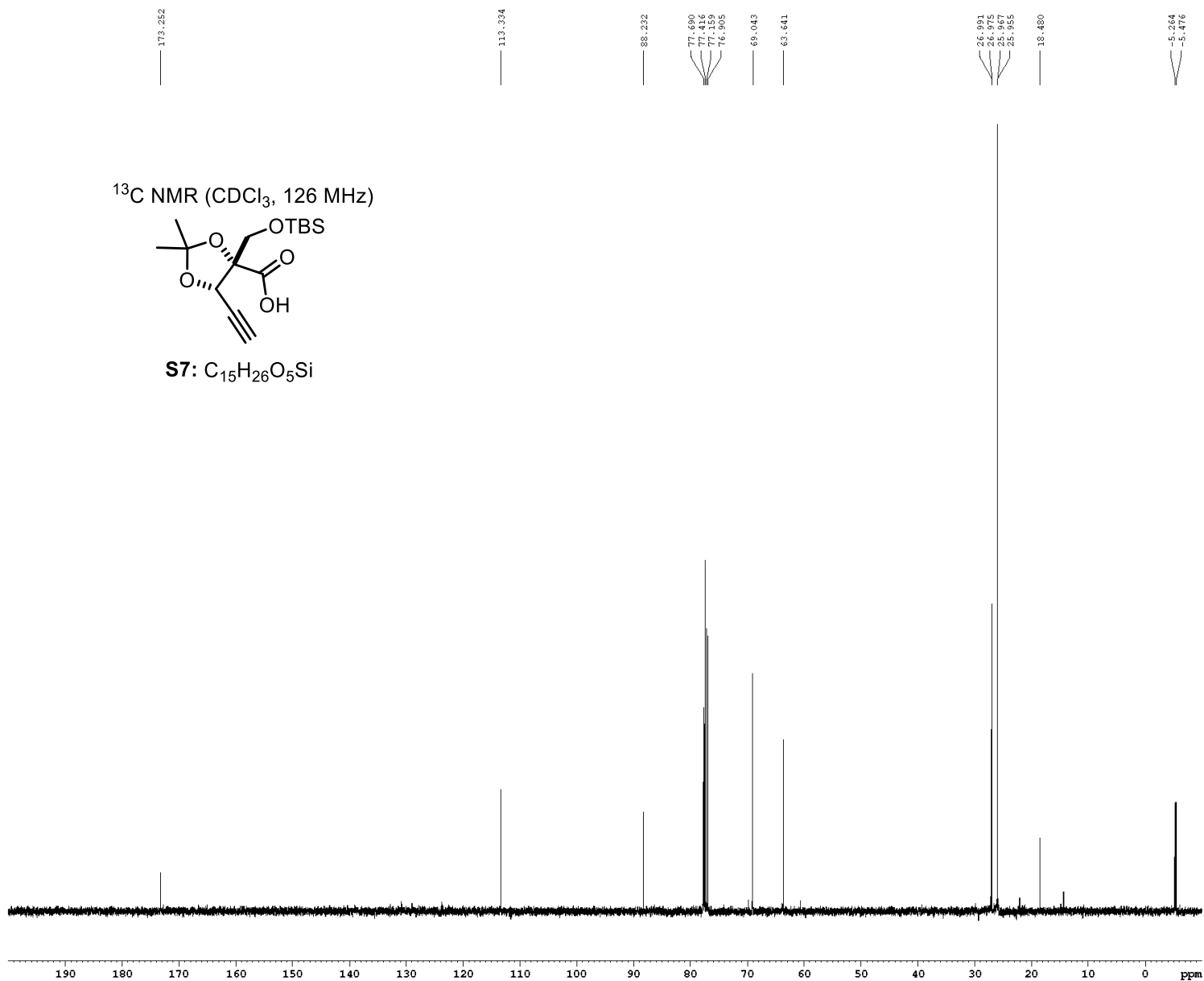
===== CHANNEL f1 =====  
 NUC1: 13  
 P1: 7.00 umax  
 PL1: 0.00 dB  
 SFO1: 100.6281619 MHz

F2 - Processing parameters  
 SI: 65536  
 SF: 500.1362600 MHz  
 DSF: 0  
 SSB: 0 Hz  
 GB: 0 Hz  
 PC: 6.00

<sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz)



S7: C<sub>15</sub>H<sub>26</sub>O<sub>5</sub>Si



```
Current Data Parameters
NAME      DJT-III-043
EXPNO     4
PROCNO    1

F2 - Acquisition Parameters
Date_     20131129
Time      13.55
INSTRUM   cryo500
PROBHD    5 mm CPTCI 1H-
PULPROG   SpinEchopg30gp.prd
TD         65536
SOLVENT   CDCl3
NS         175
DS         16
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813440 sec
RG         13004
DW         16.500 usec
DE         6.00 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
D16        0.00020000 sec
d17        0.00019600 sec
MCREST     0 sec
MCWRK     0.01500000 sec
F2         31.00 usec

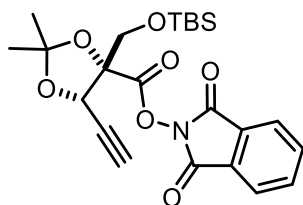
===== CHANNEL f1 =====
NUC1       13C
P1         15.50 usec
P11        500.00 usec
P12        2000.00 usec
PL0        120.00 dB
PL1        -1.00 dB
SFO1       125.7942548 MHz
SP1        3.20 dB
SP2        3.20 dB
SPNAM[1]   Crp60,0.5,20.1
SPNAM[2]   Crp60comp.4
SPOPF1     0 Hz
SPOPF2     0 Hz

===== CHANNEL F2 =====
CPDPRG[2]  waltz16
NUC2       1H
PCPD2      100.00 usec
PL2        1.60 dB
PL12       24.60 dB
SFO2       500.2225011 MHz

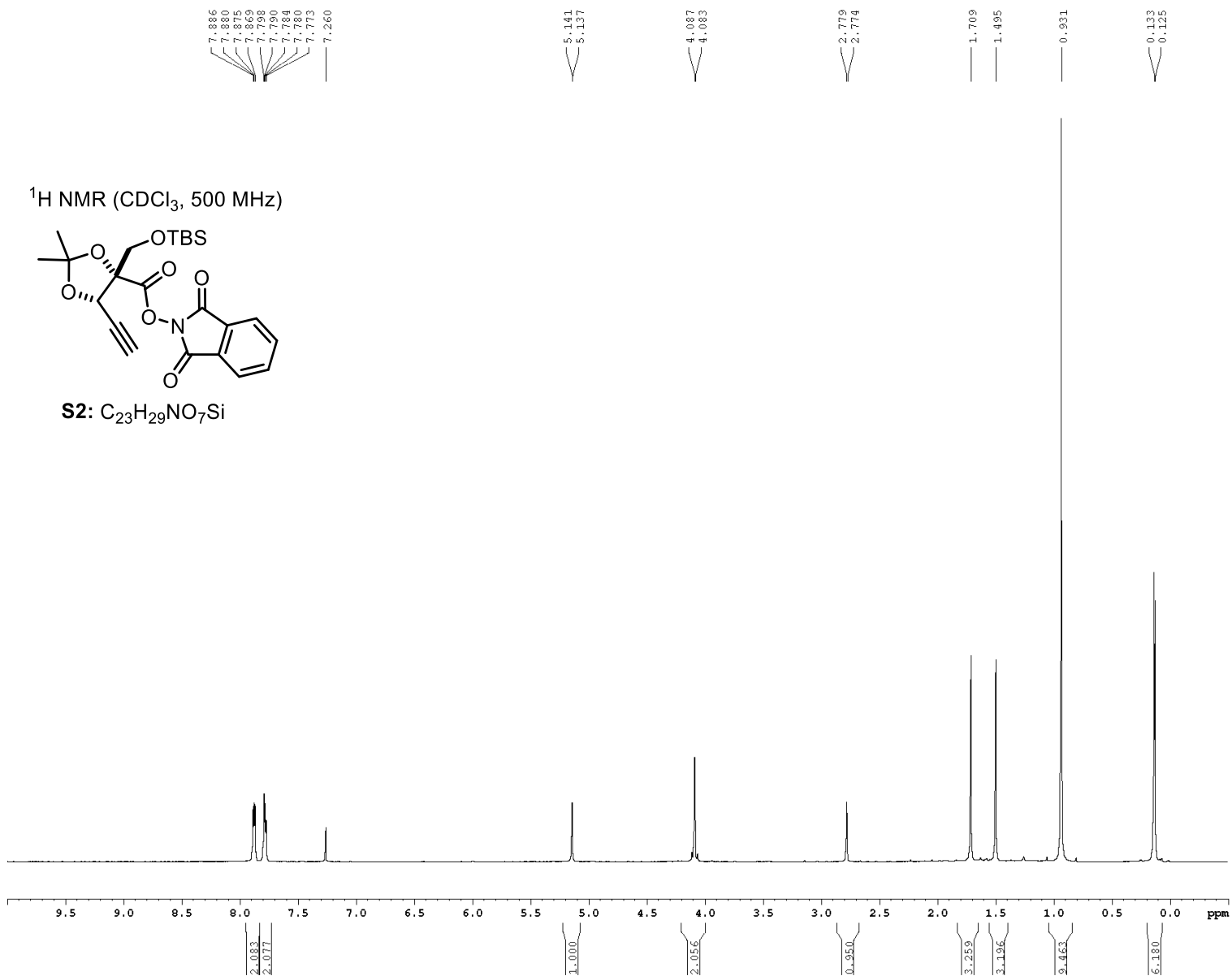
===== GRADIENT CHANNEL =====
GPNAM[1]   SINE.100
GPNAM[2]   SINE.100
GFX1       0 %
GFX2       0 %
GPY1       0 %
GPY2       0 %
GPZ1       30.00 %
GPZ2       50.00 %
p15        500.00 usec
p16        1000.00 usec

F2 - Processing parameters
SI         65536
SF         125.7804085 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00
```

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)



S2: C<sub>23</sub>H<sub>29</sub>NO<sub>7</sub>Si



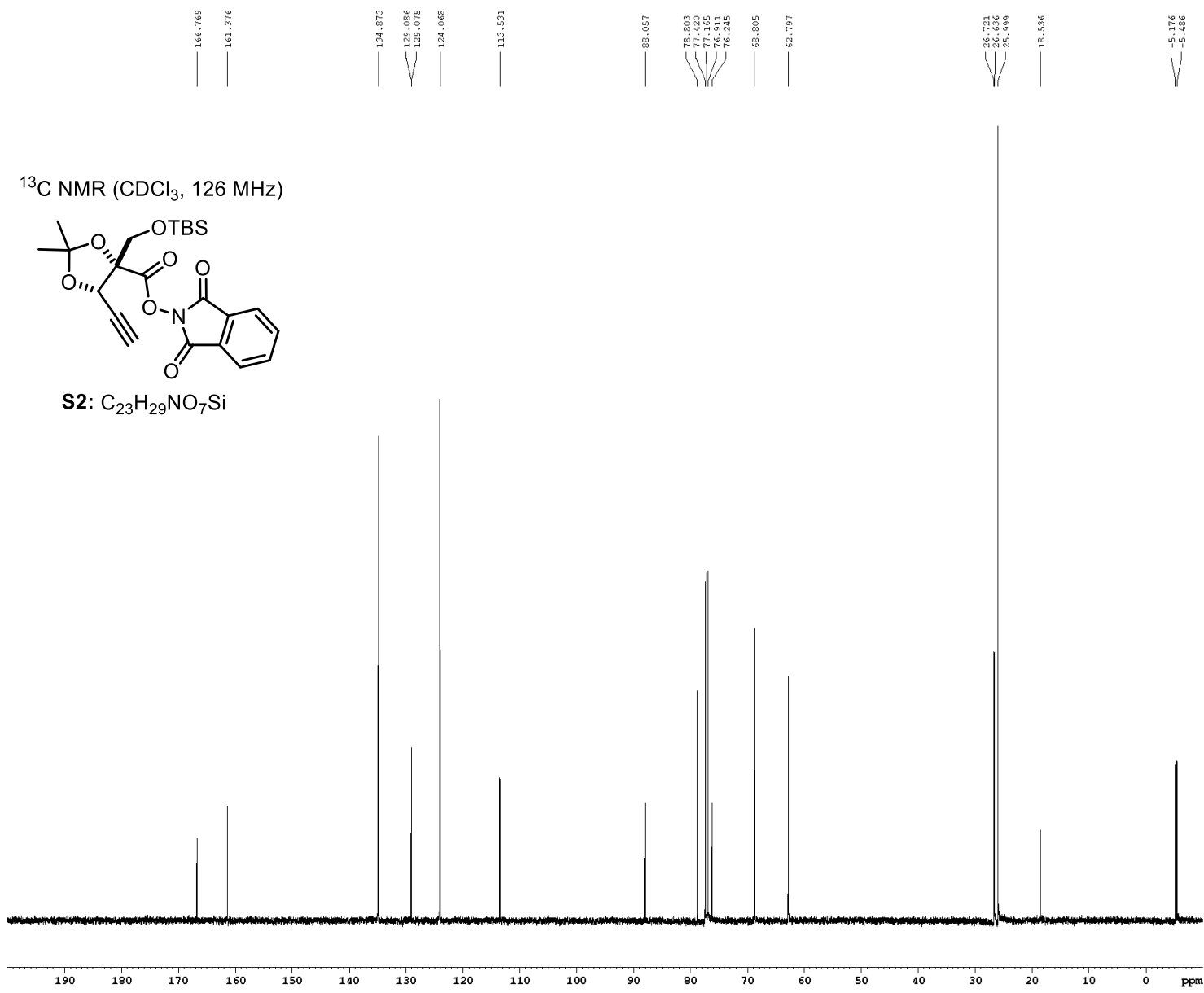
```
Current Data Parameters
NAME      D3T-III-081
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20131206
Time      13:53
INSTRUM   gm500
PROBHD    5 mm broadband
PULPROG   zgpg30
TD         61728
SOLVENT   CDCl3
NS         16
DS         0
SHI        8012.820 Hz
FIDRES    0.098043 Hz
AQ         5.0992773 sec
RG         114
EM         62.400 usec
DE         6.00 usec
TE         298.0 K
DL         0.10000000 sec
MEREST    0 sec
MEMRK     0.01500000 sec

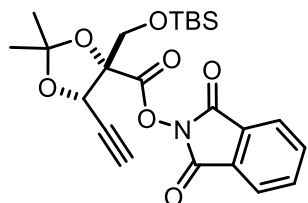
===== CHANNEL f1 =====
NUC1       1H
P1         12.20 usec
PL1        -5.00 dB
SFO1       499.2934950 MHz

F2 - Processing parameters
SI         65536
SF         499.2900294 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
```





<sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz)



S2: C<sub>23</sub>H<sub>29</sub>NO<sub>7</sub>Si

```

Current Data Parameters
NAME      DJT-III-081
EXPNO     3
PROCNO    1

F2 - Acquisition Parameters
Date_     20131207
Time      11.22
INSTRUM   cryo500
PROBHD    5 mm CPTCI 1H-
PULPROG   SpinEchoes30pp.prd
TD         65536
SOLVENT   CDCl3
NS         338
DS         16
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813440 sec
RG         7298.2
DW         16.500 usec
DE         6.00 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
D16        0.00020000 sec
d17        0.00019600 sec
MCKREST   0 sec
MCWRK     0.01500000 sec
P2         31.00 usec

===== CHANNEL f1 =====
NUC1       13C
P1         15.50 usec
P11        500.00 usec
P12        2000.00 usec
PL0        120.00 dB
PL1        -1.00 dB
SFO1       125.7942548 MHz
SP1        3.20 dB
SP2        3.20 dB
SPNAM[1]   Crp60,0.5,20.1
SPNAM[2]   Crp60comp.4
SPOFF1     0 Hz
SPOFF2     0 Hz

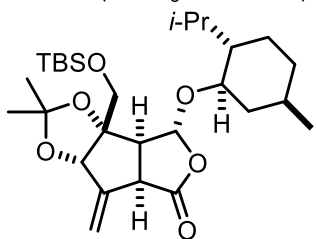
===== CHANNEL f2 =====
CPDPRG[2]  waltz16
NUC2       1H
PCPD2      100.00 usec
PL2        1.60 dB
PL12       24.60 dB
SFO2       500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM[1]   SINE.100
GPNAM[2]   SINE.100
GPX1       0 %
GPX2       0 %
GPY1       0 %
GPY2       0 %
GZ1        30.00 %
GZ2        50.00 %
p15        500.00 usec
p16        1000.00 usec

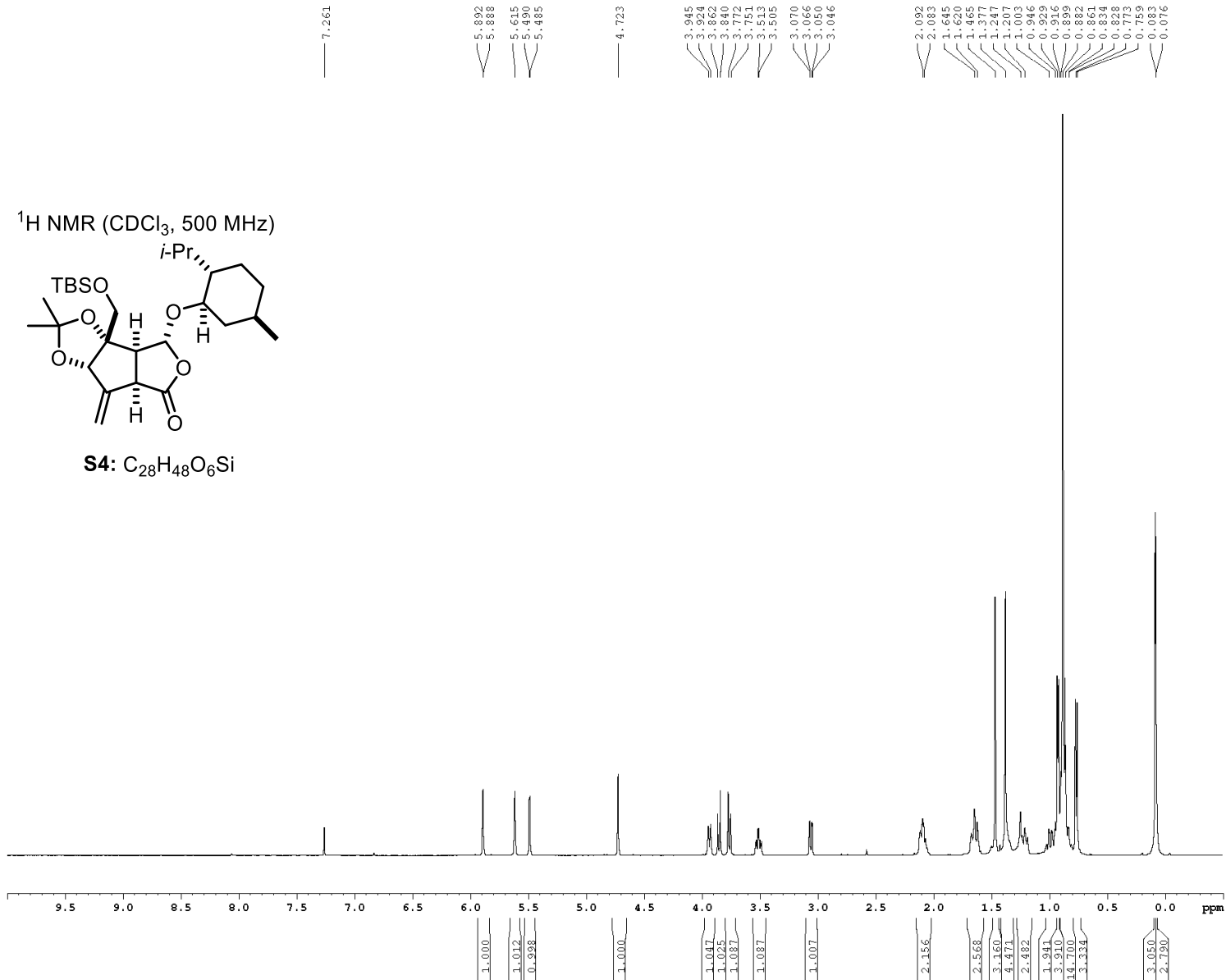
F2 - Processing parameters
SI         65536
SF         125.7804080 MHz
WDW        EM
SSE        0
LB         1.00 Hz
GB         0
PC         2.00

```

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)



**S4:** C<sub>28</sub>H<sub>48</sub>O<sub>6</sub>Si



```

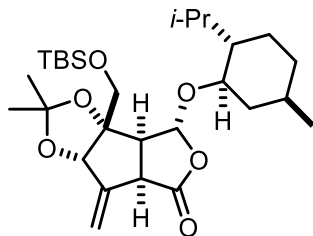
Current Data Parameters
NAME      DJ7-III-088
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20131209
Time     14.43
INSTRUM  cryo500
PROBHD   5 mm CPTCL 1H-
PULPROG  zgpg30
TD        81728
SOLVENT  CDCl3
NS        16
DS        0
SH        8012.820 Hz
FIDRES   0.098043 Hz
AQ        5.0998273 sec
RG         4
DM        62.400 usec
DE         6.00 usec
TE        298.0 K
DQ        0.10000000 sec
DELTA    0.10000000 sec
MEREST   0 sec
MWRFR    0.01500000 sec

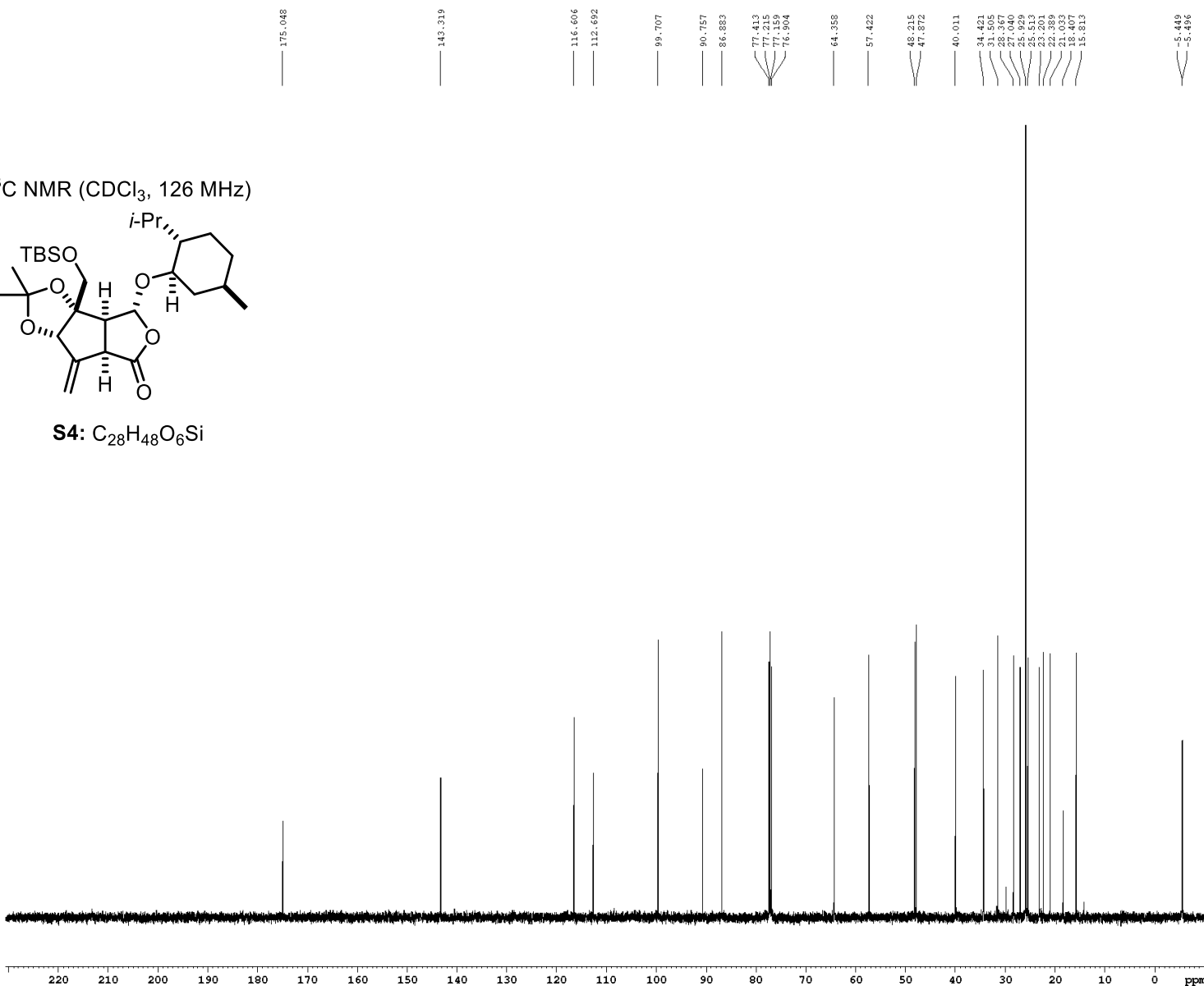
===== CHANNEL f1 =====
NUC1      1H
P1        7.50 usec
PL1       1.60 dB
SFO1     500.225015 MHz

F2 - Processing parameters
SI         65536
SF        500.2200302 MHz
RG         4096
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         4.00
    
```

<sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz)



**S4:** C<sub>28</sub>H<sub>48</sub>O<sub>6</sub>Si



```

Current Data Parameters
NAME      DJT-III-088
EXPNO    2
PROCNO   1

F2 - Acquisition Parameters
Date_    20131209
Time     14.48
INSTRUM  cryo500
PROBHD   5 mm CPTCI 1H-
PULPROG  SpinEchopg30gp.prd
TD        65536
SOLVENT  CDCl3
NS        143
DS         16
SWH       30303.031 Hz
FIDRES    0.462388 Hz
AQ        1.0813440 sec
RG         8192
DW         16.500 usec
DE         6.00 usec
TE        298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
d16        0.00020000 sec
d17        0.00019600 sec
MCREST    0 sec
MCWRK     0.01500000 sec
P2         31.00 usec

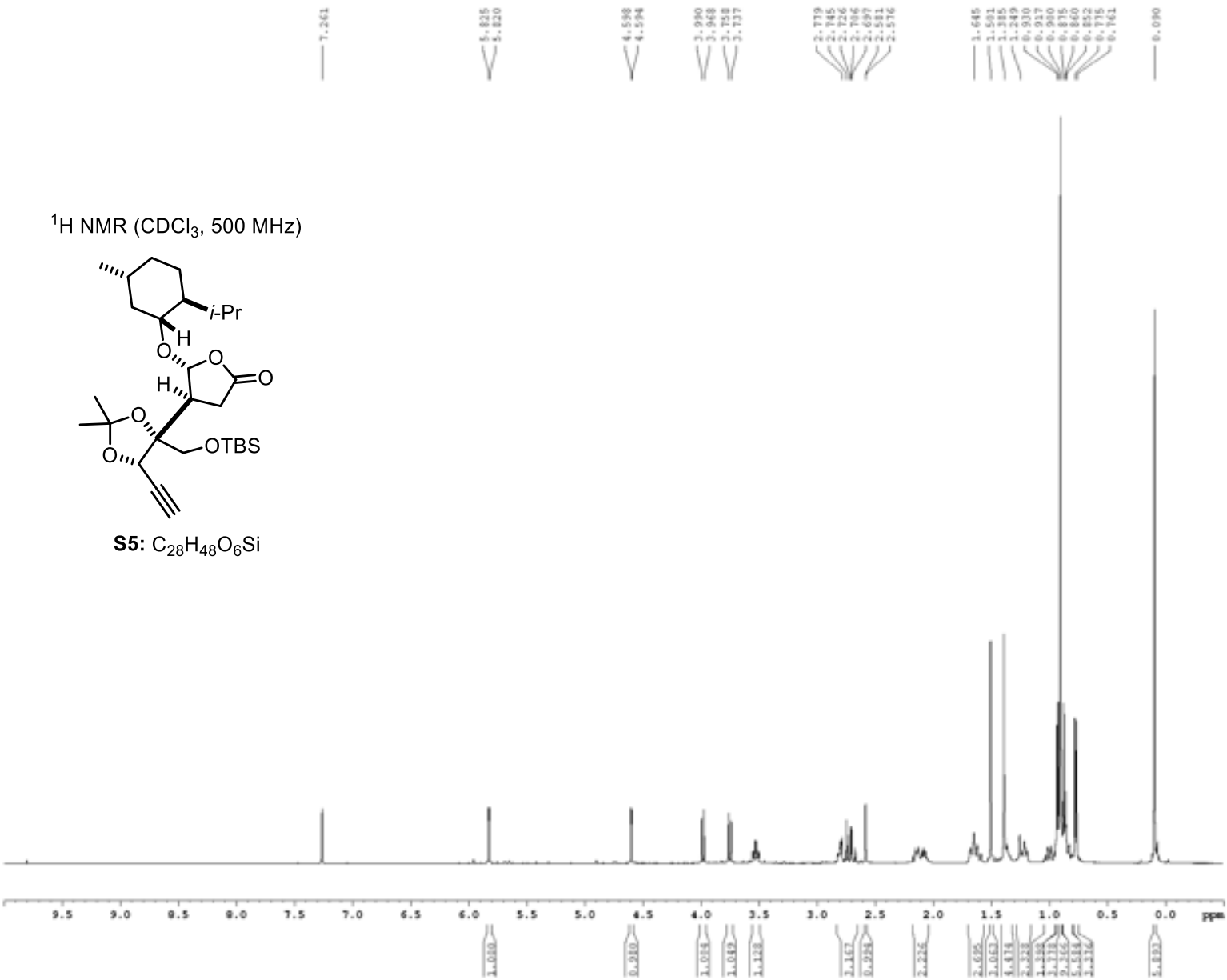
===== CHANNEL f1 =====
NUC1      13C
P1        15.50 usec
P11       500.00 usec
P12       2000.00 usec
PL0       120.00 dB
PL1       -1.00 dB
SFO1     125.7942548 MHz
SP1       3.20 dB
SP2       3.20 dB
SFOFF[1] Crp60,0.5,20.1
SFOFF[2] Crp60comp.4
SPOFF1   0 Hz
SPOFF2   0 Hz

===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2      1H
PCPD2     100.00 usec
PL2       1.60 dB
PL12      24.60 dB
SFO2     500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM[1]  SINE.100
GPNAM[2]  SINE.100
GPX1     0 %
GPX2     0 %
GPY1     0 %
GPY2     0 %
GPT1     30.00 %
GPT2     50.00 %
p15      500.00 usec
p16      1000.00 usec

F2 - Processing parameters
SI        65536
SF        125.7804090 MHz
WDW       EM
SSB       0
LB        1.00 Hz
GB        0
PC        2.00
    
```

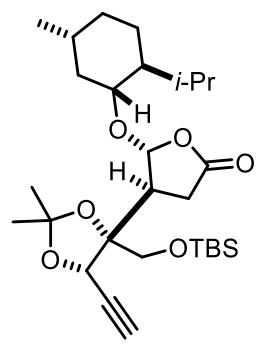
<sup>1</sup>H spectrum



```

Current Data Parameters
NAME      S77-01-009
EXPNO     2
PROCNO    1
F2 - Acquisition Parameters
Date_     20231117
Time      8.14
INSTRUM   cryo500
PROBHD    5 mm QNP1 1H-
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
AQ         14
RG         0
DS         0
SWH         8022.820 MHz
FIDRES     0.0000186 Hz
AQ         3.09942712 sec
NUC1       1
NUC2       13
NUC3       15
PC         42.000 usec
DE         4.00 usec
TE         298.0 K
SI         0.30000000 sec
SFOFFST    0 Hz
SFOFFST2   0.01500000 sec
===== CHANNEL f1 =====
NUC1       1H
PC         7.00 usec
PULPROG   zgpg30
SFOFFST    0 Hz
===== CHANNEL f2 =====
F2 - Processing parameters
IC         43334
SF         500.1300100 MHz
WDW        EM
SSB        0
GB         0.00 MHz
PC         4.00
  
```

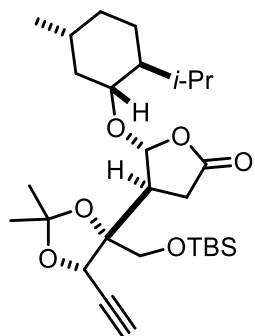
<sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)



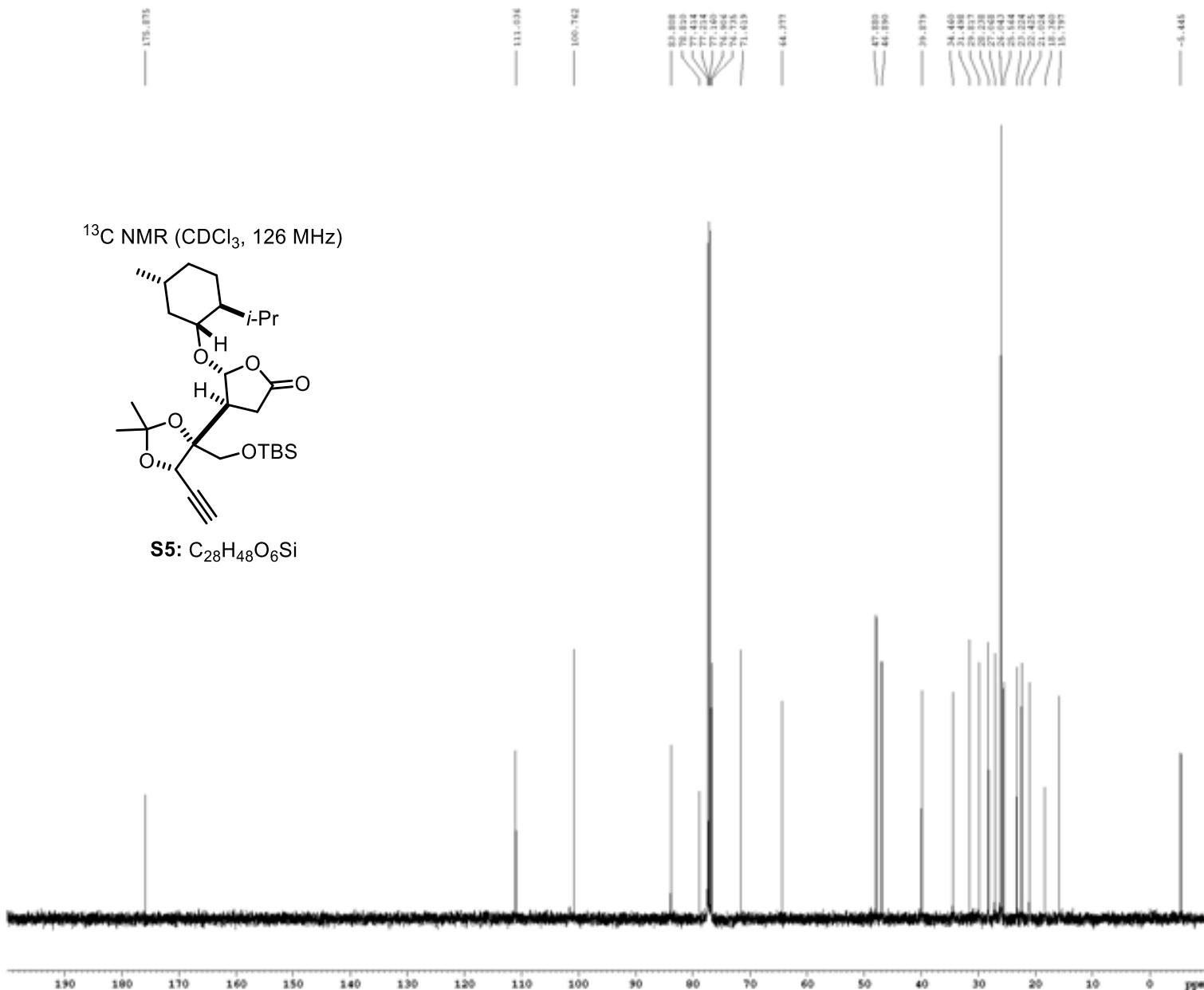
**S5:** C<sub>28</sub>H<sub>48</sub>O<sub>6</sub>Si

Z-restored spin-echo 13C spectrum with 1H decoupling

<sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz)



S5: C<sub>28</sub>H<sub>48</sub>O<sub>6</sub>Si



```

Current Data Parameters
NAME          DJT-VI-099
EXPNO        4
PROCNO       1

F2 - Acquisition Parameters
Date_        20151117
Time         9.29
INSTRUM      cryo500
PROBHD       5 mm CP131 18-
PULPROG      SpinEcho30ap-prd
TD           65536
SOLVENT      CDCl3
NS           393
DS           0
SWH          30303.031 Hz
FIDRES       0.462388 Hz
AQ           1.9813440 sec
RG           7298.2
DM           16.500 usec
DE           4.00 usec
TE           298.0 K
D1           0.25000000 sec
d11          0.03000000 sec
D16          0.00200000 sec
d17          0.00019600 sec
MCKEY        0 sec
MCWK        0.01500000 sec
F2           33.10 usec

----- CHANNEL f1 -----
NUC1         13C
F1           16.55 usec
P1           500.00 usec
P12          2000.00 usec
PL0          120.00 dB
PL1          -1.00 dB
SFO1         125.7942548 MHz
SF1          2.70 dB
SF2          2.70 dB
SFOFF[1]     Cyp60, 0.5, 20.1
SFOFF[2]     Cyp60comp, 4
SFOFF1       0 Hz
SFOFF2       0 Hz

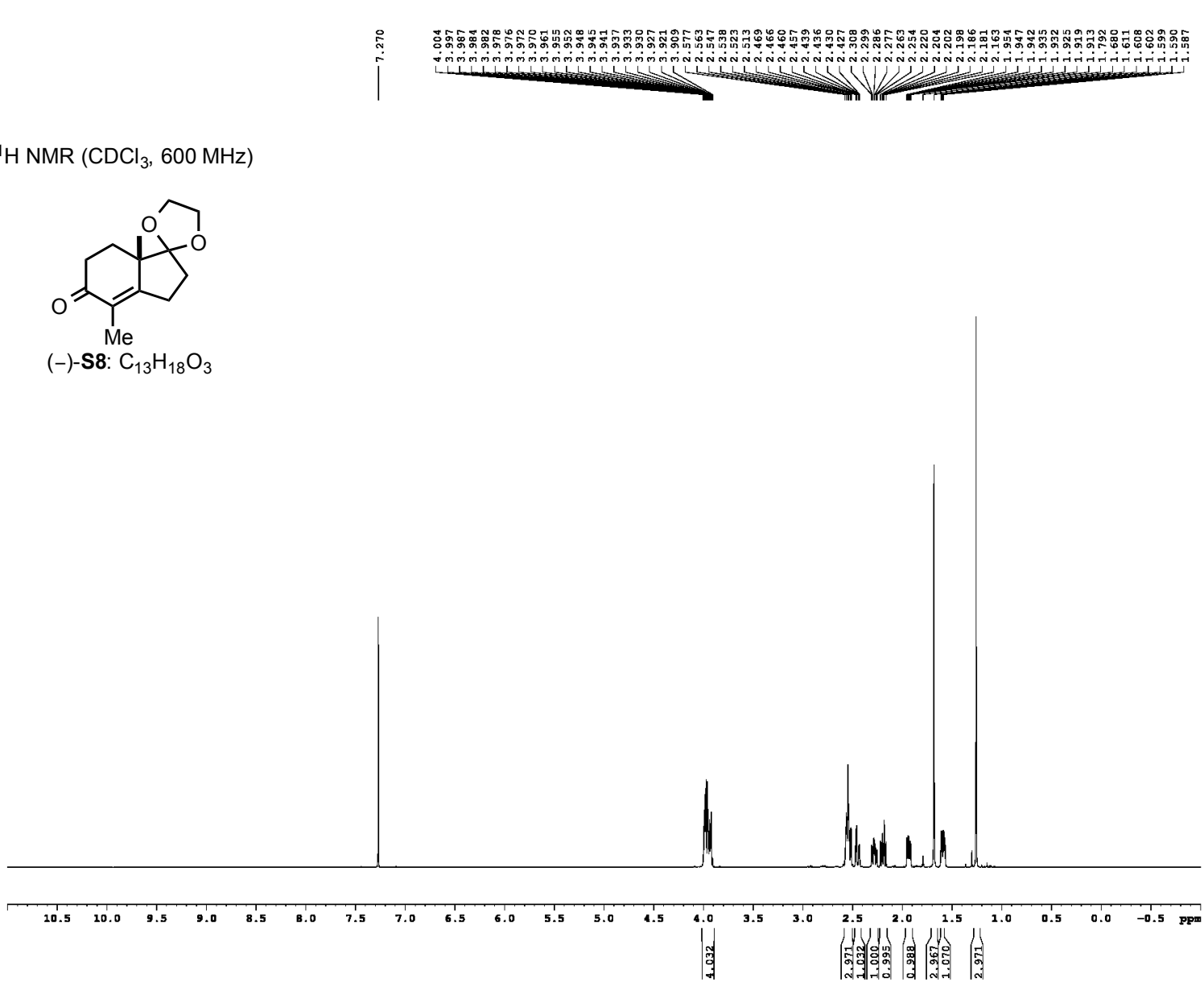
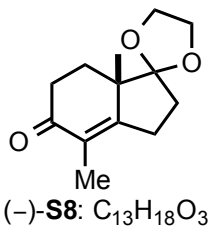
----- CHANNEL f2 -----
CPDPRG[2]   waltz16
NUC2         1H
PCPD2        100.00 usec
PL2          1.40 dB
PL12         24.50 dB
SFO2         500.2225011 MHz

----- GRADIENT CHANNEL -----
GPNAM[1]     SINE.100
GPNAM[2]     SINE.100
GPK1         0 %
GPK2         0 %
GPY1         0 %
GPY2         0 %
GPE1         30.00 %
GPE2         50.00 %
p15          500.00 usec
p16          1000.00 usec

F2 - Processing parameters
SI           65536
SF           125.7804085 MHz
MSB          RM
LSB          0
GB           1.00 Hz
FC           2.00
    
```

YS-IV-18

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz)



```

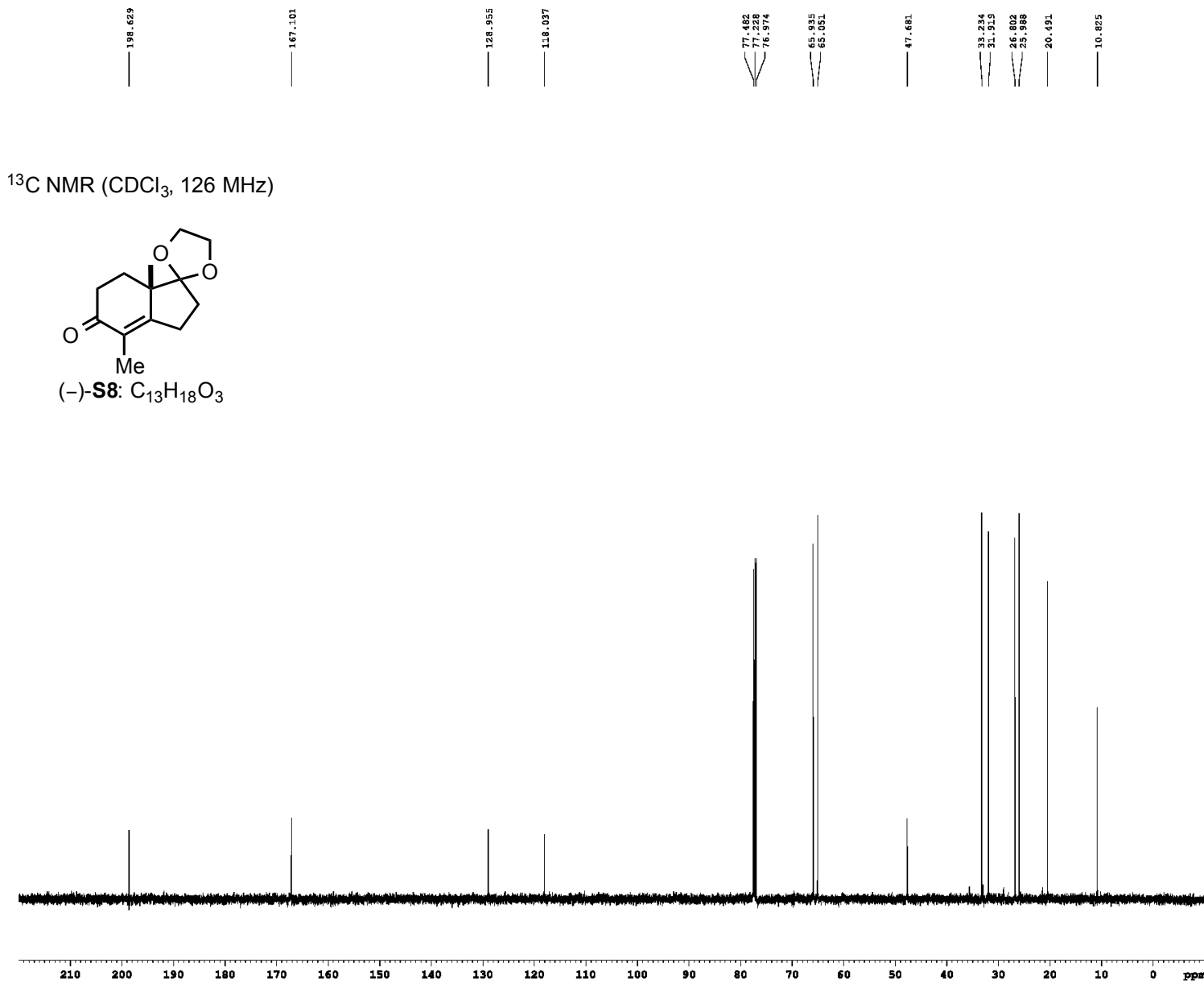
Current Data Parameters
NAME      YS-IV-18
EXPNO    4
PROCNO   1

F2 - Acquisition Parameters
Date_    20111209
Time     8.24
INSTRUM  av600
PROBHD   5 mm TBI 1H/13
PULPROG  zg30
TD        98074
SOLVENT  CDCl3
NS        8
DS        2
SWH       9615.385 Hz
FIDRES    0.098062 Hz
AQ        5.0598878 sec
RG        812
EW        52.000 use
DS        14.54 use
TS        298.0 K
D1        0.10000000 sec
TDO       1

===== CHANNEL F1 =====
SFO1     600.1342009 MHz
NUC1     13
P1       8.00 use
PLW1     24.00000000 W

F2 - Processing parameters
SI        65336
SF        600.1300294 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00
    
```

YS-IV-18



```
Current Data Parameters
NAME      YS-IV-18
EXPNO    5
PROCNO    1

F2 - Acquisition Parameters
Date_     20151216
Time      15.42
INSTRUM   cryo500
PROBHD    5 mm CPIC1 1H-
PULPROG   SpinEchopg30pp.prd
TD         65536
SOLVENT   CDCl3
NS         152
DS         2
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813440 sec
RG         2896.3
DW         16.500 usec
DE         6.00 usec
TE         298.0 K
D1         0.2500000 sec
d11        0.03000000 sec
D16        0.00020000 sec
d17        0.00019600 sec
MCREST     0 sec
MCWRK     0.01500000 sec
P2         33.10 usec

===== CHANNEL f1 =====
NUC1       13C
P1         16.55 usec
P11        500.00 usec
P12        2000.00 usec
PL0        120.00 dB
PL1        -1.00 dB
SFO1       125.7942548 MHz
SP1        2.70 dB
SP2        2.70 dB
SPNAM[1]   Crp60,0.5,20.1
SPNAM[2]   Crp60comp.4
SPOFF1     0 Hz
SPOFF2     0 Hz

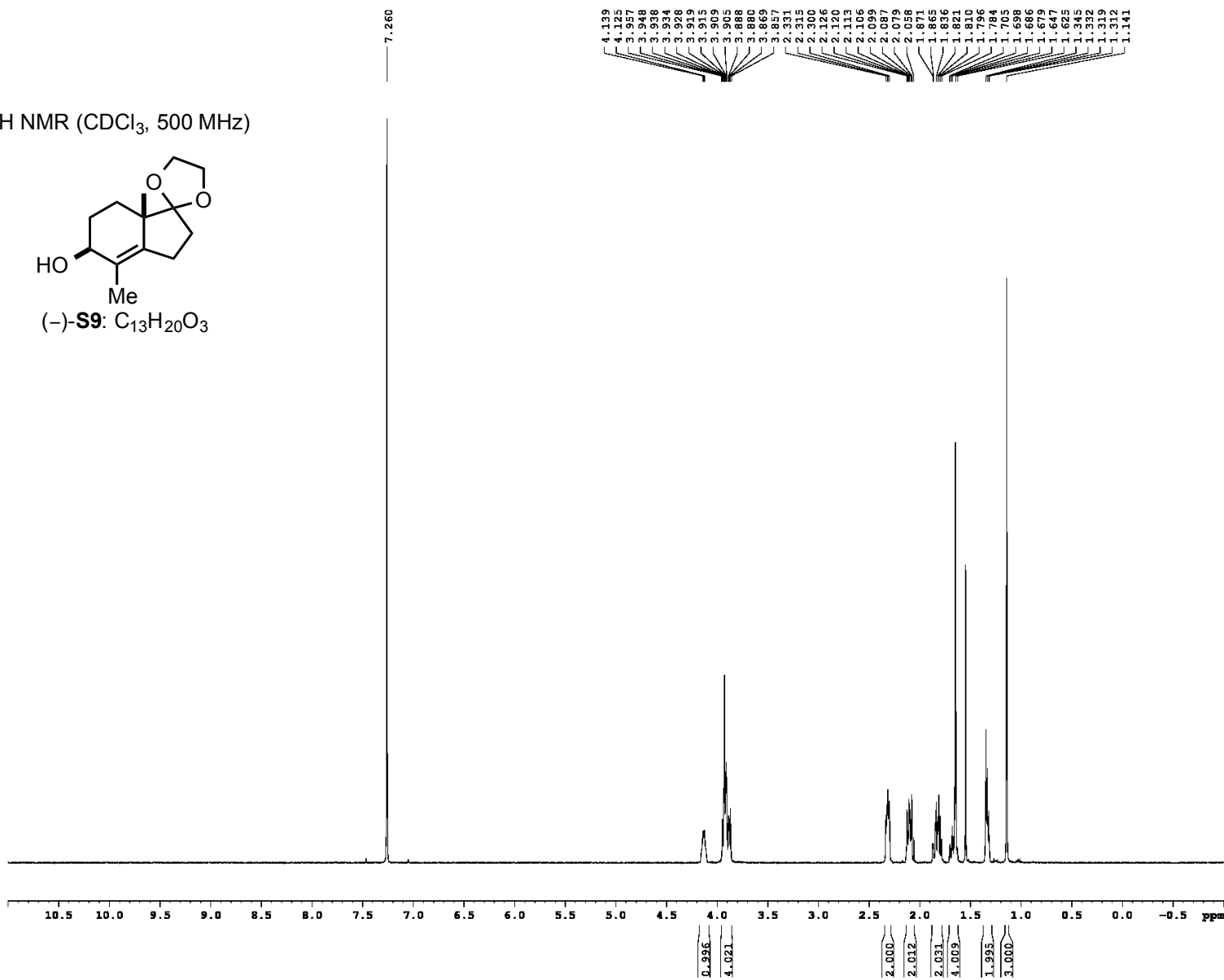
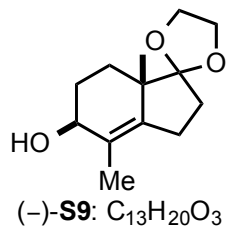
===== CHANNEL f2 =====
CPDPRG[2]  waltz16
NUC2       1H
PCPD2     100.00 usec
PL2       1.60 dB
PL12      24.50 dB
SFO2      500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM[1]   SINE.100
GPNAM[2]   SINE.100
GPX1       0 %
GPX2       0 %
GPY1       0 %
GPY2       0 %
GPZ1       30.00 %
GPZ2       50.00 %
p15        500.00 usec
p16        1000.00 usec

F2 - Processing parameters
SI         65536
SF         125.7804008 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00
```

YS-III-292

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)

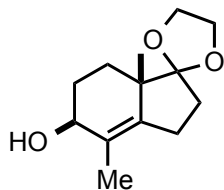


```
Current Data Parameters
NAME      YS-III-292
EXPNO    3
PROCNO   1
F2 - Acquisition Parameters
Date_    20151117
Time     8.52
INSTRUM  cryo300
PROBHD   5 mm CPYCI 1H-
PULPROG  zgpg
TD       32048
SOLVENT  CDCl3
NS       4
DS       2
SWH      8012.820 Hz
FIDRES   0.328026 Hz
AQ       1.9397952 sec
RG       7.1
DW       62.400 use
DS       6.00 use
TS       298.0 K
D1       0.1000000 sec
MCREST   0 sec
MCMXK    0.0150000 sec
----- CHANNEL f1 -----
NUC1     1H
P1       7.50 use
PI1      1.60 dB
SFO1     500.225015 MHz
F2 - Processing parameters
SI       65536
SF       500.2200324 MHz
WDW      EM
SBB      0
GB       0 0.30 Hz
PC       4.00
```

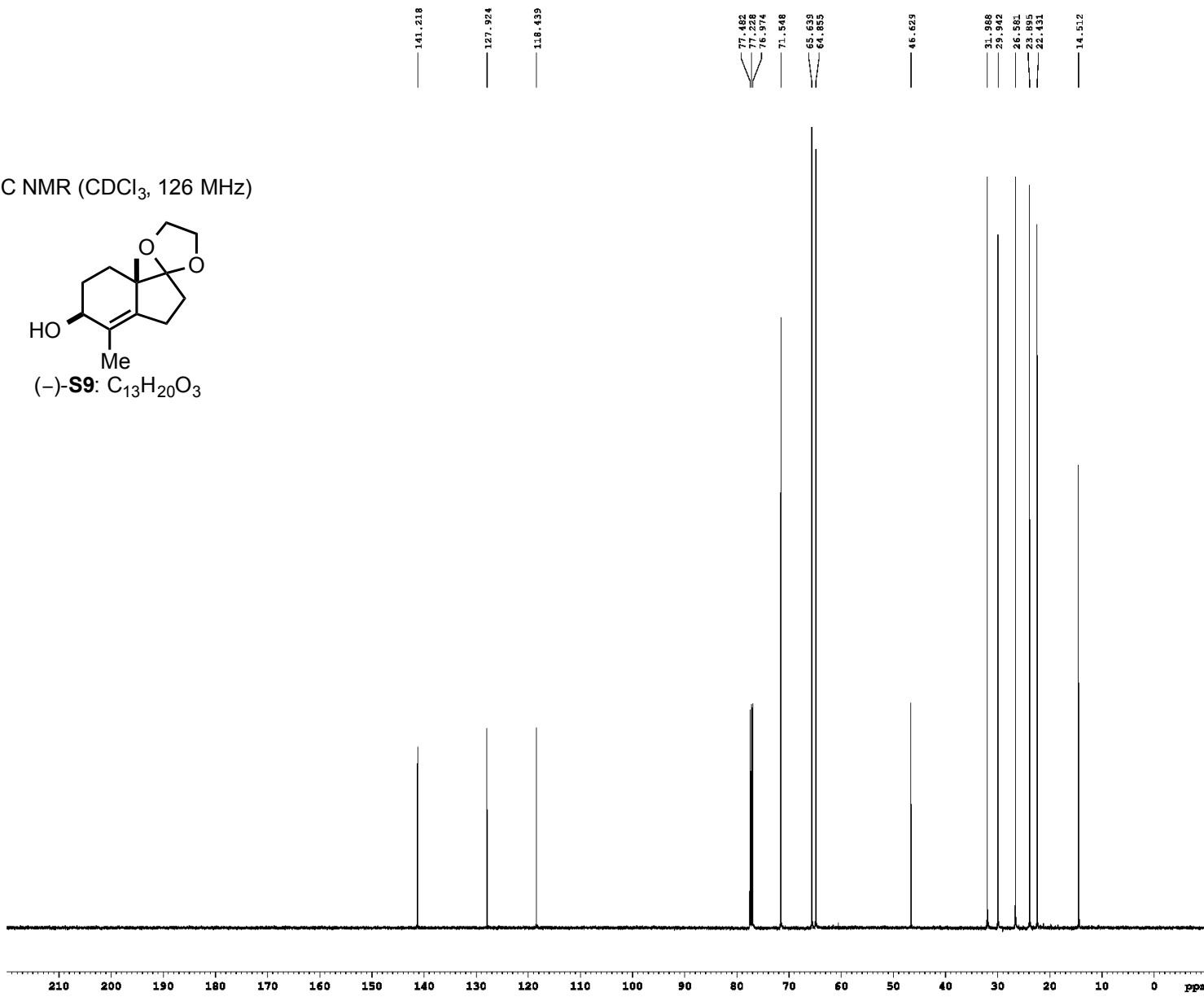


YS-IV-20

<sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz)



(-)-S9: C<sub>13</sub>H<sub>20</sub>O<sub>3</sub>



Current Data Parameters  
NAME YS-IV-20  
EXPNO 4  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20151209  
Time 17.41  
INSTRUM cryo500  
PROBED 5 mm CPIC1 1H-  
PULPROG SpinEchopg30pp.prd  
TD 65536  
SOLVENT CDCl3  
NS 640  
DS 2  
SWH 30303.031 Hz  
FIDRES 0.462388 Hz  
AQ 1.0813440 sec  
RG 7298.2  
DW 16.500 usec  
DE 6.00 usec  
TE 298.0 K  
D1 0.25000000 sec  
d11 0.03000000 sec  
D16 0.00020000 sec  
d17 0.00019600 sec  
MCREST 0 sec  
MCWRK 0.01500000 sec  
P2 33.10 usec

===== CHANNEL f1 =====  
NUC1 13C  
P1 16.55 usec  
P11 500.00 usec  
P12 2000.00 usec  
PLO 120.00 dB  
PL1 -1.00 dB  
SFO1 125.7942548 MHz  
SP1 2.70 dB  
SP2 2.70 dB  
SPNAM[1] Crp60,0.5,20.1  
SPNAM[2] Crp60comp.4  
SPOFF1 0 Hz  
SPOFF2 0 Hz

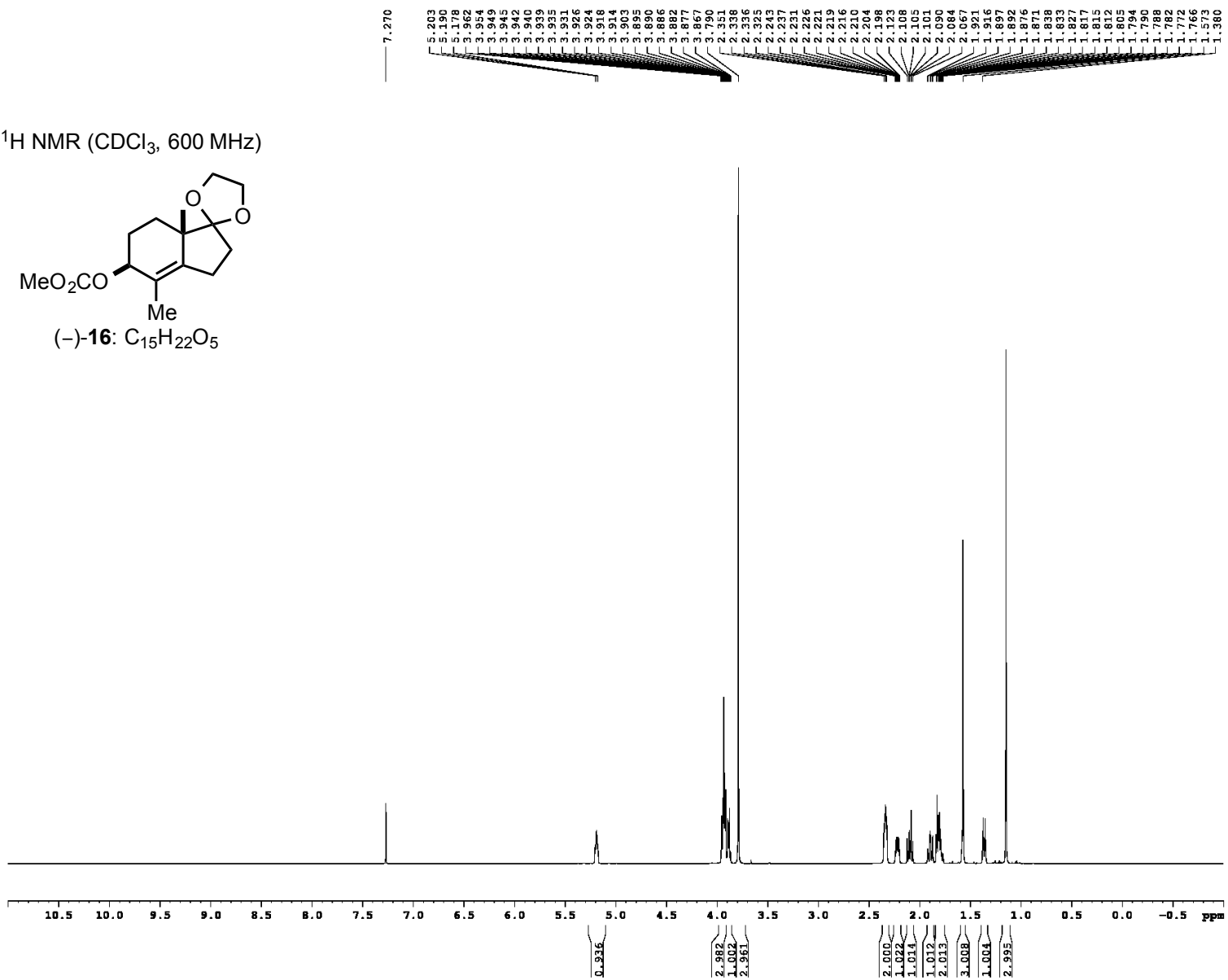
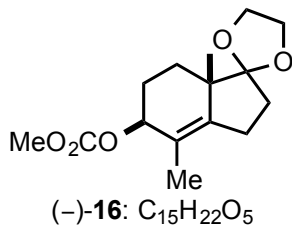
===== CHANNEL f2 =====  
CPDPRG[2] waitz16  
NUC2 1H  
PCPD2 100.00 usec  
PL2 1.60 dB  
PL12 24.50 dB  
SFO2 500.2225011 MHz

===== GRADIENT CHANNEL =====  
GPNAM[1] SINE.100  
GPNAM[2] SINE.100  
GPX1 0 %  
GPX2 0 %  
GPY1 0 %  
GPY2 0 %  
GPZ1 30.00 %  
GPZ2 50.00 %  
p15 500.00 usec  
p16 1000.00 usec

F2 - Processing parameters  
SI 65536  
SF 125.7804059 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 2.00

YS-III-293

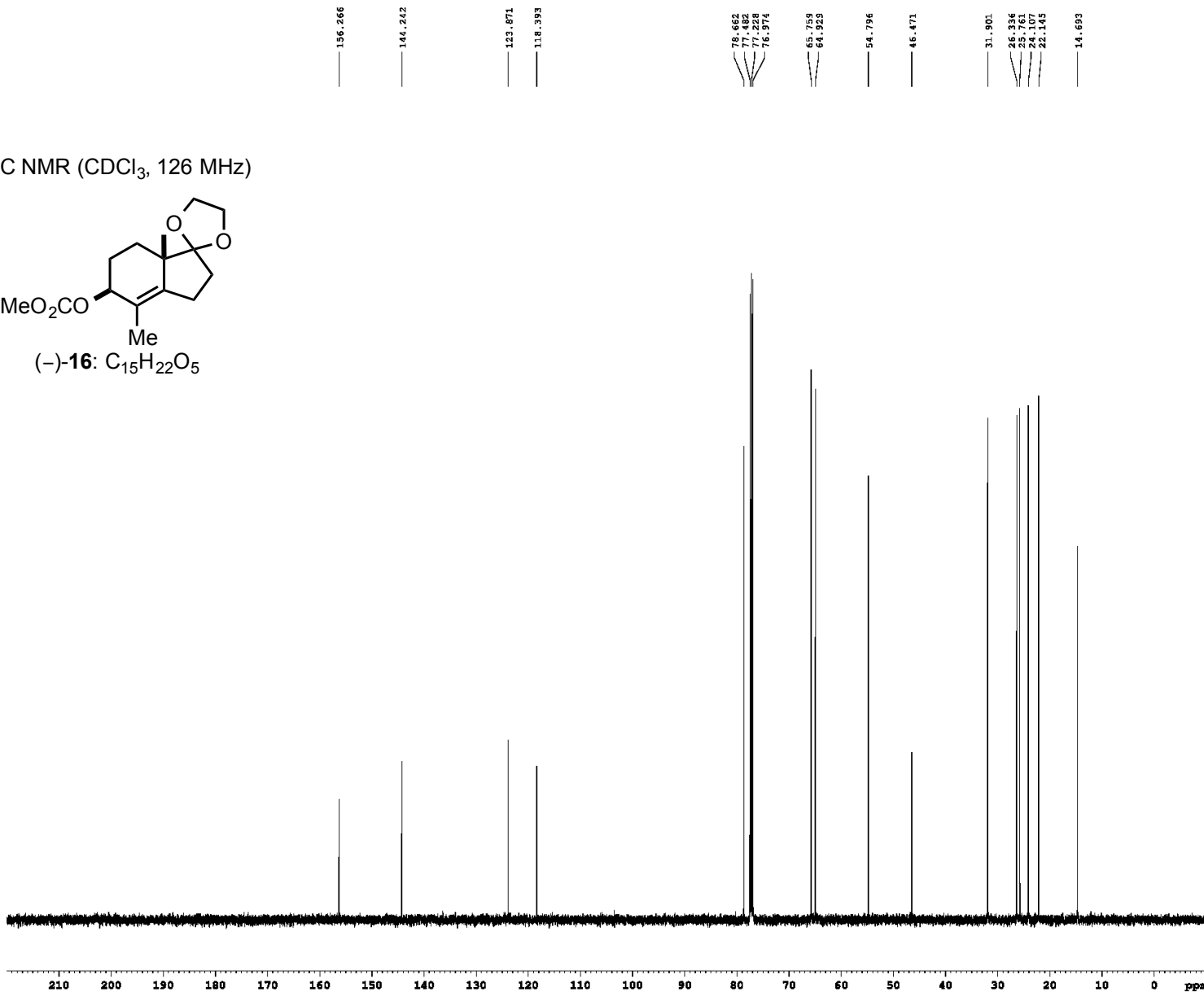
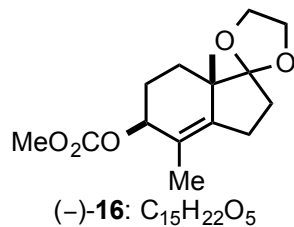
<sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz)



```
Current Data Parameters
NAME      YS-III-293
EXPNO    2
PROCNO   1
F2 - Acquisition Parameters
Date_    20151117
Time     17.10
INSTRUM  av600
PROBHD   5 mm TBI 1H/13
PULPROG  zgpg30
TD       38460
SOLVENT  CDCl3
NS       4
DS       2
SWH      9612.382 Hz
FIDRES   0.250010 Hz
AQ       1.9999200 sec
RG       161
DW       52.000 use
DS       14.54 use
TS       298.0 K
D1       0.1000000 sec
TD0      1
===== CHANNEL f1 =====
SFO1    600.1342039 MHz
NUC1     13
P1       8.00 use
PLW1    24.0000000 W
F2 - Processing parameters
SI       6536
SP       600.1300296 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00
```

YS-III-293

<sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz)



```

Current Data Parameters
NAME      YS-III-293
EXPNO     3
PROCNO    1

F2 - Acquisition Parameters
Date_     20151117
Time      17.20
INSTRUM   cryo500
PROBED    5 mm CPIC1 1H-
PULPROG   SpinEchopg30gp.prd
TD         65536
SOLVENT   CDCl3
NS         424
DS         16
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813440 sec
RG         5792.6
DW         16.500 usec
DE         6.00 usec
TE         298.0 K
d1         0.25000000 sec
d11        0.03000000 sec
d16        0.00020000 sec
d17        0.00019600 sec
MCREST    0 sec
MCWRK     0.01500000 sec
P2         33.10 usec

===== CHANNEL f1 =====
NUC1       13C
P1         16.55 usec
P11        500.00 usec
P12        2000.00 usec
PLO        120.00 dB
PL1        -1.00 dB
SFO1       125.7942548 MHz
SP1        2.70 dB
SP2        2.70 dB
SPNAM[1]   Crp60,0.5,20.1
SPNAM[2]   Crp60comp.4
SPOFF1     0 Hz
SPOFF2     0 Hz

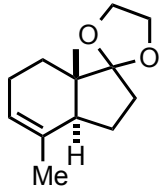
===== CHANNEL f2 =====
CPDPRG[2]  waitz16
NUC2       1H
PCPD2     100.00 usec
PL2        1.60 dB
PL12       24.50 dB
SFO2       500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM[1]   SINE.100
GPNAM[2]   SINE.100
GPX1       0 %
GPX2       0 %
GPY1       0 %
GPY2       0 %
GPZ1       30.00 %
GPZ2       50.00 %
p15        500.00 usec
p16        1000.00 usec

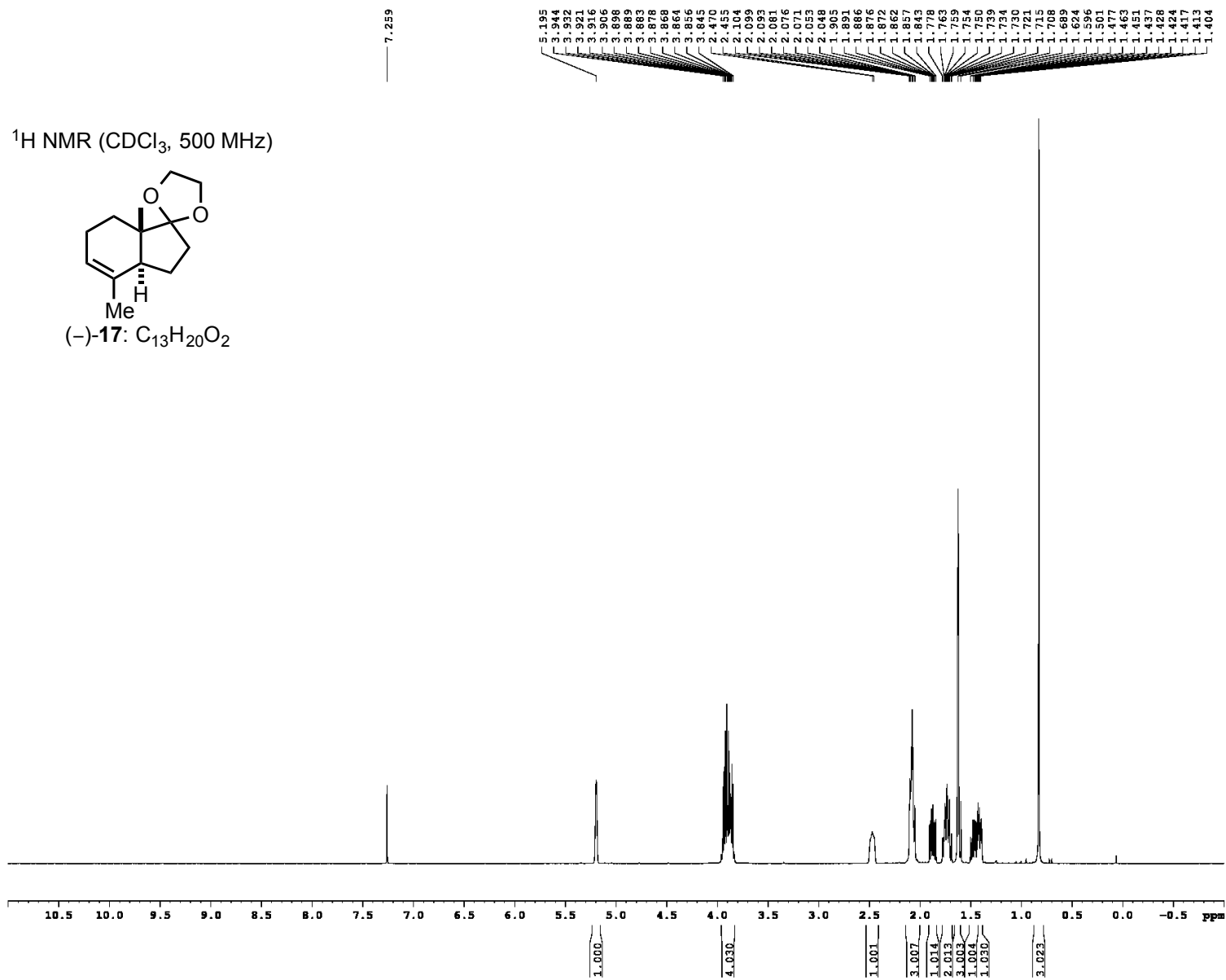
F2 - Processing parameters
SI         65536
SF         125.7804002 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00
    
```

YS-III-296

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)

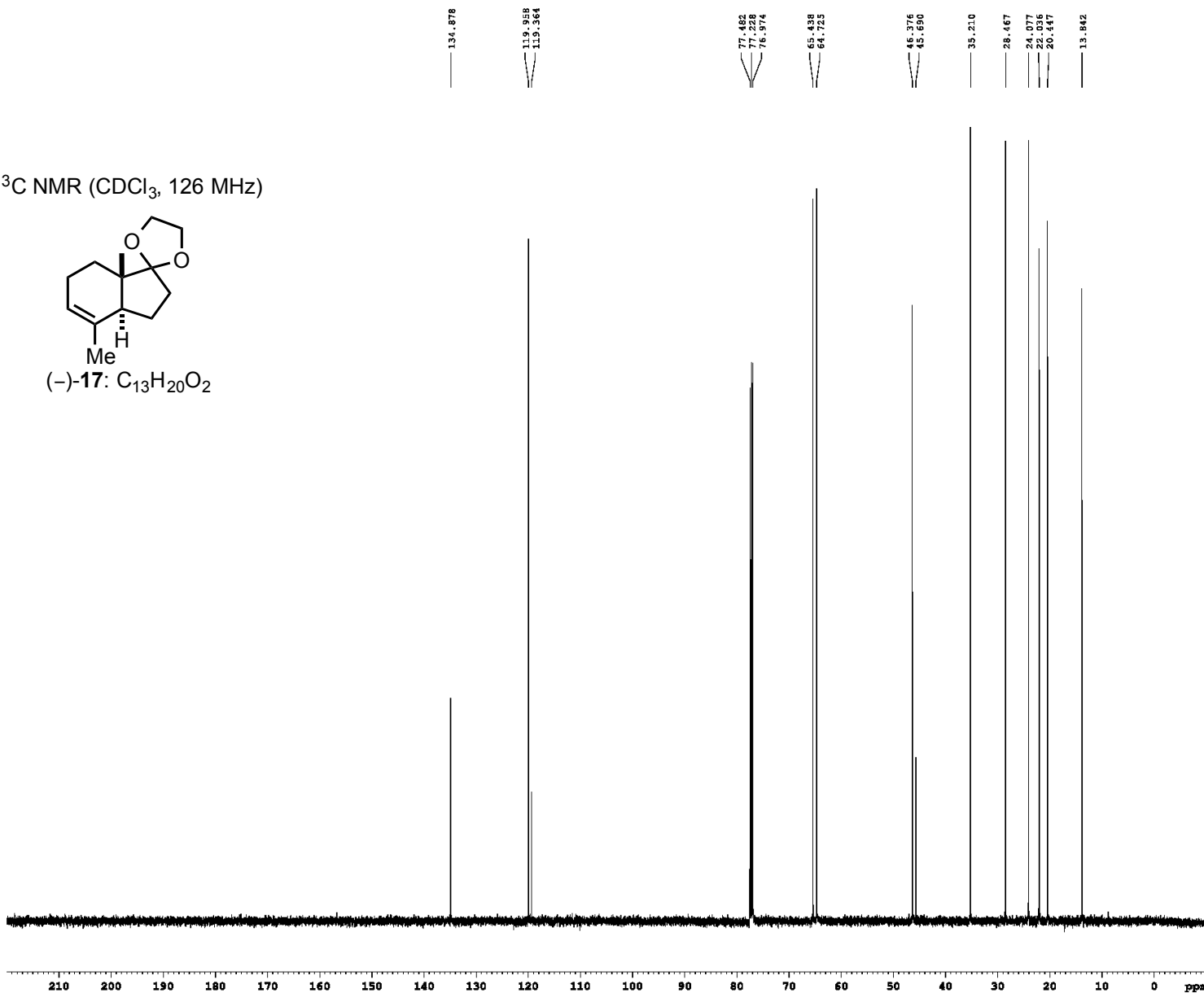
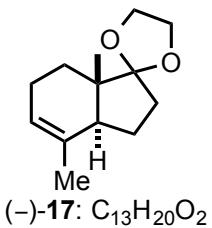


(-)-17: C<sub>13</sub>H<sub>20</sub>O<sub>2</sub>



Current Data Parameters  
NAME YS-III-296  
EXPNO 4  
PROCNO 1  
F2 - Acquisition Parameters  
Date\_ 20151119  
Time 16.51  
INSTRUM cryo300  
PROBHD 5 mm CPYCI 1H-  
PULPROG zgpg  
TD 128232  
SOLVENT CDCl3  
NS 4  
DS 2  
SWH 8012.820 Hz  
FIDRES 0.062302 Hz  
AQ 7.9988050 sec  
RG 6.3  
DW 62.400 use  
DE 6.00 use  
TE 298.0 K  
D1 0.1000000 sec  
MCRET 0 sec  
MCRM 0.0150000 sec  
CHANNEL f1  
NUC1 1H  
P1 7.50 use  
PI1 1.60 dB  
SFO1 500.225015 MHz  
F2 - Processing parameters  
SI 65536  
SF 500.2200323 MHz  
WDW EM  
SBB 0  
LB 0.30 Hz  
GB 0  
PC 4.00

<sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz)



```

Current Data Parameters
NAME      YS-III-296
EXPNO     5
PROCNO    1

F2 - Acquisition Parameters
Data_     20151119
Time      16.57
INSTRUM   cryo500
PROBED    5 mm CPIC1 1H-
PULPROG   SpinEchopg30gp.prd
TD         65536
SOLVENT   CDCl3
NS         504
DS         16
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813440 sec
RG         6502
DW         16.500 usec
DE         6.00 usec
TE         298.0 K
d1         0.25000000 sec
d11        0.03000000 sec
d16        0.00020000 sec
d17        0.00019600 sec
MCREST     0 sec
MCWRK     0.01500000 sec
P2         33.10 usec

===== CHANNEL f1 =====
NUC1       13C
P1         16.55 usec
P11        500.00 usec
P12        2000.00 usec
PLO        120.00 dB
PL1        -1.00 dB
SFO1       125.7942548 MHz
SP1        2.70 dB
SP2        2.70 dB
SPNAM[1]   Crp60,0,5,20,1
SPNAM[2]   Crp60comp.4
SPOFF1     0 Hz
SPOFF2     0 Hz

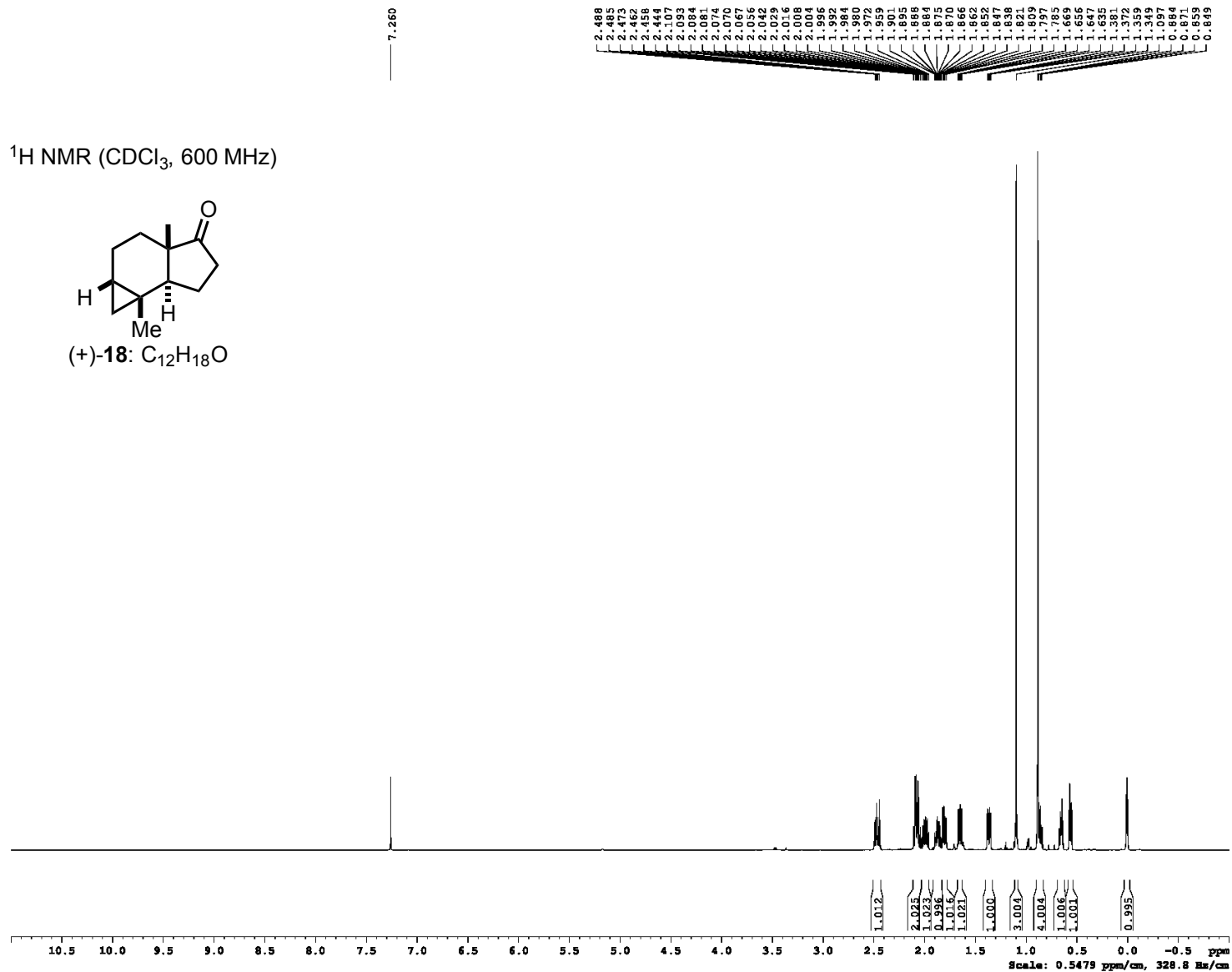
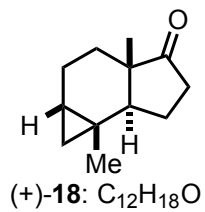
===== CHANNEL f2 =====
CPDPRG[2]  waltz16
NUC2       1H
PCPD2      100.00 usec
PL2        1.60 dB
PL12       24.50 dB
SFO2       500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM[1]   SINE.100
GPNAM[2]   SINE.100
GPX1       0 %
GPX2       0 %
GPY1       0 %
GPY2       0 %
GPZ1       30.00 %
GPZ2       50.00 %
p15        500.00 usec
p16        1000.00 usec

F2 - Processing parameters
SI         65536
SF         125.7803998 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00
    
```

YS-IV-29

$^1\text{H}$  NMR ( $\text{CDCl}_3$ , 600 MHz)



Current Data Parameters  
NAME YS-IV-29  
EXPNO 3  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20160107  
Time 15.39  
INSTRUM av600  
PROBHD 5 mm TBI 1H/13  
PULPROG zg30  
TD 98074  
SOLVENT CDCl3  
NS 8  
DS 2  
SWH 9615.385 Hz  
FIDRES 0.098042 Hz  
AQ 5.0998478 sec  
RG 362  
DW 52.000 usec  
DE 14.54 usec  
TE 298.1 K  
D1 0.10000000 sec  
TD0 1

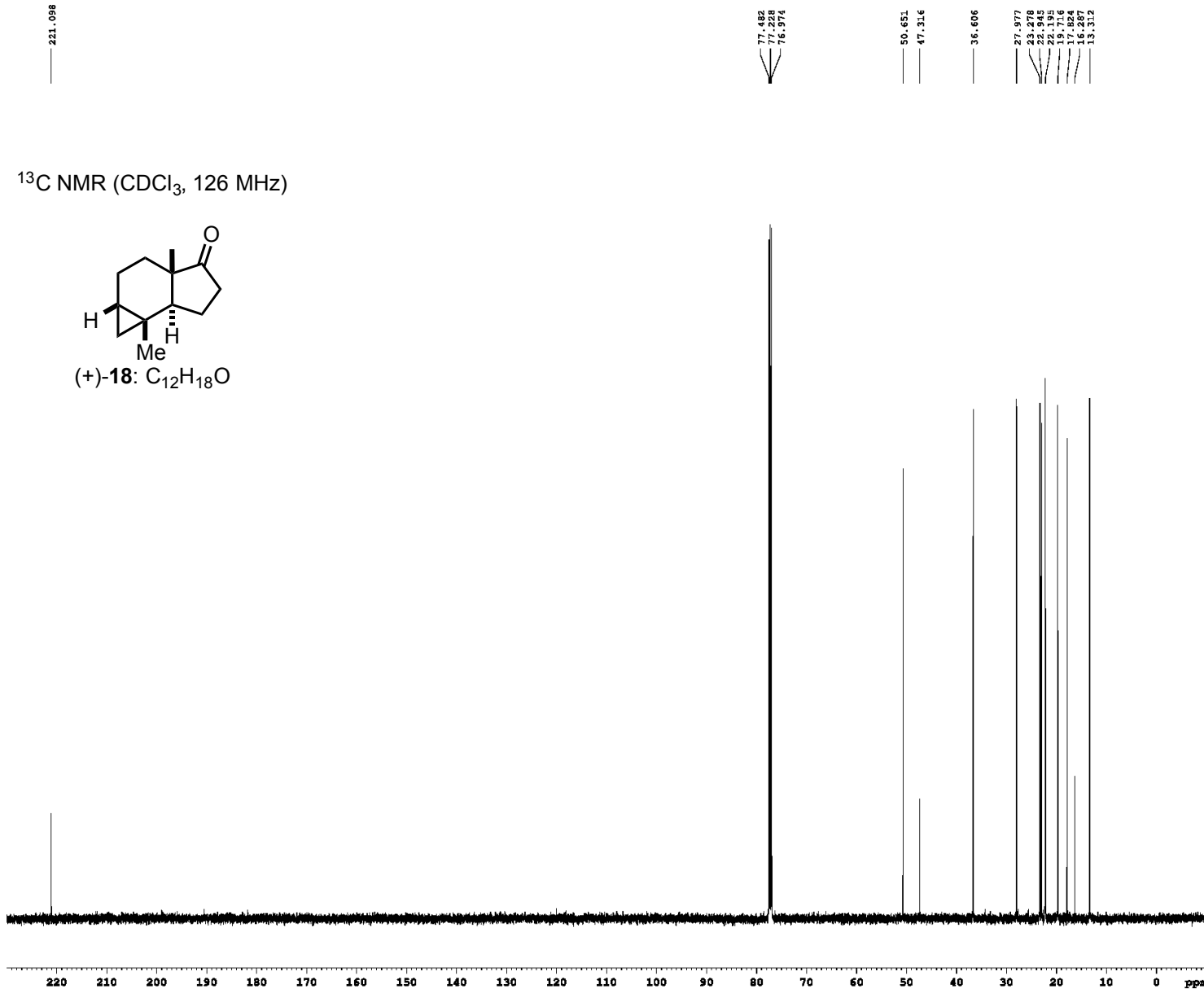
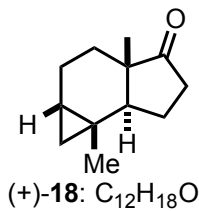
CHANNEL #1  
SFO1 600.1342009 MHz  
NUC1 1H  
P1 8.00 usec  
PIW1 24.00000000 W

F2 - Processing parameters  
SI 65536  
SF 600.1300354 MHz  
WDW SH  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

YS-IV-29

221.098

<sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz)



```
Current Data Parameters
NAME      YS-IV-29
EXPNO     4
PROCNO    1

F2 - Acquisition Parameters
Data_     20160107
Time      16.47
INSTRUM   cryo500
PROBED    5 mm CPIC1 1H-
PULPROG   SpinEchopg30pp.prd
TD         65536
SOLVENT   CDCl3
NS         832
DS         2
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ          1.0813440 sec
RG          7298.2
DW          16.500 usec
DE          6.00 usec
TE          298.0 K
d1          0.25000000 sec
d11         0.03000000 sec
d16         0.00020000 sec
d17         0.00019600 sec
MCREST     0 sec
MCWRK     0.01500000 sec
P2         33.10 usec

===== CHANNEL f1 =====
NUC1       13C
P1         16.55 usec
P11        500.00 usec
P12        2000.00 usec
PLO        120.00 dB
PL1        -1.00 dB
SFO1       125.7942548 MHz
SP1        2.70 dB
SP2        2.70 dB
SPNAM[1]   Crp60,0.5,20.1
SPNAM[2]   Crp60comp.4
SPOFF1     0 Hz
SPOFF2     0 Hz

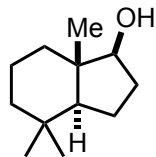
===== CHANNEL f2 =====
CDPRG[2]   waitz16
NUC2       1H
PCPD2      100.00 usec
PL2        1.60 dB
PL12       24.50 dB
SFO2       500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM[1]   SINE.100
GPNAM[2]   SINE.100
GPX1       0 %
GPX2       0 %
GPY1       0 %
GPY2       0 %
GPZ1       30.00 %
GPZ2       50.00 %
p15        500.00 usec
p16        1000.00 usec

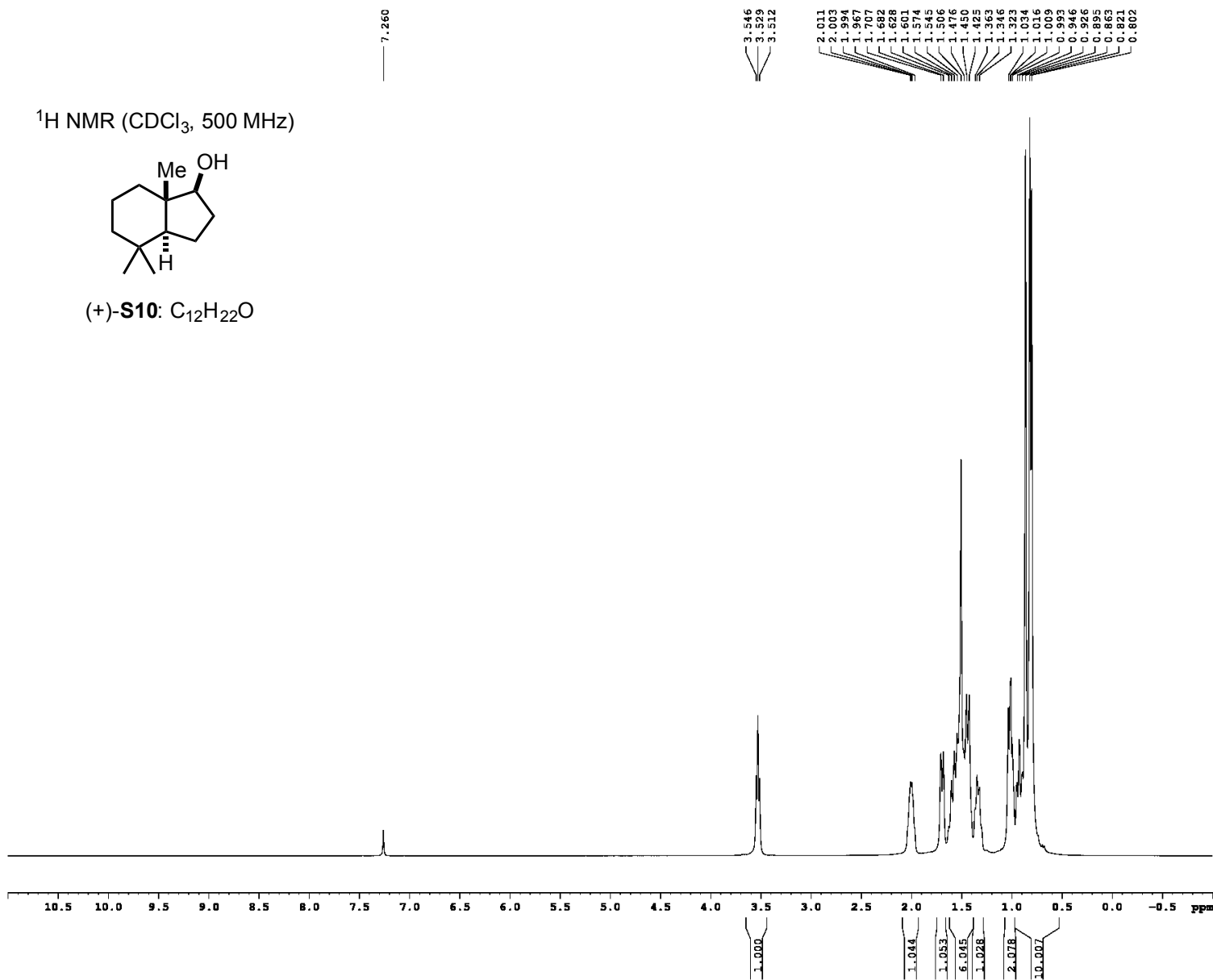
F2 - Processing parameters
SI         65536
SF         125.7803979 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00
```

YS-IV-24

$^1\text{H}$  NMR ( $\text{CDCl}_3$ , 500 MHz)



(+)-S10:  $\text{C}_{12}\text{H}_{22}\text{O}$

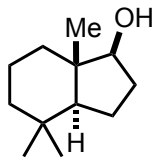


Current Data Parameters  
NAME YS-IV-24  
EXPNO 5  
PROCNO 1  
F2 - Acquisition Parameters  
Date\_ 20151216  
Time 15.49  
INSTRUM cryo300  
PROBHD 5 mm CPYCI 1H-  
PULPROG zgpg30  
TD 81728  
SOLVENT CDCl3  
NS 4  
DS 2  
SWH 8012.820 Hz  
FIDRES 0.098043 Hz  
AQ 5.0988273 sec  
RG 5  
DW 62.400 usec  
DS 6.00 usec  
TS 298.0 K  
D1 0.10000000 sec  
MCREST 0 sec  
MCMXK 0.01500000 sec  
CHANNEL F1  
NUC1 1H  
P1 7.50 usec  
PI1 1.50 dB  
SFO1 500.225015 MHz  
F2 - Processing parameters  
SI 65536  
SF 500.2200314 MHz  
WDW EM  
SBB 0  
IE 0.30 Hz  
GB 0  
PC 4.00

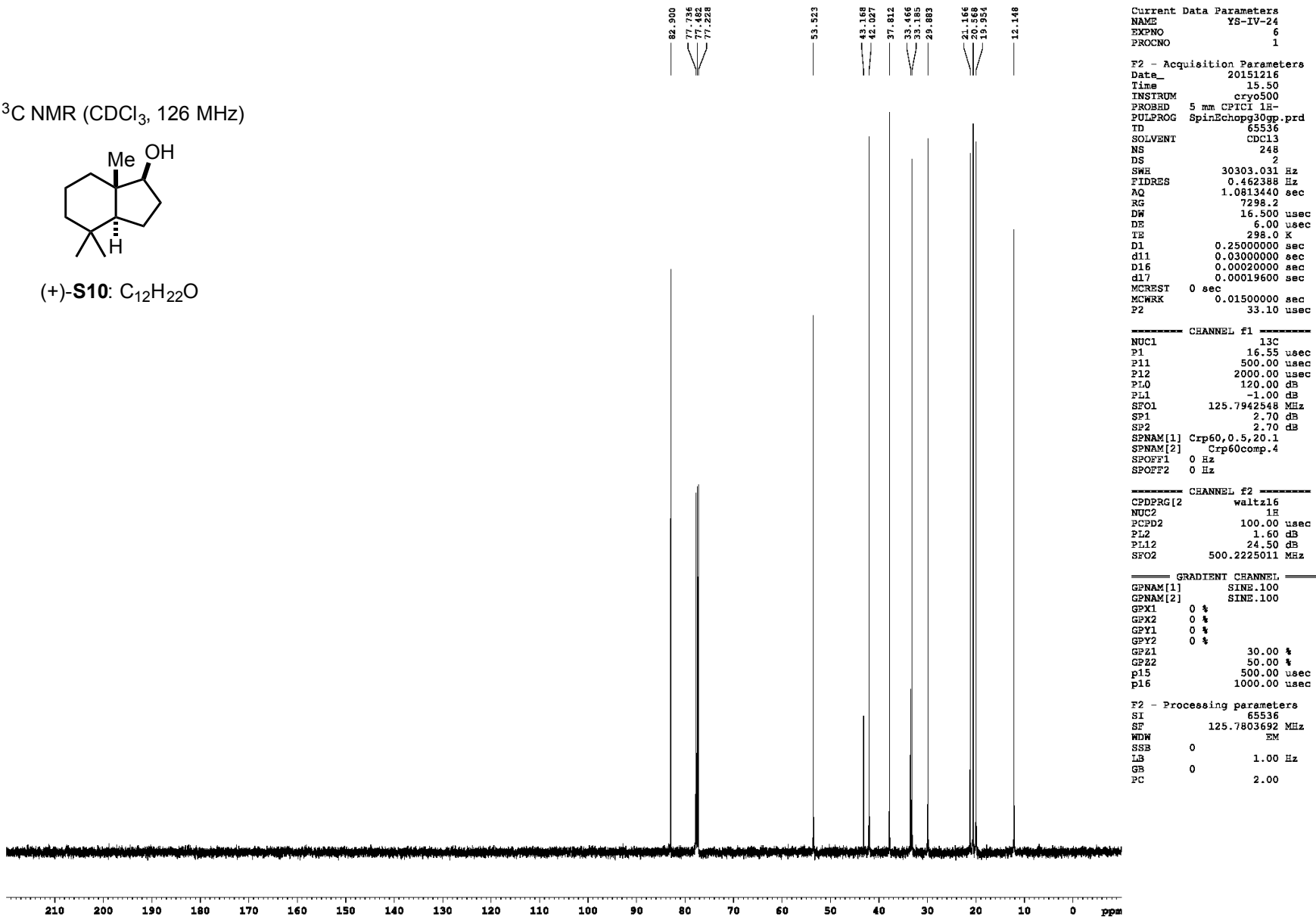


YS-IV-24

<sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz)

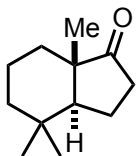


(+)-S10: C<sub>12</sub>H<sub>22</sub>O

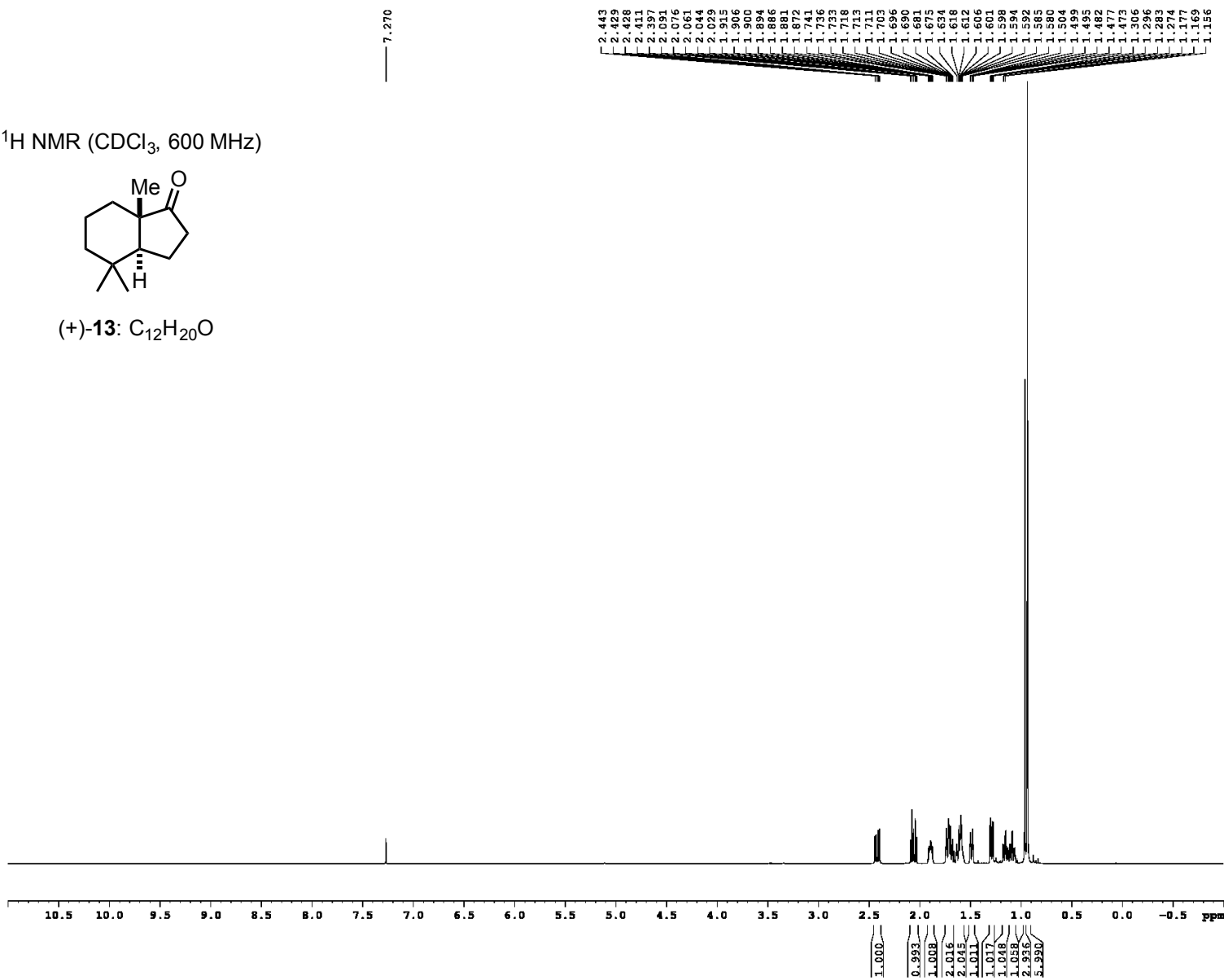


YS-IV-25

$^1\text{H}$  NMR ( $\text{CDCl}_3$ , 600 MHz)



(+)-13:  $\text{C}_{12}\text{H}_{20}\text{O}$

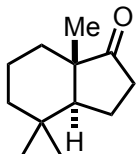


Current Data Parameters  
NAME YS-IV-25  
EXPNO 1  
PROCNO 1  
F2 - Acquisition Parameters  
Date\_ 20151217  
Time 10.44  
INSTRUM aw600  
PROBHD 5 mm TBI 1H/13  
PULPROG zg30  
TD 68074  
SOLVENT CDCl3  
NS 4  
DS 2  
SWH 9613.385 Hz  
FIDRES 0.098042 Hz  
AQ 5.0988478 sec  
RG 228  
DW 52.000 usec  
DE 14.54 usec  
TE 293.0 K  
D1 0.1000000 sec  
TD0 1  
===== CHANNEL f1 =====  
SFO1 600.1342039 MHz  
NUC1 13  
P1 8.00 usec  
PLW1 24.0000000 W  
F2 - Processing parameters  
SI 6536  
SF 600.1300296 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

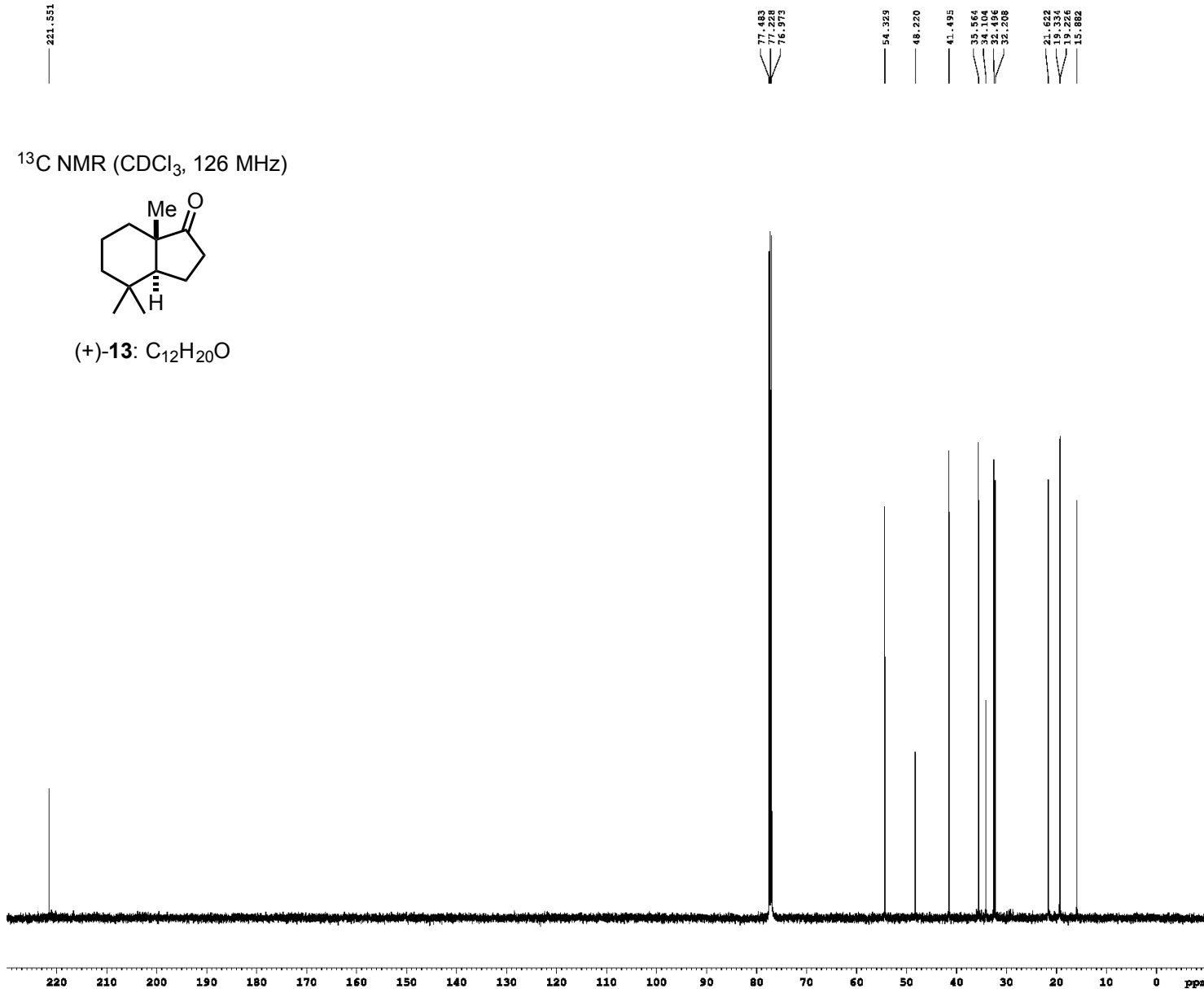
YS-IV-25

221.551

$^{13}\text{C}$  NMR ( $\text{CDCl}_3$ , 126 MHz)



(+)-13:  $\text{C}_{12}\text{H}_{20}\text{O}$



Current Data Parameters  
NAME YS-IV-25  
EXPNO 2  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20151217  
Time 10.53  
INSTRUM gn500  
PROBED 5 mm broadband  
PULPROG zgdc30  
TD 65536  
SOLVENT CDCl3  
NS 800  
DS 2  
SWH 30303.031 Hz  
FIDRES 0.462388 Hz  
AQ 1.0813440 sec  
RG 18390.4  
DW 16.500 usec  
DE 4.50 usec  
TE 298.0 K  
D1 0.25000000 sec  
d11 0.03000000 sec  
MCREST 0 sec  
MCWRK 0.01500000 sec

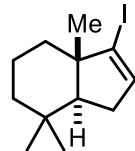
CHANNEL f1  
NUC1 13C  
P1 9.00 usec  
PL1 -0.60 dB  
SFO1 125.5327181 MHz

CHANNEL f2  
CPDPRG[2] waltz16  
NUC2 1H  
PCPD2 80.00 usec  
PL2 -3.00 dB  
PL12 12.80 dB  
SFO2 499.1824959 MHz

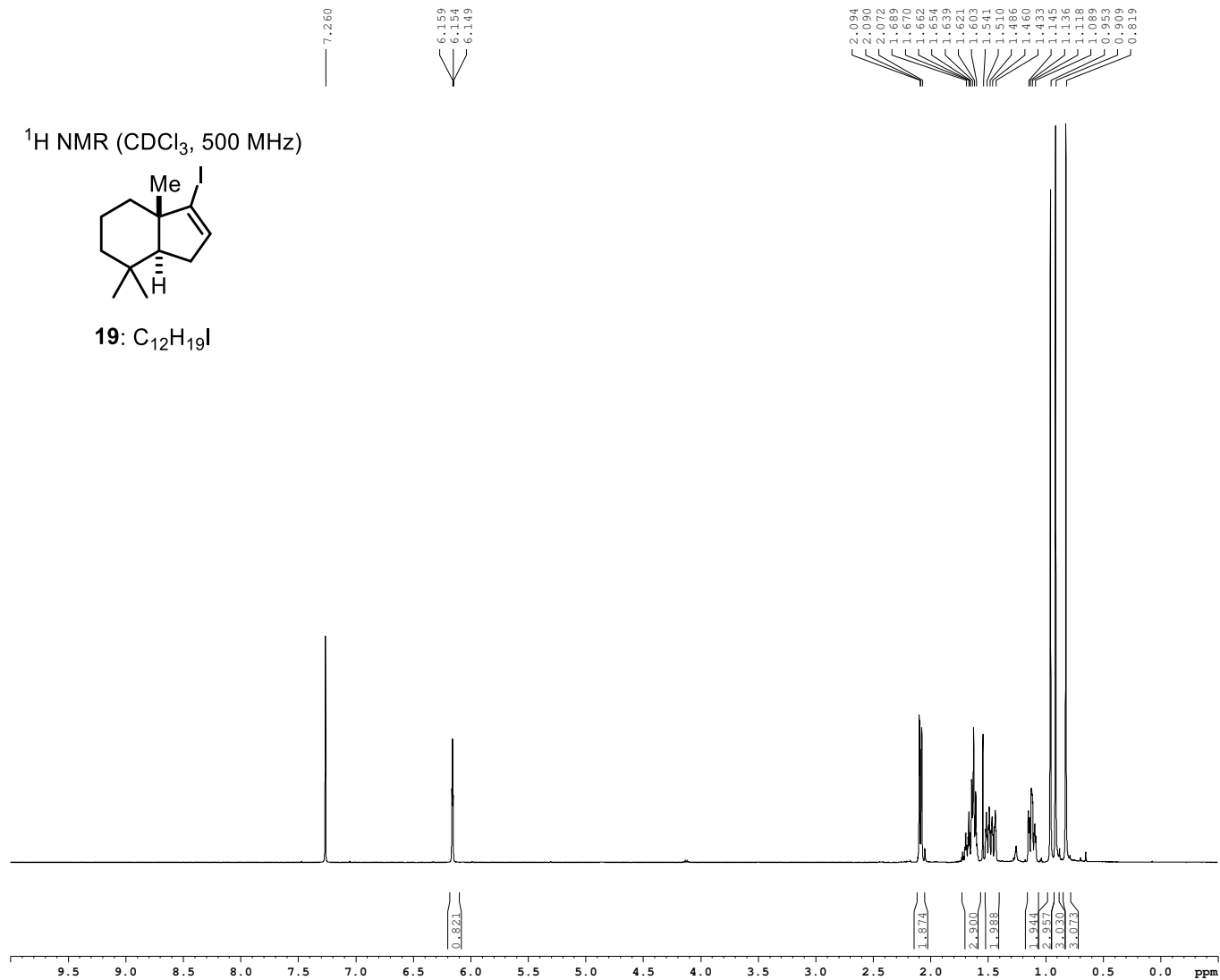
F2 - Processing parameters  
SI 65536  
SF 125.5188906 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
GB 0  
PC 2.00

**<sup>1</sup>H spectrum**

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)



**19: C<sub>12</sub>H<sub>19</sub>I**



Current Data Parameters  
 NAME DJT-V-036  
 EXPNO 1  
 PROCNO 1

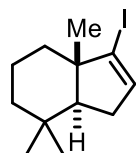
F2 - Acquisition Parameters  
 Date\_ 20150203  
 Time 18.19  
 INSTRUM cryo500  
 PROBHD 5 mm CPTCI 1H-  
 PULPROG zg30  
 TD 16022  
 SOLVENT CDCl3  
 NS 60  
 DS 0  
 SWH 8012.820 Hz  
 FIDRES 0.500114 Hz  
 AQ 0.9997728 sec  
 RG 5.7  
 DW 62.400 usec  
 DE 6.00 usec  
 TE 298.0 K  
 D1 0.10000000 sec  
 MCREST 0 sec  
 MCWRK 0.01500000 sec

===== CHANNEL f1 =====  
 NUC1 1H  
 P1 7.50 usec  
 PL1 1.60 dB  
 SFO1 500.2235015 MHz

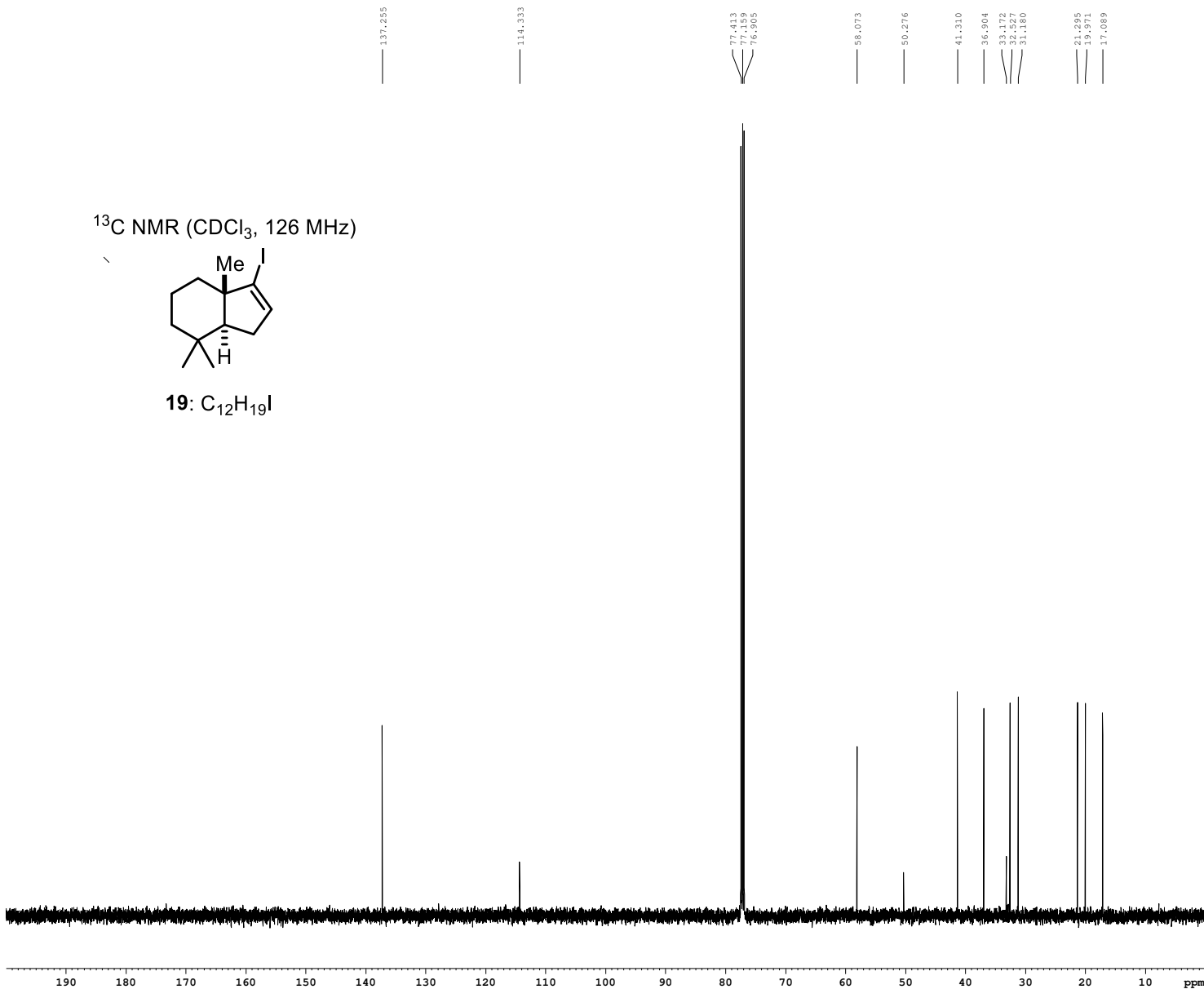
F2 - Processing parameters  
 SI 65536  
 SF 500.2200311 MHz  
 WDW no  
 SSB 0  
 LB 0 Hz  
 GB 0  
 PC 4.00

Z-restored spin-echo <sup>13</sup>C spectrum with <sup>1</sup>H decoupling

<sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz)



19: C<sub>12</sub>H<sub>19</sub>I



```

Current Data Parameters
NAME          DJT-V-036
EXPNO         2
PROCNO        1

F2 - Acquisition Parameters
Date_         20150203
Time          18.24
INSTRUM       cryo500
PROBHD        5 mm CPTCI 1H-
PULPROG       SpinEchopg30gp.prd
TD            65536
SOLVENT       CDCl3
NS            391
DS            0
SWH           30303.031 Hz
FIDRES        0.462388 Hz
AQ            1.0813440 sec
RG            13004
DW            16.500 usec
DE            6.00 usec
TE            298.0 K
D1            0.25000000 sec
d11           0.03000000 sec
D16           0.00020000 sec
d17           0.00019600 sec
MCREST        0 sec
MCWRK         0.01500000 sec
P2            33.10 usec

===== CHANNEL f1 =====
NUC1           13C
P1             16.55 usec
P11            500.00 usec
P12            2000.00 usec
PL0            120.00 dB
PL1            -1.00 dB
SFO1           125.7942548 MHz
SP1            2.70 dB
SP2            2.70 dB
SPNAM[1]      Crp60,0.5,20.1
SPNAM[2]      Crp60comp.4
SPOFF1         0 Hz
SPOFF2         0 Hz

===== CHANNEL f2 =====
CPDPRG[2]     waltz16
NUC2           1H
PCPD2         100.00 usec
PL2            1.60 dB
PL12           24.50 dB
SFO2           500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM[1]      SINE.100
GPNAM[2]      SINE.100
GPX1          0 %
GPX2          0 %
GPY1          0 %
GPY2          0 %
GPZ1          30.00 %
GPZ2          50.00 %
p15           500.00 usec
p16           1000.00 usec

F2 - Processing parameters
SI            65536
SF            125.7804074 MHz
WDW           EM
SSB           0
LB            1.00 Hz
GB            0
PC            2.00
    
```

**<sup>1</sup>H spectrum**

7.328  
7.313  
7.307  
7.299  
7.268  
7.260  
7.256  
7.245

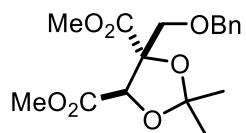
5.118

4.527  
4.503  
4.486  
4.462

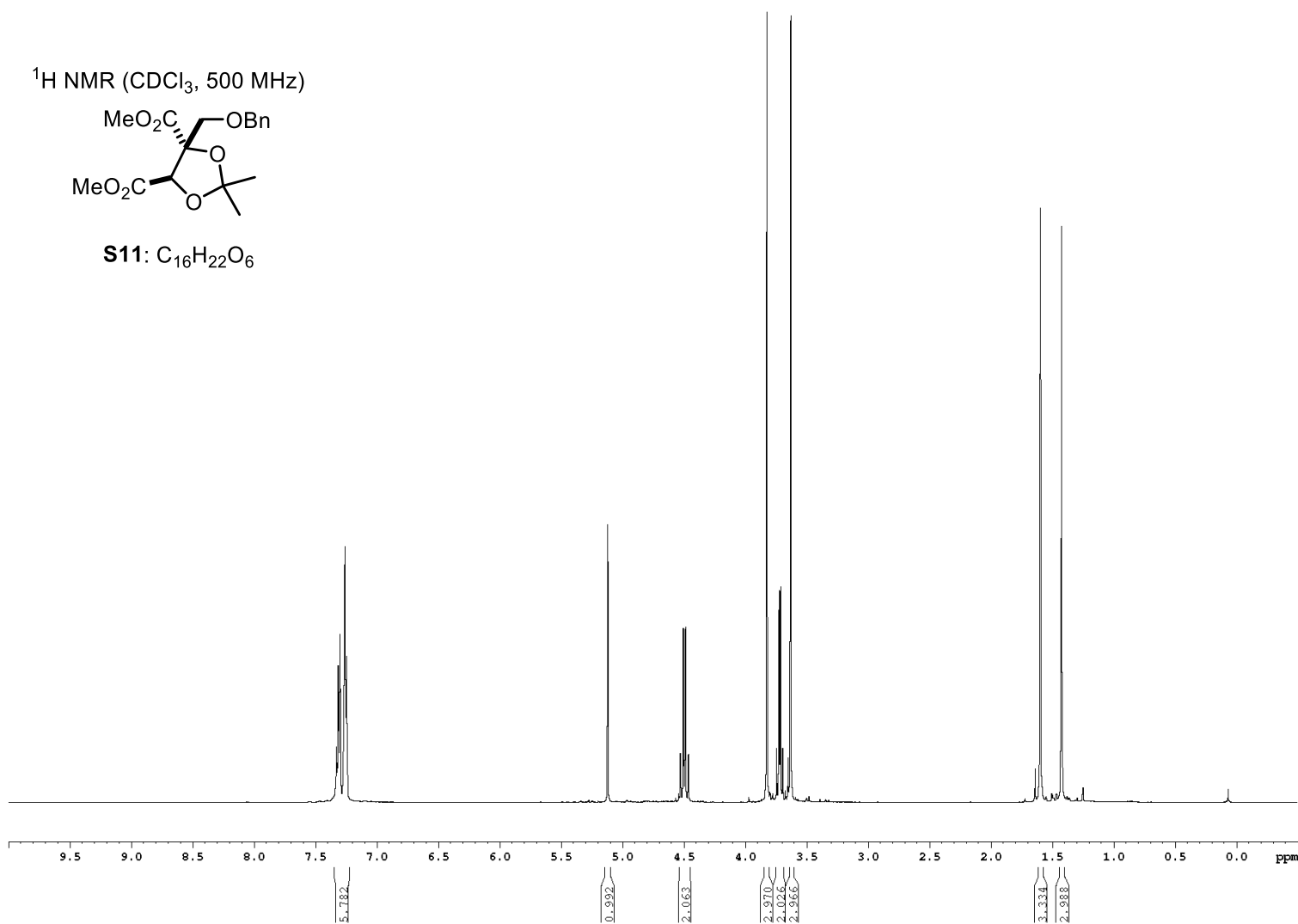
3.822  
3.742  
3.723  
3.712  
3.692  
3.689  
3.629

1.637  
1.594  
1.422

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)



**S11: C<sub>16</sub>H<sub>22</sub>O<sub>6</sub>**



```

Current Data Parameters
NAME      DJT-VI-104
EXPNO     1
PROCNO    1

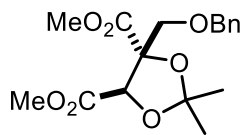
F2 - Acquisition Parameters
Date_     20151118
Time      7.36
INSTRUM   cryo500
PROBHD    5 mm CPTCI 1H-
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         30
DS         0
SWH        8012.620 Hz
FIDRES     0.100003 Hz
AQ          4.9998622 sec
RG          8
DM          62.400 usec
DE          6.00 usec
TE          298.0 K
D1          0.10000000 sec
SFO1       500.225015 MHz

===== CHANNEL f1 =====
NUC1        1H
P1          7.50 usec
PL1         1.00 dB
SFO1       500.225015 MHz

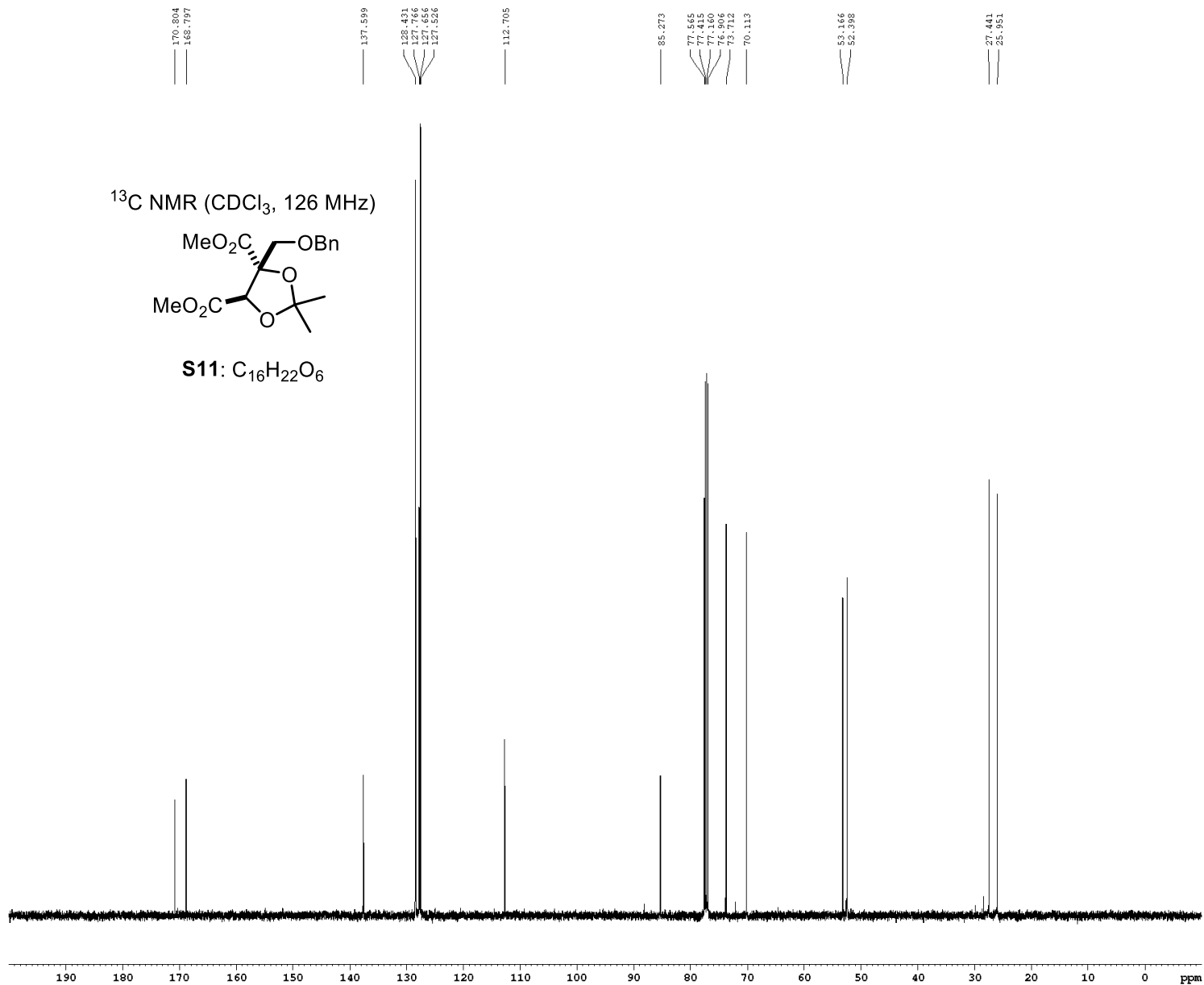
F2 - Processing parameters
SI          65536
SF          500.2200325 MHz
WDW         EM
SSB         0
LB          0.30 Hz
GB          0
PC          4.00
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling

<sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz)



S11: C<sub>16</sub>H<sub>22</sub>O<sub>6</sub>



```

Current Data Parameters
NAME      DJT-VI-104
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20151118
Time      7.42
INSTRUM   cryo500
PROBHD    5 mm CPTCI 1H-
PULPROG   SpinEchopg30gp.prd
TD         65536
SOLVENT   CDCl3
NS         374
DS         0
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813440 sec
RG         6502
DW         16.500 usec
DE         6.00 usec
TE         298.0 K
D1         0.25000000 sec
d11        0.03000000 sec
D16        0.00020000 sec
d17        0.00019600 sec
MCREST     0 sec
MCWRK     0.01500000 sec
F2         33.10 usec

===== CHANNEL f1 =====
NUC1       13C
P1         16.55 usec
P11        500.00 usec
P12        2000.00 usec
PL0        120.00 dB
PL1        -1.00 dB
SF01       125.7942548 MHz
SP1        2.70 dB
SP2        2.70 dB
SPNAM[1]   Crp60,0.5,20.1
SPNAM[2]   Crp60comp.4
SPOFF1     0 Hz
SPOFF2     0 Hz

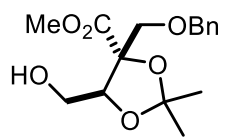
===== CHANNEL f2 =====
CPDPRG[2]  waltz16
NUC2       1H
PCPD2      100.00 usec
PL2        1.60 dB
PL12       24.50 dB
SF02       500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM[1]   SINE.100
GPNAM[2]   SINE.100
GPX1       0 %
GPX2       0 %
GPY1       0 %
GPY2       0 %
GPZ1       30.00 %
GPZ2       50.00 %
p15        500.00 usec
p16        1000.00 usec

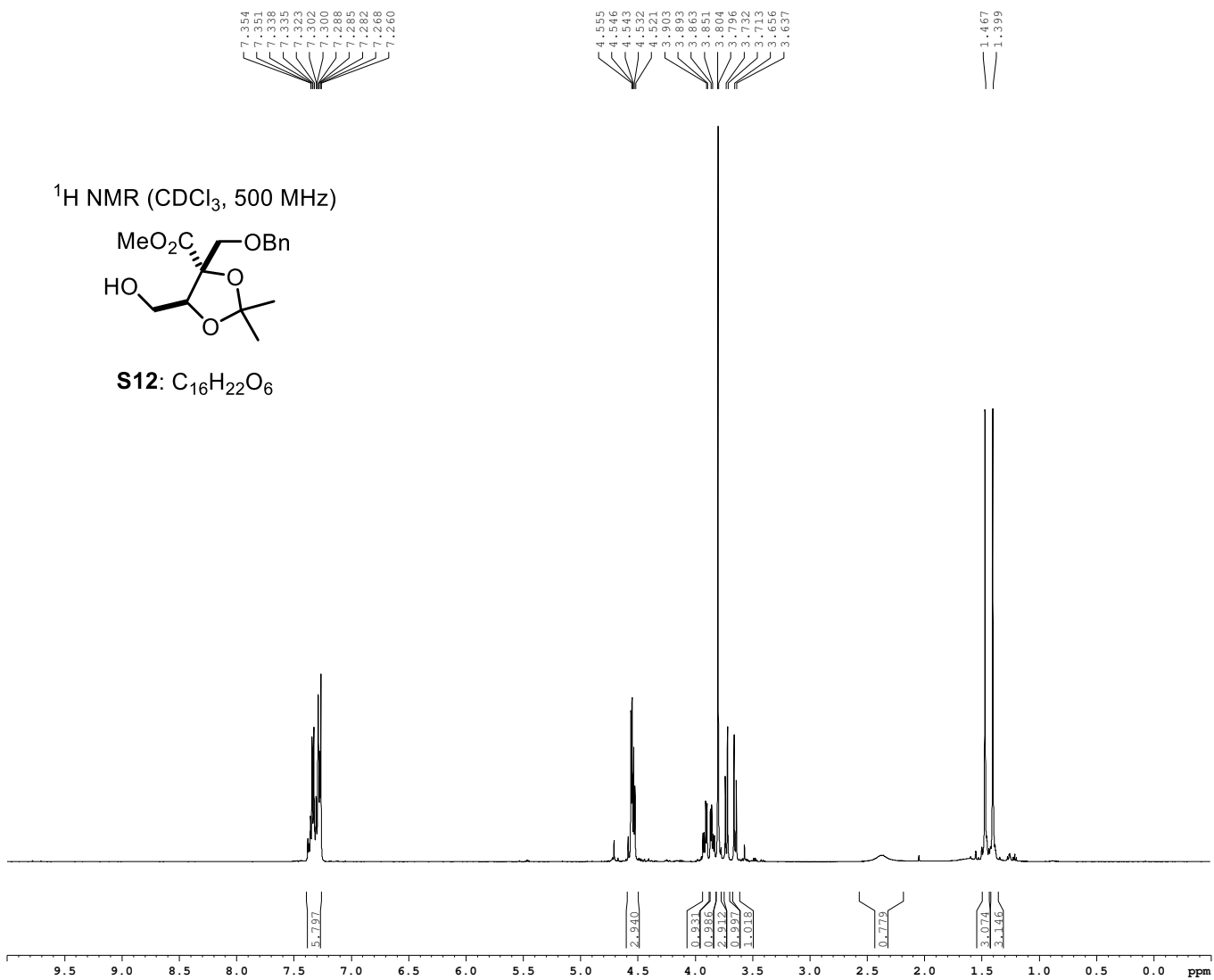
F2 - Processing parameters
SI         65536
SF         125.7804103 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00
    
```

**<sup>1</sup>H spectrum**

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)



**S12: C<sub>16</sub>H<sub>22</sub>O<sub>6</sub>**



```

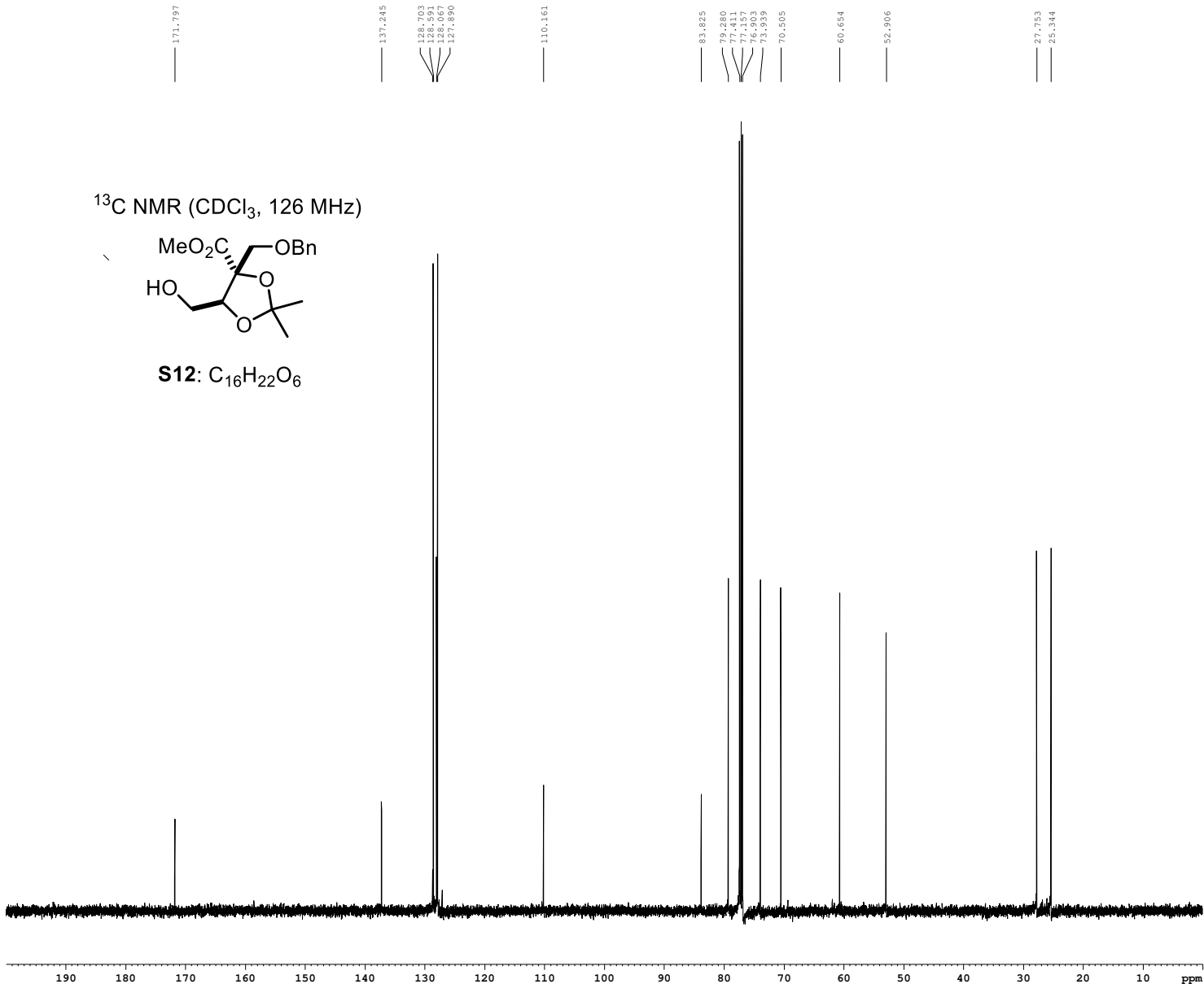
Current Data Parameters
NAME      DJT-IV-110
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20140813
Time     14.46
INSTRUM  cryo500
PROBHD   5 mm CPTCI 1H-
PULPROG  zg30
TD        81728
SOLVENT  CDCl3
NS        8
DS        2
SWH       8012.820 Hz
FIDRES    0.098043 Hz
AQ        5.0998273 sec
RG        7.1
DW        62.400 usec
DE        6.00 usec
TE        298.0 K
D1        0.10000000 sec
MCREST   0 sec
MCWRK    0.01500000 sec

===== CHANNEL f1 =====
NUC1      1H
P1        7.50 usec
PL1       1.60 dB
SF01      500.2235015 MHz

F2 - Processing parameters
SI        65536
SF        500.2200307 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        4.00
    
```





```

Current Data Parameters
NAME      DJT-IV-110
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20140814
Time      11.08
INSTRUM   cryo500
PROBHD    5 mm CPTCI 1H-
PULPROG   SpinEchopg30gp.prd
TD         65536
SOLVENT   CDCl3
NS         562
DS         16
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ          1.0813440 sec
RG          7298.2
DW          16.500 usec
DE          6.00 usec
TE          298.0 K
D1          0.25000000 sec
d11         0.03000000 sec
D16         0.00020000 sec
d17         0.00019600 sec
MCREST     0 sec
MCWRK      0.01500000 sec
P2          31.00 usec

===== CHANNEL f1 =====
NUC1        13C
P1          15.50 usec
P11         500.00 usec
P12         2000.00 usec
PL0         120.00 dB
PL1         -1.00 dB
SFO1        125.7942548 MHz
SP1         3.20 dB
SP2         3.20 dB
SPNAM[1]    Crp60,0.5,20.1
SPNAM[2]    Crp60comp.4
SPOFF1      0 Hz
SPOFF2      0 Hz

===== CHANNEL f2 =====
CPDPRG[2]   waltz16
NUC2         1H
PCPD2       100.00 usec
PL2         1.60 dB
PL12        24.60 dB
SFO2        500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM[1]     SINE.100
GPNAM[2]     SINE.100
GPX1         0 %
GPX2         0 %
GPY1         0 %
GPY2         0 %
GPZ1         30.00 %
GPZ2         50.00 %
p15          500.00 usec
p16          1000.00 usec

F2 - Processing parameters
SI          65536
SF          125.7804089 MHz
WDW         EM
SSB         0
LB          1.00 Hz
GB          0
PC          2.00

```

YS-III-303

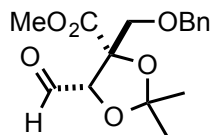
9.696  
9.695

7.354  
7.351  
7.342  
7.340  
7.329  
7.304  
7.292  
7.279  
7.270  
7.260

4.900  
4.898  
4.491  
4.471  
4.457  
4.437  
3.843  
3.834  
3.676  
3.659  
3.650  
3.634

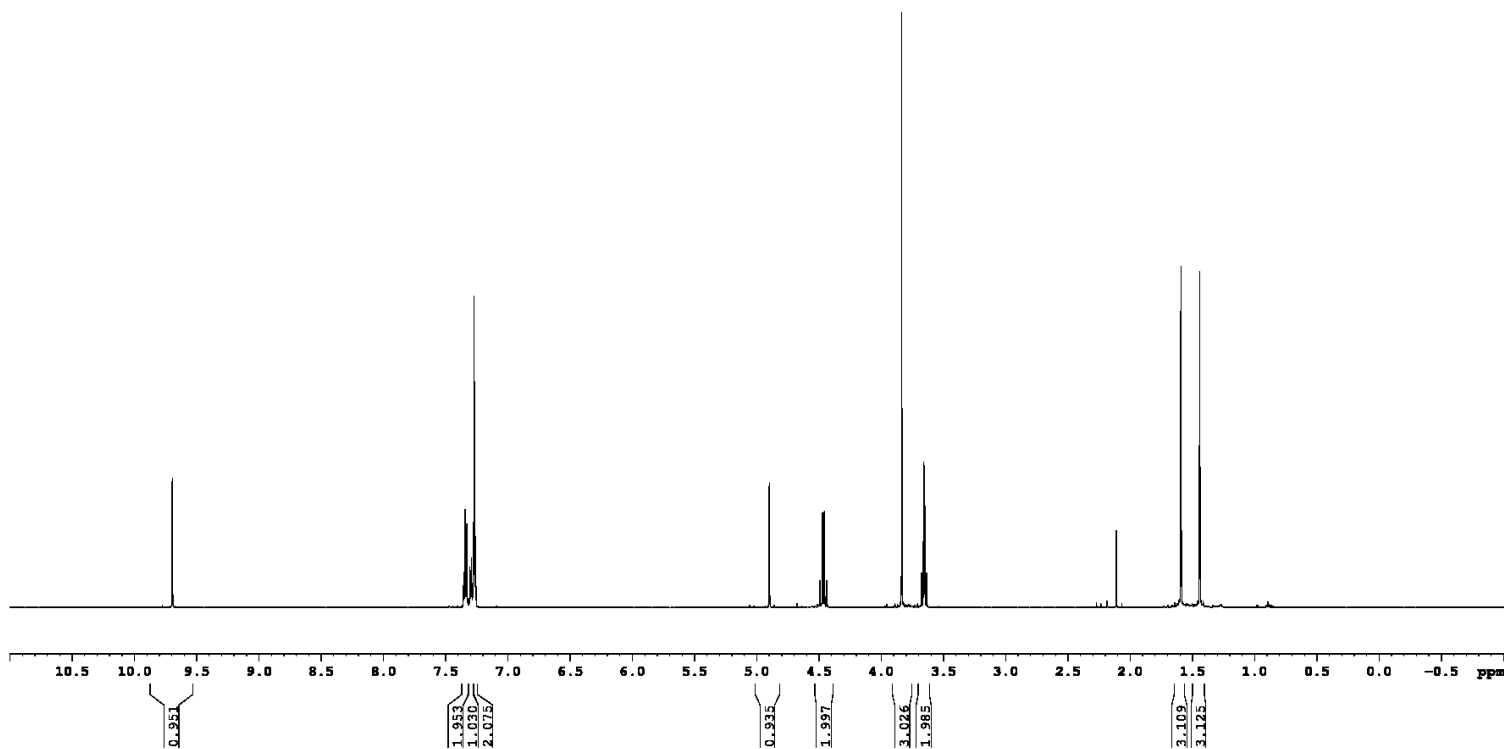
2.110  
1.591  
1.440

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz)

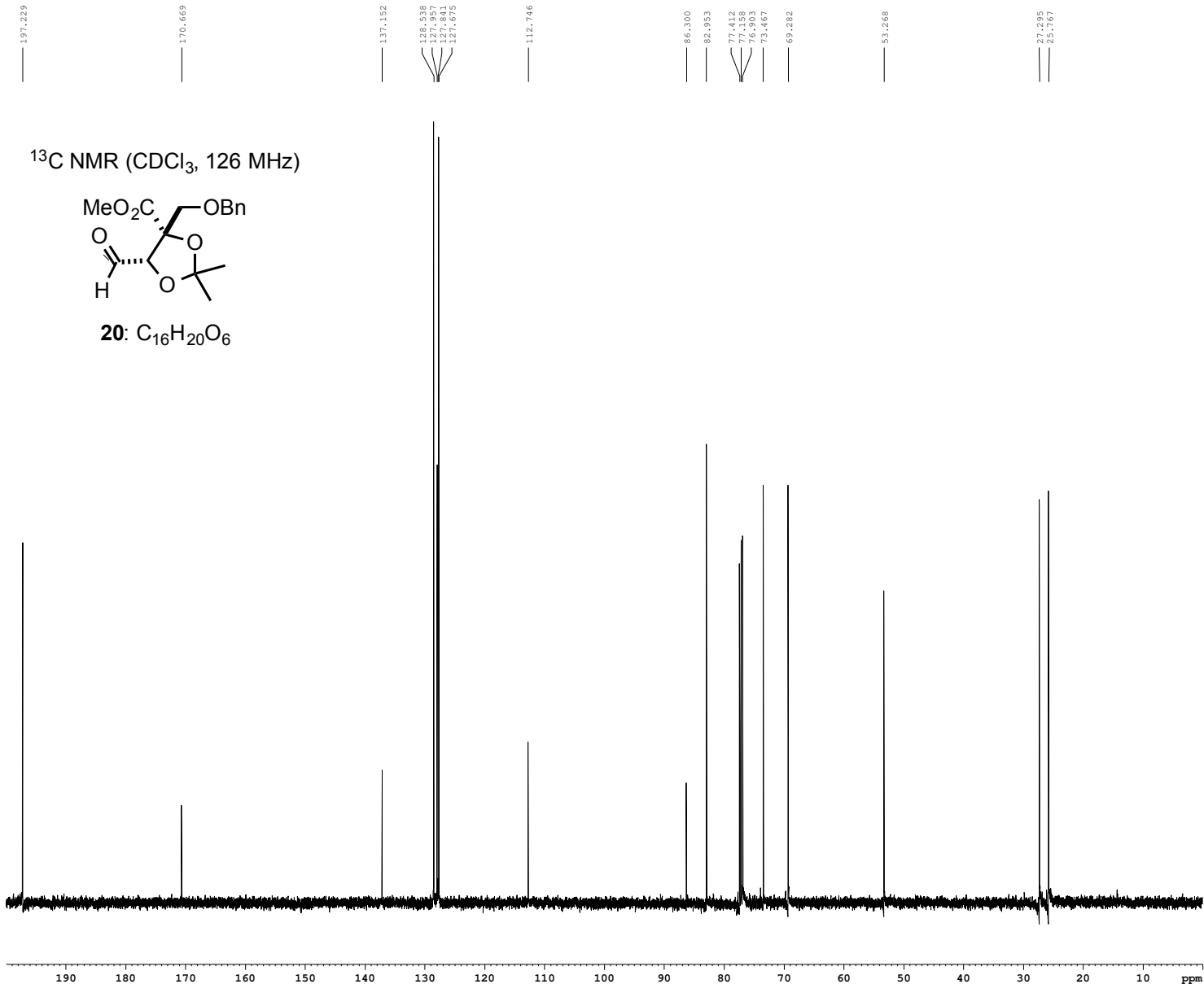


20: C<sub>16</sub>H<sub>20</sub>O<sub>6</sub>

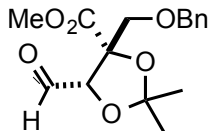
```
Current Data Parameters
NAME      YS-III-303
EXPNO     1
PROCNO    1
F2 - Acquisition Parameters
Date_     20151128
Time      13.45
INSTRUM   av600
PROBHD    5 mm TBI 1H/13
PULPROG   zgpg
TD         38400
SOLVENT   CDCl3
NS         4
DS         2
SWH       9612.382 Hz
FIDRES    0.250010 Hz
AQ         1.9999200 sec
RG         1620
DW         52.000 use
DS         14.54 use
TS         298.0 K
D1         0.1000000 sec
ZD0        1
===== CHANNEL f1 =====
SFO1     600.1342039 MHz
NUC1      13
P1         8.00 use
P1W1     24.00000000 W
F2 - Processing parameters
SI         6536
SF         600.1300297 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
```



Z-restored spin-echo 13C spectrum with 1H decoupling



<sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz)



20: C<sub>16</sub>H<sub>20</sub>O<sub>6</sub>

```

Current Data Parameters
NAME      DJT-IV-040
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20140616
Time      16.32
INSTRUM   cryo500
PROBHD    5 mm CPTCI 1H-
PULPROG   SpinEchopg30gp.prd
TD         65536
SOLVENT   CDCl3
NS         101
DS         0
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ          1.0813440 sec
RG          6502
DW          16.500 usec
DE          6.00 usec
TE          298.0 K
D1          0.25000000 sec
d11         0.03000000 sec
D16         0.00020000 sec
d17         0.00019600 sec
MCREST     0 sec
MCWRK      0.01500000 sec
P2          31.00 usec

===== CHANNEL f1 =====
NUC1        13C
P1          15.50 usec
P11         500.00 usec
P12         2000.00 usec
PL0         120.00 dB
PL1         -1.00 dB
SFO1        125.7942548 MHz
SP1         3.20 dB
SP2         3.20 dB
SPNAM[1]    Crp60,0.5,20.1
SPNAM[2]    Crp60comp.4
SPOFF1      0 Hz
SPOFF2      0 Hz

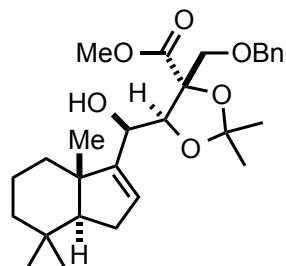
===== CHANNEL f2 =====
CPDPRG[2]   waltz16
NUC2         1H
PCPD2       100.00 usec
PL2         1.60 dB
PL12        24.60 dB
SFO2        500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM[1]    SINE.100
GPNAM[2]    SINE.100
GPX1        0 %
GPX2        0 %
GPY1        0 %
GPY2        0 %
GPZ1        30.00 %
GPZ2        50.00 %
p15         500.00 usec
p16         1000.00 usec

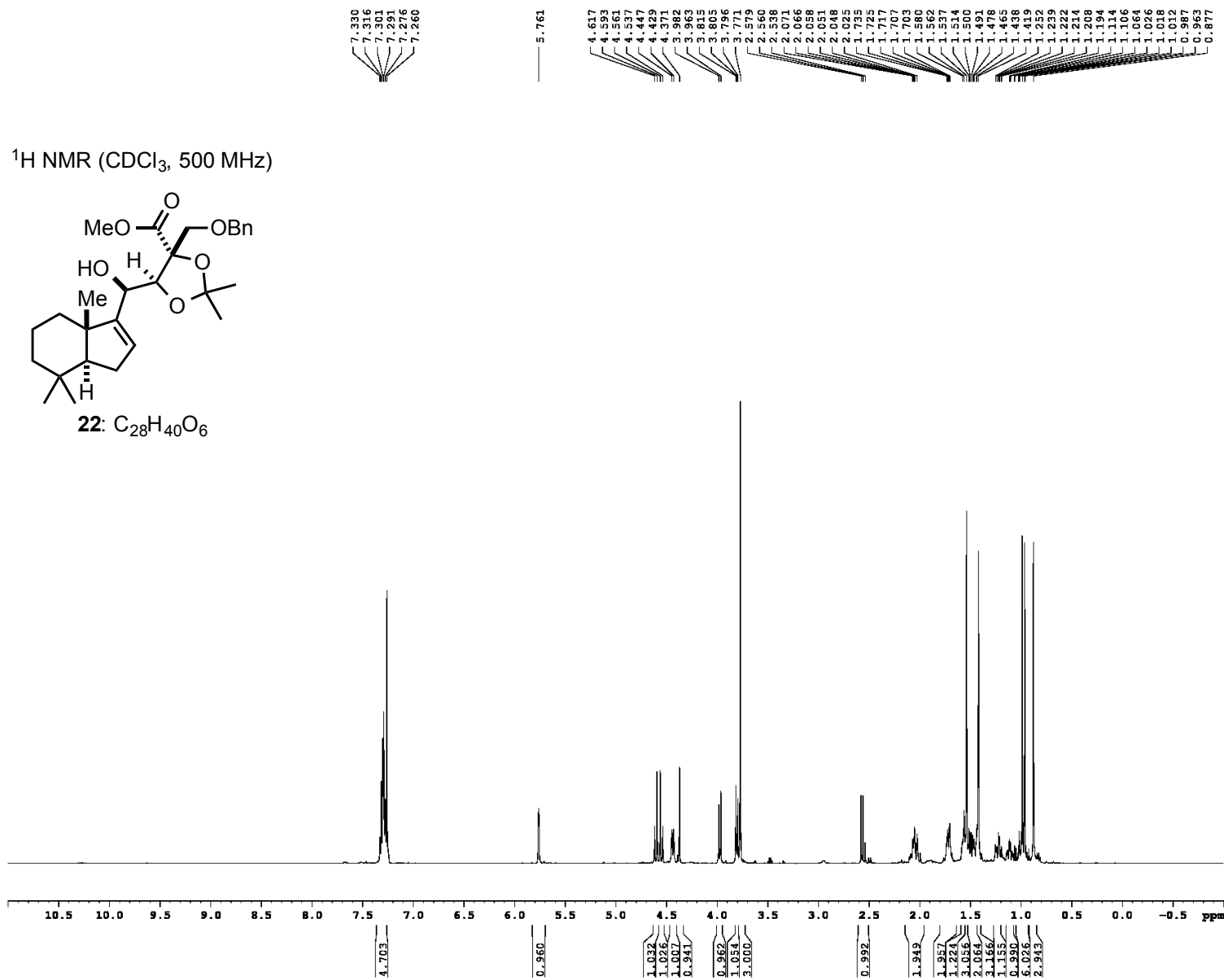
F2 - Processing parameters
SI          65536
SF          125.7804122 MHz
WDW         EM
SSB         0
LB          1.00 Hz
GB          0
PC          2.00
    
```

YS-III-303

$^1\text{H}$  NMR ( $\text{CDCl}_3$ , 500 MHz)



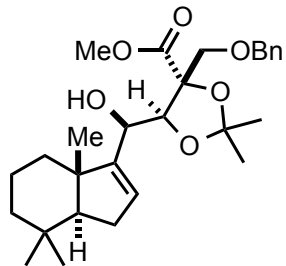
22:  $\text{C}_{28}\text{H}_{40}\text{O}_6$



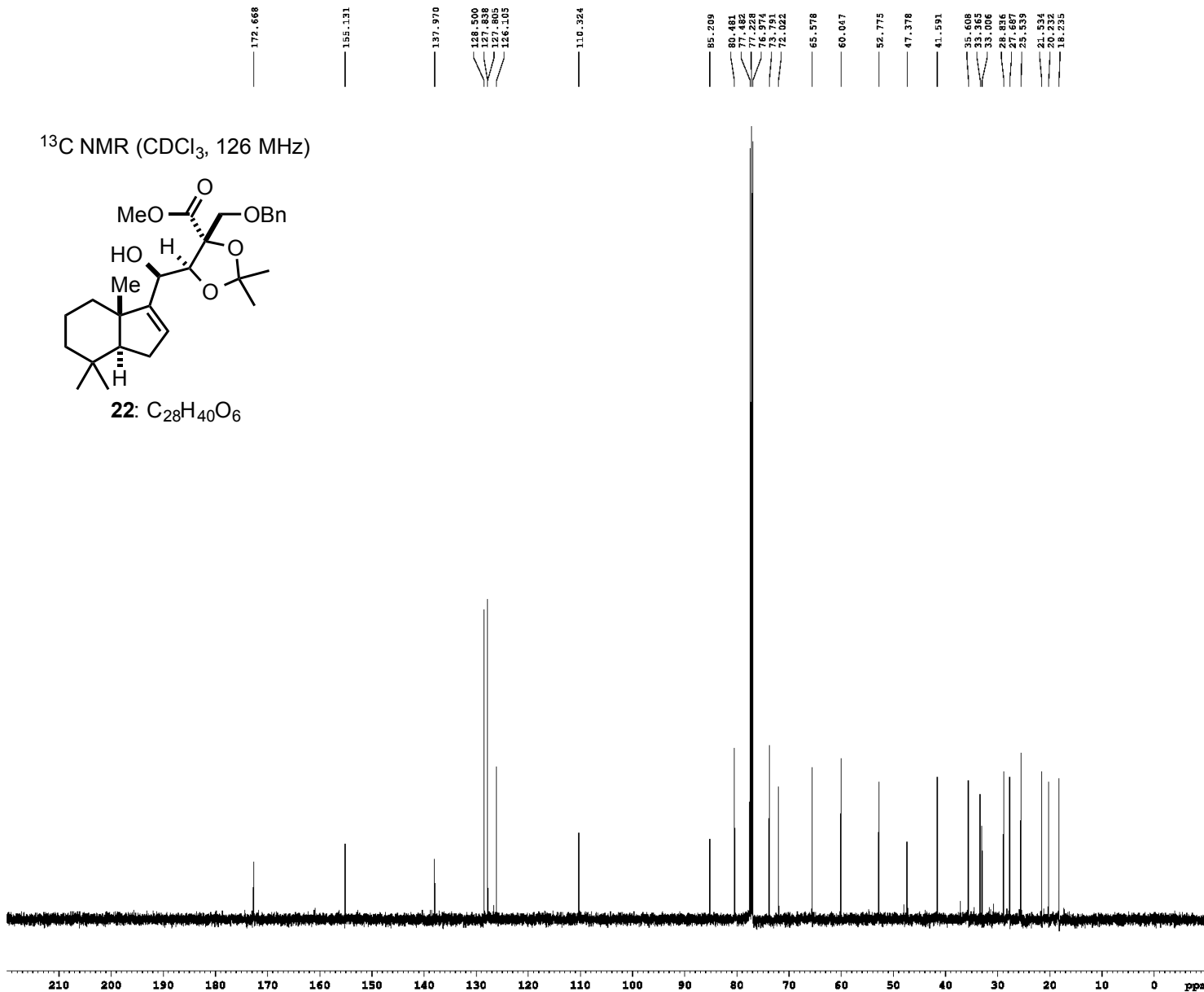
Current Data Parameters  
NAME YS-III-303  
EXPNO 5  
PROCNO 1  
F2 - Acquisition Parameters  
Date\_ 20151202  
Time 13.46  
INSTRUM cryo300  
PROBHD 5 mm CPXI 1H-  
PULPROG zgpg30  
TD 81728  
SOLVENT CDCl3  
NS 4  
DS 2  
SWH 8012.820 Hz  
FIDRES 0.098043 Hz  
AQ 5.0988273 sec  
RG 4  
DW 62.400 use  
DS 6.00 use  
TS 298.0 K  
D1 0.1000000 sec  
MCRET 0 sec  
MCRBK 0.0150000 sec  
CHANNEL F1  
NUC1 1H  
P1 7.50 use  
PI1 1.60 dB  
SFO1 500.225015 MHz  
F2 - Processing parameters  
SI 65536  
SF 500.2200320 MHz  
WDW EM  
SBB 0  
LB 0.30 Hz  
GB 0  
PC 4.00

YS-III-303

<sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz)



22: C<sub>28</sub>H<sub>40</sub>O<sub>6</sub>



```

Current Data Parameters
NAME      YS-III-303
EXPNO     6
PROCNO    1

F2 - Acquisition Parameters
Data_     20151202
Time      13.57
INSTRUM   cryo500
PROBED    5 mm CPIC1 1H-
PULPROG   SpinEchopg30gp.prd
TD         65536
SOLVENT   CDCl3
NS         496
DS         2
SWH        30303.031 Hz
FIDRES     0.462388 Hz
AQ         1.0813440 sec
RG         2580.3
DW         16.500 usec
DE         6.00 usec
TE         298.0 K
d1         0.25000000 sec
d11        0.03000000 sec
d16        0.00020000 sec
d17        0.00019600 sec
MCREST    0 sec
MCWRK     0.01500000 sec
P2         33.10 usec

===== CHANNEL f1 =====
NUC1       13C
P1         16.55 usec
P11        500.00 usec
P12        2000.00 usec
PLO        120.00 dB
PL1        -1.00 dB
SFO1       125.7942548 MHz
SP1        2.70 dB
SP2        2.70 dB
SPNAM[1]   Crp60,0.5,20.1
SPNAM[2]   Crp60comp.4
SPOFF1     0 Hz
SPOFF2     0 Hz

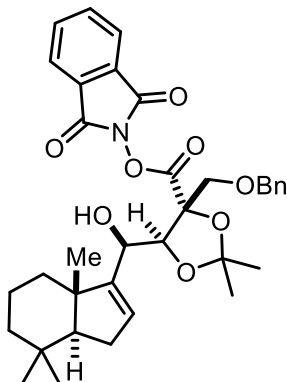
===== CHANNEL f2 =====
CDPRG[2]   waitz16
NUC2       1H
PCPD2     100.00 usec
PL2        1.60 dB
PL12       24.50 dB
SFO2       500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM[1]   SINE.100
GPNAM[2]   SINE.100
GPX1       0 %
GPX2       0 %
GPY1       0 %
GPY2       0 %
GPZ1       30.00 %
GPZ2       50.00 %
p15        500.00 usec
p16        1000.00 usec

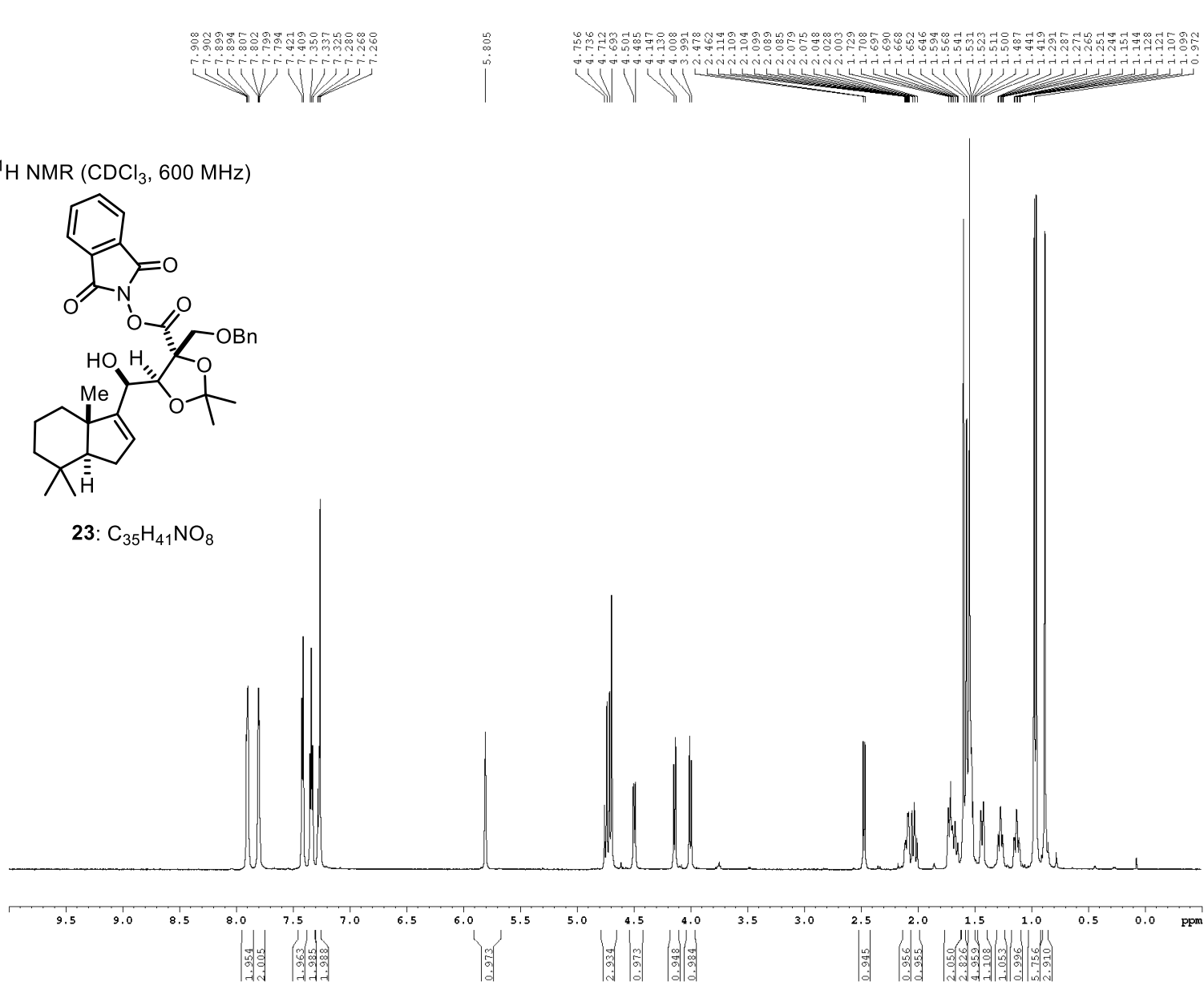
F2 - Processing parameters
SI         65536
SF         125.7803996 MHz
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         2.00
    
```

<sup>1</sup>H spectrum

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz)



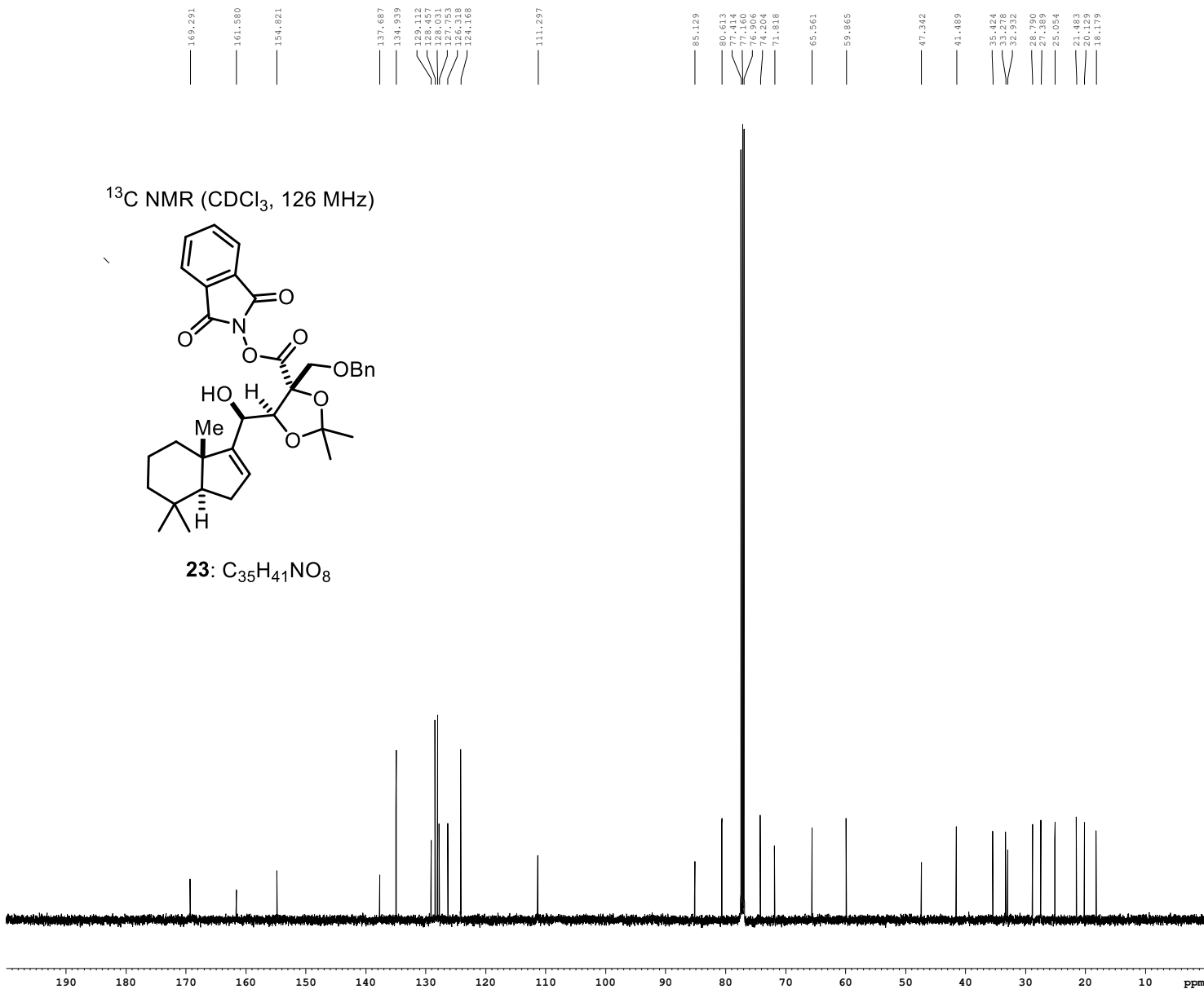
23: C<sub>35</sub>H<sub>41</sub>NO<sub>8</sub>



```

Current Data Parameters
NAME          D:\7-17-20
EXPNO         1
PROCNO        1
F2 - Acquisition Parameters
Date_         20141013
Time          16.31
INSTRUM       svt600
PROBHD        5 mm TBI IN13
PULPROG       zg30
TD            66874
SOLVENT       CDCl3
NS            12
DS            0
SHH           9615.385 Hz
FIDRES        0.098042 Hz
AQ            5.0998478 sec
RG            322
DM            52.0000 usec
DE            14.54 usec
TE            298.0 K
D2            0.10000000 sec
TDO           1
===== CHANNEL f1 =====
SFO1          600.1342009 MHz
NUC1          1H
P1            9.00 usec
PLW1          23.01441956 W
F2 - Processing parameters
SI            65536
SF            600.1300943 MHz
WDW           EM
SSB           0
LB            0.30 Hz
GB            0
PC            1.00
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
NAME      DJT-IV-203
EXPNO    8
PROCNO   1

F2 - Acquisition Parameters
Date_    20150414
Time     8.55
INSTRUM  cryo500
PROBHD   5 mm CPTCI 1H-
PULPROG  SpinEchopg30gp.prd
TD       65536
SOLVENT  CDCl3
NS       857
DS       0
SWH      30303.031 Hz
FIDRES   0.462388 Hz
AQ       1.0813440 sec
RG       7298.2
DW       16.500 usec
DE       6.00 usec
TE       298.0 K
D1       0.25000000 sec
d11      0.03000000 sec
D16      0.00020000 sec
d17      0.00019600 sec
MCREST   0 sec
MCWRK    0.01500000 sec
P2       33.10 usec

===== CHANNEL f1 =====
NUC1     13C
P1       16.55 usec
P11      500.00 usec
P12      2000.00 usec
PL0      120.00 dB
PL1      -1.00 dB
SFO1     125.7942548 MHz
SP1      2.70 dB
SP2      2.70 dB
SPNAM[1] Crp60,0.5,20.1
SPNAM[2] Crp60comp.4
SPOFF1   0 Hz
SPOFF2   0 Hz

===== CHANNEL f2 =====
CPDPRG[2] waltz16
NUC2     1H
PCPD2    100.00 usec
PL2      1.60 dB
PL12     24.50 dB
SFO2     500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM[1] SINE.100
GPNAM[2] SINE.100
GPX1     0 %
GPX2     0 %
GPY1     0 %
GPY2     0 %
GPZ1     30.00 %
GPZ2     50.00 %
p15      500.00 usec
p16      1000.00 usec

F2 - Processing parameters
SI       65536
SF       125.7804076 MHz
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
PC       2.00
    
```

<sup>1</sup>H spectrum

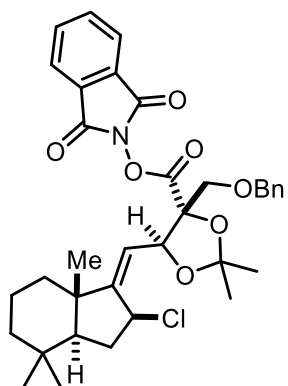
7.925  
7.919  
7.915  
7.911  
7.815  
7.809  
7.800  
7.390  
7.345  
7.330  
7.315  
7.282  
7.260

5.605  
5.586  
5.193  
5.174  
4.972  
4.957  
4.942  
4.715  
4.691  
4.659  
4.635

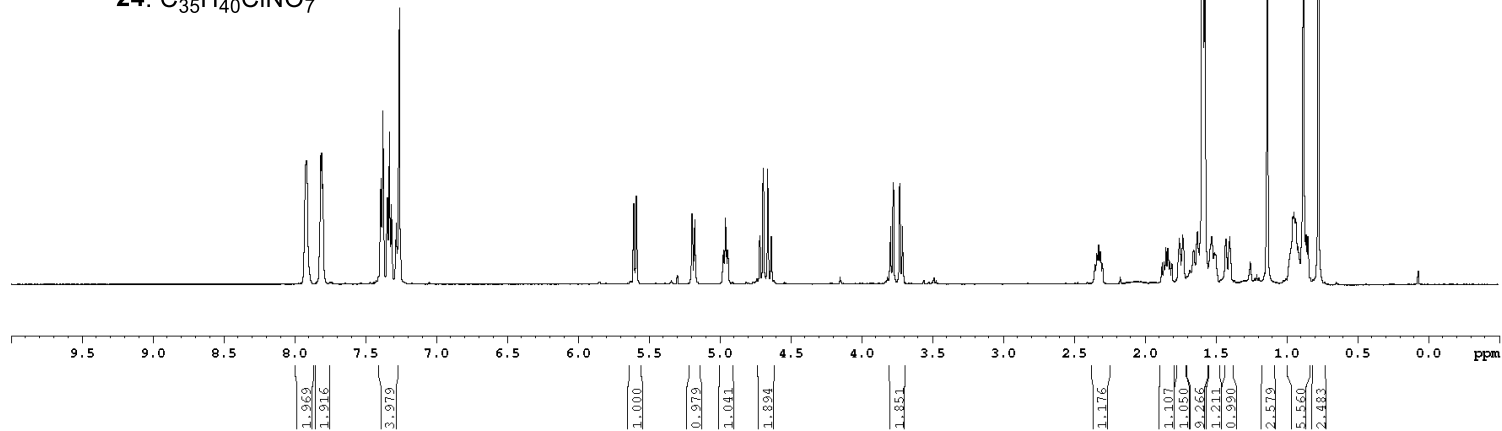
3.793  
3.774  
3.727  
3.708

2.337  
2.324  
2.312  
1.848  
1.834  
1.784  
1.729  
1.697  
1.689  
1.575  
1.527  
1.507  
1.425  
1.398  
1.134  
0.945  
0.945  
0.931  
0.868  
0.860  
0.807  
0.773

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)



24: C<sub>35</sub>H<sub>40</sub>ClNO<sub>7</sub>

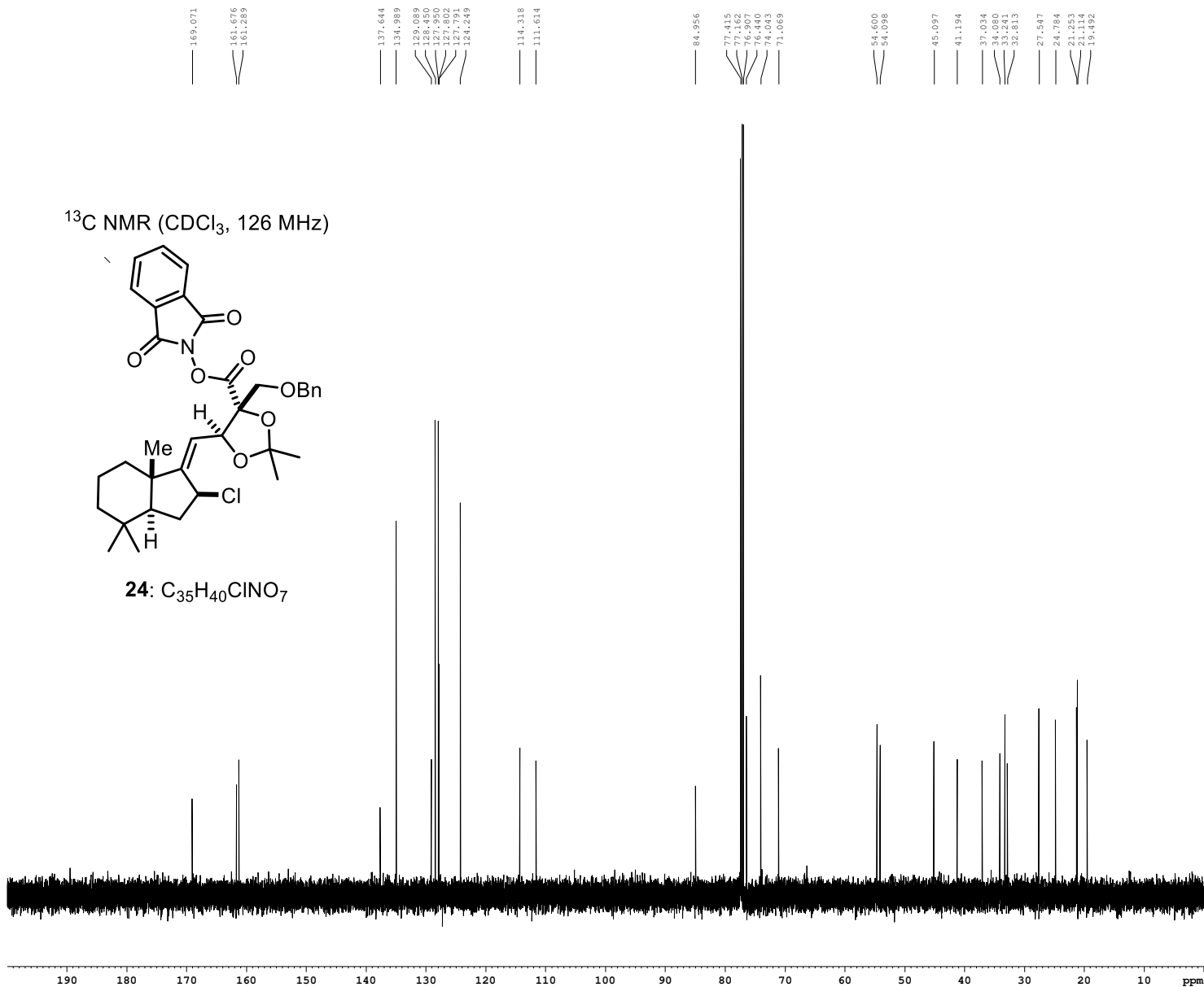


```

Current Data Parameters
NAME      DJY-IV-271
EXPNO    3
PROCNO   1
F2 - Acquisition Parameters
Date_    20141206
Time     11.39
INSTRUM  cryo500
PROBHD   5 mm CPTCI 1H-
PULPROG  zgpg30
TD        81728
SOLVENT  cdcl3
NS        48
DS        2
SWH       8012.820 Hz
FIDRES    0.098043 Hz
AQ        5.0998273 sec
RG         5.7
DW        62.400 usec
TE        6.00 usec
TR        298.0 K
DE        0.10000000 sec
REFREST   0 sec
CHECK     0.01500000 sec
===== CHANNEL f1 =====
NUC1      1H
P1        7.50 usec
PL1       1.60 dB
SFO1     500.225015 MHz
F2 - Processing parameters
SI        65536
SF        500.225015 MHz
WDW       no
SSB       0
LB        0 Hz
GB        0
PC        4.00
    
```



Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
NAME      DJT-IV-271
EXPNO    4
PROCNO   1

F2 - Acquisition Parameters
Date_    20141206
Time     11.49
INSTRUM  cryo500
PROBHD   5 mm CPTCI 1H-
PULPROG  SpinEchopg30gp.prd
TD       65536
SOLVENT  CDCl3
NS       269
DS       16
SWH      30303.031 Hz
FIDRES   0.462388 Hz
AQ       1.0813440 sec
RG       7298.2
DW       16.500 usec
DE       6.00 usec
TE       298.0 K
D1       0.25000000 sec
d11      0.03000000 sec
D16      0.00020000 sec
d17      0.00019600 sec
MCREST   0 sec
MCWRK    0.01500000 sec
P2       33.10 usec

===== CHANNEL f1 =====
NUC1     13C
P1       16.55 usec
P11      500.00 usec
P12      2000.00 usec
PL0      120.00 dB
PL1      -1.00 dB
SFO1     125.7942548 MHz
SP1      2.70 dB
SP2      2.70 dB
SPNAM[1] Crp60,0.5,20.1
SPNAM[2]  Crp60comp.4
SPOFF1   0 Hz
SPOFF2   0 Hz

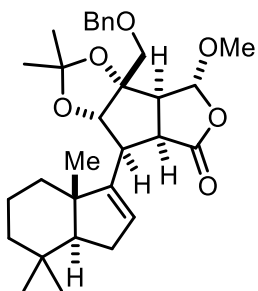
===== CHANNEL f2 =====
CPDPRG[2] waltz16
NUC2     1H
PCPD2    100.00 usec
PL2      1.60 dB
PL12     24.50 dB
SFO2     500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM[1] SINE.100
GPNAM[2] SINE.100
GPX1     0 %
GPX2     0 %
GPY1     0 %
GPY2     0 %
GPZ1     30.00 %
GPZ2     50.00 %
p15      500.00 usec
p16      1000.00 usec

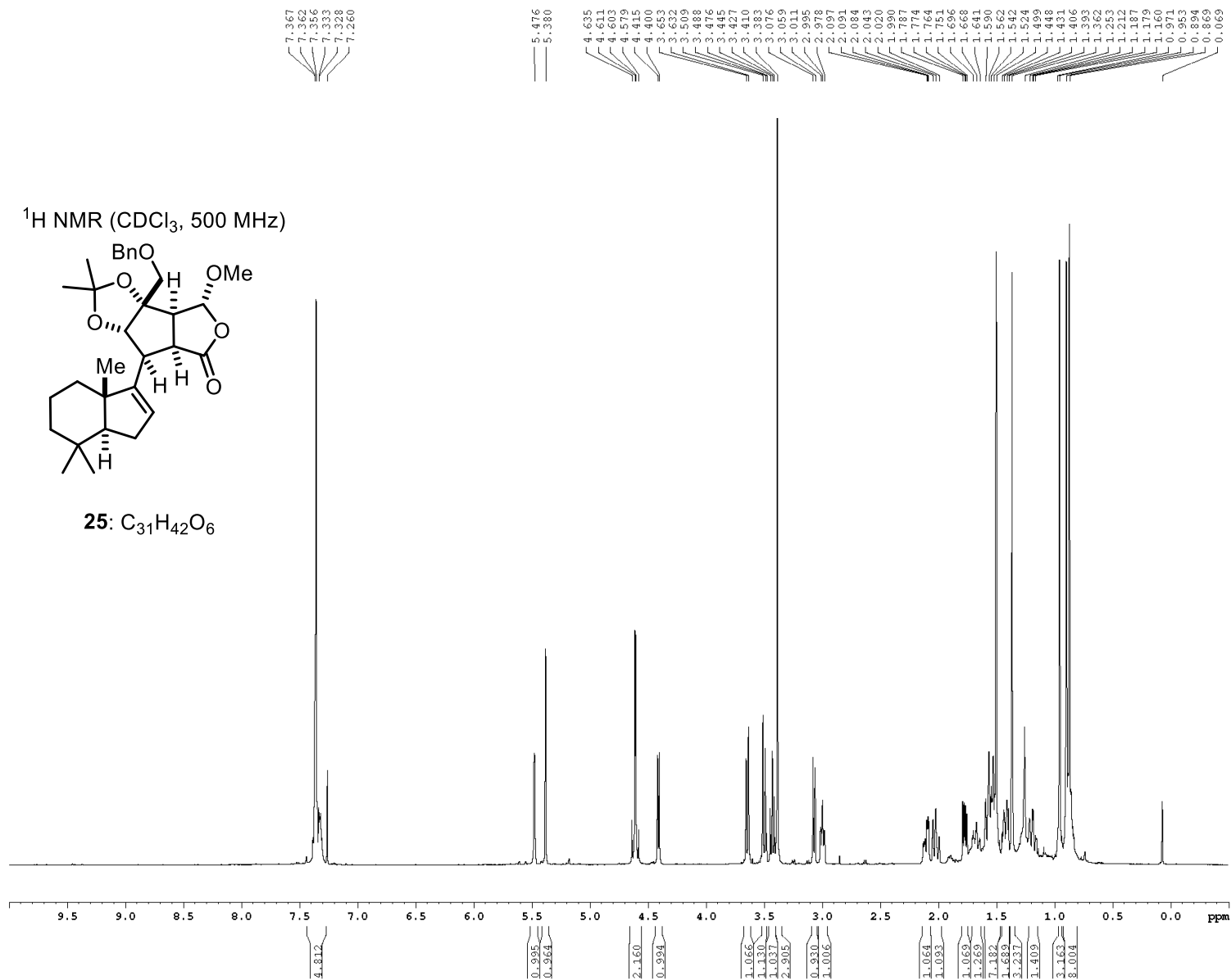
F2 - Processing parameters
SI       65536
SF       125.7804099 MHz
WDW      no
SSB      0
LB       0 Hz
GB       0
PC       2.00
    
```

<sup>1</sup>H spectrum

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)



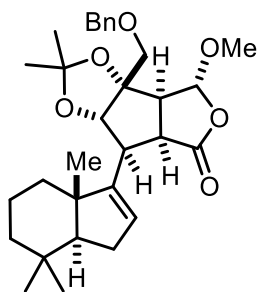
25: C<sub>31</sub>H<sub>42</sub>O<sub>6</sub>



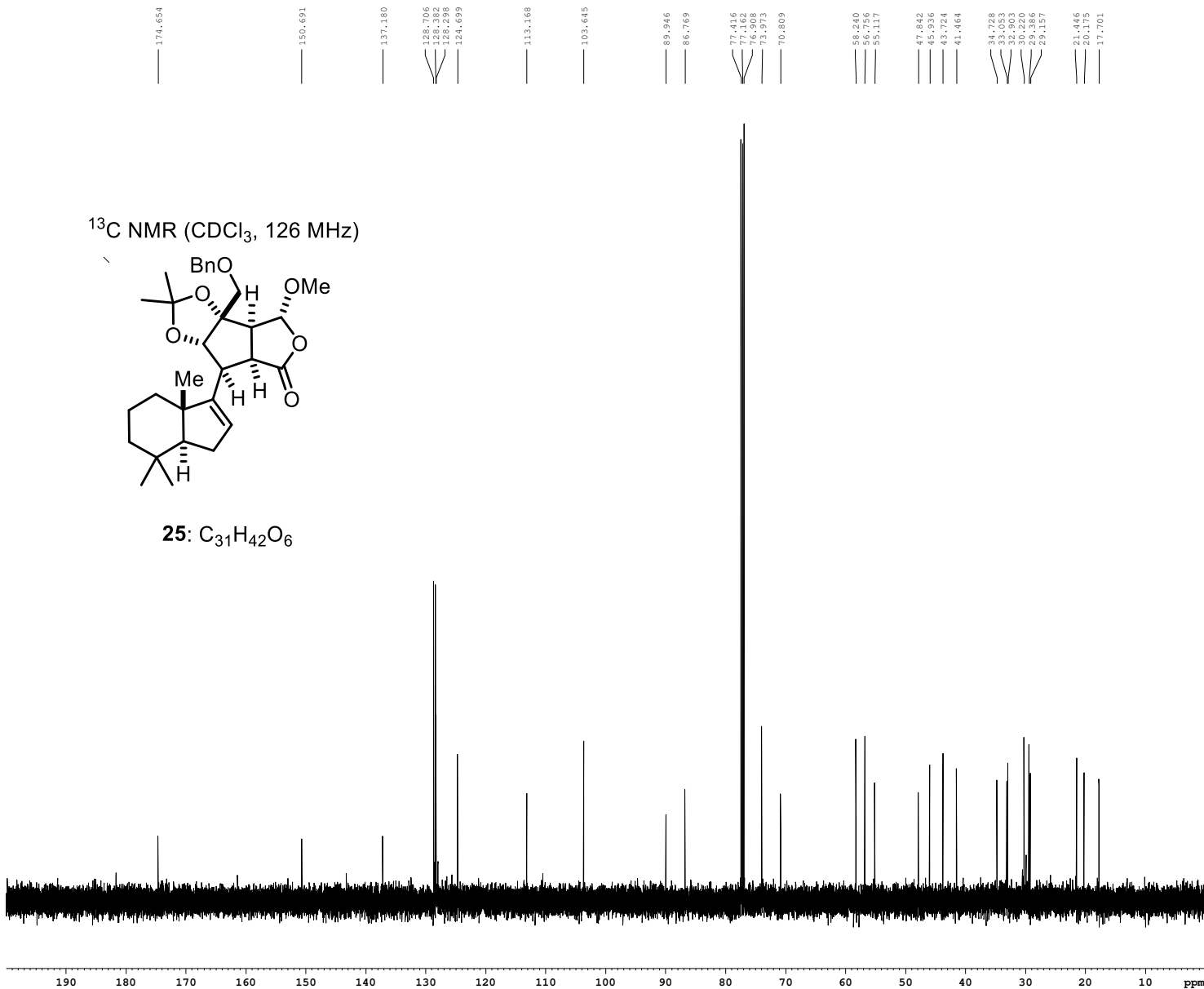
Current Data Parameters  
 NAME DJT-V-103  
 EXPNO 1  
 PROCNO 1  
 F2 - Acquisition Parameters  
 Date\_ 20150328  
 Time 14.16  
 INSTRUM cryo500  
 PEOBHD 5 mm CPTCI 1H-  
 PULPROG zg30  
 TD 16022  
 SOLVENT CDCl3  
 NS 30  
 DS 0  
 SWH 8012.820 Hz  
 FIDRES 0.590114 Hz  
 AQ 0.9997728 sec  
 RG 3.6  
 DW 62.400 usec  
 DE 6.00 usec  
 TE 298.0 K  
 D1 0.10000000 sec  
 REFRET 0 sec  
 MEXX 0.01500000 sec  
 ----- CHANNEL f1 -----  
 NU01 1H  
 P1 7.50 usec  
 PL1 1.60 dB  
 SFO1 500.2235015 MHz  
 F2 - Processing parameters  
 SI 65536  
 SF 500.2200000 MHz  
 MDW EM  
 SSB 0 0.30 Hz  
 GB 0  
 PC 4.00

Z-restored spin-echo 13C spectrum with 1H decoupling

<sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz)



25: C<sub>31</sub>H<sub>42</sub>O<sub>6</sub>



Current Data Parameters  
 NAME DJT-V-027  
 EXPNO 6  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20150128  
 Time 8.37  
 INSTRUM cryo500  
 PROBHD 5 mm CPTCI 1H-  
 PULPROG SpinEchopg30gp.prd  
 TD 65536  
 SOLVENT CDCl3  
 NS 260  
 DS 16  
 SWH 30303.031 Hz  
 FIDRES 0.462388 Hz  
 AQ 1.0813440 sec  
 RG 6502  
 DW 16.500 usec  
 DE 6.00 usec  
 TE 298.0 K  
 D1 0.25000000 sec  
 d11 0.03000000 sec  
 D16 0.00020000 sec  
 d17 0.00019600 sec  
 MCREST 0 sec  
 MCWRK 0.01500000 sec  
 P2 33.10 usec

===== CHANNEL f1 =====  
 NUC1 13C  
 P1 16.55 usec  
 P11 500.00 usec  
 P12 2000.00 usec  
 PL0 120.00 dB  
 PL1 -1.00 dB  
 SFO1 125.7942548 MHz  
 SP1 2.70 dB  
 SP2 2.70 dB  
 SPNAM[1] Crp60,0.5,20.1  
 SPNAM[2] Crp60comp.4  
 SPOFF1 0 Hz  
 SPOFF2 0 Hz

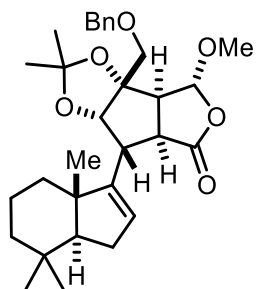
===== CHANNEL f2 =====  
 CPDPRG[2] waltz16  
 NUC2 1H  
 PCPD2 100.00 usec  
 PL2 1.60 dB  
 PL12 24.50 dB  
 SFO2 500.2225011 MHz

===== GRADIENT CHANNEL =====  
 GPNAM[1] SINE.100  
 GPNAM[2] SINE.100  
 GPX1 0 %  
 GPX2 0 %  
 GPY1 0 %  
 GPY2 0 %  
 GPZ1 30.00 %  
 GPZ2 50.00 %  
 p15 500.00 usec  
 p16 1000.00 usec

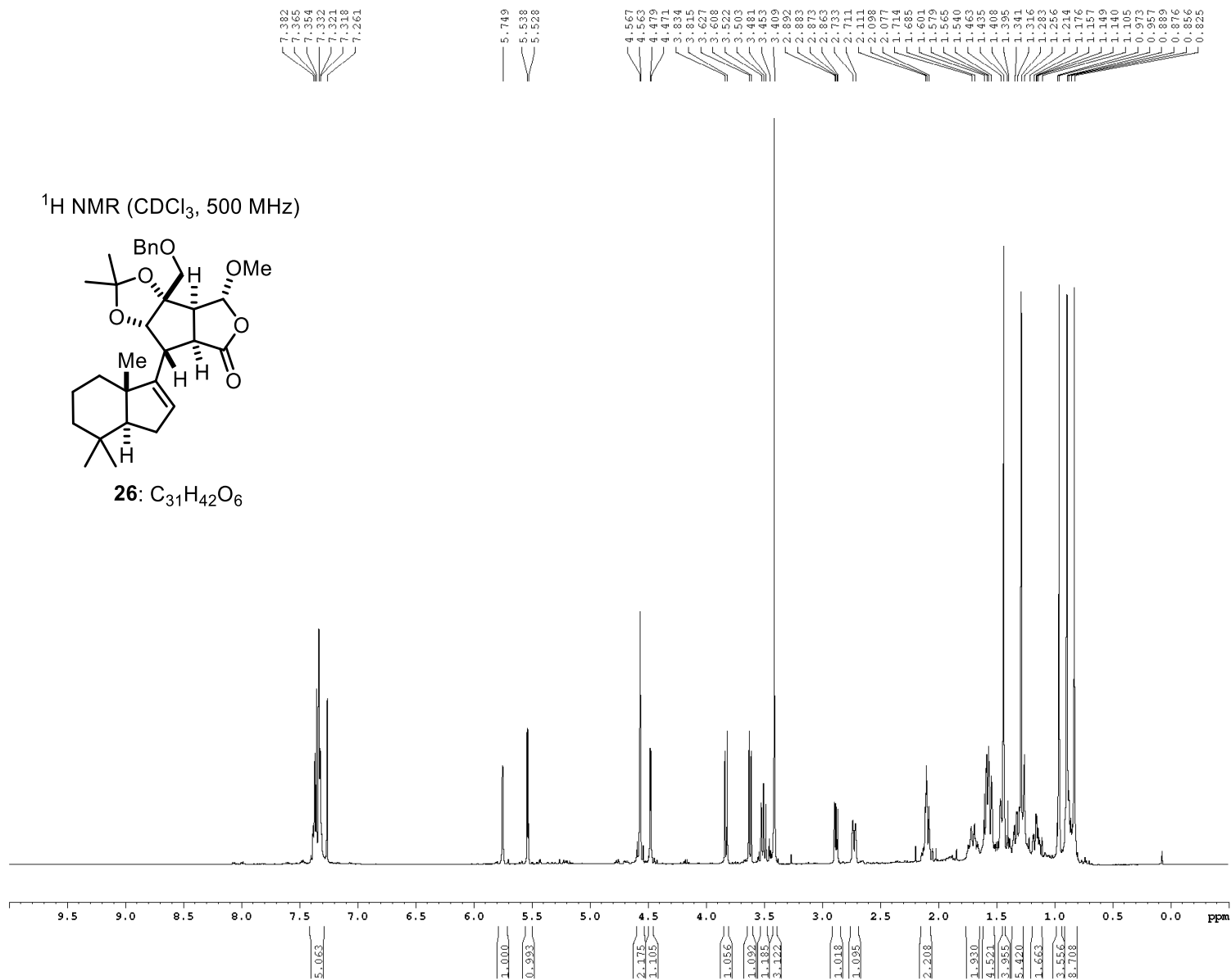
F2 - Processing parameters  
 SI 65536  
 SF 125.7804076 MHz  
 WDW no  
 SSB 0  
 LB 0 Hz  
 GB 0  
 PC 2.00

<sup>1</sup>H spectrum

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)



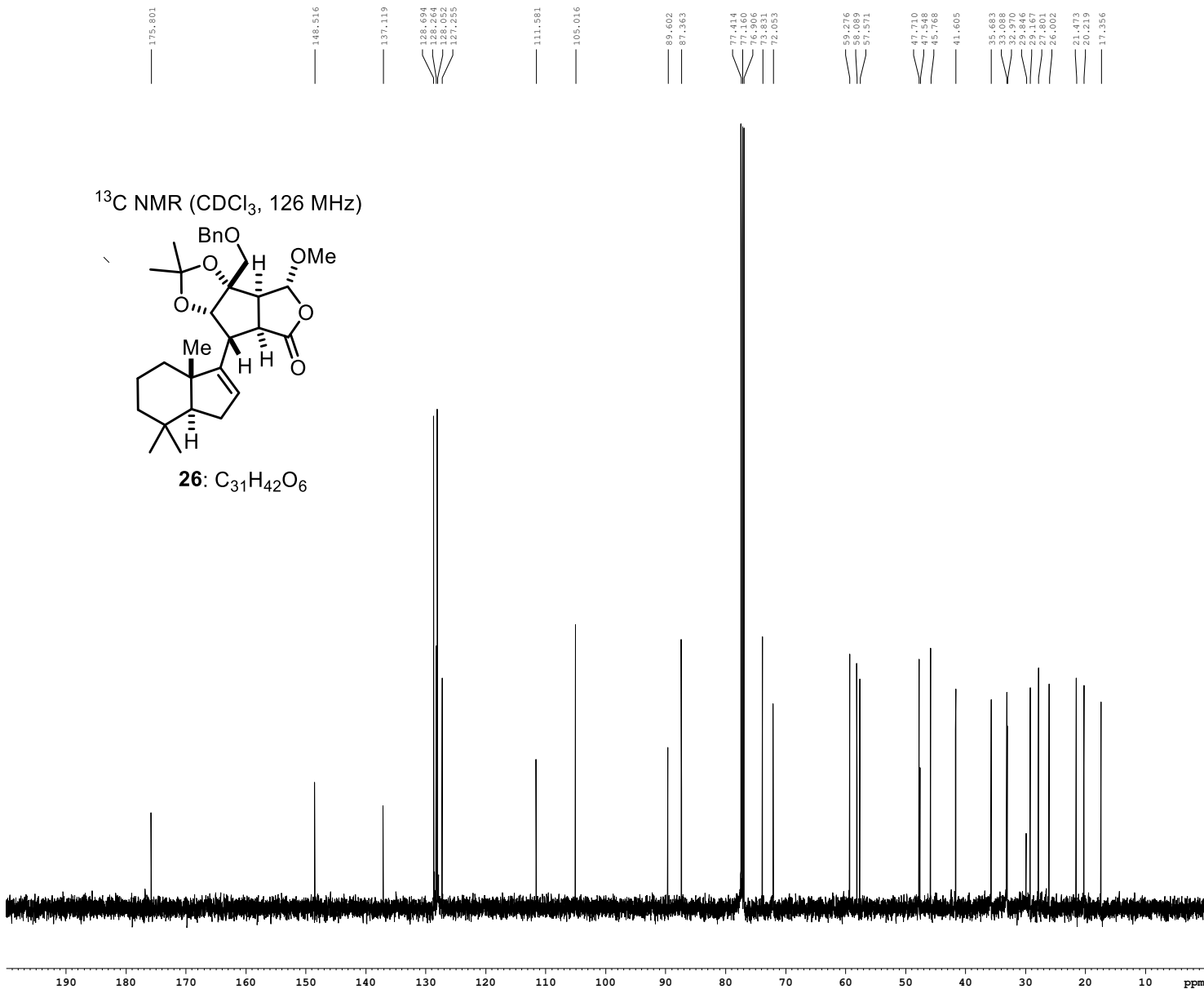
26: C<sub>31</sub>H<sub>42</sub>O<sub>6</sub>



```

Current Data Parameters
NAME          DJT-V-103
EXPNO         2
PROCNO        1
F2 - Acquisition Parameters
Date_         20150328
Time          14.19
INSTRUM       cryo500
PROBHD        5 mm CPTCI 1H-
PULPROG       zg30
TD            16022
SOLVENT       CDCl3
NS            60
DS            0
SWH           8012.820 Hz
FIDRES       0.500114 Hz
AQ           0.9997728 sec
RG            5
AQ           62.400 usec
DE           6.00 usec
TE           298.0 K
D1           0.10000000 sec
REFRESH      0 sec
SFO1         0.01500000 sec
----- CHANNEL f1 -----
NUC1          1H
P1            7.50 usec
PL1           1.60 dB
SFO1         500.2235015 MHz
F2 - Processing parameters
SI            65536
SF           500.2200004 MHz
WDW           EM
SSB           0
GB           0.30 Hz
CB            0
PC            4.00
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
NAME          DJT-V-056
EXPNO         6
PROCNO        1

F2 - Acquisition Parameters
Date_         20150225
Time          16.33
INSTRUM       cryo500
PROBHD        5 mm CPTCI 1H-
PULPROG       SpinEchopg30gp.prd
TD            65536
SOLVENT       CDCl3
NS            135
DS            0
SWH           30303.031 Hz
FIDRES        0.462388 Hz
AQ            1.0813440 sec
RG            3251
DW            16.500 usec
DE            6.00 usec
TE            298.0 K
D1            0.25000000 sec
d11           0.03000000 sec
D16           0.00020000 sec
d17           0.00019600 sec
MCREST        0 sec
MCWRK         0.01500000 sec
P2            33.10 usec

===== CHANNEL f1 =====
NUC1           13C
P1             16.55 usec
P11            500.00 usec
P12            2000.00 usec
PL0            120.00 dB
PL1            -1.00 dB
SFO1           125.7942548 MHz
SP1            2.70 dB
SP2            2.70 dB
SPNAM[1]       Crp60,0.5,20.1
SPNAM[2]       Crp60comp.4
SPOFF1         0 Hz
SPOFF2         0 Hz

===== CHANNEL f2 =====
CPDPRG[2]      waltz16
NUC2           1H
PCPD2          100.00 usec
PL2            1.60 dB
PL12           24.50 dB
SFO2           500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM[1]       SINE.100
GPNAM[2]       SINE.100
GPX1           0 %
GPX2           0 %
GPY1           0 %
GPY2           0 %
GPZ1           30.00 %
GPZ2           50.00 %
p15            500.00 usec
p16            1000.00 usec

F2 - Processing parameters
SI             65536
SF             125.7804085 MHz
WDW            EM
SSB            0
LB             1.00 Hz
GB             0
PC             2.00
    
```

**<sup>1</sup>H spectrum**

```

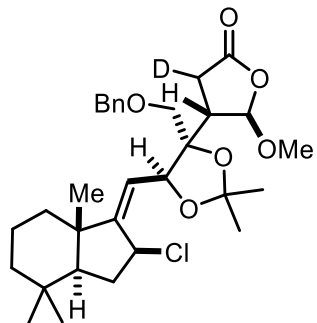
Current Data Parameters
NAME      DJT-V-263
EXPNO    19
PROCNO   1

F2 - Acquisition Parameters
Date_    20151009
Time     15:47
INSTRUM  av600
PROBHD   5 mm TBI 1H/13
PULPROG  zg30
TD       98074
SOLVENT  CDCl3
NS       89
DS       0
SWH      9615.385 Hz
FIDRES   0.098882 Hz
AQ       5.0998478 sec
RG       1030
LW       52.000 usec
DE       14.54 usec
TE       296.2 K
D1       0.10000000 sec
TDO      1

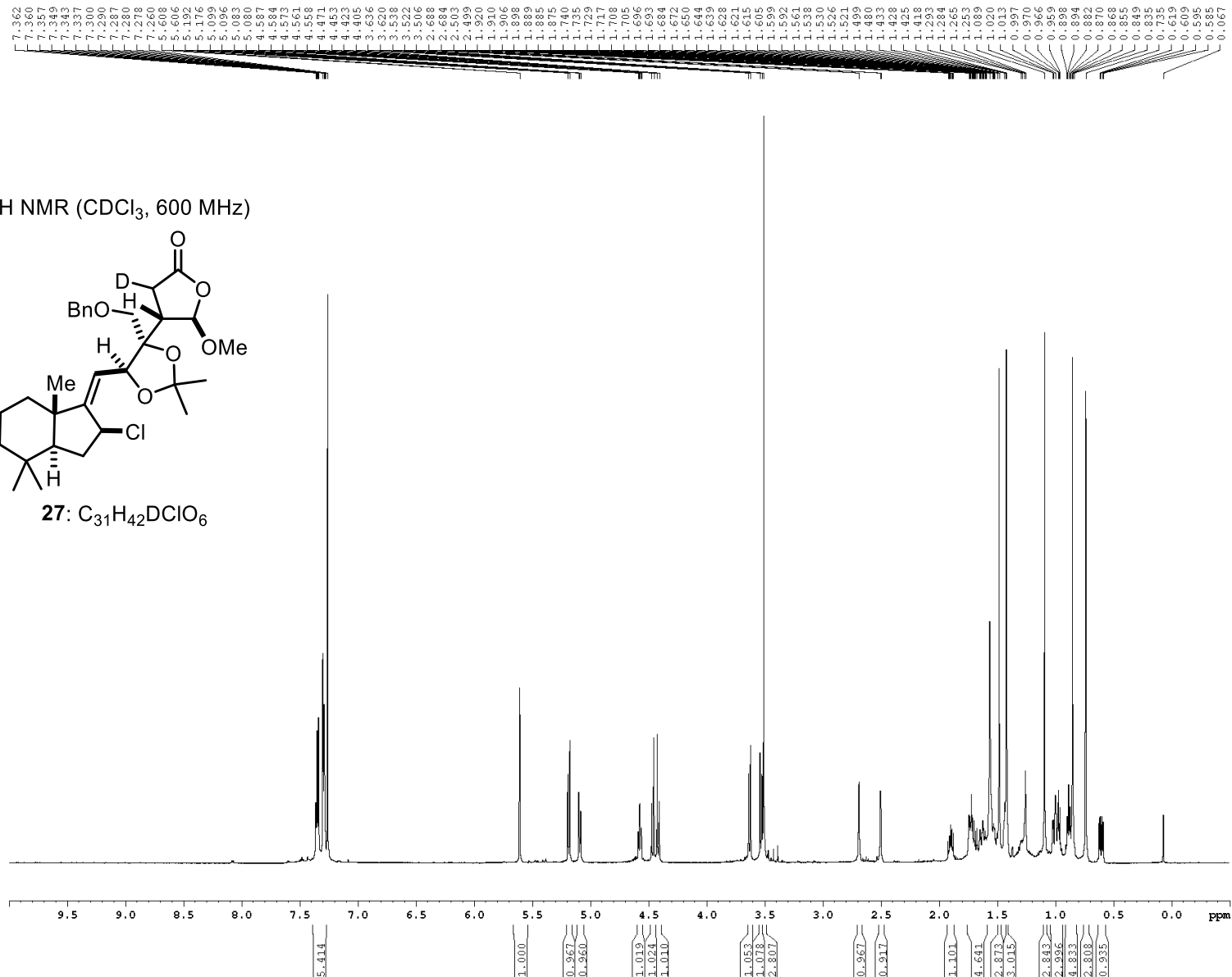
===== CHANNEL f1 =====
SFO1     600.1342000 MHz
NUC1     1H
P1       8.00 usec
PLW1     24.00000000 W

F2 - Processing parameters
SI       65536
SF       600.1300357 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00
    
```

**<sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz)**

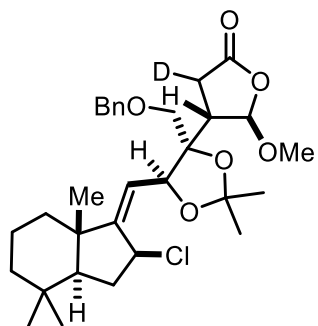


**27: C<sub>31</sub>H<sub>42</sub>DClO<sub>6</sub>**

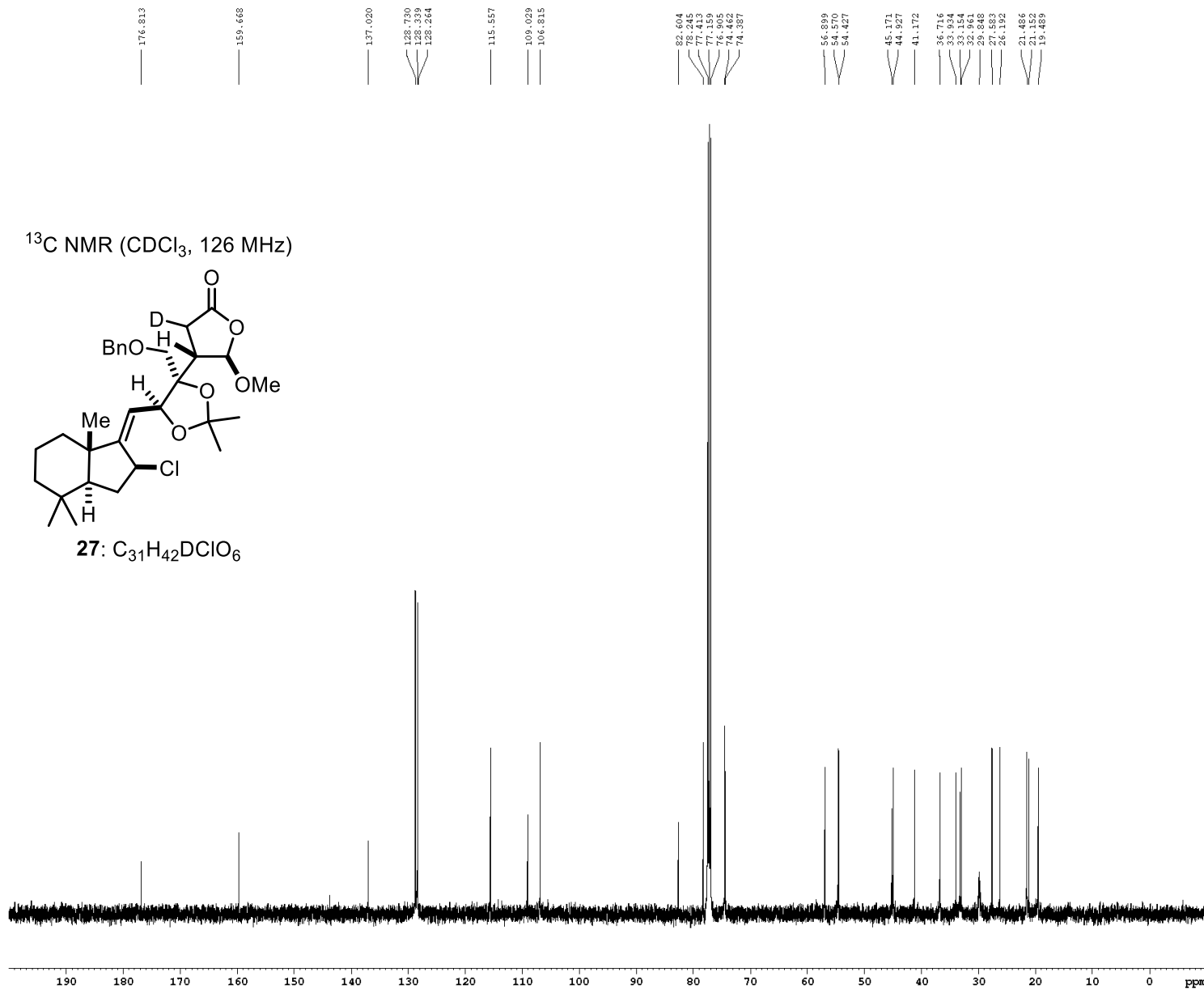


Z-restored spin-echo <sup>13</sup>C spectrum with <sup>1</sup>H decoupling

<sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz)



27: C<sub>31</sub>H<sub>42</sub>DClO<sub>6</sub>



```

Current Data Parameters
NAME          DJT-V-263
EXPNO         21
PROCNO        1

F2 - Acquisition Parameters
Date_         20151010
Time          9.52
INSTRUM       cryo500
PROBHD        5 mm CPTCI 1H-
PULPROG       SpinEchopg30gp.prd
TD            65536
SOLVENT       CDCl3
NS            842
DS            0
SWH           30303.031 Hz
FIDRES        0.462388 Hz
AQ            1.0813440 sec
RG            5792.6
DW            16.500 usec
DE            6.00 usec
TE            298.0 K
D1            0.25000000 sec
d11           0.03000000 sec
D16           0.00020000 sec
d17           0.00019600 sec
MCREST        0 sec
MCWRK         0.01500000 sec
F2            33.10 usec

===== CHANNEL f1 =====
NUC1           13C
P1            16.55 usec
P11           500.00 usec
P12           2000.00 usec
PL0           120.00 dB
PL1           -1.00 dB
SFO1          125.7942548 MHz
SP1           2.70 dB
SP2           2.70 dB
SPNAM[1]      Crp60,0.5,20.1
SPNAM[2]      Crp60comp.4
SPOFF1        0 Hz
SPOFF2        0 Hz

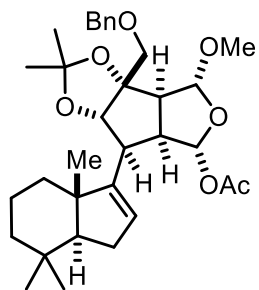
===== CHANNEL f2 =====
CPDPRG[2]     waltz16
NUC2           1H
PCPD2         100.00 usec
PL2           1.60 dB
PL12          24.50 dB
SFO2          500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM[1]      SINE.100
GPNAM[2]      SINE.100
GPX1          0 %
GPX2          0 %
GPY1          0 %
GPY2          0 %
GPZ1          30.00 %
GPZ2          50.00 %
p15           500.00 usec
p16           1000.00 usec

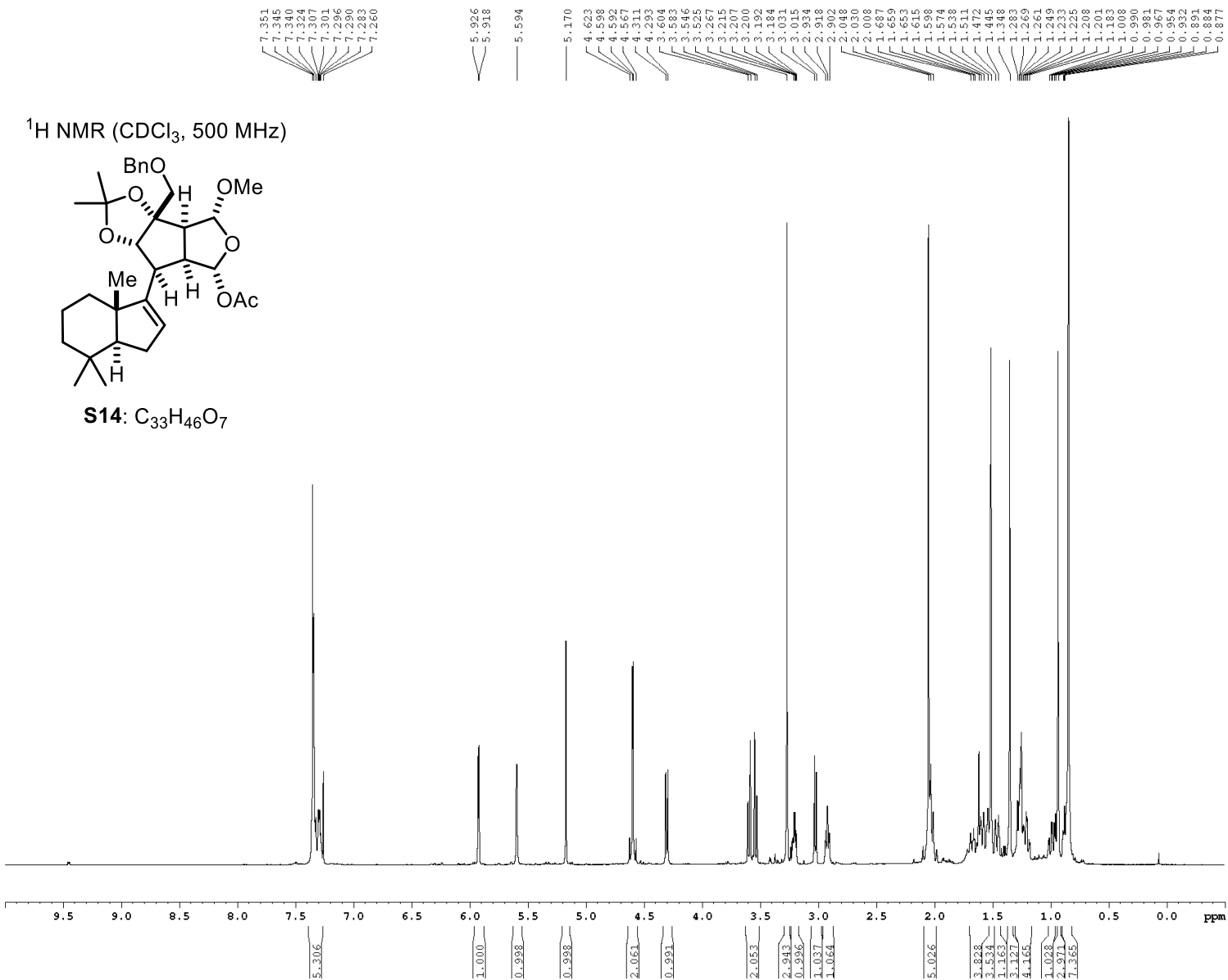
F2 - Processing parameters
SI            65536
SF            125.7804080 MHz
WDW           EM
SSB           0
LB            1.00 Hz
GB            0
PC            2.00
    
```

<sup>1</sup>H spectrum

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)



S14: C<sub>33</sub>H<sub>46</sub>O<sub>7</sub>



```

Current Data Parameters
NAME      DJT-V-268
EXPNO     1
PROCNO    1

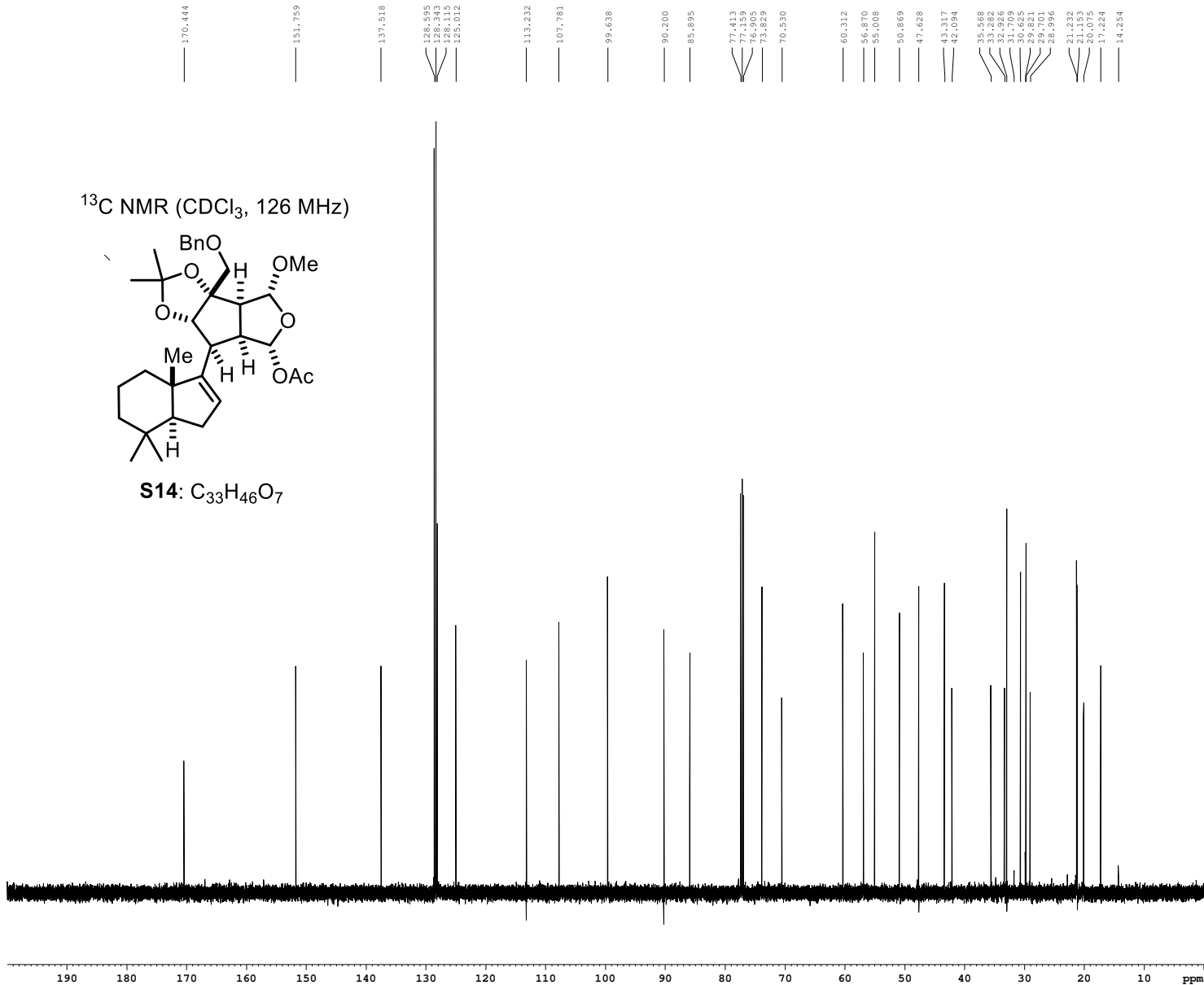
F2 - Acquisition Parameters
Date_     20150821
Time      11.02
INSTRUM   cryo500
PROBHD    5 mm CPTCI 1H-
PULPROG   zgpg30
TD         81728
SOLVENT   CDCl3
NS         20
DS         0
SWH        8012.820 Hz
FIDRES     0.096043 Hz
AQ         5.0996273 sec
RG         6.3
RW         62.400 usec
DE         6.00 usec
TE         298.0 K
D1         0.10000000 sec
MORPH     0 sec
MEMPROG   0.01500000 sec

***** CHANNEL f1 *****
NUC1       1H
P1         7.50 usec
PL1        1.60 dB
SFO1       500.2235015 MHz

F2 - Processing parameters
SI         65536
SF         500.200313 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         4.00
    
```



Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
NAME          DJT-V-268
EXPNO         2
PROCNO        1

F2 - Acquisition Parameters
Date_         20150821
Time          11.07
INSTRUM       cryo500
PROBHD        5 mm CPTCI 1H-
PULPROG       SpinEchopg30gp.prd
TD            65536
SOLVENT       CDCl3
NS            171
DS            0
SWH           30303.031 Hz
FIDRES        0.462388 Hz
AQ            1.0813440 sec
RG            5792.6
DW            16.500 usec
DE            6.00 usec
TE            298.0 K
D1            0.25000000 sec
d11           0.03000000 sec
D16           0.00020000 sec
d17           0.00019600 sec
MCREST        0 sec
MCWRK         0.01500000 sec
P2            33.10 usec

===== CHANNEL f1 =====
NUC1           13C
P1             16.55 usec
P11            500.00 usec
P12            2000.00 usec
PL0            120.00 dB
PL1            -1.00 dB
SFO1           125.7942548 MHz
SP1            2.70 dB
SP2            2.70 dB
SPNAM[1]       Crp60,0.5,20.1
SPNAM[2]       Crp60comp.4
SPOFF1         0 Hz
SPOFF2         0 Hz

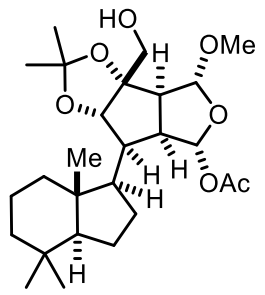
===== CHANNEL f2 =====
CPDPRG[2]     waltz16
NUC2           1H
PCPD2         100.00 usec
PL2            1.60 dB
PL12           24.50 dB
SFO2           500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM[1]       SINE.100
GPNAM[2]       SINE.100
GPX1           0 %
GPX2           0 %
GPY1           0 %
GPY2           0 %
GPZ1           30.00 %
GPZ2           50.00 %
p15            500.00 usec
p16            1000.00 usec

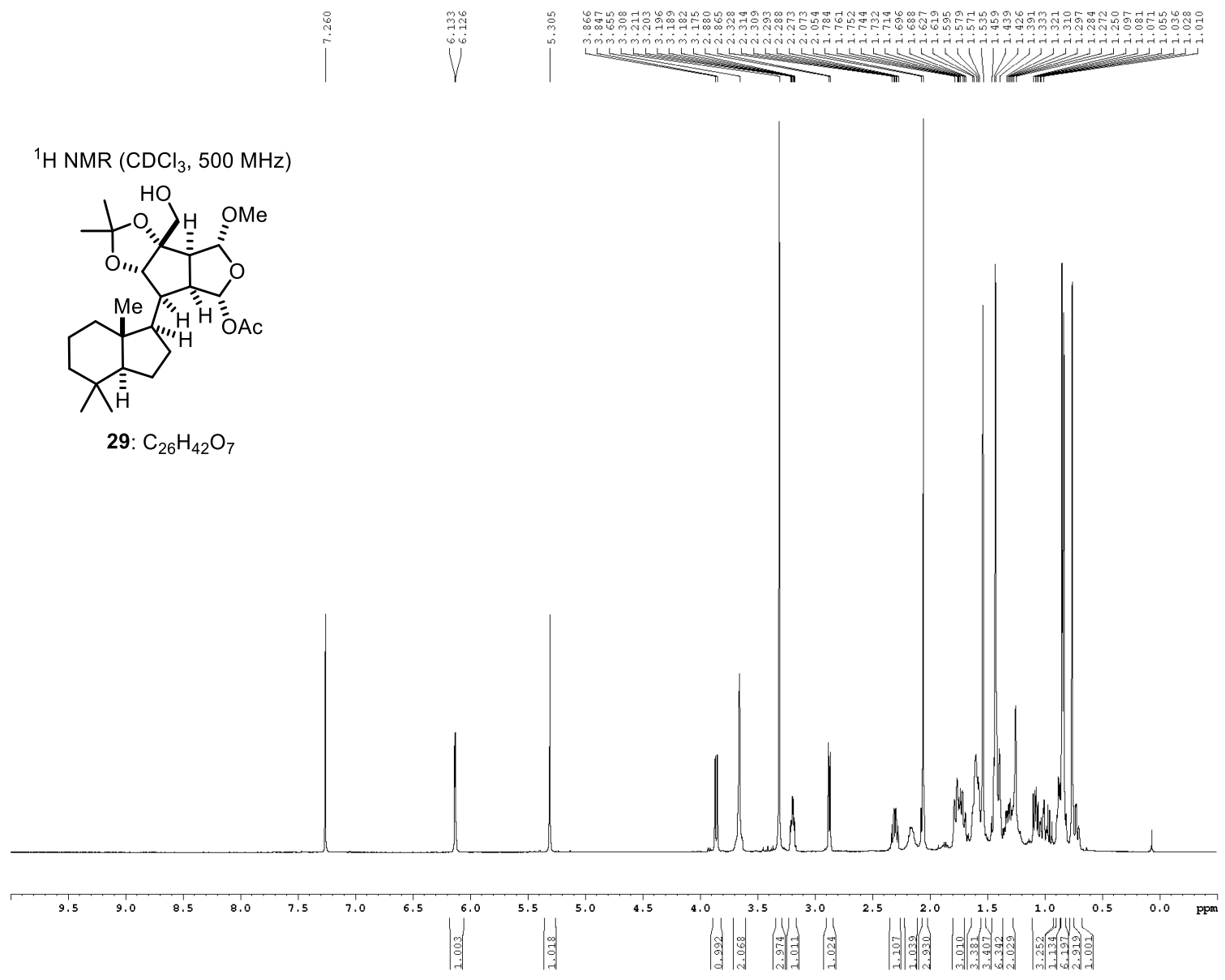
F2 - Processing parameters
SI            65536
SF            125.7804108 MHz
WDW           no
SSB           0
LB            0 Hz
GB            0
PC            2.00
    
```

<sup>1</sup>H spectrum

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)



29: C<sub>26</sub>H<sub>42</sub>O<sub>7</sub>



```

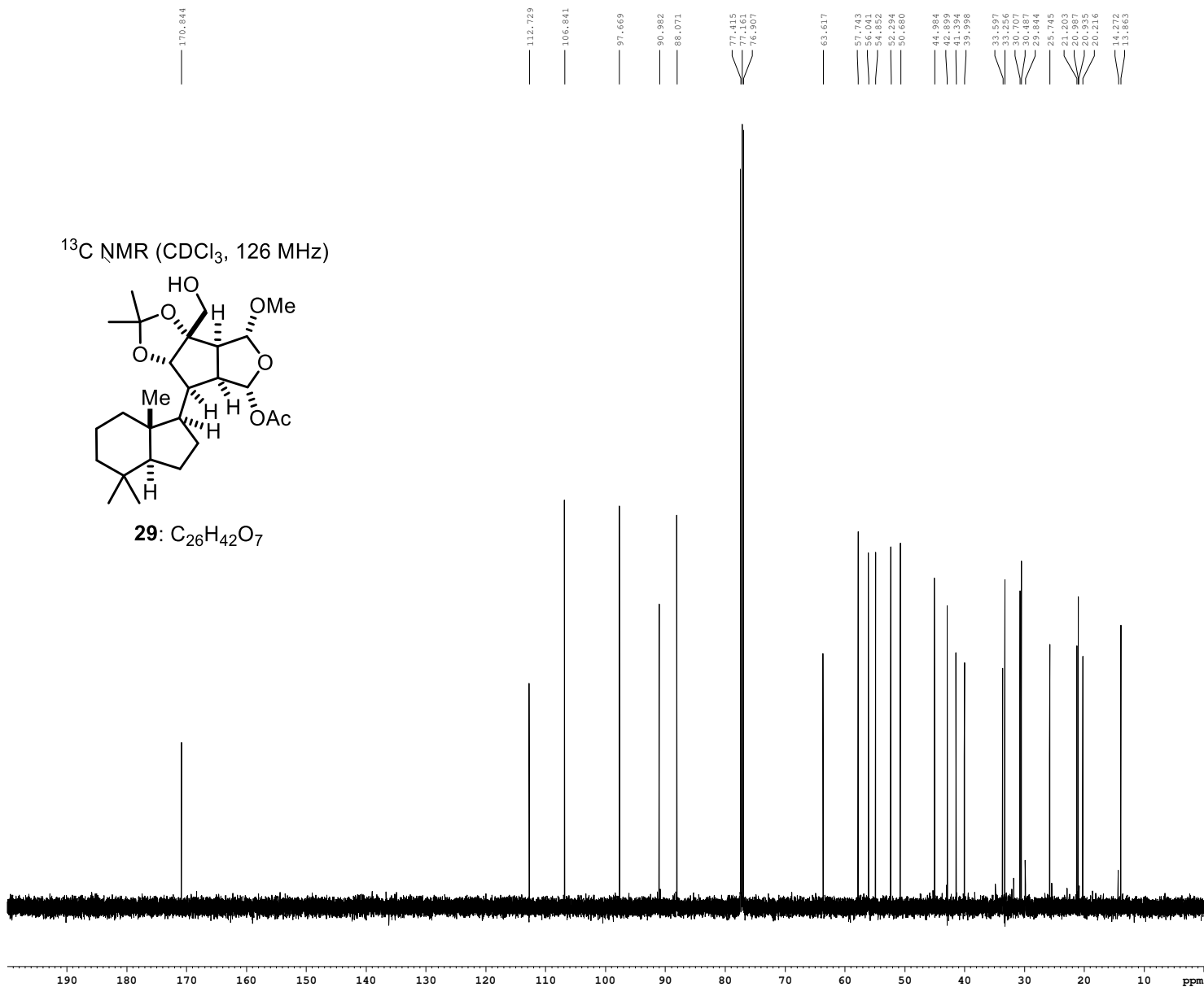
Current Data Parameters
NAME      DJ-V-270
EXPNO     5
PROCNO    1

F2 - Acquisition Parameters
Date_     20150824
Time      13.07
INSTRUM   crys500
PROBHD    5 mm CPTCL 1H-
PULPROG   zg30
TD         61728
SOLVENT   CDCl3
NS         20
DS         0
SHH        8012.820 Hz
FIDRES     0.098043 Hz
AQ         5.0998273 sec
RG         5
DH         62.400 usec
DE         6.00 usec
TE         298.0 K
D1         0.10000000 sec
d11        0 sec
MORST      0.01500000 sec

===== CHANNEL f1 =====
NUC1       1H
P2         7.50 usec
PL1        1.60 dB
SFO1       500.225015 MHz

F2 - Processing parameters
SI         65536
SF         500.2200212 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         4.00
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling



```

Current Data Parameters
NAME          DJT-V-270
EXPNO         6
PROCNO        1

F2 - Acquisition Parameters
Date_         20150824
Time          13.11
INSTRUM       cryo500
PROBHD        5 mm CPTCI 1H-
PULPROG       SpinEchopg30gp.prd
TD            65536
SOLVENT       CDCl3
NS            498
DS            0
SWH           30303.031 Hz
FIDRES        0.462388 Hz
AQ            1.0813440 sec
RG            4096
DW            16.500 usec
DE            6.00 usec
TE            298.0 K
D1            0.25000000 sec
d11           0.03000000 sec
D16           0.00020000 sec
d17           0.00019600 sec
MCREST        0 sec
MCWRK         0.01500000 sec
P2            33.10 usec

===== CHANNEL f1 =====
NUC1           13C
P1             16.55 usec
P11            500.00 usec
P12            2000.00 usec
PL0            120.00 dB
PL1            -1.00 dB
SFO1           125.7942548 MHz
SP1             2.70 dB
SP2             2.70 dB
SPNAM[1]       Crp60,0.5,20.1
SPNAM[2]       Crp60comp.4
SPOFF1         0 Hz
SPOFF2         0 Hz

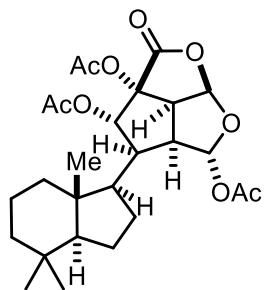
===== CHANNEL f2 =====
CPDPRG[2]      waltz16
NUC2            1H
PCPD2          100.00 usec
PL2             1.60 dB
PL12           24.50 dB
SFO2           500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM[1]       SINE.100
GPNAM[2]       SINE.100
GPX1           0 %
GPX2           0 %
GPY1           0 %
GPY2           0 %
GPZ1           30.00 %
GPZ2           50.00 %
p15            500.00 usec
p16            1000.00 usec

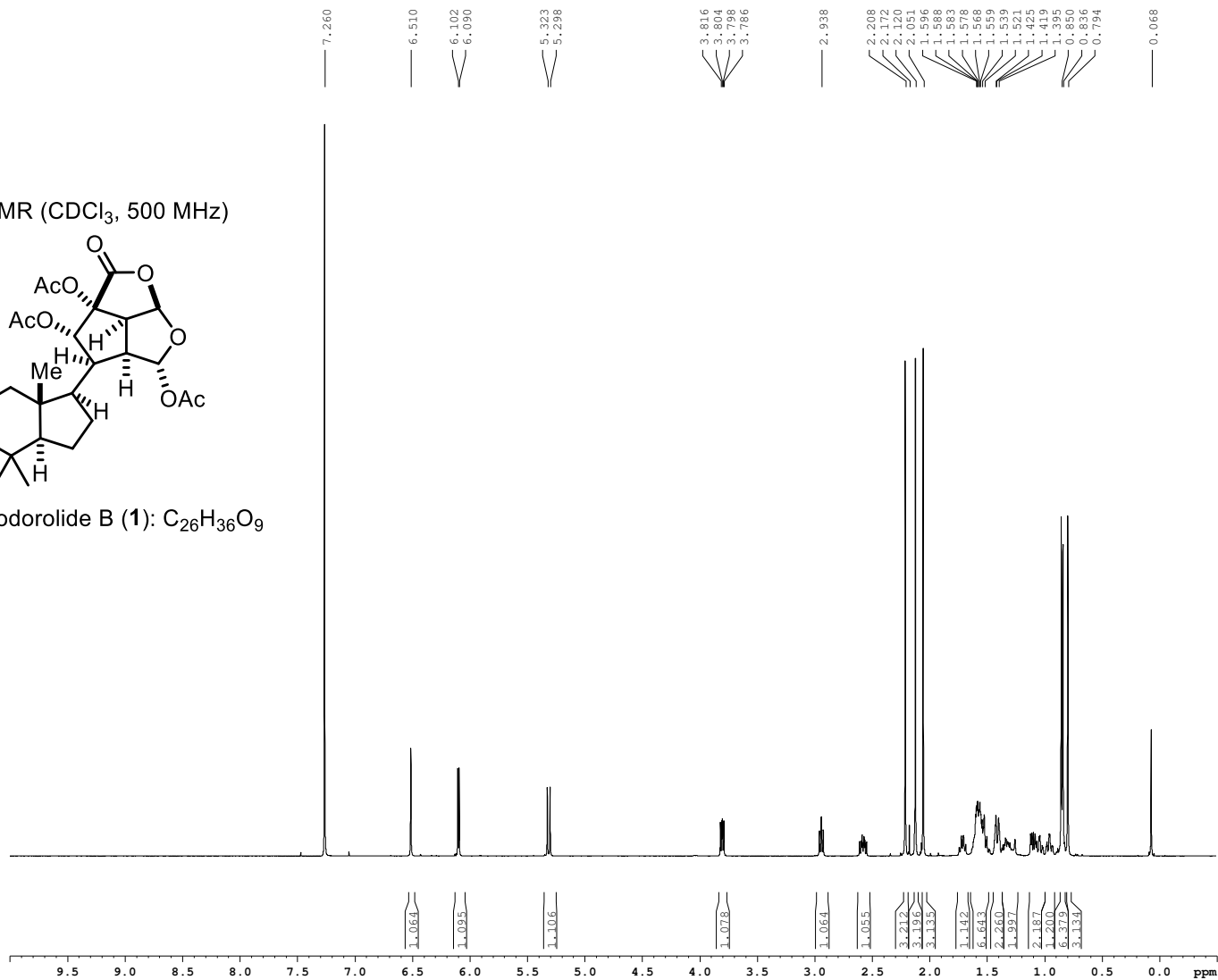
F2 - Processing parameters
SI            65536
SF            125.7804080 MHz
WDW           no
SSB           0
LB            0 Hz
GB            0
PC            2.00
    
```

**<sup>1</sup>H spectrum**

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)



chromodorolide B (1): C<sub>26</sub>H<sub>36</sub>O<sub>9</sub>



```

Current Data Parameters
NAME          DJT-V-272
EXPNO        5
PROCNO       1

F2 - Acquisition Parameters
Date_        20150907
Time         9.43
INSTRUM      cryo500
PROBHD       5 mm CPTCI 1H-
PULPROG      zg30
TD           81728
SOLVENT      CDCl3
NS           140
DS           0
SWH          8012.820 Hz
FIDRES       0.098043 Hz
AQ           5.0998273 sec
RG           8
DW           62.400 usec
DE           6.00 usec
TE           298.0 K
D1           0.10000000 sec
MCREST       0 sec
MCWRK        0.01500000 sec

===== CHANNEL f1 =====
NUC1          1H
P1           7.50 usec
PL1          1.60 dB
SFO1         500.2235015 MHz

F2 - Processing parameters
SI           65536
SF           500.2200312 MHz
WDW          EM
SSB          0
LB           0.30 Hz
GB           0
PC           4.00
    
```

Z-restored spin-echo 13C spectrum with 1H decoupling

170.391  
169.329  
169.242

103.505  
97.872

81.466  
77.412  
77.188  
74.052

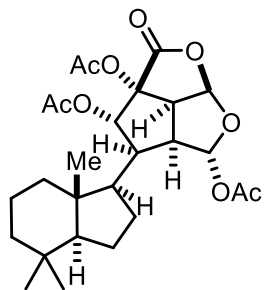
57.086

50.588  
49.744  
48.122  
45.729  
43.985  
41.054  
39.250

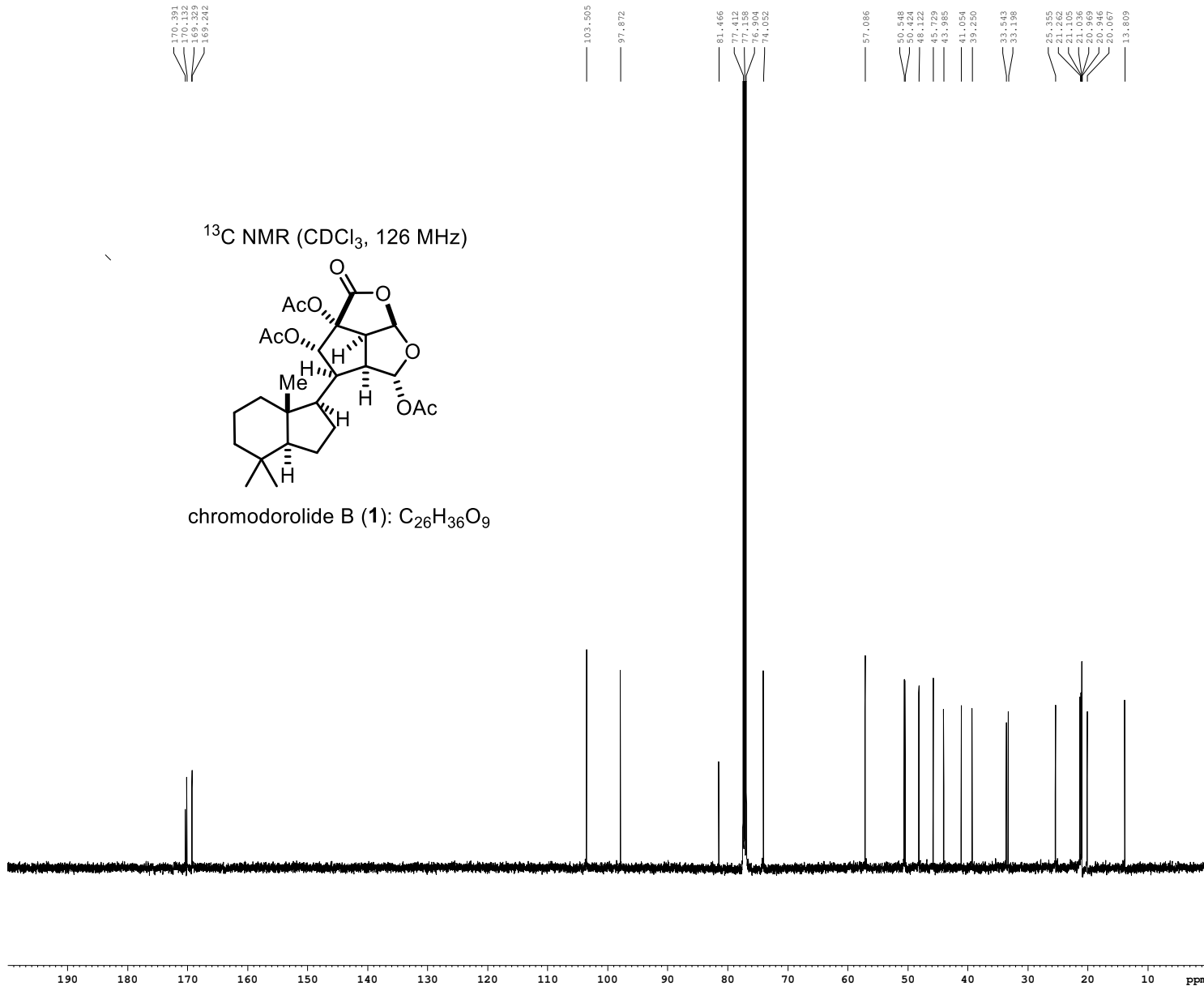
33.543  
31.198

25.355  
21.262  
21.105  
20.989  
20.946  
20.067  
13.809

<sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz)



chromodorolide B (1): C<sub>26</sub>H<sub>36</sub>O<sub>9</sub>



```

Current Data Parameters
NAME          DJT-V-272
EXPNO         10
PROCNO        1

F2 - Acquisition Parameters
Date_         20151012
Time          7.21
INSTRUM       cryo500
PROBHD        5 mm CPTCI 1H-
PULPROG       SpinEchopg30gp.prd
TD            65536
SOLVENT       CDCl3
NS            2116
DS            0
SWH           30303.031 Hz
FIDRES        0.462388 Hz
AQ            1.0813440 sec
RG            2298.8
DW            16.500 usec
DE            6.00 usec
TE            298.0 K
D1            0.25000000 sec
d11           0.03000000 sec
D16           0.00020000 sec
d17           0.00019600 sec
MCREST        0 sec
MCWRK         0.01500000 sec
P2            33.10 usec

===== CHANNEL f1 =====
NUC1           13C
P1            16.55 usec
P11           500.00 usec
P12           2000.00 usec
PL0           120.00 dB
PL1           -1.00 dB
SFO1          125.7942548 MHz
SP1           2.70 dB
SP2           2.70 dB
SPNAM[1]      Crp60,0.5,20.1
SPNAM[2]      Crp60comp.4
SPOFF1        0 Hz
SPOFF2        0 Hz

===== CHANNEL f2 =====
CPDPRG[2]     waltz16
NUC2           1H
PCPD2         100.00 usec
PL2           1.60 dB
PL12          24.50 dB
SFO2          500.2225011 MHz

===== GRADIENT CHANNEL =====
GPNAM[1]      SINE.100
GPNAM[2]      SINE.100
GPX1          0 %
GPX2          0 %
GPY1          0 %
GPY2          0 %
GPZ1          30.00 %
GPZ2          50.00 %
p15           500.00 usec
p16           1000.00 usec

F2 - Processing parameters
SI            65536
SF            125.7804080 MHz
WDW           EM
SSB           0
LB            1.00 Hz
GB            0
PC            2.00
    
```