

Supplemental Materials

Molecular Biology of the Cell

Senoo et al.

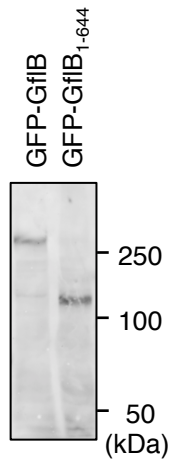


Figure S2. Whole-cell lysates analyzed using immunoblotting with anti-GFP antibodies to detect the indicated PTEN-GFP fusions.

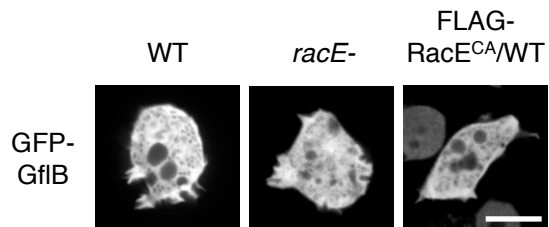


Figure S3. GFP-Gf1B was examined in WT cells, *racE*⁻ cells, and WT cells expressing constitutively active RacE_{G20V} using fluorescence microscopy. Bar, 10 μ m.

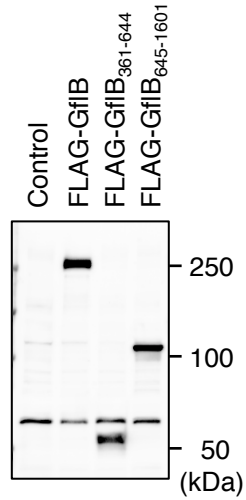


Figure S4. Whole-cell lysates analyzed using immunoblotting with anti-FLAG antibodies.

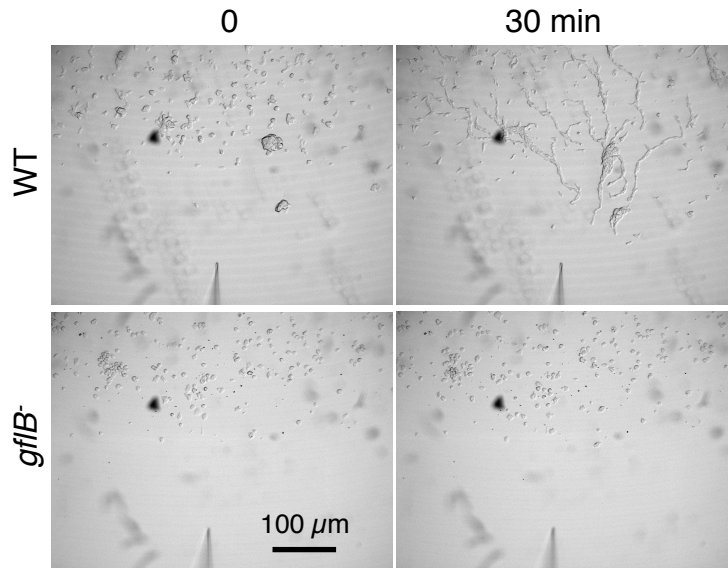


Figure S5. After 8 hours starvation, WT and *gflB*⁻ cells were placed in a chemoattractant gradient generated by a micropipette that released cAMP and observed for 20 min using phase contrast microscopy.

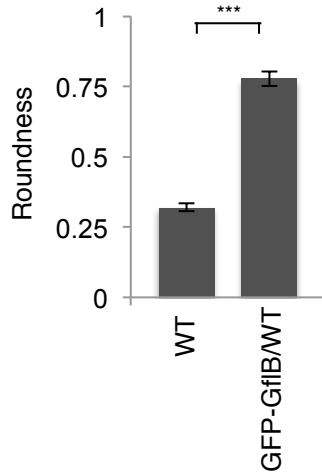


Figure S6. Roundness was determined by calculating the ratio between the short axis (A_s) and long axis (A_l) of cells (A_s/A_l) in WT cells in the presence or absence of GFP-GfIB expression. Values represent mean \pm SEM. At least 30 cells were analyzed for each group.

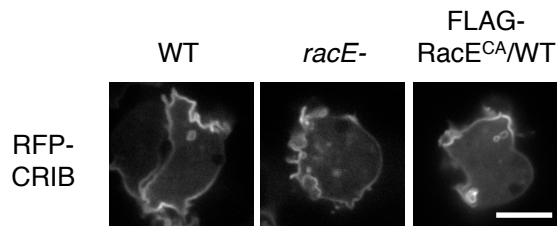


Figure S7. CRIB-RFP was examined in WT cells, *racE*⁻ cells, and WT cells expressing constitutively active RacE_{G20V} using fluorescence microscopy. Bar, 10 μ m.

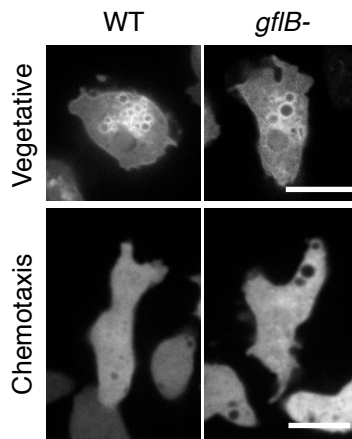


Figure S8. The PS biosensor GFP-LactC2 was observed in WT and *gflB*⁻ cells during growth (vegetative) and after differentiation (chemotaxis).

Table S1. Plasmids used in this study

Plasmids	Primers used	Drug for selection	References
<i>Dictyostelium</i> expression			
pJK1-GFP: pIS1		G418 (20 µg/ml)	Zhang et.al. (2011)
pJK1-GFP- <i>racIA</i>	10 11	G418 (20 µg/ml)	This Study
pJK1-GFP- <i>racB</i>	12 13	G418 (20 µg/ml)	This Study
pJK1-GFP- <i>racE</i>		G418 (20 µg/ml)	Wang et.al. (2013)
pJK1-GFP- <i>racE</i> (G20V)		G418 (20 µg/ml)	Wang et.al. (2013)
pJK1-GFP- <i>racE</i> (T25N)		G418 (20 µg/ml)	Wang et.al. (2013)
pDRH-FLAG- <i>RasC</i>		Hygromycin (50 µg/ml)	Gift from Dr. Devreotes (JHMI)
pDRH-FLAG- <i>RasG</i>		Hygromycin (50 µg/ml)	Gift from Dr. Devreotes (JHMI)
pDRH- <i>LimE</i> Δ <i>coli</i> -mRFP		Hygromycin (50 µg/ml)	Gift from Dr. Devreotes (JHMI)
pDM323-RBD(<i>Raf</i>)-GFP		G418 (20 µg/ml)	Xiong et.al. (2010)
pDRH-PH <i>crac</i> -RFP		Hygromycin (50 µg/ml)	Gift from Dr. Devreotes (JHMI)
pTX-GFP		G418 (20 µg/ml)	Levi et.al. (2000)
pTX-GFP-GfIB	14 15	G418 (20 µg/ml)	This Study
pTX-GFP-GfIB 1-644	14 16	G418 (20 µg/ml)	This Study
pTX-GFP-GfIB 361-644	16 17	G418 (20 µg/ml)	This Study
pTX-GFP-GfIB 645-1601	15 18	G418 (20 µg/ml)	This Study
pTX-FLAG		G418 (20 µg/ml)	Levi et.al. (2000)
pTX-FLAG-GfIB	14 15	G418 (20 µg/ml)	This Study
pTX-FLAG-GfIB 361-644	16 17	G418 (20 µg/ml)	This Study
pTX-FLAG-GfIB 645-1601	15 18	G418 (20 µg/ml)	This Study
pDRH-CRIB-RFP	19 20	Hygromycin (50 µg/ml)	This Study
<i>Bacterial</i> expression			
pGEX 4T-1			Amersham
pGEX-Byr2-RBD			Kae et.al. (2004)

Table S2. Primers used in this study

Primers for gene disruption in Dictyostelium cells

1	A15P	CCAACCCAAGTTTTTTTAAACC
2	gflB-2	GACTCGAGTTATTCGGCATTGTTGAAGGAAC
3	gflB-3	GTGTACCAAATTTCTCTATACTTCG
4	gflB-4	CACCACCACTACCTCAACTACAACC
5	gflB-5	GAGCGGCCGCGTTGTAGTACTACTAGTTAC
6	gflB salI 1-19	GAGTCGACATGACAGATTTAAATTCAG
7	gflB smaI 800-781	GACCCGGGAATTGATTAGCCTGTAAAAC
8	gflB smaI 4116-4134	GACCCGGGAATCATCAGAATTGGAGAC
9	gflB NotI 4875-4856	GAGCGGCCGCGTTGTAGTACTACTAGTTAC

Primers for expression plasmids in Dictyostelium cells

10	rac1A-1	GGAAGATCTTCCATGCAAGCAATTAATG
11	rac1A-2	GGAAGATCTTCCTTATAAAATGTTGC
12	racB-1	GGAAGATCTTCCATGCAATCAATTAATTGG
13	racB-2	GGAAGATCTTCCTTATAAAATTGAACATTTTG
14	gflB-Start	CCCGAGCTCATGACAGATTTAAATTC
15	gflB-Stop	CCCCTCGAGTTATTCGGCATTGTTGAAGGAAC
16	gflB 1932-1909	GACTCGAGTTAATATGGTGAATCTAAAATTTGTTG
17	gflB 1081-1101	GAGAGCTCTTACAAGTAGTTTCCAATGCC
18	gflB 2042-2066	GAGAGCTCGGAATTTATGGTGTAGATTAACAC
19	CRIB 943	CCCGAGCTCACTACATCACCACCATCC
20	CRIB 1233	CCCCTCGAGTTAATGGAAATCTAAAACATC