

Supplemental Materials

Molecular Biology of the Cell

Senoo et al.

A

1	MTDLNSESFSALNFWKSVEKKNISTNFQGTKVVAPSCKINSFPPLLAPPAPPPPTEQEINIGSGNSTFISSNNNSNNNNNNNSNNNNNN	90
91	NLNNSNNNNNNLNSNNNNNNNNNNNNNNNNNNFLRQDSSTQKEWDEQNVTAEFGFWKQAVQLQKETERYNARRNARQTIDLNTI	180
181	LRKSTSSDLLIKPPVESPLTPVGQDDEGEQQQQQQQQQKQSSPSTSNDTDTETTAAAVTTTTTTTTTSTTTTETVLQANOLEIK	270
271	YGETIAVVDSGTTPRDYRSRSISCEIPKINGVITSPQRVTTTTTPSTGGVVVADEESESDSSEESEDEYTDDESETE	360
361	LQVVSNATPRRSRSDFTPTIVESPPLTSVNSNDNTSGTVVAPIDLNSSTGGNTSQOPQPSSQOKPDQASENTAVAASSISATTNVTS	450
451	AASTTVAPDSIINTKDVTVVSSLTTSATSSTQSIPAPPSPSQORAAQSISTSSVTPAATKPTKDADKKDPAKKSIGATL	540
541	TRTVTKTFIRDSENNKVPVTGTPSVSSSTSISSTGKDKVQLSKEEKDRIKEKSACKKEKDEKKQQKTNKAUTKNNSSTDVKGK	630
631	FVSPQQQIILDPSYRIYGVRLTLVLSNDGDLPAITQTTIVLNSNKLDVNSVFGVAENEPAVREIRRSRSLERIDF11GDPRVVAGL	720
721	LILFFAELPOPLFNNSKFFGDLVBEINDITNPQVKLNDLKQLINSLSQLRRSSLQILVTFFTSKYINGNSVTTRAIAIOSIAQSFGPQFFRG	810
811	TSSSDSDIOVGIELKLIDNYVFLFEKTNEPDVKYKNIDGKMIISEGSIDKLIDKATDQYYPYNEKYFSLTFITHLFFFQPHELADKL	900
901	ITLYRENLDTLETKKWKKHRRSKASFINEAVKLWVDYCYKEMREDKELSKKILKGFPHLEAQLASRLSHRTTINDFLKLKPVRVHSRTR	990
991	SASFSDTLLSTGGIGSTSGGIGGVNNCLLSAMEIAEQCTLVDYDIFTNVRLSDWVRLVQGSVPQTAPSLSLALKRSTIWAQWAMGEIL	1080
1081	STEDKSQRVAIINLLVDVAINCKDLANFNTAISHTALTNNHHIKRLQQTWDSPVKETLNKIQTQLEQSLQVWLKPDAATNFGVVICQSINSA	1170
1171	CVPNFSILRTIILSQIDQKIPFTSNDGSMVNVEKLRTIFGIVVEIQLQOORNNTMPTKLF1QLODINTVSMDELADLSLKCEPPVSKAK	1260
1261	KYNAPADIVEDDWRLKITKTFNKPPLATTSGVIDLPLRASSFTFNTGHKTTPEEKSYAGGNKIQDIFHVLVSLAQIESSELETDVREKFTT	1350
1351	YIPMSTDPSDFSKRELLKFLDEVCHADNSKLVRVLKCCNQAIIAPIVIIETLTNIAKGVPFMADGGWRILISNNNSNSVILDKIDEINE	1440
1441	DSSNVEKEKLSSSQEQEEQKQQEQQQQEPQLFLIRHYKKQRSRAQSOKDFEFEWFLIQNLNDADCKNLISFDLKISNLVFSTDTS	1530
1531	PNIRDOLLESFKSYLVSOECVOYINFENKOPVAAPVTATTIVTTKEESTTVTSSTTVVOESVPSTNAE	1601

- Asparagine rich region (72-126)
 - Region require for plasma membrane localization (361-644)
 - Lysine rich region (520-534), (579-616)
 - RhoGAP (649-836)
 - RasGEF_N (851-983)
 - RasGEF (1021-1255)

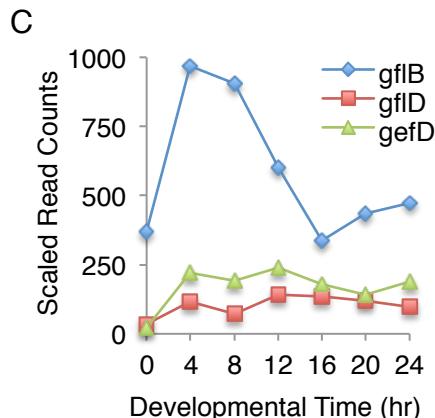
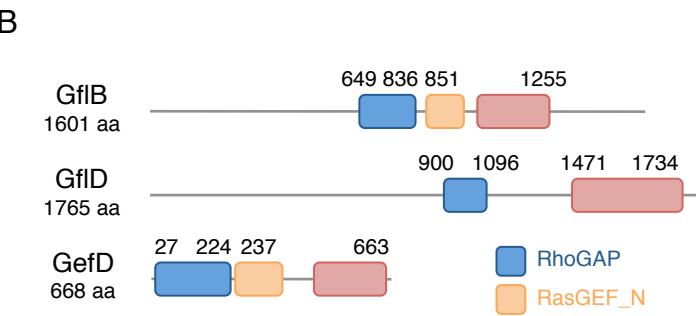


Figure S1. Amino acid sequence, domain structure, and expression profile of GflB. (A) Amino acid sequence of GflB. (B) Domain structures of GflB, GflD, and GefD. (C) Expression profiles of GflB, GflD, and GefD during development. The values are obtained from dictyExpress in the *Dictyostelium* database (www.dictyexpress.org) as described elsewhere (Parikh et al., 2010).

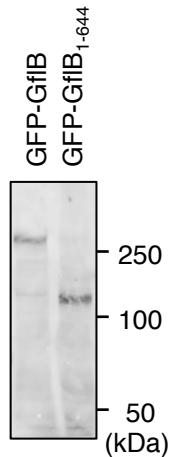


Figure S2. Whole-cell lysates analyzed using immunoblotting with anti-GFP antibodies to detect the indicated PTEN-GFP fusions.

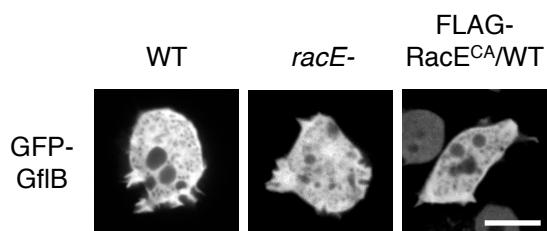


Figure S3. GFP-GflB was examined in WT cells, *racE*⁻ cells, and WT cells expressing constitutively active RacE_{G20V} using fluorescence microscopy. Bar, 10 μ m.

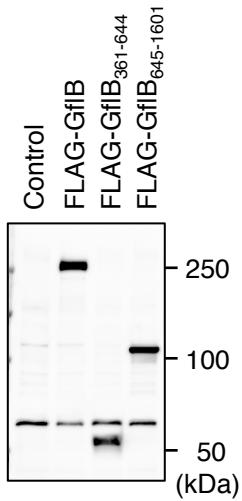


Figure S4. Whole-cell lysates analyzed using immunoblotting with anti-FLAG antibodies.

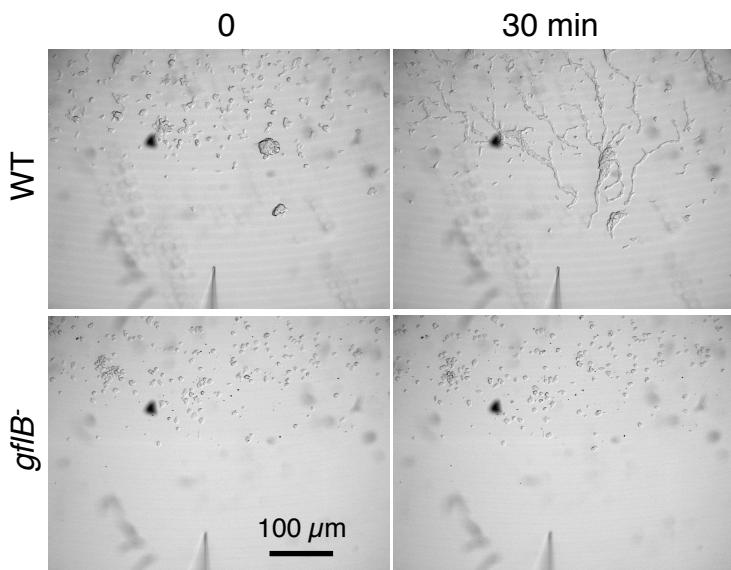


Figure S5. After 8 hours starvation, WT and *gflB*⁻ cells were placed in a chemoattractant gradient generated by a micropipette that released cAMP and observed for 20 min using phase contrast microscopy.

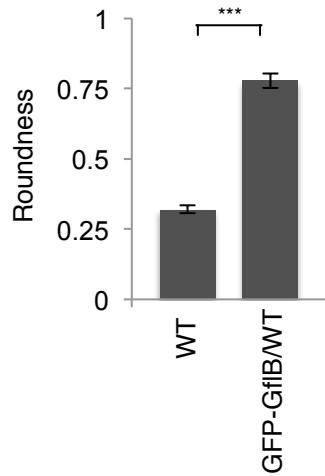


Figure S6. Roundness was determined by calculating the ratio between the short axis (A_s) and long axis (A_l) of cells (A_s/A_l) in WT cells in the presence or absence of GFP-GflB expression. Values represent mean \pm SEM. At least 30 cells were analyzed for each group.

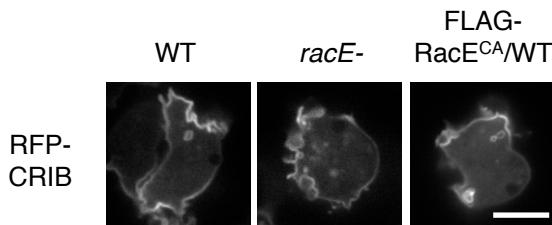


Figure S7. CRIB-RFP was examined in WT cells, *racE*⁻ cells, and WT cells expressing constitutively active RacE_{G20V} using fluorescence microscopy. Bar, 10 μ m.

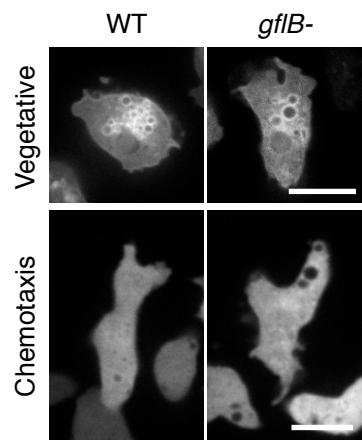


Figure S8. The PS biosensor GFP-LactC2 was observed in WT and *gflB*⁻ cells during growth (vegetative) and after differentiation (chemotaxis).

Table S1. Plasmids used in this study

Plasmids	Primers used	Drug for selection	References
<i>Dictyostelium expression</i>			
pJK1-GFP: pIS1		G418 (20 µg/ml)	Zhang et.al. (2011)
pJK1-GFP-racIA	10 11	G418 (20 µg/ml)	This Study
pJK1-GFP-racB	12 13	G418 (20 µg/ml)	This Study
pJK1-GFP-racE		G418 (20 µg/ml)	Wang et.al. (2013)
pJK1-GFP-racE (G20V)		G418 (20 µg/ml)	Wang et.al. (2013)
pJK1-GFP-racE (T25N)		G418 (20 µg/ml)	Wang et.al. (2013)
pDRH-FLAG-RasC		Hygromycin (50 µg/ml)	Gift from Dr. Devreotes (JHMI)
pDRH-FLAG-RasG		Hygromycin (50 µg/ml)	Gift from Dr. Devreotes (JHMI)
pDRH-LimEΔcoli-mRFP		Hygromycin (50 µg/ml)	Gift from Dr. Devreotes (JHMI)
pDM323-RBD(Raf)-GFP		G418 (20 µg/ml)	Xiong et.al. (2010)
pDRH-PHcrac-RFP		Hygromycin (50 µg/ml)	Gift from Dr. Devreotes (JHMI)
pTX-GFP		G418 (20 µg/ml)	Levi et.al. (2000)
pTX-GFP-GfIB	14 15	G418 (20 µg/ml)	This Study
pTX-GFP-GfIB 1-644	14 16	G418 (20 µg/ml)	This Study
pTX-GFP-GfIB 361-644	16 17	G418 (20 µg/ml)	This Study
pTX-GFP-GfIB 645-1601	15 18	G418 (20 µg/ml)	This Study
pTX-FLAG		G418 (20 µg/ml)	Levi et.al. (2000)
pTX-FLAG-GfIB	14 15	G418 (20 µg/ml)	This Study
pTX-FLAG-GfIB 361-644	16 17	G418 (20 µg/ml)	This Study
pTX-FLAG-GfIB 645-1601	15 18	G418 (20 µg/ml)	This Study
pDRH-CRIB-RFP	19 20	Hygromycin (50 µg/ml)	This Study

Bacterial expression

pGEX 4T-1

Amersham

pGEX-Byr2-RBD

Kae et.al. (2004)

Table S2. Primers used in this study

Primers for gene disruption in Dictyostelium cells

1	A15P	CCAACCCAAGTTTTAAACC
2	gfIB-2	GACTCGAGTTATCGGCATTTGTTGAAGGAAC
3	gfIB-3	GTGTACCAAATTCTCTATACTTCG
4	gfIB-4	CACCACCACTACCTCAACTACAACC
5	gfIB-5	GAGCGGCCGCGTTGTAGTACTACTAGTTAC
6	gfIB sall 1-19	GAGTCGACATGACAGATTAAATTCAG
7	gfIB smal 800-781	GACCCGGGAATTGATTAGCCTGTAAAAC
8	gfIB smal 4116-4134	GACCCGGGAATCATCAGAATTGGAGAC
9	gfIB notI 4875-4856	GAGCGGCCGCGTTGTAGTACTACTAGTTAC

Primers for expression plasmids in Dictyostelium cells

10	rac1A-1	GGAAGATCTTCCATGCAAGCAATTAAATG
11	rac1A-2	GGAAGATCTCCTTATAAAATGTTGC
12	racB-1	GGAAGATCTTCCATGCAATCAATTAAATTGG
13	racB-2	GGAAGATCTCCTTATAAAATTGAACATTTG
14	gfIB-Start	CCCGAGCTCATGACAGATTAAATT
15	gfIB-Stop	CCCCTCGAGTTATCGGCATTTGTTGAAGGAAC
16	gfIB 1932-1909	GACTCGAGTTAATATGGTGAATCTAAATTGTTG
17	gfIB 1081-1101	GAGAGCTCTTACAAGTAGTTCCAATGCC
18	gfIB 2042-2066	GAGAGCTCGGAATTATGGTGTAGATTAACAC
19	CRIB 943	CCCGAGCTCACTACATCACCACCATCC
20	CRIB 1233	CCCCTCGAGTTAATGGAAATCTAAACATC