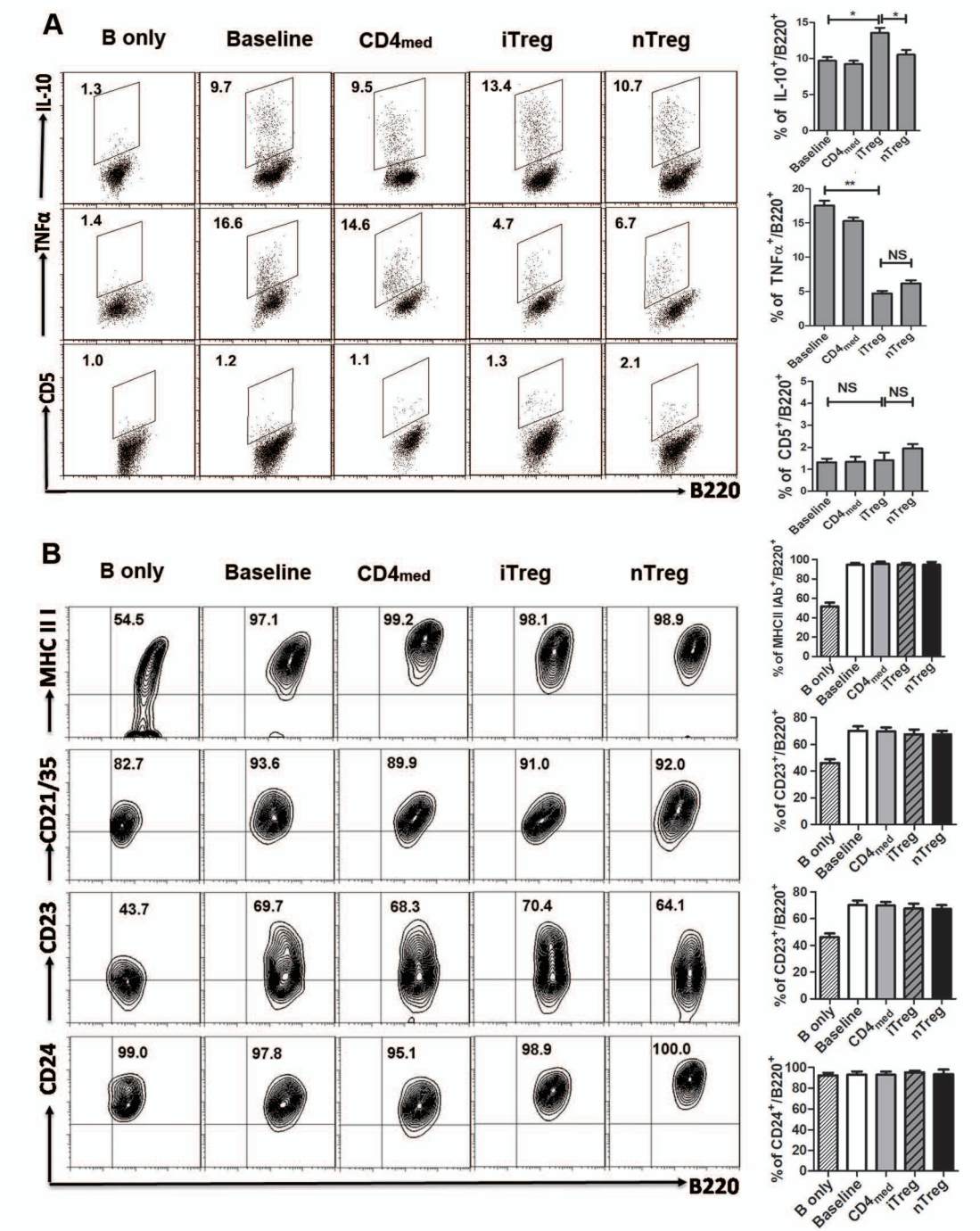

FIGURE S1. Implicate of Treg subsets on the phenotypes and cytokine production by B cells. **(A and B)** B cells were stimulated with (Baseline) or without LPS (2 μg/ml) (B only) in the presence or absence of nTreg, iTreg and CD4med (ratio of T to B cells was 1:2 to 1:4). IL-10, TNFα, CD5, MHC-II, CD21/35, CD23 and CD24 expression by B cells was determined by flow cytometry after culture days 2. The data indicate the Mean ± SEM of 3 separated experiments (*p<0.05, **p<0.01, baseline vs. Treg group).

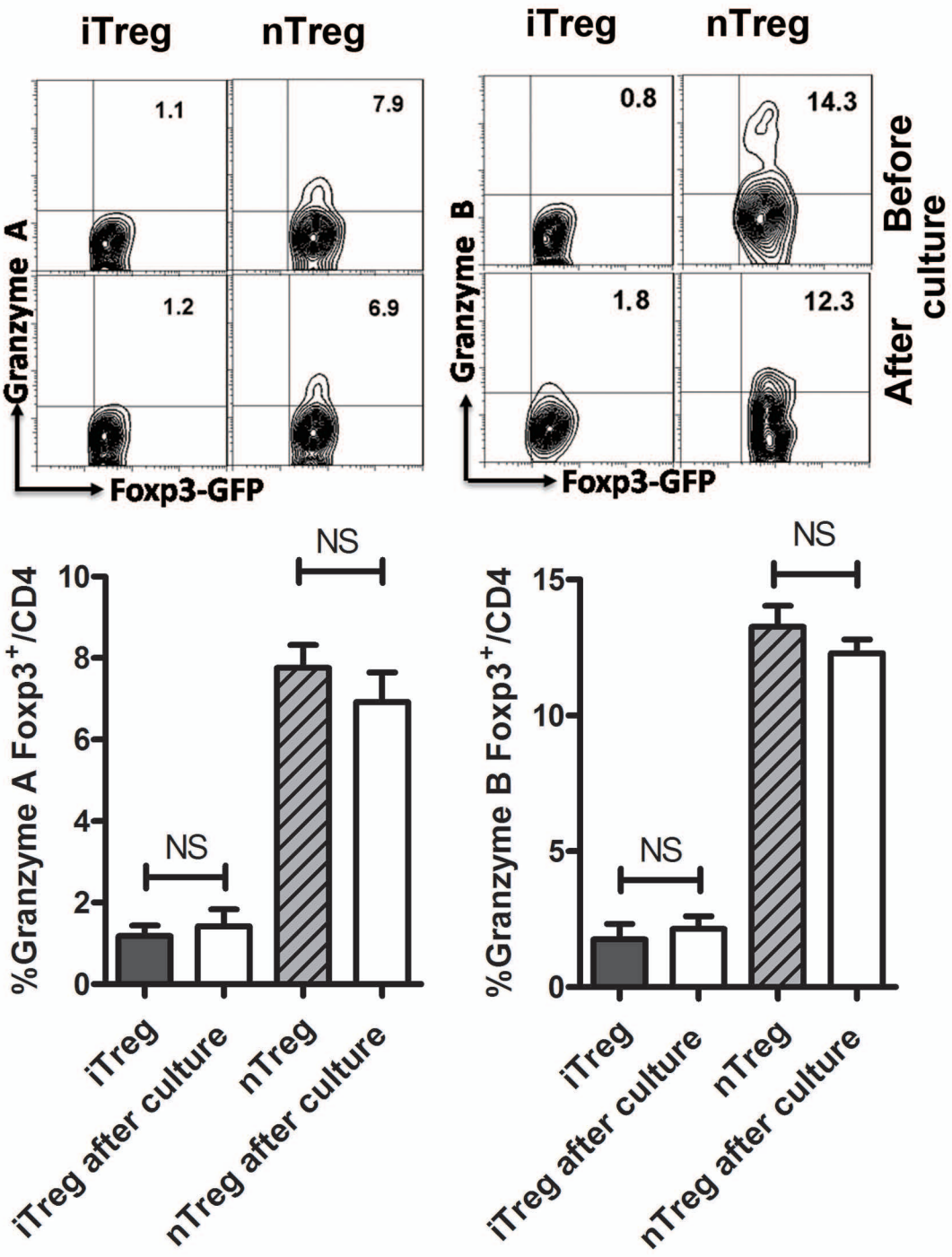

FIGURE S2. Granzyme A and Granzyme B expression was not altered after co-cultures in Treg subsets. B cells were stimulated with (Baseline) or without LPS (2 μg/ml) (B only) in the presence or absence of nTreg, iTreg and CD4med (ratio of T to B cells was 1:2 to 1:4). Granzyme A and Granzyme B were analyzed on nTreg, iTreg and CD4med before or after co-cultures by flow cytometry. The data indicated Mean ± SEM of three separate experiments.

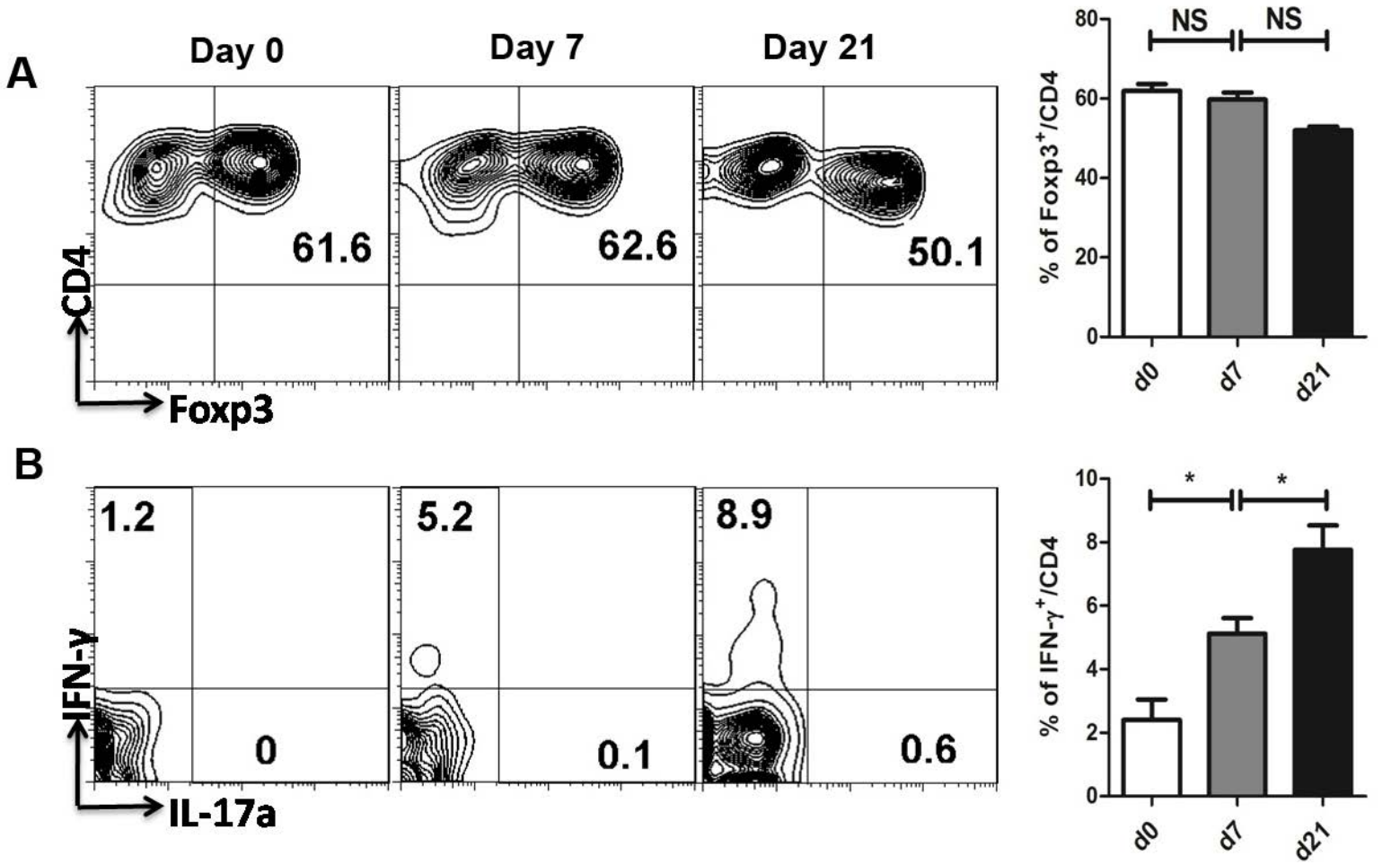

FIGURE S3. Stability of iTreg in vivo. **(A and B)** iTreg were generated from splenic naive CD4+ T cells from C57BL/6 (Foxp3-GFP reporter) mice, and then i.v injected into Rag1-/- mice. The mice were sacrificed and spleen cells were harvested at days 7 and 21. The expression of Foxp3, IFN-γ and IL-17A in CD4+ T cells was determined by flow cytometry. There were five mice in each group. The data indicated Mean ± SEM of three separate experiments (*p<0.05).

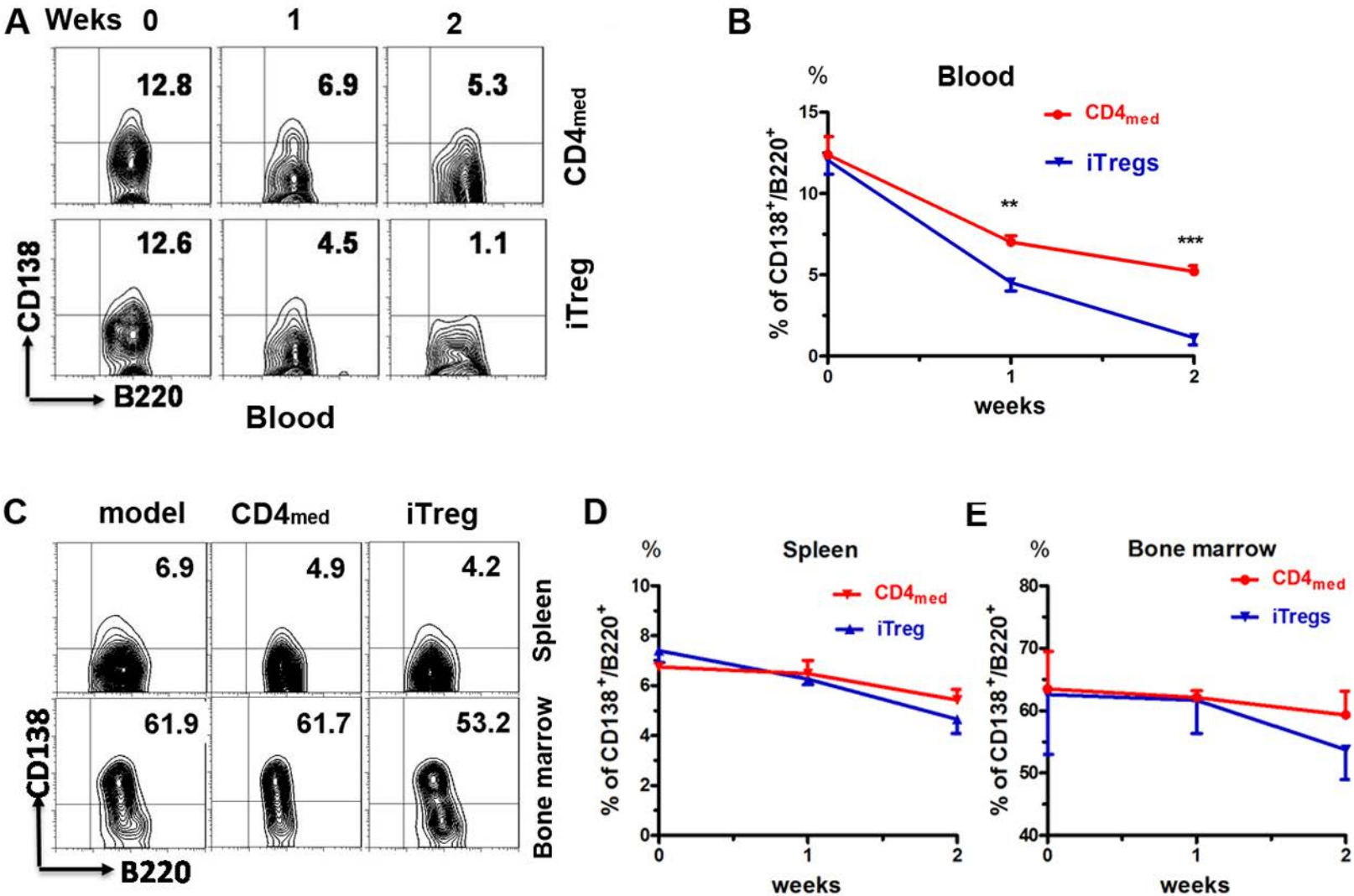

FIGURE S4. iTreg subset reduced the frequency of circulating CD138+ plasma cell in lupus mice. iTreg or CD4med cells induced from BWF1 lupus mice with evident disease syndromes were adoptively transferred to established lupus mice, the percentages of CD138+ plasma cell in blood **(A and B)**, spleen and bone marrow **(C-E)** were calculated by flow cytometry. **p<0.01, ***p<0.001 means CD4med control cell group versus iTreg group at weeks 1, 2 after cell transfer.