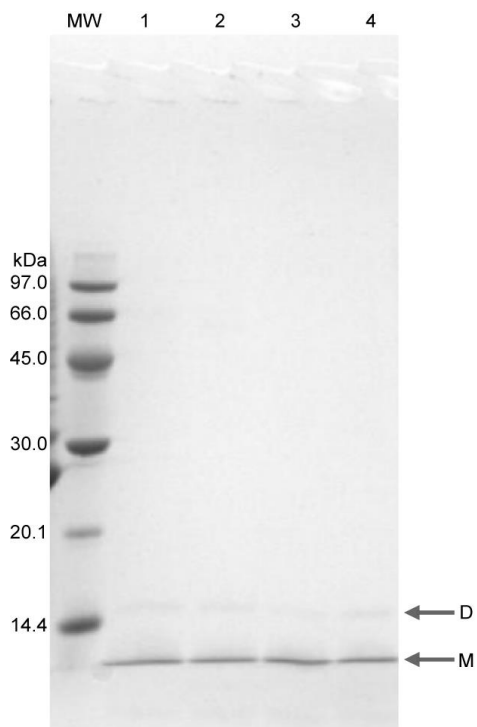
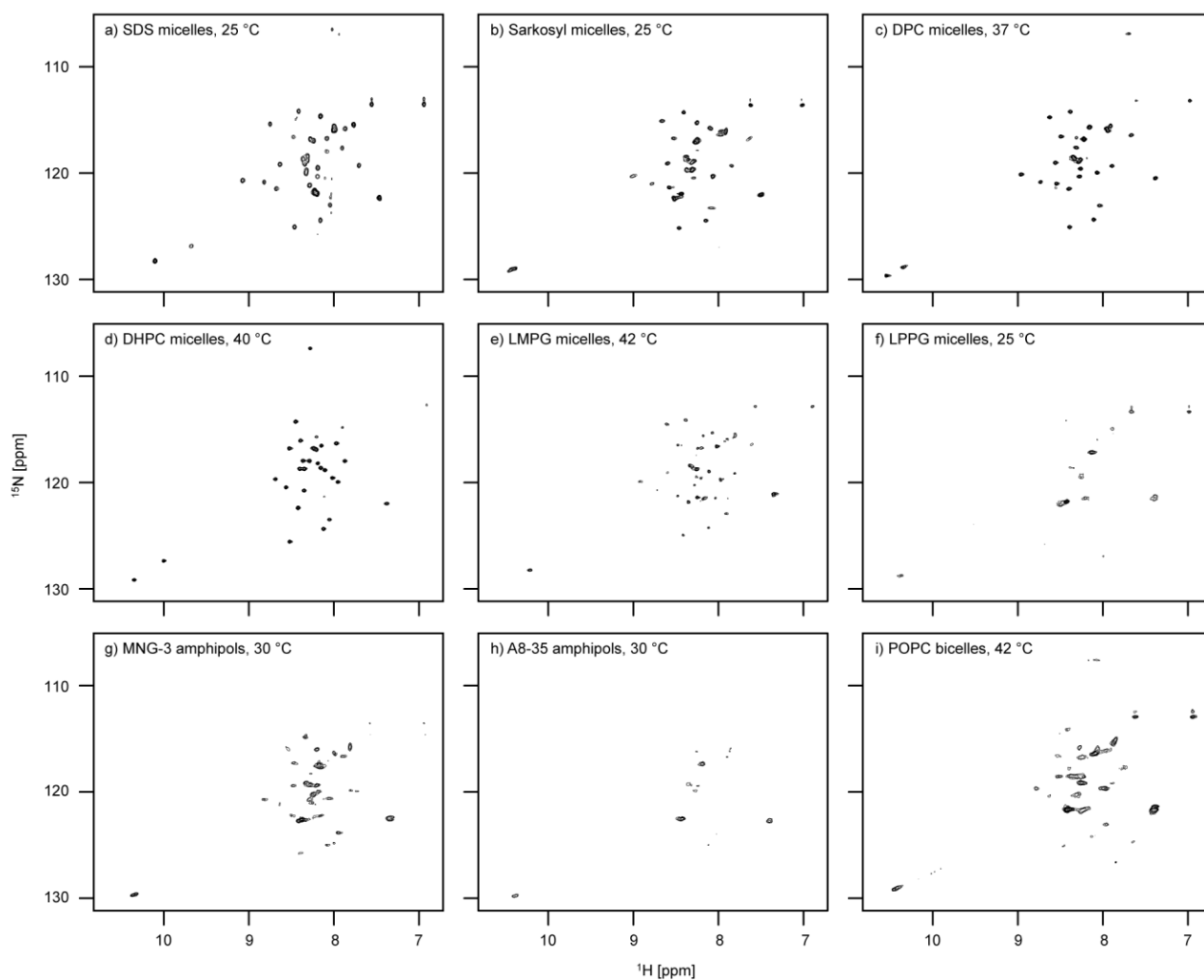


Supplementary Information

Supplementary Figures

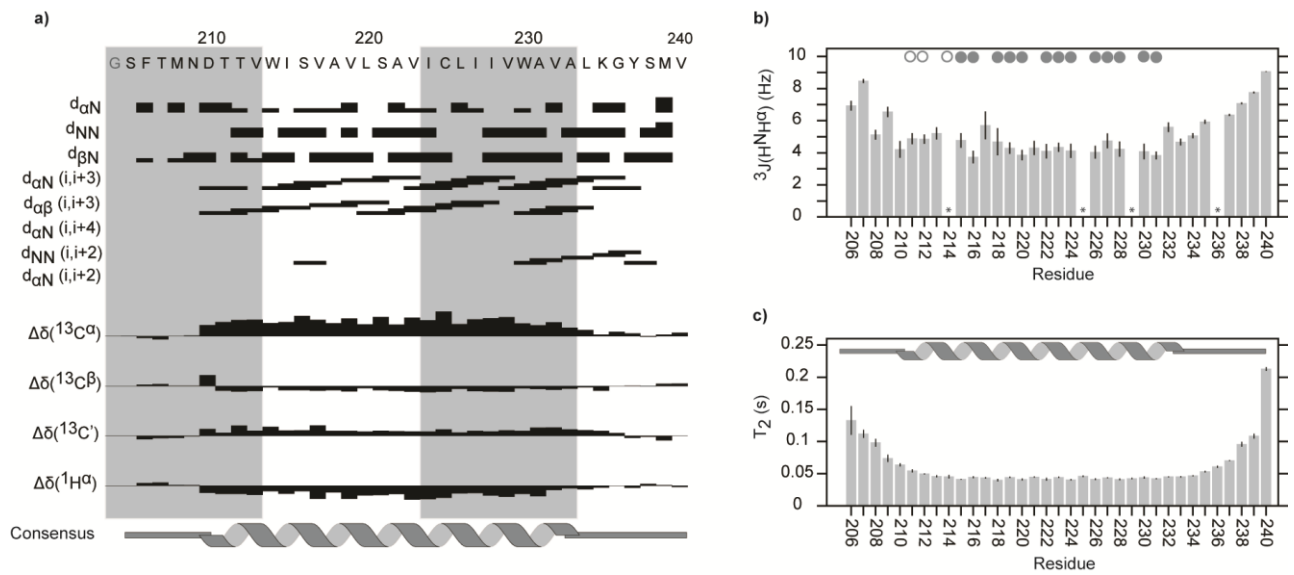


Supplementary Figure 1. SDS-PAGE analysis of hPRLR-TMD. A 15% (w/v) SDS-PAGE run under reducing conditions. Right: Molecular weight (MW) markers. Left (lane 1-4): hPRLR-TMD. The bands corresponding to monomeric and dimeric hPRLR-TMD are indicated by the arrows marked M and D, respectively.

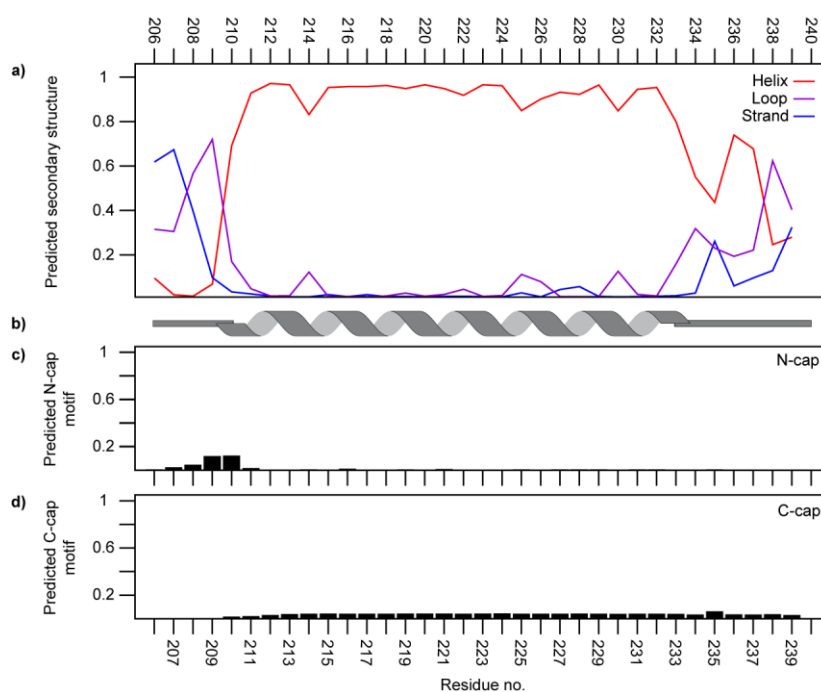


Supplementary Figure 2. Screening of NMR-suitable membrane mimetics for hPRLR-TMD structure determination. To obtain optimal conditions for structural studies with solution state NMR spectroscopy, hPRLR-TMD was reconstituted in various membrane mimicking solvents. A total of 13 different membrane mimetic solvents were screened: HFIP, SDS micelles, 1:2 SDS:DPC micelles, DPC micelles, sarkosyl micelles, DHPC micelles, LMPG micelles, LPPG micelles, MNG-3 amphipols, A8-35 amphipols, DHPC/POPC bicelles ($q=0.3$), DHPC/POPC:POPS (3:1) bicelles ($q=0.3$), and DHPC/DMPC bicelles ($q=0.3$). Furthermore, DPC micelles, DHPC micelles, LMPG micelles, LPPG micelles, and the bicelles were tested at two different concentrations. The majority of the membrane mimetics readily solubilized the TMD, but especially 60% (v/v) HFIP, and to some extent LPPG, LMPG, and the amphipols, resulted in visible aggregates. The potential for high-quality NMR data of the solubilized TMDs were evaluated from ^1H - ^{15}N -HSQC spectra at varying temperatures (25°C-40°C, not all temperatures were tested for all conditions), and a protein concentration of 0.4 mM in 50 mM NaCl, 20 mM $\text{Na}_2\text{HPO}_4/\text{NaH}_2\text{PO}_4$ (pH 7.2), and 10% (v/v) D_2O . Here nine selected ^1H - ^{15}N -HSQC spectra are shown of 0.4 mM hPRLR-TMD reconstituted in different membrane mimetics and at various temperatures. **a)** 80 mM SDS micelles, 25°C. **b)** 0.5% (w/v) sarkosyl micelles, 25°C. **c)** 200 mM DPC micelles, 37°C. **d)** 500 mM DHPC micelles, 40°C. **e)** 200 mM LMPG micelles, 42°C. **f)** 20 mM LPPG micelles, 25°C. **g)** 2

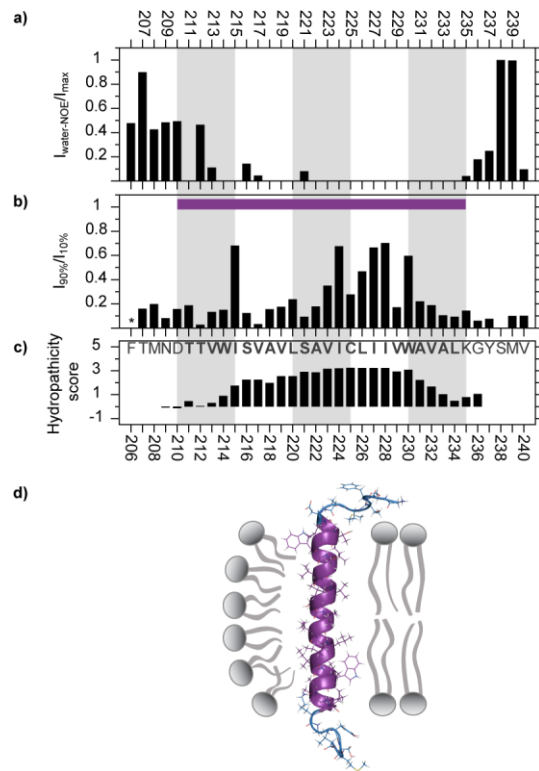
mM MNG-3 amphipols, 30°C. h) 2 mM A8-35 amphipols, 30°C. i) 15% (w/v) DHPC/POPC bicelles (q=0.3), 42°C. Reconstitution in some membrane mimetics resulted in broad line widths and/or a significant lower peak count than expected, revealing a high degree of variability in the quality of the resulting NMR spectra. DPC micelles and DHPC micelles provided the ^1H - ^{15}N -HSQC spectra of highest quality both resulting in well-dispersed peaks with narrow line widths. A closer inspection revealed that 31 out of the expected 35 peaks could be identified in the ^1H - ^{15}N -HSQC spectrum of hPRLR-TMD in DPC micelles, while all 35 expected peaks were resolved in DHPC micelles. Temperatures above 30 °C were found to provide the highest quality data. Based on these results, a high concentration of DHPC micelles (1:700) in 50 mM NaCl, 20 mM $\text{Na}_2\text{HPO}_4/\text{NaH}_2\text{PO}_4$ (pH 7.2) at 37°C was selected for the structural studies of hPRLR-TMD with solution state NMR spectroscopy. Under these conditions, the sample appeared stable for at least four weeks at 37 °C.



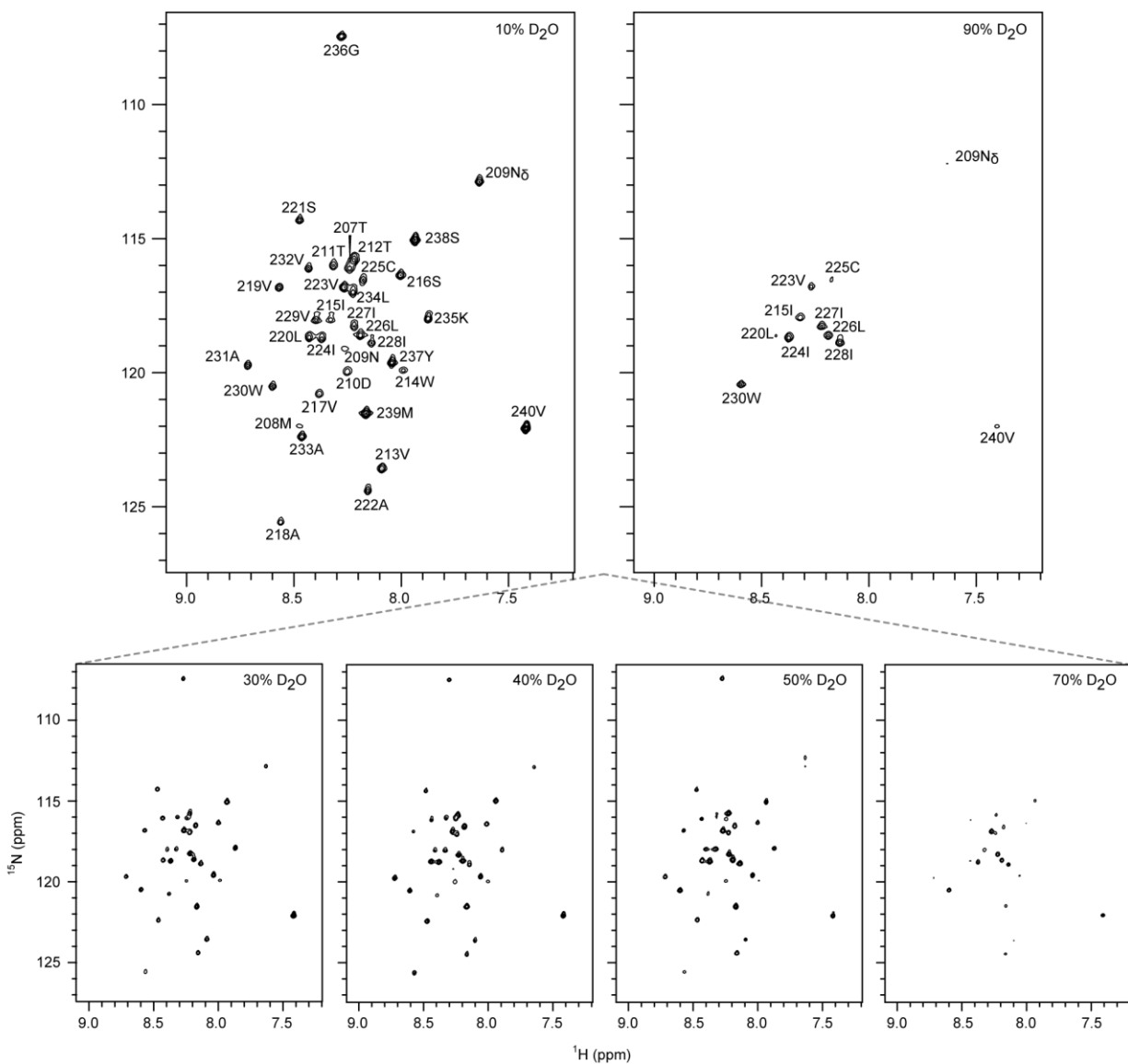
Supplementary Figure 3. Summary of structural data collected on hPRLR-TMD in DHPC micelles. a) Overview of secondary chemical shifts (SCSs) for C^α , C^β , C' , and H^α resonances and NOE connectivities within hPRLR-TMD. Positive deviations from random coil chemical shifts¹ for C^α and C' are representative of α -helical conformation, while negative deviations suggest β -sheet or extended conformations. The signs are opposite for C^β and H^α SCSs. The figure has been adapted from CcpNmr Analysis software output². **b)** $^3J(H^N-H^\alpha)$ coupling constants extracted from an HNHA spectrum plotted against residue number. The vertical black lines represent the estimated error of the fits for each scalar coupling constant, and “*” represents glycines or residues where the coupling constant could not be determined. The measured scalar coupling values were used to estimate the ϕ backbone dihedral angles using the Karplus equation³. Filled grey circles at the top highlight residues for which the $^3J(H^N-H^\alpha)$ coupling constants along with characteristic NOE patterns provided the basis for inclusion of a hydrogen bond restraint in the structure calculations, while empty grey circles highlight residues only receiving the hydrogen bond restraint on their C' . **c)** T_2 -relaxation times for hPRLR-TMD backbone amides plotted against residue number. The vertical black lines represent the estimated error of each fit.



Supplementary Figure 4. Identification of structural motifs in hPRLR-TMD using C^α , C^β , C' , N^H , H^N , and H^α chemical shifts and the motif identification from chemical shifts (MICS) program⁴. *a)* Statistical probability calculated by MICS of finding helix (red), loop (purple), or strand (blue) elements for each residue of hPRLR-TMD. *b)* Cartoon representation of the secondary structure in the lowest energy structure of hPRLR-TMD. *c)* Statistical probability calculated by MICS of a residue in hPRLR-TMD forming an N-terminal capping motif (N-cap) plotted against residue number. *d)* Statistical probability calculated by MICS of a residue in hPRLR-TMD forming a C-terminal capping motif (C-cap) plotted against residue number.



Supplementary Figure 5. DHPC-embedding of hPRLR-TMD. *a)* Normalized intensities of NOEs between backbone amides and water plotted against residue number, extracted from a 3D ^{15}N -NOESY-HSQC spectrum. *b)* Intensities of backbone amide peaks in 90% D_2O normalized to peak intensities in 10% D_2O , plotted against residue number. The purple bar highlights the residues in helical conformation. Relatively high peak intensity indicates that the backbone amide has low exchange with water. *c)* Kyte-Doolittle hydropathy plot of hPRLR-TMD⁵. Positive values indicate hydrophobic stretches. *d)* Schematic model of hPRLR-TMD embedding in a DHPC micelle (left) and a lipid bilayer (right), illustrating the organizational differences of the two solvents.



Supplementary Figure 6. ^1H - ^{15}N -HSQC spectra from H-D exchange experiments. ^1H - ^{15}N -HSQC spectra of hPRLR-TMD in DHPC micelles at different levels of D_2O , acquired following establishment of a quasi-stationary state.

Supplementary References

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