

CIRCADIAN RHYTHM

Timing of expression of the core clock gene *Bmal1* influences its effects on aging and survival

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ABSTRACT

The absence of *Bmal1*, a core clock gene, results in a loss of circadian rhythms, an acceleration of aging, and a shortened life span in mice. To address the importance of circadian rhythms in the aging process, we generated conditional *Bmal1* knockout mice that lacked the BMAL1 protein during adult life and found that wild-type circadian variations in wheel-running activity, heart rate, and blood pressure were abolished. Ocular abnormalities and brain astrogliosis were conserved irrespective of the timing of *Bmal1* deletion. However, life span, fertility, body weight, blood glucose levels, and age-dependent arthropathy - which are altered in standard *Bmal1* knockout mice - remained unaltered, while atherosclerosis and hair growth improved, in the conditional adult-life *Bmal1* knockout mice, despite abolition of clock function in both groups. Hepatic RNA-Seq revealed that expression of oscillatory genes was dampened in the adult-life *Bmal1* knockout mice, while overall gene expression was largely unchanged. Thus, many phenotypes in conventional *Bmal1* knockout mice, hitherto attributed to disruption of circadian rhythms, reflect the loss of properties of BMAL1 that are independent of its role in the clock. These findings prompt re-evaluation of the systemic consequences of disruption of the molecular clock.

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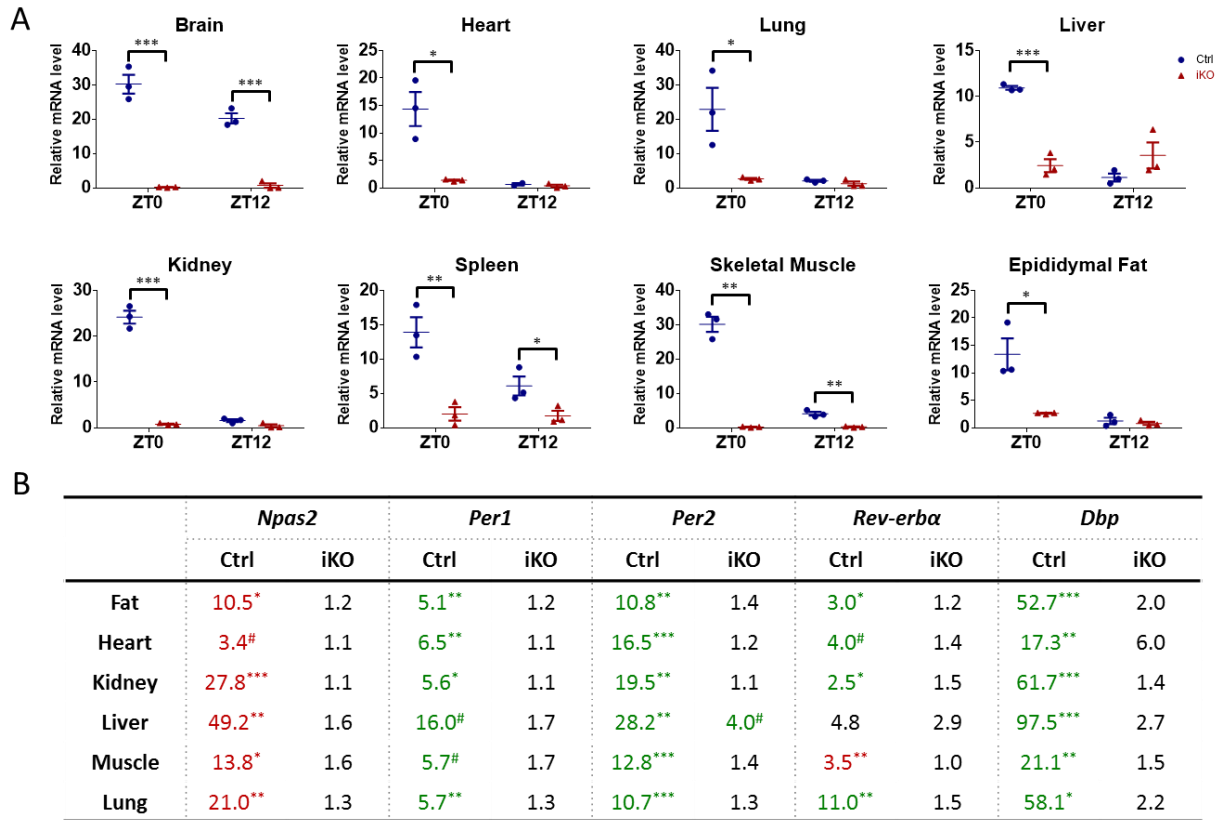


Fig. S1. Validation of *Bmal1* deletion and dampening effect of other clock genes. (A) Brain, heart, lung, liver, kidney, spleen, skeletal muscle, and epididymal fat were collected at ZT0 and ZT12. The *Bmal1* mRNA levels were determined using qRT-PCR and compared between Ctrl and iKOs (2-way ANOVA; *, $P < 0.05$; **, $P < 0.01$; ***, $P < 0.001$). (B) The fold change of clock gene expression levels in various tissues in Ctrl and iKO mice between ZT0 and ZT12 (Student's t-test, ZT0 vs. ZT12; #, $P < 0.1$; *, $P < 0.05$; **, $P < 0.01$; ***, $P < 0.001$; Red, ZT0 > ZT12; Green, ZT0 < ZT12; Black, no significant difference).

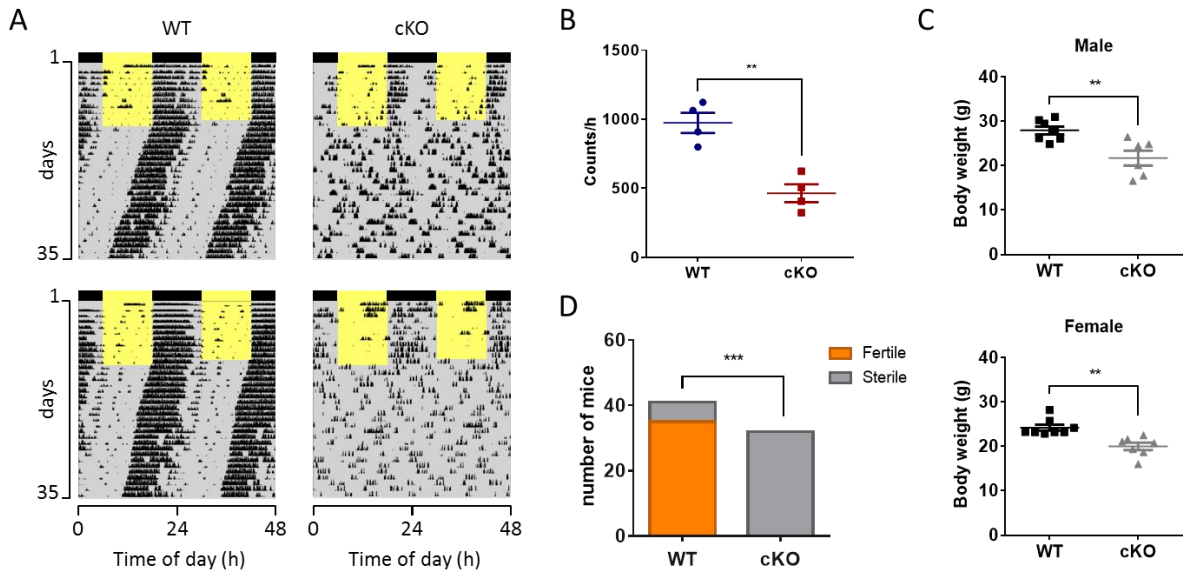


Fig. S2. Wheel-running activity, body weight, and fertility in cKO mice. (A) Representative activity records of individual WT and cKO littermates aged 3m are presented in double-plotted format. (B) Individual value plot of wheel running activity from WT and cKO mice under DD (Student's t-test; **, $P < 0.01$). (C) Body weight of 6m old male and female mice (Student's t-test; **, $P < 0.01$). (D) Fertility analysis of WT and cKO mice (male and female combined, χ^2 test; ***, $P < 0.001$).

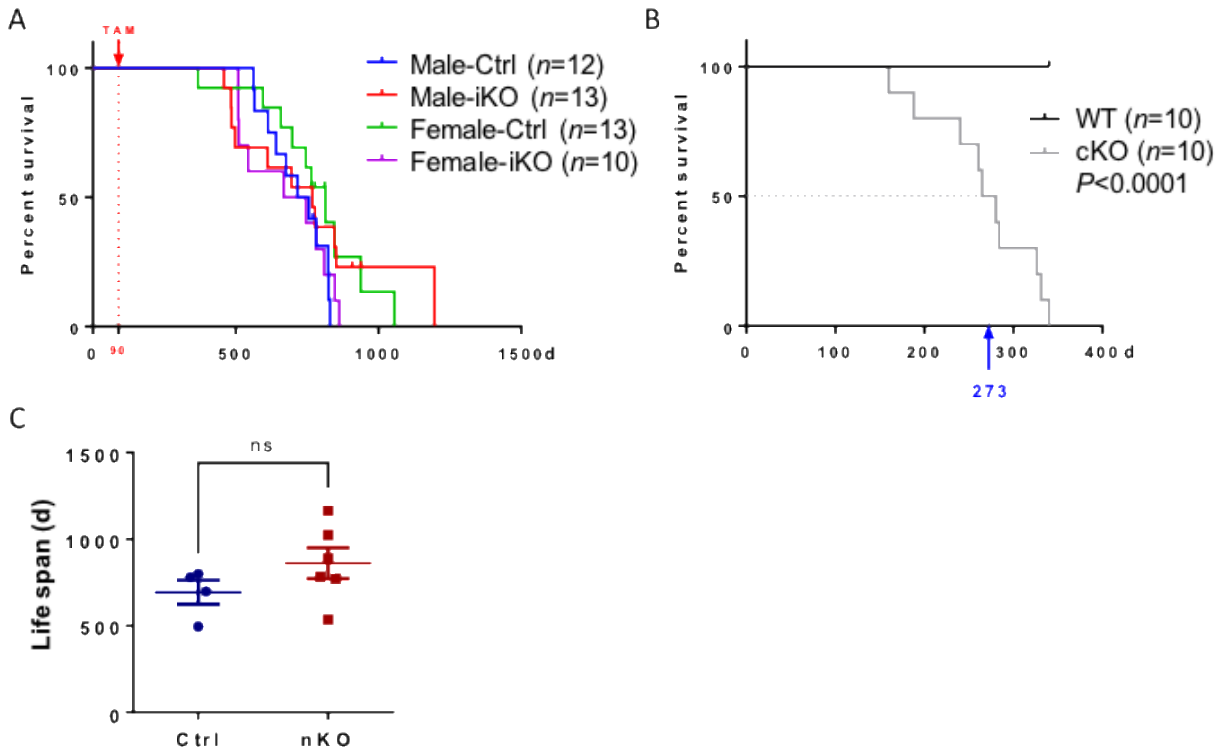


Fig. S3. Life span of iKO, cKO, and nKO mice. (A) Both male and female mice at 3 m old were treated with TAM (red arrow) and their life span was monitored for comparison (log-rank test; no significant difference between any 2 groups). (B) life span of cKOs. All WT mice at 340 d old were euthanized when last cKO mouse died (log-rank test; $P < 0.0001$). (C) The life span of nKOs and their littermate controls (Student's t-test; ns, no significant difference).

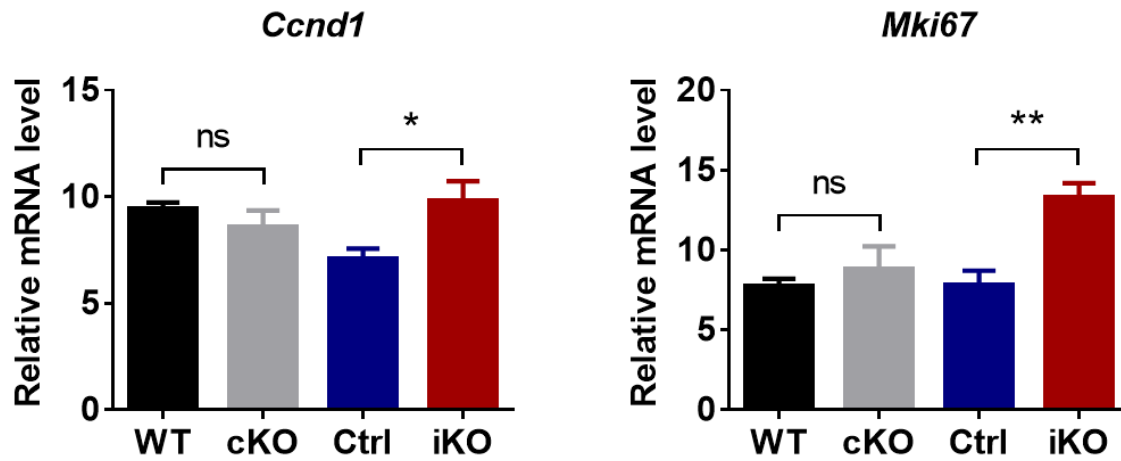


Fig. S4. mRNA levels of *Ccnd1* and *Mki67* in skin. Skin samples from both cKO and iKO and their littermates were used for qRT-PCR ($N=6$, Student's t-test; ns, no significant difference; *, $P<0.05$; **, $P<0.01$).

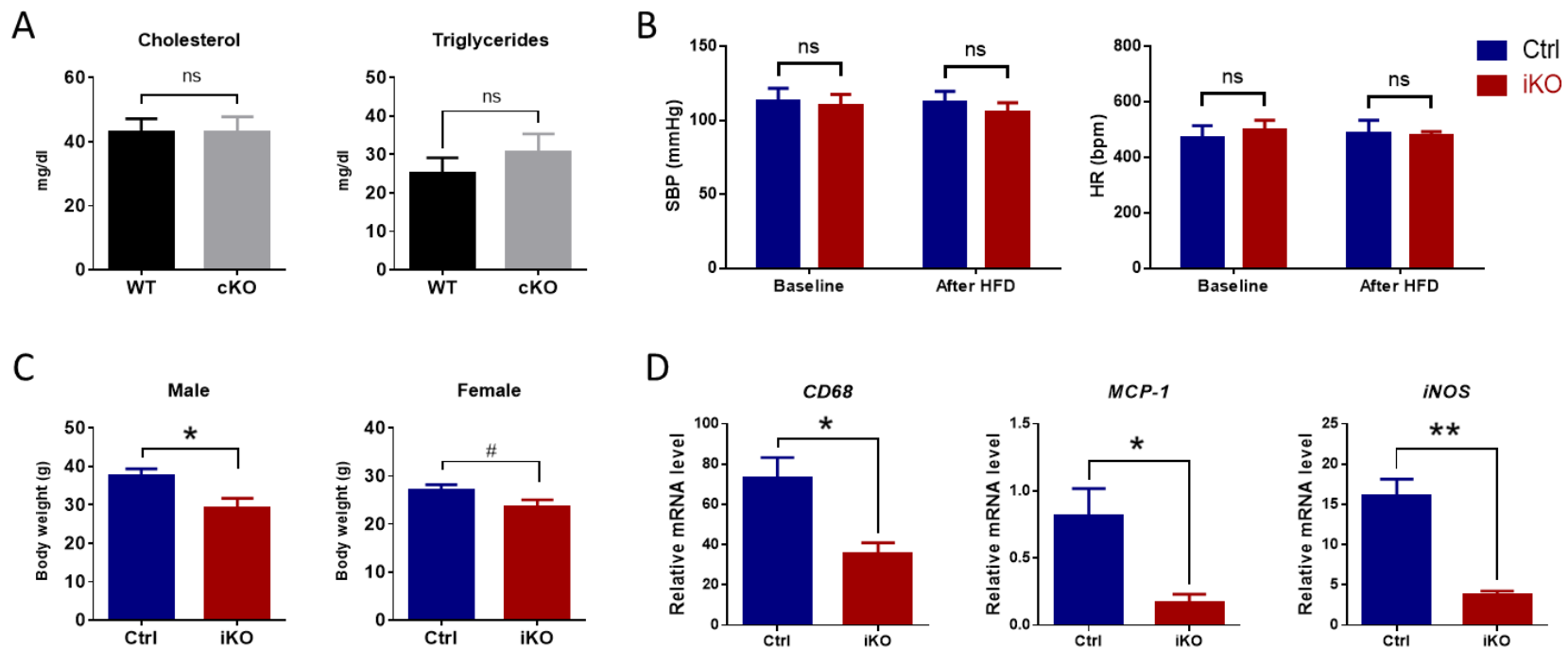


Fig. S5. Other parameters in HFD-fed mice. (A) Total cholesterol and triglycerides in cKO plasmas ($N=5$). (B) SBP and HR of Ctrl and iKO mice were measured by tail cuff at ZT8-9 before or after 2 m HFD treatment ($N=6$ to 8). (C) Body weights of both male ($N=14$) and female ($N=8$ to 12) mice. (D) qRT-PCR analysis of *CD68*, *MCP-1* and *iNOS* expression in aortas from iKO mice ($N=4$). Student's t-test was used for all comparisons (#, $P<0.1$; *, $P<0.05$; **, $P<0.01$).

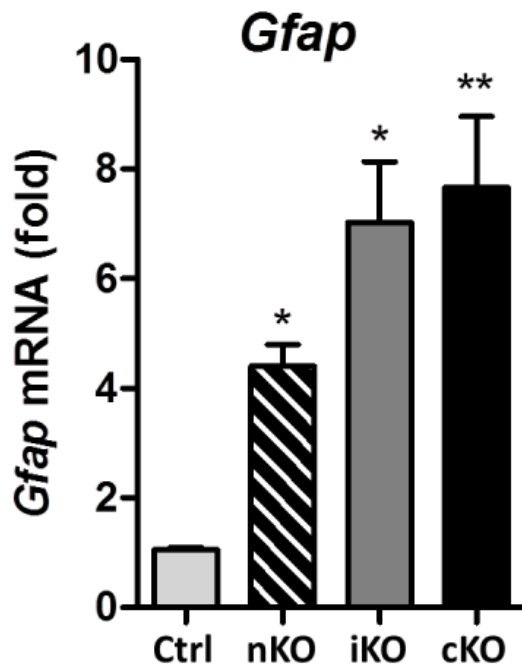


Fig. S6. iKO mice display similar *Gfap* induction in the brain as cKO. Cerebral cortex samples harvested from Ctrl, nKO, iKO, and cKO mice. *Gfap* mRNA level was quantified by qRT-PCR ($N=4$ to 12, 1-way ANOVA with Bonferroni posttest analysis, as compared to Ctrl group; *, $P<0.05$, **, $P<0.01$).

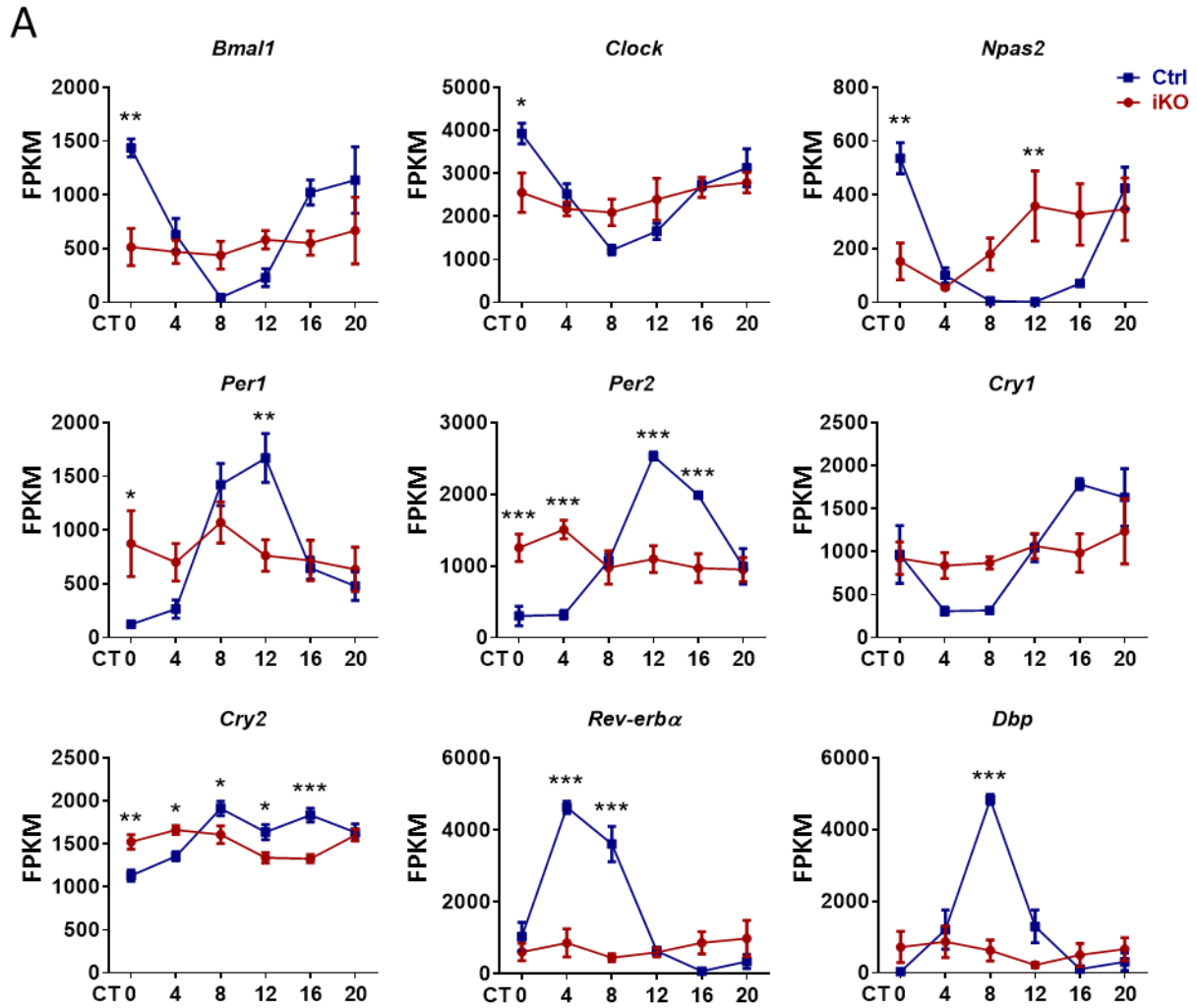


Fig. S7. RNA-Seq results revealed dampening

effect in core clock genes in iKO mice. (A)

Expression of core clock genes in Ctrl and iKO

mice ($N=4/\text{genotype}/\text{time point}$, 2-way ANOVA;

*, $P<0.05$; **, $P<0.01$; ***, $P<0.001$). **(B)**

JTK_CYCLE q -values of core clock genes in Ctrl

and iKO mice.

Table S1. Top 20 oscillating hepatic genes (JTK_CYCLE *q*-value) in Ctrl mice show no circadian pattern in iKO mice

Genes	Ctrl	iKO
<i>Tars</i>	1.76e-07	0.5649
<i>Igfbp4</i>	1.49e-06	0.6630
<i>Hnrnpl</i>	1.49e-06	1.0000
<i>Ptprk</i>	1.49e-06	0.9546
<i>Blcap</i>	1.49e-06	1.0000
<i>Npas2</i>	1.49e-06	0.3044
<i>Cald1</i>	2.18e-06	0.4950
<i>Rfxank</i>	2.18e-06	1.0000
<i>Gde1</i>	2.36e-06	0.1105
<i>Spata22</i>	2.36e-06	0.8014
<i>Crot</i>	2.36e-06	1.0000
<i>Slc9a3r1</i>	2.36e-06	0.2826
<i>Chka</i>	2.36e-06	1.0000
<i>St3gal1</i>	2.36e-06	1.0000
<i>Nr1d2</i>	3.14e-06	1.0000
<i>Lasp1</i>	3.22e-06	1.0000
<i>Arntl</i>	3.22e-06	1.0000
<i>Pitpnb</i>	3.22e-06	1.0000
<i>Mdfic</i>	3.22e-06	1.0000
<i>Rnf125</i>	4.59e-06	1.0000

Table S2. The ratio of differentially expressed gene numbers in cKO strain to the numbers in iKO strain

<i>q</i> -value	cKO	iKO	Fold (cKO/iKO)
0.01	234	3	78
0.05	462	3	154
0.1	656	9	73
0.2	998	15	67

Table S3. Summary of phenotypes of cKO and iKO mice

Phenotypes	cKO	iKO	Consistence
Circadian rhythm	Loss	Loss	√
Life span	↓	-	×
Fertility	↓	-	×
Body weight	↓	- ²	×
Organ weight	↓ ¹	-	×
Arthropathy	+	-	×
Astrogliosis	+	+	√
Eye pathologies	+ ³	+	√
Hair growth	↓	↑	××
Atherogenesis	↑	↓	××

↑, increased; ↓, decreased; +, positive; -, negative/no change; √, yes; ×, no; ××, opposite

1, experiment was not performed in the current study; 2, lower body weight seen in HFD fed mice;

3, histology was not studied, but the ocular abnormalities can be easily seen.

Data file S1. Circadian transcriptome. [Excel file]

Data file S2. Differentially expressed genes irrespective of time points. [Excel file]

Data file S3. Phenotype enrichment analysis of differentially expressed genes. [Excel file]