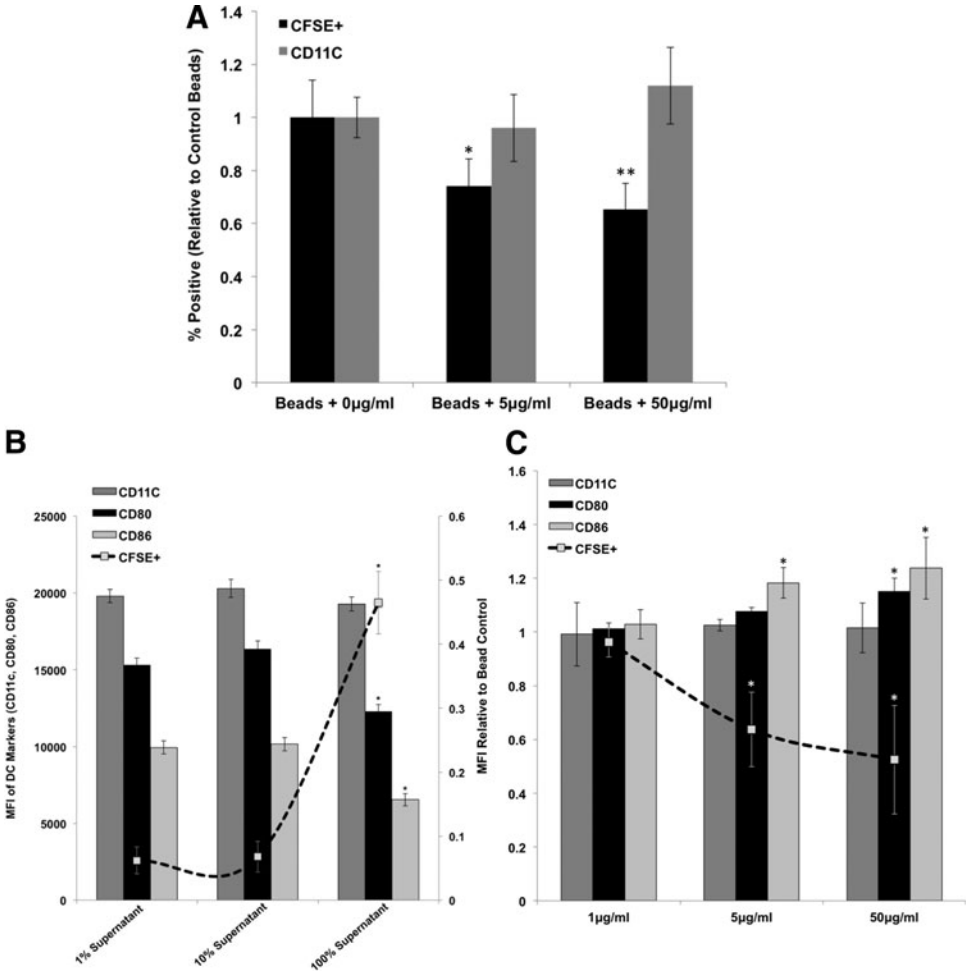

SUPPLEMENTARY FIG. S7. **(A)** Quantitation of Fig. 4D—Incubation of DC culture supernatants with αCTLA-4-coated beads blocks subsequent uptake of CFSE-labeled EV by unlabeled DC. Incubation of DC culture supernatants with αCTLA-4-coated beads blocked the subsequent uptake of CFSE-labeled EV by unlabeled DC in a titratable manner. At an antibody concentration of 5 µg/mL, 26% fewer recipient DC internalized $CFSE^+$ EV ($^*P < 0.05$), whereas at an antibody concentration of 50 µg/mL, 35% fewer recipient DC internalized $CFSE^+$ EV ($^{**}P < 0.01$). Staining of CD11c internal control was not statistically different between groups. Error bars = ±SD of three independent experiments. **(B)** Quantitation of S6A—DC-EV CTLA-4 physiologically binds B7. In three experiments, incubation of unlabeled recipient DC with cell culture supernatants derived from CFSE-labeled DC for 20 min at 4°C along with αCD80 and αCD86 flow-qualified antibodies and 0.1% sodium azide resulted in no change in B7 MFI using 1% or 10% supernatant (as well as no detectable uptake of $CFSE^+$ vesicles); however, in 100% supernatant, CD80 MFI and CD86 MFI were both significantly reduced as CFSE uptake quintupled. There was no statistical difference in CD11c MFI among any of the dilutions. $^*P < 0.05$. Error bars = ±SD. **(C)** Quantitation of S6B—DC-EV CTLA-4 physiologically binds B7. In three experiments, preclearance of CFSE-labeled DC culture supernatants with αCTLA-4 beads for 1 h before incubation of unlabeled DC for 20 min at 4°C along with αCD80 and αCD86 flow-qualified antibodies and 0.1% sodium azide resulted in a titratable and statistically significant increase in B7 MFI with no concomitant increase in CD11c MFI, while percentage of $CFSE^+$ cells dropped 45%. $^*P < 0.05$. Error bars = ±SD. **(B, C)** *Dashed line*, percentage of $CFSE^+$ cells.