

## Human melanoma immunotherapy using tumor antigen-specific T cells generated in humanized mice

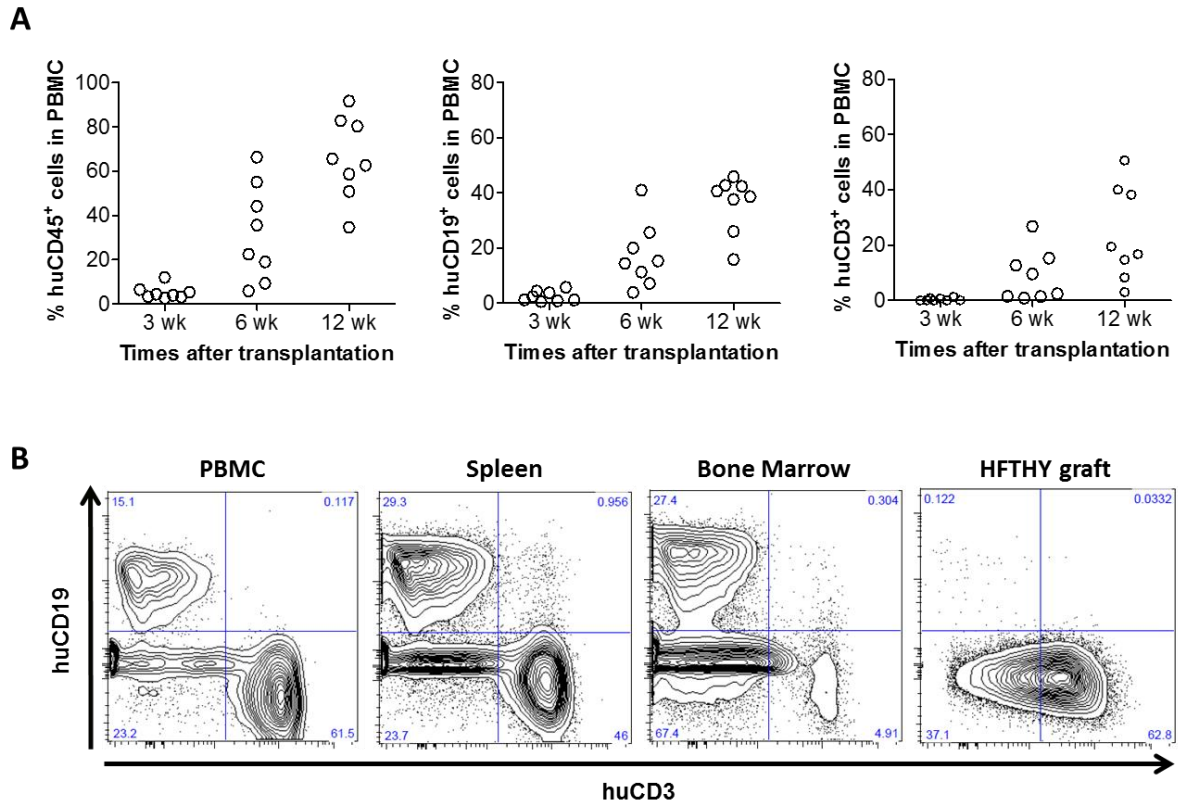
### Supplementary Material

**Table S1. Numbers of tetramer<sup>+</sup> CD8 T cells in hu-mouse spleens**

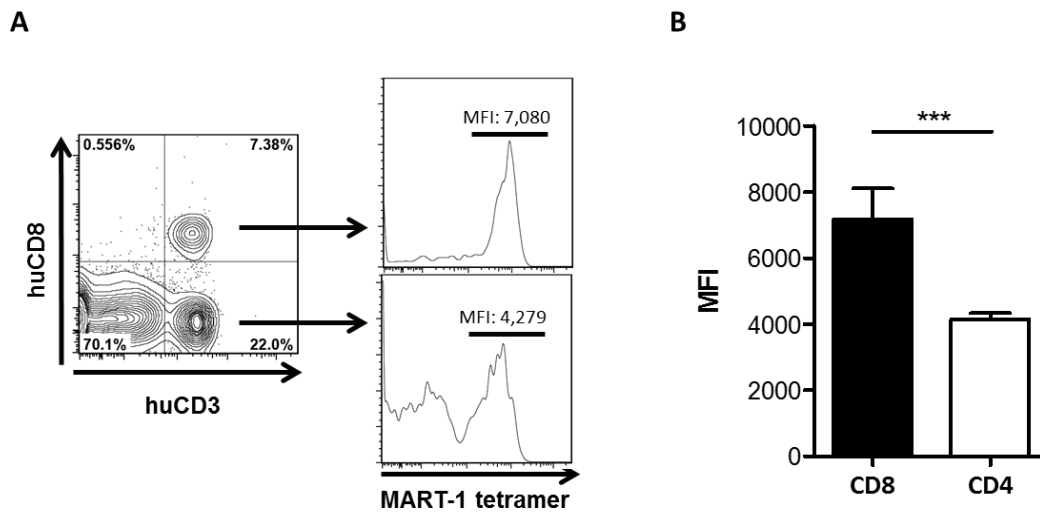
<b>Hu-mouse<sup>1</sup></b> (Eartag)	<b>Time of analysis</b> (Weeks post-Tx)	<b>Treatment<sup>2</sup></b>	<b>Tetramer<sup>+</sup> CD8 cells</b> (x10 <sup>6</sup> /spleen)
359/360	19	PBS	1.1
351/352	19	MART-1	9.3
361/362	20	PBS	1.2
355/356	21	PBS	1.8
353/354	21	MART-1	8.0
171/172	34	None	6.0
173/174	34	None	9.8
175/176	34	None	12.0

<sup>1</sup> Hu-mice from two experiments were analyzed: first 5 mice were from one experiment and the rest were from another one.

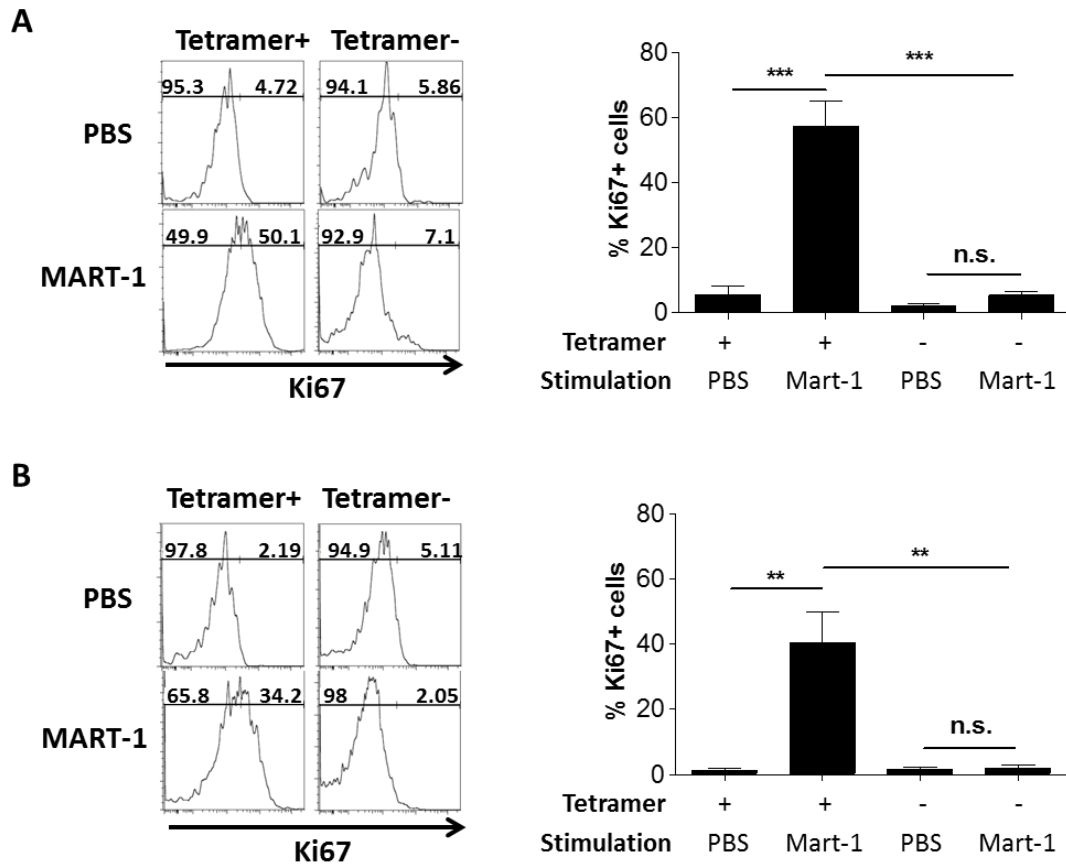
<sup>2</sup> Hu-mice were immunized with PBS or MART-1 peptides 3 weeks prior to analysis, or received no treatment.



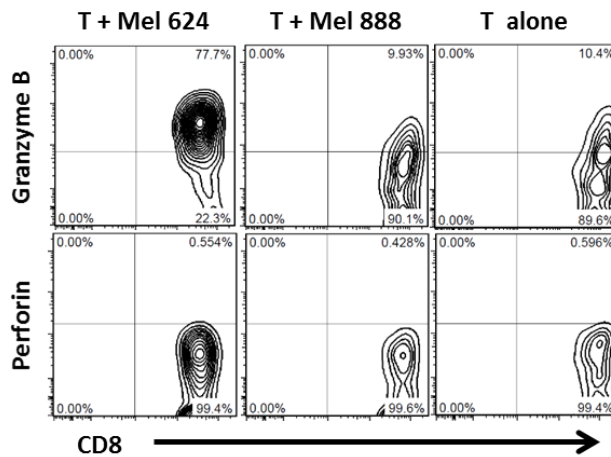
**Fig. S1. Human lymphohematopoietic cell reconstitution in humanized mice.** Flow cytometric profiles showing multilineages of human lymphohematopoietic cell reconstitution in blood at the indicated times (**A**), and in various tissues prepared from a representative hu-mouse 22 weeks (**B**) after transplantation of human FTHY and CD34<sup>+</sup> FLCs virally-transduced with MART-1-specific TCR gene.



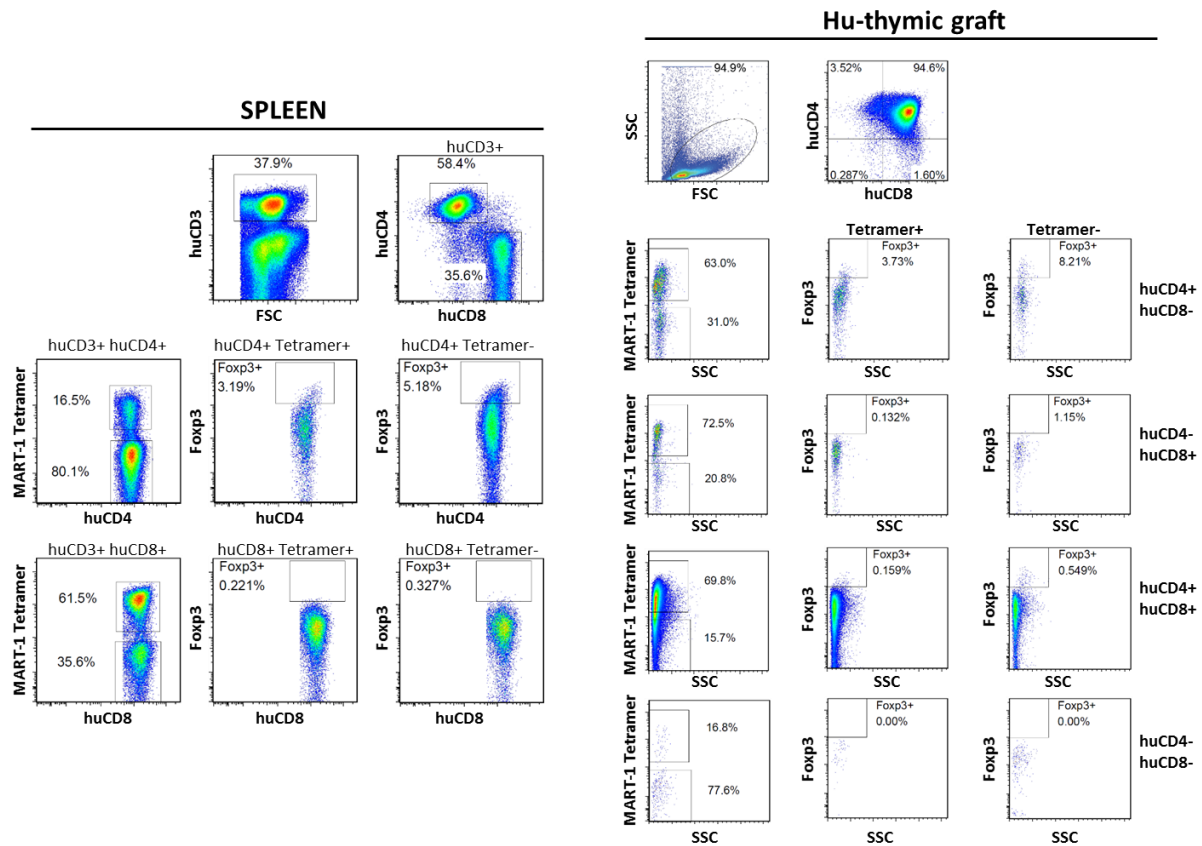
**Fig. S2. CD4 T cells express a lower level of MART-1-specific TCR than CD8 T cells. (A)** Representative flow cytometric profiles showing reconstitution of MART-1-specific TCR<sup>+</sup> cells in peripheral blood CD8<sup>+</sup> and CD8<sup>-</sup> (i.e., CD4<sup>+</sup> confirmed by staining with anti-CD4 mAb) and fluorescence intensity of MART-1 tetramer staining 14 weeks after humanization. **(B)** Median fluorescence intensity (MFI; mean±SEMs; n=6) of MART tetramer staining in CD8<sup>+</sup> and CD4<sup>+</sup> T cells. \*\*\*, p<0.001.



**Fig. S3. Proliferation of tetramer<sup>+</sup> T cells in response to MART-1 stimulation.** Spleen cells harvested from hu-mice were incubated with MART-1 peptides (10 $\mu$ g/mL) or PBS as control for 3 days. Proliferation of CD8<sup>+</sup> (**A**) or CD4<sup>+</sup> (**B**) T cells was then determined by measuring Ki67 expression. N=7 per group. \*, P<0.05; \*\*, P<0.01; \*\*\*, P<0.001; n.s., not significant.



**Fig. S4. Expression of granzyme B and perforin in MART-1-TCR<sup>+</sup> human CD8 T cells following stimulation with melanoma cells.** MART-1-TCR<sup>+</sup> human CD8 T cells were cultured alone or stimulated with Mel 624 (HLA-A2<sup>+</sup>MART-1<sup>+</sup>) or Mel 888 (HLA-A2<sup>-</sup>MART-1<sup>-</sup>) melanoma cells for 4-6 hours, and the secretion of granzyme B and perforin was determined by flow cytometric analysis. Results from a representative of two independent experiments are shown.



**Fig. S5. MART-1-TCR<sup>+</sup> human regulatory T cell development in humanized mice.** FACS analysis was performed 17 weeks after human thymus/CD34<sup>+</sup> cell transplantation. Shown are representative flow cytometric profiles of Fxp3 expression in MART-1 TCR<sup>+</sup> or MART-1 TCR<sup>-</sup> human T cells in the spleen (A) or thymocyte subsets in the human thymic graft (B).