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# Main Figures:	3
# Supplementary Figures:	15
# Supplementary Tables:	
# Supplementary Videos:	2

Reporting Checklist for Nature Neuroscience

This checklist is used to ensure good reporting standards and to improve the reproducibility of published results. For more information, please read Reporting Life Sciences Research.

Please note that in the event of publication, it is mandatory that authors include all relevant methodological and statistical information in the manuscript.

▶ Statistics reporting, by figure

- Please specify the following information for each panel reporting quantitative data, and where each item is reported (section, e.g. Results, & paragraph number).

- Each figure legend should ideally contain an exact sample size (n) for each experimental group/condition, where n is an exact number and not a range, a clear definition of how n is defined (for example x cells from x slices from x animals from x litters, collected over x days), a description of the statistical test used, the results of the tests, any descriptive statistics and clearly defined error bars if applicable.

- For any experiments using custom statistics, please indicate the test used and stats obtained for each experiment.

- Each figure legend should include a statement of how many times the experiment shown was replicated in the lab; the details of sample collection should be sufficiently clear so that the replicability of the experiment is obvious to the reader.

- For experiments reported in the text but not in the figures, please use the paragraph number instead of the figure number.

Note: Mean and standard deviation are not appropriate on small samples, and plotting independent data points is usually more informative. When technical replicates are reported, error and significance measures reflect the experimental variability and not the variability of the biological process; it is misleading not to state this clearly.

	TEST USED		n			DESCRIPTIVE STATS (AVERAGE, VARIANCE)		P VALUE		DEGREES OF FREEDOM & F/t/z/R/ETC VALUE	
FIGURE NUMBER	WHICH TEST?	SECTION & PARAGRAPH #	EXACT VALUE	DEFINED?	SECTION & PARAGRAPH #	REPORTED?	SECTION & PARAGRAPH #	EXACT VALUE	SECTION & PARAGRAPH #	VALUE	SECTION & PARAGRAPH #
example 1a	one-way ANOVA	Fig. legend	9, 9, 10, 15	mice from at least 3 litters/group	Methods para 8	error bars are mean +/- SEM	Fig. legend	p = 0.044	Fig. legend	$F(3, 36) = 2.97$	Fig. legend
example results, para 6	unpaired t-test	Results para 6	15	slices from 10 mice	Results para 6	error bars are mean +/- SEM	Results para 6	p = 0.0006	Results para 6	$t(28) = 2.808$	Results para 6
+/-											

	FIGURE NUMBER	TEST USED		n			DESCRIPTIVE STATS (AVERAGE, VARIANCE)		P VALUE		DEGREES OF FREEDOM & F/t/z/R/ETC VALUE	
		WHICH TEST?	SECTION & PARAGRAPH #	EXACT VALUE	DEFINED?	SECTION & PARAGRAPH #	REPORTED?	SECTION & PARAGRAPH #	EXACT VALUE	SECTION & PARAGRAPH #	VALUE	SECTION & PARAGRAPH #
+ -	1b top	one-way ANOVA	Figure legend	4	mice/group	Figure legend	error bars are mean +/- SEM	Figure legend	p=4.99517E-06	Figure legend	F (3, 29) = 14.79	
+ -	1b bottom	one-way ANOVA	Figure legend	4	mice/group	Figure legend	error bars are mean +/- SEM	Figure legend	p=2.45183E-06	Figure legend	F (3, 44) = 13.37	
+ -	1c	Repeated measures ANOVA, Bonferroni post-hoc multiple comparisons	Figure legend	7 wt; 8 GFAP-TGFb; 6 GFAP-TGFb:p75 -/-	mice/group	Figure legend	error bars are mean +/- SEM	Figure legend	p=0.0025	Figure legend	F (4, 8) = 10.98	
+ -	1d	one-way ANOVA, Bonferroni post-hoc multiple comparisons	Figure legend	7 wt; 8 GFAP-TGFb; 6 GFAP-TGFb:p75 -/-	mice/group	Figure legend	error bars are mean +/- SEM	Figure legend	1-5 mov/min: p=0.0091 6-25 mov/min: p=9.56752E-05 26-100 mov/min: p=4.79311E-17 >100 mov/min: p=0.0002	Figure legend	1-5 mov/min: F (2, 373) = 4.764 6-25 mov/min: F (2, 298) = 9.548 26-100 mov/min: F (2, 282) = 43.06 >100 mov/min: F (2, 35) = 10.93	
+ -	3g	one-way ANOVA, Bonferroni post-hoc multiple comparisons	Figure legend	16	nuclear pores	Figure legend	error bars are mean +/- SEM	Figure legend	p=0.0003(wt: - vs +) p=3.7942E-07 (-: wt vs KO) p=8.7739E-10 (+:wt vs KO) p=0.0544 (KO:- vs +)	Figure legend	t=3.797 df=71 (wt: - vs +) t=5.600 df=71 (-: wt vs KO) t=7.069 df=71 (+:wt vs KO) t=1.956 df=71 (KO:- vs +)	
+ -	Supplementary Fig. 1b	one-way ANOVA, Bonferroni post-hoc multiple comparisons	Figure legend	3	mice/group	Figure legend	error bars are mean +/- SEM	Figure legend	p=0.0001 (wt vs GFAP-TGFbeta) p=3.8877E-05 (KO vs GFAP-TGFbeta) p=0.0007 (GFAP-TGFbeta:wt vs KO)	Figure legend	t=7.592 df=8 (wt vs GFAP-TGFbeta) t=8.130 df=8 (KO vs GFAP-TGFbeta) t=5.364 df=8 (GFAP-TGFbeta:wt vs KO)	
+ -	Supplementary Fig. 1c	unpaired t-test	Figure legend	3	mice/group	Figure legend	error bars are mean +/- SEM	Figure legend	p=0.2841	Figure legend	t=1.236 df=4	
+ -	Supplementary Fig. 2d top	unpaired t-test	Figure legend	8	mice/group	Figure legend	error bars are mean +/- SEM	Figure legend	p=0.026	Figure legend	t=2.490 df=14	

+ -	Supplementary Fig. 2d bottom	unpaired t-test	Figure legend	8	mice/group	Figure legend	error bars are mean +/- SEM	Figure legend	p=0.0277	Figure legend	t=2.456 df=14	
+ -	Supplementary Fig. 3a right	two-way ANOVA	Figure legend	2	independent experiments	Figure legend	error bars are mean +/- SEM	Figure legend	p=0.0416 (6h) p=0.0033 (12h)	Figure legend	t=2.424 df=8 (6h) t=4.134 df=8 (12h)	
+ -	Supplementary Fig. 3b right	one-way ANOVA Bonferroni post-hoc multiple comparisons	Figure legend	≥ 3	independent experiments	Figure legend	error bars are mean +/- SEM	Figure legend	p=4.6977E-07 (wt: - vs +) p=1.3455E-07 (wt+ vs KO-) p=4.6977E-07 (wt+ vs KO+) p=0.5681 (-: wt vs KO) p=1.0000 (wt- vs KO+)	Figure legend	t=7.155 df=21 (wt: - vs +) t=7.757 df=21 (wt+ vs KO-) t=7.155 df=21 (wt+ vs KO+) t=0.5800 df=21 (-: wt vs KO) p=0.0 df=21 (wt- vs KO+)	
+ -	Supplementary Fig. 3c	unpaired t-test	Figure legend	3	independent experiments	Figure legend	error bars are mean +/- SEM	Figure legend	p=0.0004	Figure legend	t=10.63 df=4	
+ -	Supplementary Fig. 3d	two-way ANOVA Bonferroni post-hoc multiple comparisons	Figure legend	3	independent experiments	Figure legend	error bars are mean +/- SEM	Figure legend	p=0.0098 (GAT1) p=0.0039 (S100b)	Figure legend	t=3.727 df=6 (GAT1) t=4.555 df=6 (S100b)	
+ -	Supplementary Fig. 4	two-way ANOVA Bonferroni post-hoc multiple comparisons	Figure legend	3	independent experiments	Figure legend	error bars are mean +/- SEM	Figure legend	p=7.8871E-05 (wt:- vs +) p=0.0002 (wt+ vs KO +) p=0.7181 (KO:- vs +)	Figure legend	t=7.364 df=8 (wt:- vs +) t=6.396 df=8 (wt + vs KO +) t=0.3740 df=8 (KO:- vs +)	
+ -	Supplementary Fig. 5a	unpaired t-test	Figure legend	3	independent experiments	Figure legend	error bars are mean +/- SEM	Figure legend	p=0.0106 (untreated vs TGFbeta) p=0.0456 (TGFbeta vs TGFbeta + secretase)	Figure legend	t=4.529 df=4 (untreated vs TGFbeta) t=2.867 df=4 (TGFbeta vs TGFbeta + secretase)	
+ -	Supplementary Fig. 8c	unpaired t-test	Figure legend	6	independent experiments	Figure legend	error bars are mean +/- SEM	Figure legend	p=0.0497 (Nup358) p=6.4055E-06 (Nup153)	Figure legend	t=2.232 df=10 (Nup358) t=8.571 df=10 (Nup153)	

▶ Representative figures

1. Are any representative images shown (including Western blots and immunohistochemistry/staining) in the paper?

   If so, what figure(s)?

   > Yes.
   > - Fig. 1a
   > - Fig. 2a-2f
   > - Fig. 3b, d, g
   > - Suppl. Fig. 1a, 1d
   > - Suppl. Fig. 2b, 2c
   > - Suppl. Fig. 3a, 3b
   > - Suppl. Fig. 4
   > - Suppl. Fig. 5b
   > - Suppl. Fig. 6a-e
   > - Suppl. Fig. 7
   > - Suppl. Fig. 8c
   > - Suppl. Fig. 9b, 9c
   > - Suppl. Fig. 11a-c
   > - Suppl. Fig. 12a, 12b
   > - Suppl. Fig. 13

2. For each representative image, is there a clear statement of how many times this experiment was successfully repeated and a discussion of any limitations in repeatability?

   If so, where is this reported (section, paragraph #)?

   > - Fig. 1a: Yes, Fig. legend of Fig. 1a (n=4 mice)
   > - Fig. 2a, 2b: Yes, Fig. legend (n=3 independent experiments)
   > - Fig. 2c-f: Yes, (Fig. 2c, n=3; Fig. 2d, n=2; Fig. 2e, n=3, Fig. 2f, n=2 All n independent experiments)
   > - Fig. 3b: Yes, Fig. legend (n=3 independent experiments)
   > - Fig. 3d: Yes, Fig. legend (n=5 independent experiments)
   > - Fig. 3g: Yes, Fig. legend (n=16 nuclear pores)
   > - Suppl. Fig. 1a: Yes, Fig. legend 1 (n=4 mice)
   > - Suppl. Fig. 1d: Yes, Fig. legend (n=4 mice)
   > - Suppl. Fig. 2b: Yes, (n=2 mice)
   > - Suppl. Fig. 2c: Yes, Fig. legend of Fig 2d (n=8 mice)
   > - Suppl. Fig. 3a, 3b: Yes, Fig. legend (n=3 independent experiments)
   > - Suppl. Fig. 4: Yes, Fig. legend (n=3 independent experiments)
   > - Suppl. Fig. 5b: Yes, Fig. legend (n=2 independent experiments)
   > - Suppl. Fig. 6a-e: Yes, Fig. legend (n=2 independent experiments)
   > - Suppl. Fig. 7: Yes, Fig. legend (n=3 independent experiments)
   > - Suppl. Fig. 8c: Yes, (n=6 independent experiments)
   > - Suppl. Fig. 9b, 9c: Yes, Fig. legend (n=2 independent experiments)
   > - Suppl. Fig. 11a-c: Yes, Fig. legend (n=3 (11a, 11b) n=2 (11c) independent experiments)
   > - Suppl. Fig. 12a, 12b: Yes, Fig. legend (n=3 (12a) n=2 (12b) independent experiments)
   > - Suppl. Fig. 13: Yes, Fig. legend (n=2 independent experiments)

▶ Statistics and general methods

1. Is there a justification of the sample size?

   If so, how was it justified?

   Where (section, paragraph #)?

   Even if no sample size calculation was performed, authors should report why the sample size is adequate to measure their effect size.

   > Sample size calculations are based on previously published data as stated for the different experiments in the Methods or Results sections. No statistical methods were used to predetermine sample sizes.

2. Are statistical tests justified as appropriate for every figure?

   Where (section, paragraph #)?

   | We described the statement for the application of statistical test in Figure Legends |

   a. If there is a section summarizing the statistical methods in the methods, is the statistical test for each experiment clearly defined?

   | We described the statistical methods in the Materials and Methods section and for each experiment in the respective Figure Legend. |

   b. Do the data meet the assumptions of the specific statistical test you chose (e.g. normality for a parametric test)?

   Where is this described (section, paragraph #)?

   | The data meet the assumptions of the specific statistical test chosen. Normal distribution was not tested. |

   c. Is there any estimate of variance within each group of data?

   Is the variance similar between groups that are being statistically compared?

   Where is this described (section, paragraph #)?

   | There is no estimate of variance within each group of data. |

   d. Are tests specified as one- or two-sided?

   | We used two-sided Student's t-test. |

   e. Are there adjustments for multiple comparisons?

   | For one-way ANOVA and two-way ANOVA: Bonferroni post-hoc multiple comparisons test. |

3. Are criteria for excluding data points reported?

   Was this criterion established prior to data collection?

   Where is this described (section, paragraph #)?

   | No data points were excluded. |

4. Define the method of randomization used to assign subjects (or samples) to the experimental groups and to collect and process data.

   If no randomization was used, state so.

   Where does this appear (section, paragraph #)?

   | The study does not contain pharmacologic treatments in vivo. Mice were age-matched for the different genotypes. Cells were all plated at the same time and wells were randomly selected for treatments with growth factors or inhibitors. |

5. Is a statement of the extent to which investigator knew the group allocation during the experiment and in assessing outcome included?

   If no blinding was done, state so.

   Where (section, paragraph #)?

   | Analyses were performed in a blinded manner for all histopathological evaluations. |

6. For experiments in live vertebrates, is a statement of compliance with ethical guidelines/regulations included?

   Where (section, paragraph #)?

   | Yes, this is stated in the Methods section. |

7. Is the species of the animals used reported?

   Where (section, paragraph #)?

   | Yes, this is stated in the Methods section. |

8. Is the strain of the animals (including background strains of KO/ transgenic animals used) reported?

   Where (section, paragraph #)?

   | Yes, this is stated in the Methods section. |

9. Is the sex of the animals/subjects used reported?

    Where (section, paragraph #)?

    No. Animals of both sex have been used.

10. Is the age of the animals/subjects reported?

    Where (section, paragraph #)?

    Yes, this is stated in the Figure Legends and in the Methods section.

11. For animals housed in a vivarium, is the light/dark cycle reported?

    Where (section, paragraph #)?

    No. Mice were housed under a 12 h light/dark cycle.

12. For animals housed in a vivarium, is the housing group (i.e. number of animals per cage) reported?

    Where (section, paragraph #)?

    No. Up to 5 animals per cage were housed and were fed standard chow, and had access to food and water ad libitum.
    For EEG recordings mice were individually housed.

13. For behavioral experiments, is the time of day reported (e.g. light or dark cycle)?

    Where (section, paragraph #)?

    No. Experiments were performed during the light cycle.

14. Is the previous history of the animals/subjects (e.g. prior drug administration, surgery, behavioral testing) reported?

    Where (section, paragraph #)?

    N/A

    a. If multiple behavioral tests were conducted in the same group of animals, is this reported?

        Where (section, paragraph #)?

        N/A

15. If any animals/subjects were excluded from analysis, is this reported?

    Where (section, paragraph #)?

    We did not exclude any mouse data value from analysis.

    a. How were the criteria for exclusion defined?

        Where is this described (section, paragraph #)?

    b. Specify reasons for any discrepancy between the number of animals at the beginning and end of the study.

        Where is this described (section, paragraph #)?

        There is no discrepancy. All animals were present at the end of the study.

▶ Reagents

1. Have antibodies been validated for use in the system under study (assay and species)?

    We used validated antibodies in this study.

    a. Is antibody catalog number given?

        Where does this appear (section, paragraph #)?

        Catalog number of antibodies are provided in the Methods section.

b. Where were the validation data reported (citation, supplementary information, Antibodypedia)?

   Where does this appear (section, paragraph #)?

> Literature is cited in the Methods section or can be found on the companies website.

2. Cell line identity
   a. Are any cell lines used in this paper listed in the database of commonly misidentified cell lines maintained by ICLAC and

   NCBI Biosample?

   Where (section, paragraph #)?

> Cell lines used in this paper are not listed in the database of commonly misidentified cell lines maintained by ICLAC and NCBI Biosample.
>
> We used NIH3T3 cells and HEK293T cells (ATCC) to validate a reporter construct (Suppl Fig. 12b) and perform co-immunoprecipitations of deletion mutants (Suppl Fig. 9b, c). We included these cell lines in the Materials and Methods.

b. If yes, include in the Methods section a scientific justification of their use--indicate here in which section and paragraph the justification can be found.

c. For each cell line, include in the Methods section a statement that specifies:

   - the source of the cell lines

   - have the cell lines been authenticated? If so, by which

     method?

   - have the cell lines been tested for mycoplasma

     contamination?

   Where (section, paragraph #)?

▶ Data deposition

Data deposition in a public repository is mandatory for:

   a. Protein, DNA and RNA sequences
   b. Macromolecular structures
   c. Crystallographic data for small molecules
   d. Microarray data

Deposition is strongly recommended for many other datasets for which structured public repositories exist; more details on our data policy are available here. We encourage the provision of other source data in supplementary information or in unstructured repositories such as Figshare and Dryad.

We encourage publication of Data Descriptors (see Scientific Data) to maximize data reuse.

1. Are accession codes for deposit dates provided?

   Where (section, paragraph #)?

> N/A

▶ Computer code/software

Any custom algorithm/software that is central to the methods must be supplied by the authors in a usable and readable form for readers at the time of publication. However, referees may ask for this information at any time during the review process.

1. Identify all custom software or scripts that were required to conduct the study and where in the procedures each was used.

> N/A

2. If computer code was used to generate results that are central to the paper's conclusions, include a statement in the Methods section under "**Code availability**" to indicate whether and how the code can be accessed. Include version information as necessary and any restrictions on availability.

> N/A

▶ Human subjects

1. Which IRB approved the protocol?

   Where is this stated (section, paragraph #)?

   > N/A

2. Is demographic information on all subjects provided?

   Where (section, paragraph #)?

   > N/A

3. Is the number of human subjects, their age and sex clearly defined?

   Where (section, paragraph #)?

   > N/A

4. Are the inclusion and exclusion criteria (if any) clearly specified?

   Where (section, paragraph #)?

   > N/A

5. How well were the groups matched?

   Where is this information described (section, paragraph #)?

   > N/A

6. Is a statement included confirming that informed consent was obtained from all subjects?

   Where (section, paragraph #)?

   > N/A

7. For publication of patient photos, is a statement included confirming that consent to publish was obtained?

   Where (section, paragraph #)?

   > N/A

▶ fMRI studies

For papers reporting functional imaging (fMRI) results please ensure that these minimal reporting guidelines are met and that all this information is clearly provided in the methods:

1. Were any subjects scanned but then rejected for the analysis after the data was collected?

   > N/A

   a. If yes, is the number rejected and reasons for rejection described?

   Where (section, paragraph #)?

   > N/A

2. Is the number of blocks, trials or experimental units per session and/or subjects specified?

   Where (section, paragraph #)?

   N/A

3. Is the length of each trial and interval between trials specified?

   N/A

4. Is a blocked, event-related, or mixed design being used? If applicable, please specify the block length or how the event-related or mixed design was optimized.

   N/A

5. Is the task design clearly described?

   Where (section, paragraph #)?

   N/A

6. How was behavioral performance measured?

   N/A

7. Is an ANOVA or factorial design being used?

   N/A

8. For data acquisition, is a whole brain scan used?

   If not, state area of acquisition.

   N/A

   a. How was this region determined?

   N/A

9. Is the field strength (in Tesla) of the MRI system stated?

   N/A

   a. Is the pulse sequence type (gradient/spin echo, EPI/spiral) stated?

   N/A

   b. Are the field-of-view, matrix size, slice thickness, and TE/TR/flip angle clearly stated?

   N/A

10. Are the software and specific parameters (model/functions, smoothing kernel size if applicable, etc.) used for data processing and pre-processing clearly stated?

    N/A

11. Is the coordinate space for the anatomical/functional imaging data clearly defined as subject/native space or standardized stereotaxic space, e.g., original Talairach, MNI305, ICBM152, etc? Where (section, paragraph #)?

    N/A

12. If there was data normalization/standardization to a specific space template, are the type of transformation (linear vs. nonlinear) used and image types being transformed clearly described? Where (section, paragraph #)?

    N/A

13. How were anatomical locations determined, e.g., via an automated labeling algorithm (AAL), standardized coordinate database (Talairach daemon), probabilistic atlases, etc.?

    N/A

14. Were any additional regressors (behavioral covariates, motion etc) used?

> N/A

15. Is the contrast construction clearly defined?

> N/A

16. Is a mixed/random effects or fixed inference used?

> N/A

    a. If fixed effects inference used, is this justified?

> N/A

17. Were repeated measures used (multiple measurements per subject)?

> N/A

    a. If so, are the method to account for within subject correlation and the assumptions made about variance clearly stated?

> N/A

18. If the threshold used for inference and visualization in figures varies, is this clearly stated?

> N/A

19. Are statistical inferences corrected for multiple comparisons?

> N/A

    a. If not, is this labeled as uncorrected?

> N/A

20. Are the results based on an ROI (region of interest) analysis?

> N/A

    a. If so, is the rationale clearly described?

> N/A

    b. How were the ROI's defined (functional vs anatomical localization)?

> N/A

21. Is there correction for multiple comparisons within each voxel?

> N/A

22. For cluster-wise significance, is the cluster-defining threshold and the corrected significance level defined?

> N/A

▶ Additional comments

Additional Comments