

A *Phytophthora sojae* effector PsCRN63 forms homo-/hetero-dimers to suppress plant immunity via an inverted association manner

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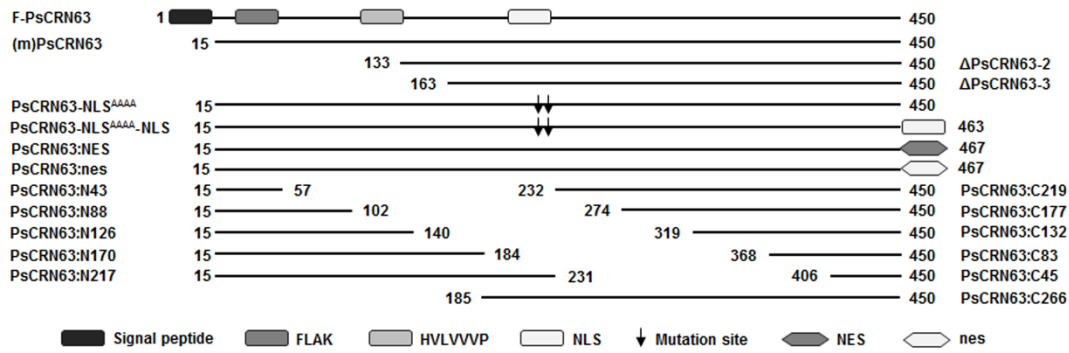
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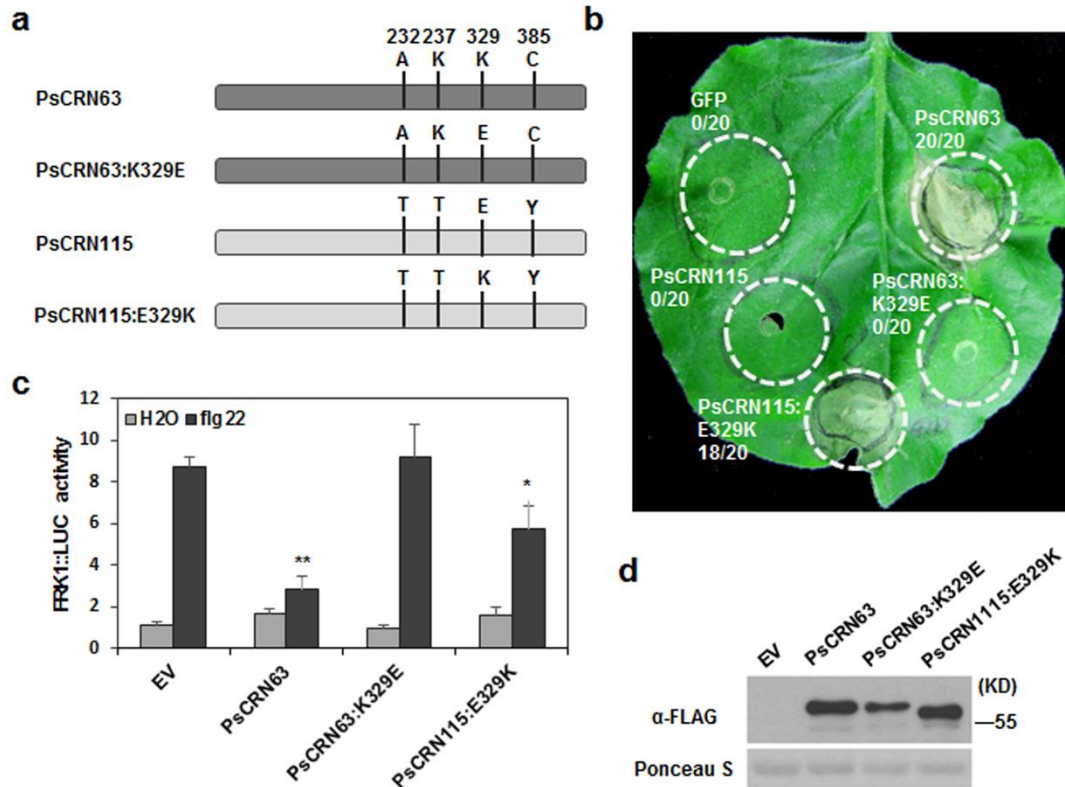
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Supplementary Figure S1. Schematic view of a succession of artificial mutant constructs derived from PsCRN63.

PsCRN63 carries a N-terminal signal peptide (black rectangle), and has a conserved motif “FLAK” in its N-termini (dark grey rectangle) followed by another conserved motif “HVLVVVP” (light gray rectangle), then a nuclear localization signal (NLS) in addition (white rectangle). Little arrows represent mutation sites and flat hexagon (dark grey) symbolize a nuclear exclusion signal (NES) while the white hexagon means a nonfunctional NES (nes). The C-terminal domain carries the module with biochemical effector activity.



Supplementary Figure S2. PTI suppression of PsCRN63 strongly in correlation with its cell death-inducing ability.

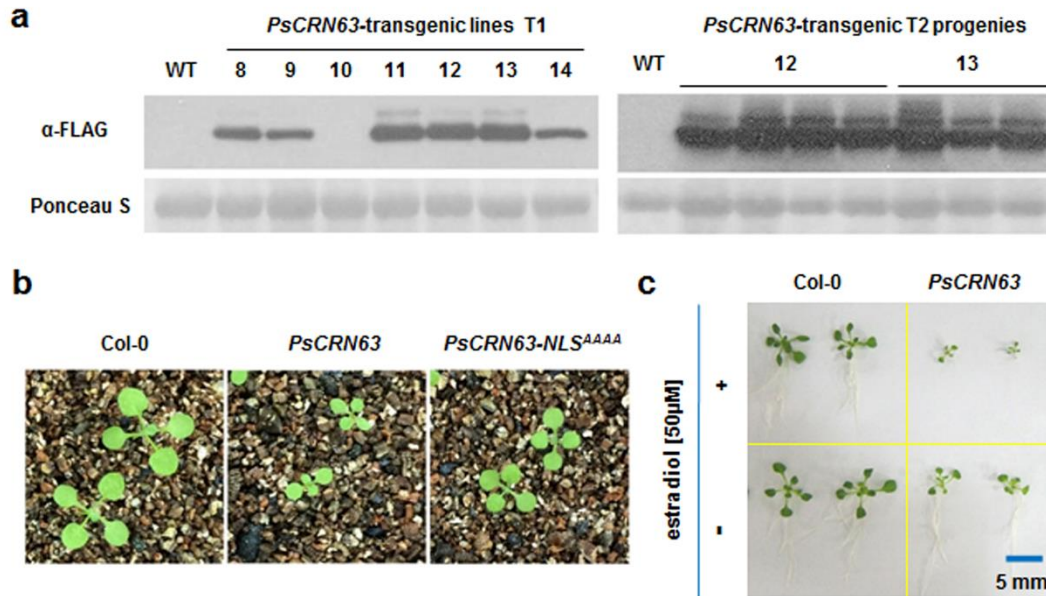
(a) Amino acid-swapping experiment in C-terminus between PsCRN63 and PsCRN115. The upper two strips represent PsCRN63 and PsCRN63:K329E (dark grey), while the lower ones symbolize PsCRN115 and PsCRN115:E329K (light grey).

(b) Cell death induction experiments in *N. benthamiana* leaves. *N. benthamiana* was transformed with the indicated constructs or GFP (negative control) by agroinfiltration. The white circle indicates infiltration sites and the numbers show the ratio of necrotic responses to the total infiltration sites. The experiments had three biological repeats with similar results.

(c) Flg22-induced *FRK1::LUC* expression. PsCRN63, PsCRN63:K329E, PsCRN115:E329K or empty vector were transfected into Col-0 protoplasts, along with FRK1::LUC and 35S::RLUC. Samples were treated with H₂O or flg22, and the *LUC* reporter activity was determined. Each data point represents the

mean of three replicates. Error bars indicate standard deviation(*, $P < 0.05$; **, $P < 0.01$; Student's *t*-test). The experiments were repeated three times with similar results.

(d) Protein expression levels determined by Western blot. Proteins coded by the constructs indicated as (C) were detected with an anti-FLAG antibody. Equal loading is indicated by ponceau staining of Rubisco protein.



Supplementary Figure S3. Phenotype alteration of *PsCRN63*-transgenic *Arabidopsis* plants.

(a) Protein expression levels determination in *PsCRN63*-transgenic plants.

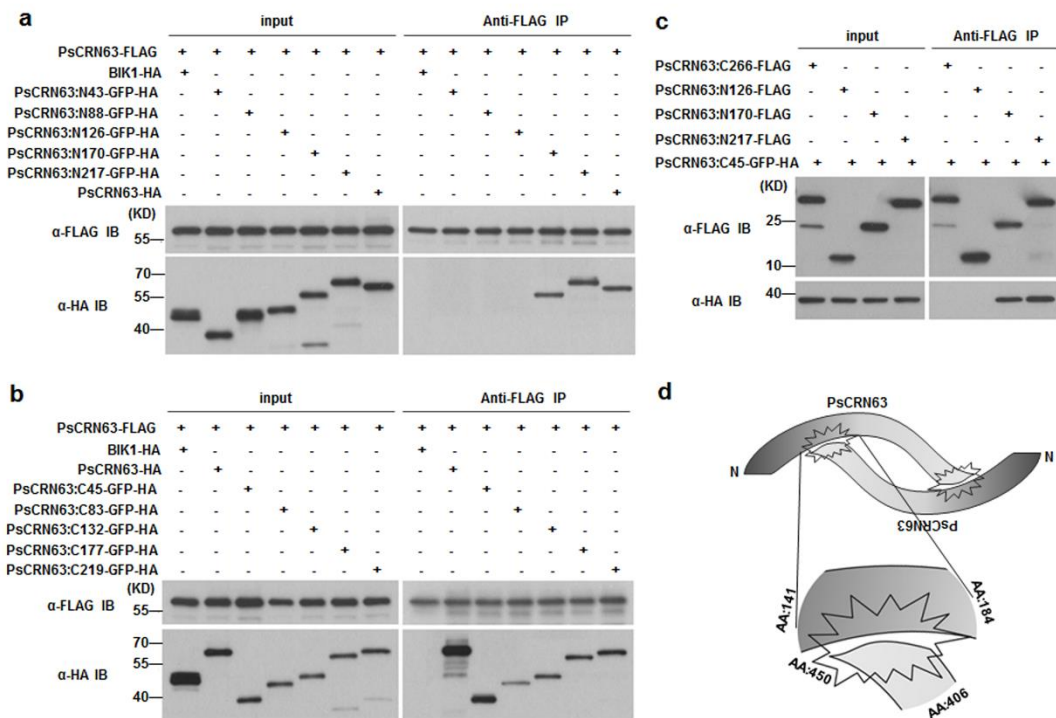
Total protein isolated and detected by immunoblot with an anti-FLAG antibody. Equal loading is indicated by ponceau staining of Rubisco protein. Left panel, T1 generation of the independent *PsCRN63*-transgenic lines (8#-14#). Right panel, T2 generation of *PsCRN63*-transgenic line 12# and 13#.

(b) Phenotype of *PsCRN63* and *PsCRN63-NLS^{AAAA}* transgenic seedlings grown in roseite.

Two-week-old *Arabidopsis* of the indicated genotypes under no oestrogen treatment were photographed.

(c) Phenotype of *PsCRN63* transgenic lines grown in 1/2 MS medium.

Three-week-old wild type Col-0 or *PsCRN63*-transgenic *Arabidopsis* with continuous oestrogen treatment or under no oestrogen induction were photographed.

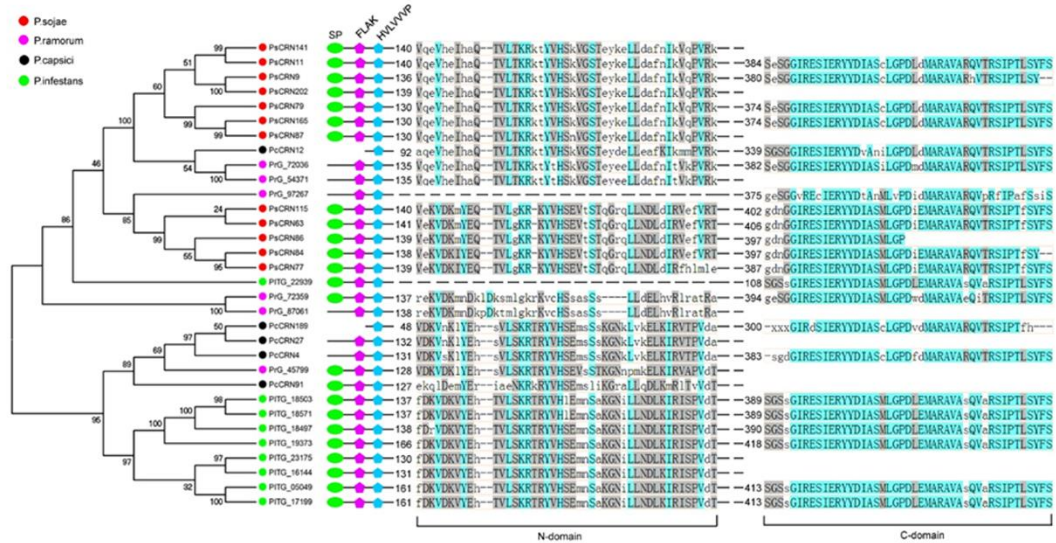


Supplementary Figure S4. Spontaneous dimerization of PsCRN63 through an inverted association manner.

(a-b) Testing essential segments of PsCRN63 for dimerization. BIK1-HA, PsCRN63-HA and its truncated derivatives were co-expressed with PsCRN63-FLAG in Col-0 protoplasts and subjected to an Anti-FLAG co-immunoprecipitation.

(c) Association between N-terminal segment and C-terminal segment of PsCRN63. Indicated plasmids combination were co-expressed in WT *Arabidopsis* protoplasts, subjected to a α -FLAG IP, and the bound protein was detected by immunoblot with the indicated antibodies.

(d) A sketch map for inverted association of PsCRN63. The dark grey end of the ribbon represents N-terminal of PsCRN63 and the shape of the explosion indicates where the association happens. The lower picture enlarges happened association as a schematic plot including combination sites. This diagrammatic sketch portrays a dimer association of PsCRN63.



Supplementary Figure S5. Construction of phylogenetic tree for *Phytophthora* CRN effectors.

Thirty-two *Phytophthora* effectors were identified by similarity searches using BLAST analysis with an E-value cut-off 1E-10 in *P. sojae*, *P. ramorum*, *P. capsici* and *P. infestans* genome sequences. A phylogenetic tree of these effector proteins was constructed using MEGA 5.1 by the neighbor joining method. The predicted SP (Signal peptide), FLAK motif and HVLVVVP motif are indicated using corresponding geometric figure.

Supplementary Table S1. Suppression activities of flg22-induced *FRK1::LUC* expression of *Phytophthora sojae* effectors.

Effectors	relative <i>FRK1::LUC</i> expression	STDEV	Student's <i>t</i> test	Suppression of <i>FRK1::LUC</i>
Empty Vector	22.82	4.79		
HopA1	3.23	0.47	0.01	YES
PsAvr1K	17.76	2.63	0.12	NO
PsAvr3C	15.74	6.06	0.15	NO
PsAvr4/6	24.50	6.65	0.27	NO
PsCRN63	2.11	0.48	0.02	YES
PsCRN115	23.38	2.28	0.84	NO
PsCRN124	17.38	5.53	0.75	NO
PsCRN127	21.73	0.48	0.28	NO

Supplementary Table S2. Oligonucleotides used for plasmids construction and gene expression.

Primers for gene cloning			
CRN63- XhoI-F	GCACTCGAGatgTTTCCTGTGGA CATTGAC	CRN63- Csp45I-R	CGCTTCGAAGCTGAAATAACTA AACGTGGG
CRN115- XhoI-F	CATCTCGAGatgGCGTTCCCAGT GGACATC	CRN115- Csp45I-R	CGCTTCGAAGCTGAAATAACTA AACGTGGG
CRN127- XhoI-F	CATCTCGAGgtgTTTGACGTGGA AGTCGAC	CRN127- Csp45I-R	GCATTCGAA TAGCCTCACTTTCGTCAAAGA
ΔCRN63- 2-XhoI-F	CATCTCGAGatgGAGACTTCCAA GATGGAT	CRN63:NE S-Csp45I-R	CGCTTCGAACTTGTTAATATCAA GTCCAGC
ΔCRN63- 3-XhoI-F	TATCTCGAGatgGTAACCTCGAC CCAGGGC	CRN63:nes -Csp45I-R	GCATTCGAACTTGTTAGCATCT GCTCCAGC
		CRN63:NLS -Csp45I-R	CGCTTCGAACTTTACCTTTCGT TTCTTCTT
CRN63- NLS ^{AAAA} -F	GAGGAGCAGCAGGCCAAAGCA TATGCCGCATACGTGGAACAC	CRN63- NLS ^{AAAA} -R	GTGTTCCACGTATGCGGCATAT GCTTTGGCCTGCTGCTCCTC
CRN63:Δ4 3-F	ATACTCGAGatgGGCGCGTGGCT GCCAGAC	CRN63- N217-BstBI- R	GCATTCGAAGCCAATGTTGTGT TCCACGTA
CRN115:Δ 43-F	ATACTCGAGatgGGCGCGTGGCT GCCAGAC	CRN115- N217-BstBI- R	GCATTCGAAGCCAATGTTGTGT TCCACGTA
CRN63- C45-XhoI- F	ATACTCGAGatgGGTGACAATGG CGGGATT	CRN63- N43-BstBI- R	ATATTCGAAGTCCGCCGTCTTC GCCAGGGA
CRN63- C83-XhoI-	ATACTCGAGatgCAGCCACCGAC TGCACCC	CRN115- N43-BstBI-	ATATTCGAAGCCCTCCGTCTTC GCCAGGAA

F		R	
CRN63- C132- XhoI-F	ATACTCGAGatgGACCCAGTAAT GGCGCTC	CRN63- N88-BstBI- R	GCATTCGAAGTTAGCTGTCAAC ACGTCCTC
CRN63- C177- XhoI-F	CATCTCGAGatgCTTGTTGCGAT GAGGCCG	CRN63- N126-BstBI- R	CGCTTCGAAGAGTTGATCCATC TTGGAAGT
CRN63- C219- XhoI-F	ATACTCGAGatgGCTGTGCTCAA GGAGAAG	CRN63- N170-BstBI- R	GCATTCGAATGTGCGCACAAAT TCCACCCG
CRN63- C266- XhoI-F	CTCGAGatgGTTCCATTCGATGC TGGA		
GFP- Csp45I-F	TCGTTCGAATACCCATACGATGT TCCTGAC	GFP-SpeI-R	CGAACTAGTTTACTTGTACAGC TCGTCCATGCC
CRN79- C45-XhoI- F	CATCTCGAGatgAGTGAAAGTGG CGGGATT	CRN79- N129-BstBI- R	CGCTTCGAACACTTGATCCATC CTGGAAGT
CRN79- C244- XhoI-F	ATACTCGAGatgGCTGCTGCTAA GTGGGGT	CRN79- N174-BstBI- R	CGCTTCGAATTTACGCACAGGT TGACCTT
		CRN79- N221-BstBI- R	GCATTCGAAACCGAACACGCA AAGCTTCTC
CRN63- NcoI-F	GCACCATGGatgTTTCCTGTGGA CATTGAC	CRN63- EcoRI-R	CGCGAATTCGCTGAAATAACTA AACGTGGG
CRN115- NcoI-F	ATACCATGGatgGACATCGATGG GGGCCAG	CRN115- EcoRI-R	CGCGAATTCGCTGAAATAACTA AACGTGGG
CRN63-	GCAGGTACatgTTTCCTGTGGA	CRN63-	GCAGTCGACGCTGAAATAACTA

KpnI-F	CATTGAC	Sall-R	AACGTGGG
CRN115- KpnI-F	TATGGTACCatgGACATCGATGG GGGCCAG	CRN115- Sall-R	GCAGTCGACGCTGAAATAACTA AACGTGGG
PcCRN4- KpnI-F	CATGGTACCATGGTGAAGCTTT CACTACAG	PcCRN4- XhoI-R	GCACTCGAGACTGAAATAACTC AACGTGGG
PsCRN79- KpnI-F	GCAGGTACCATGGTCAAGCTTT TTTGTGCT	PsCRN79- Sall-R	GCAGTCGACGCTGAAATAACTC AACGTGGG
RxLR172- KpnI-F	tacctGGTACCATGTACAGCGGTG CAGTCGAGTC	RxLR172- Sall-R	TcgcGTCGACCCGCCCGTTGTC GCCCGTA
ΔCRN63- 2-KpnI-F	CATGGTACCatgGAGACTTCCAA GATGGAT	ΔCRN63-3- KpnI-F	TATGGTACCatgGTAACCTCGAC CCAGGGC

Primers for RT-PCR			
FRK1-F	GCCAACGGAGACATTAGAG	FRK1-R	CCATAACGACCTGACTCATC
NHL10-F	TTCCTGTCCGTAACCCAAAC	NHL10-R	CCCTCGTAGTAGGCATGAGC
WRKY53-F	CACCAGAGTCAAACCAGCCAT TAC	WRKY53-R	CTTTACCATCATCAAGCCCAT CGG
CBP60g-F	AAGAAGAATTGTCCGAGAGGA G	CBP60g-R	GGCGAGTTTATGAAGCACAG
PR1-F	GAAAACCTTAGCCTGGGGTAGC	PR1-R	TTCATTAGTATGGCTTCTCGTT CA
PR2-F	GCAATGCAGAACATCGAGAA	PR2-R	TCATCCCTGAACCTTCCTTG
PDF1.2-F	CCATCATCACCCCTTATCTTCGC	PDF1.2-R	TGTCCCACTTGGCTTCTCG
ERF1-F	TTAATTCAGTCCCCATTCTC	ERF1-R	CCAAGTCCCACTATTTTCAG
ORA59-F	CTCTGCTTCTACAATTTTATG	ORA59-R	CTACACATCTATACATGTTTCC
ACT1-F	TCTTGATCTTGCTGGTCGTG	ACT1-R	GAGCTGGTTTTGGCTGTCTC