Supplementary material for:

Identification and analysis of integrons and cassette arrays in bacterial genomes.

Jean Cury^{1,2}, Thomas Jové³, Marie Touchon^{1,2}, Bertrand Néron⁴, Eduardo PC Rocha^{1,2}

¹ Microbial Evolutionary Genomics, Institut Pasteur, 28, rue Dr Roux, Paris, 75015, France

² CNRS, UMR3525, 28, rue Dr Roux, Paris, 75015, France

³ Univ. Limoges, INSERM, CHU Limoges, UMR_S 1092, F-87000 Limoges, France.

⁴ Centre d'Informatique pour la Biologie, C3BI, Institut Pasteur, Paris, France

Table of contents

Figure S1 – Phylogenetic tree of tyrosine recombinases.

Figure S2 – Phylogenetic tree of Intl.

Figure S3 – Distribution of the proportion of integron-integrase in a species

Figure S4 – Histogram of the distribution of the number of *attC* sites per integron

Figure S5 – Distribution of hits of PF00589 and intl_Cterm in function of each other and the existence of neighboring *attC* sites.

Figure S6 – Taxonomic distribution of integrons in clades with less than 50 genomes fully sequenced.

Figure S7 – Frequency of integrons and related elements as a function of the genome size when the analysis is restricted to Gammaproteobacteria.

Figure S8 – Histogram of the distribution of the number of *attC* sites in CALIN elements.

Figure S9 – Comparisons between attC sites.

Table S1 – Correspondence table between replicon's names in the following table and real name and NC numbers.

Table S2a/b – List of expected (a) and observed (b) position of the 596 *attC* sites from INTEGRALL (attached as text file)

Table S3 – Sequence of the promoter of the integrase (Pint), the promoter of the cassette (Pc), and of the attl site in 3 classes of integron when available.

Table S4 – List of Intl protein from tree of Figure S2 with the data (attached as a text file).

Table S5 – List of genome used to create the 6 pseudo-genomes with different GC% background composition.

Table S6 – List of all integrons identified in bacterial genomes per element (attached as text file)

Table S7 – List of all integrons identified in bacterial genomes per integron (attached as text file)

Table S8 – List of all integrons identified in bacterial genomes per genome (attached as text file)

File S1 – List of 291 *attC* sites used to train and test the model (fasta file)

File S2 – intl_Cterm HMM profile.

File S3 – Covariance Model of the *attC* site.

File S4 – Alignment of 96 attC sites to build the covariance model (Stockholm format)

Tree S1 – Phylogenetic tree of tyrosine recombinases, corresponding to Figure S1 (attached as a nexus file).

Tree S2 – Phylogenetic tree of integron integrases, corresponding to Figure S2 (attached as a newick file)



Figure S1 – Phylogenetic tree of tyrosine recombinases including the 21 proteins, which match the profile PF00589 but not intl_Cterm (red) and Intl from complete integron (blue).



Figure S2 - Phylogenetic tree of Intl. The tree was made using IQ-Tree on the multiple alignments of the integrases carrying the two profiles (intl_Cterm and PF00589) using XerC and XerD as outgroups (see Methods for details). Bootstrap values are indicated if lower than 60%. The matrix on the right represents traits associated with the integron. Column (A) represents whether the corresponding

integron is on a plasmid (dark grey) or on a chromosome (grey). Lighter grey corresponds to integrons of classes 1 to 5, but whose replicon types are unknown (unavailable genomes). The column (B) represents whether the corresponding integron is a sedentary chromosomal integron (green) or a mobile integron (dark orange), or other (not determinable) (grey). Column (C) represents the integrases associated with at least one attC site (grey). The column (D) represents the number of *attC* sites with a gradient of grey (the darker the more *attC* sites, with saturation from 20 *attC* sites). The column (E) represents the frequency of resistance genes among cassettes (darker blue indicates higher values). The last column (F) represents the integron with inverted integrase (red). The leaves of the tree are colored according to their clade. For the names corresponding to the colors, see Figure 5 and Figure S6. See Table S3 for full data.



Figure S3 – Histogram of the proportion of genomes in a species carrying orthologs for a given integron-integrase. Only species with more than 4 genomes were used for the analysis.



Figure S4. Histogram of the distribution of the number of *attC* sites per integron. Mobile integrons are depicted in dark orange; sedentary chromosomal integrons (present in more than 60% of the genomes of a species, actually 100%) are depicted in green; undetermined elements are depicted in grey. The largest mobile integron has 15 *attC* sites.



Figure S5 - Distribution of hits of PF00589 and intl_Cterm in function of each other and the existence of neighboring *attC* sites. The y-axis represents percentage in a log10 scale. Numbers on the bar represent the actual quantity for the underlying bar.



Figure S6 – Taxonomic distribution of integrons in clades with less than 50 genomes available in our dataset. The grey bar represents the number of genome sequenced for a given clade. The blue bar represents the number of complete integron, the red bar number of In0 and the yellow bar the number of CALIN.



Figure S7 - Frequency of integrons and related elements as a function of the genome size when the analysis is restricted to Gammaproteobacteria. Vertical bar

represents standard error of the mean. The sample sizes of each bin are: 68 [0-2], 108 [2-4], 339 [4-6], and 34 [>6].



Figure S8 - Histogram of the distribution of the number of *attC* sites in CALIN elements. One point was a clear outlier and is not represented on the figure for sake of clarity (its value was 114).



Figure S9 Comparison of *attC* sites. A column represents comparisons of *attC* sites between (inter) or within (intra) element(s) (Complete integron or CALIN) depending on the type of element (mobile or sedentary chromosomal integron (SCI)). (A) Distribution of the sequence distance between two R-UCS-L boxes of two *attC* sites. (B) Distribution of the difference in VTS size. All distributions are significantly different from each other in any given panel (Mann-Whitney rank test, all p-values < 10^{-3})

Table S3 – Sequence of the promoter of the integrase (Pint), the promoter of the cassette (Pc), and of the attl site in 3 classes of integron when available. Sequences were provided by INTEGRALL. Square brackets indicate that it can be one of the letters at that position. Curly brackets indicate that the square brackets before is repeated a number of times comprised between a minimum and a maximum.

Class	P _{int}	Pc	attl	
1	TTGCTGCTTGGATGCCCGAGG CATAGACTGTACA	T[GT]G[AG][CT]ATAAGCCTGTT CGGTT[CG]GT[AG]A[AG]CTGTA ATCGCA	TGATGTTATGGAGCAGCAACG ATGTTACGCAGCAGGGCAGTC	
		TTGTTATGACTGTTTTTT[G-]{1,4}[GT]ACA[GCA][AT]	GCCCTAAAACAAAGTT	
2	ND	ND	TTAATTAACGGTAAGCATCAGC GGGTGACAAAACGAGCATGCT TACTAATAAAATGTT	
3	ND	TAGACATAAGCTTTCTCGGTCT GTAGG[CA]TGTAATG	CTTTGTTTAACGACCACGGTTG TGGGTATCCGGTGTTTGGTCA GATAAACCACAAGTT	

Table S5 – List of genome used to create the 6 pseudo-genomes with different GC%background composition.

Genome	Size (pb)	%GC
Mycoplasma hyorhinis GDL-1	837480	25.91
Anaerococcus prevotii DSM 20548	1883067	36.07
Bacilllus subtilis subsp. subtilis str. 168	4215606	43.51
Escherichia coli str. K-12 substr. MG1655	4639675	50.79
Arthrobacter aurescens TC1	4597686	62.34
Clavibacter michiganensis subsp. michiganensis NCPPB 382	3297891	72.66