Supplementary information

Nutrient shortage triggers the hexosamine biosynthetic pathway *via* the GCN2-ATF4 signalling pathway.

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Supplementary Figure S1: Tunicamycin treatment increases GFAT1 abundance in HBEC 3KT-RL cells. Immunoblot analysis of the abundance of GFAT1 and ATF4 from HBEC 3KT-RL cells treated with DMSO or tunicamycin (0.5 µg/mL) for 16 hours. Tubulin serves as a loading control. Corresponding mRNAs were analyzed to assess XBP1 splicing by RT-PCR followed by Pst1 restriction: bold s- spliced XBP1, h- hydrid XBP1, u1 and u2- products from Pst1 cleavage of unspliced XBP1.



Supplementary Figure S2: GFAT1 silencing impairs proteins O-GlcNAcylation in glucose-deprived cells. Immunoblot analysis of the abundance of O-GlcNAcylated proteins and GFAT1 from HBEC 3KT-RL transfected with a control siRNA or GFAT1 siRNA and incubated in a medium containing 0.1 mM of glucose for 24 hours. Tubulin serves as a loading control.



Supplementary Figure S3: OGT and OGA protein analysis in HBEC 3KT-RL cells silenced for ATF4 and glucose-deprived for 24 hours. Immunoblot analysis of the abundance of OGT and OGA, from HBEC 3KT-RL cells transfected with a control siRNA or an ATF4 siRNA. Cells were incubated with DMEM containing 25 mM (+) or 0.1 mM (-) of glucose for 24 hours. Tubulin serves as a loading control.



Supplementary Figure S4: GCN2 contributes to the ATF4-mediated transcription in response to glucose deprivation. (a) Immunoblot analysis of the abundance of ATF4, CHOP, and GCN2 in mouse embryonic fibroblasts wild type (+/+) or knockout (-/-) for GCN2 incubated in medium with 25 mM (+) or no (-) glucose for 4, 8 and 16 hours. Tubulin serves as a loading control. (b) Luciferase assays from HeLa cells stably expressing the ATF4 reporter system: CARE-Luc. Cells were prior transfected with control or GCN2 siRNAs. 72 hours following transfection, cells were incubated in a medium with 25 mM (+) or no (-) glucose. Data were normalized to the total protein content for each sample. *** p < 0.001. (c) RT-qPCR expression measurements of three ATF4-target model genes: *Chop, Trib3* and *Gadd34* in HBEC 3KT-RL transfected with control or GCN2 siRNAs and incubated for 4, 8 and 16 hours either in medium containing 25 mM (+) or 0.1 mM (-) of glucose. Data are expressed as fold inductions * p < 0.05 and *** p < 0.001.



Supplementary Figure S5: OGT and OGA protein analysis in HBEC 3KT-RL cells silenced for GCN2 and glucose-deprived for 24 hours. Immunoblot analysis of the abundance of OGT and OGA, from HBEC 3KT-RL cells transfected with control or GCN2 siRNAs. Cells were incubated with DMEM containing 25 mM (+) or 0.1 mM (-) of glucose for 24 hours. Tubulin serves as a loading control.



Supplementary Figure S6: PGM3 abundance in HBEC 3KT-RL cells silenced for ATF4 or XBP1 and glucose-deprived for 24 hours. Immunoblot analysis of the abundance of PGM3 and UAP1, from HBEC 3KT-RL cells transfected with control, ATF4 or XBP1 siRNAs. Cells were incubated with DMEM containing 25 mM (+) or 0.1 mM (-) of glucose for 24 hours. Tubulin serves as a loading control. Corresponding mRNAs were analyzed to assess XBP1 splicing by RT-PCR and Pst1 restriction: bold s- spliced XBP1, h- hydrid XBP1, u1 and u2-products from Pst1 cleavage of unspliced XBP1.



ATF4

P-elF2a

ATF6

p90

p50

р58^{ірк}

BiP

Tubulin 🖡

- Glc

O-GIcNAc

Tubulin

0 4

siCtrl

Relative to Figure 3a

130 kDa

72 kDa

55 kDa

95 kDa

72 kDa

55 kDa

Glo

GCN2

ATF4

GFAT1

Tubulin



43 kDa

34 kDa



AP20187	0	2	4	6	8	10	16	24 (h)	
									- 170 kDa
								İ	- 130 kDa
			-						- 95 kDa
			Alexander -	-	See.		8078		- 72 kDa
O-GICINAC	-	-	-	-	-				- 55 kDa
		-	-	-	-	-	-		- 43 kDa
		-			-				- 34 kDa
								1	

Relative to Figure 2c

ATF4

O-GIcNAc

GFAT1

Tubulin

Relative to Figure 3d

Ctrl -Leu HF

- 72 kDa

55 kDa

170 kDa

130 kDa

95 kDa

72 kDa

55 kDa

43 kDa

34 kDa

95 kDa

- 72 kDa

55 kDa

55 kDa

Relative to Figure 1c

ATF4	- 95 kDa - 72 kDa - 55 kDa
р58 ^{ірк}	
BiP	- 95 kDa - 72 kDa
GFAT1	- 95 kDa - 72 kDa - 72 kDa
UAP1	- 55 kDa
ubulin	- 55 kDa

siCtrl siATF4 siXBP1

Glc + - + - + -

AP20187	0 2 4 6 8 10 16 24 (h)	
FKBP	_======	130 kD 95 kDa
P-elF2α		43 kDa 34 kDa
ATF4		95 kDa 72 kDa 55 kDa
		55 KDa
р58 ^{ірк}	- 72 kD)a)a
BiP		95 kDa
GFAT1		72 kDa 95 kDa 72 kDa
Tubulin		55 kDa

Supplementary Figure S7: Uncropped blots corresponding to main Figures 1, 2, and 3.

O-GIcNAc

Tubulin

All SDS-PAGE were run under the same experimental conditions. Following transfer, the membranes were generally cut into strips to minimize the required amount of antibody. Black boxes with dashed lines indicate how blots were cropped. (a) Uncropped blots for the images shown in Fig. 1a, 1b and 1c. (b) Uncropped blots for the images shown in Fig. 2a, 2b and 2c. (c) Uncropped blots for the images shown in Fig. 3a, 3c and 3d.

Relative to Figure 3c

95 kDa

72 kDa

55 kDa 43 kDa

34 kDa

55 kDa

b

С

Human primers used for XBP1s RT-PCR

Gene name	Primer sequence			
XBP1	Forward: AAACAGAGTAGCAGCTCAGACTGC Reverse: TCCTTCTGGGTAGACCTCTGGGAG			
<i>Human RT-qPCR primers</i> Gene name	Primer sequence			
Chop	Forward: AGCTGTGCCACTTTCCTTTC Reverse: CAGAACCAGCAGAGGTCACA			
Gadd34	Forward: CTGTGATCGCTTCTGGCA Reverse: GGAAGAAAGGGTGGGCATC			
Gfat1	Forward: CGGCTGCCTGATTTGATT Reverse: GATAGCCTCGTCCCATTA			
Hprt	Forward: TGACCTTGATTTATTTTGCATACC Reverse: CGAGCAAGACGTTCAGTCCT			
Trib3	Forward: TGGTACCCAGCTCCTCTACG Reverse: GACAAAGCGACACAGCTTGA			
Human primers used for ATF4 ChIP qPCR				
Gene name	Genome location of the amplicon			

Gfat1	Forward: GTAATTCTCCTGCCTTGGCC
	Reverse: GGTGTTTCATGTCCTGGGTG

Chr2:69318472+69318604

Human primers used for GFAT1 siRNA

Target

Primer sequence

Gfat1

Forward: GCAGAUACUUUGAUGGGUCUU Reverse: AAGACCCAUCAAAGUAUCUGC

Supplementary Table S1: Primers sequences used for RT-PCR, RT-qPCR, ChIP experiments and GFAT1 siRNA.